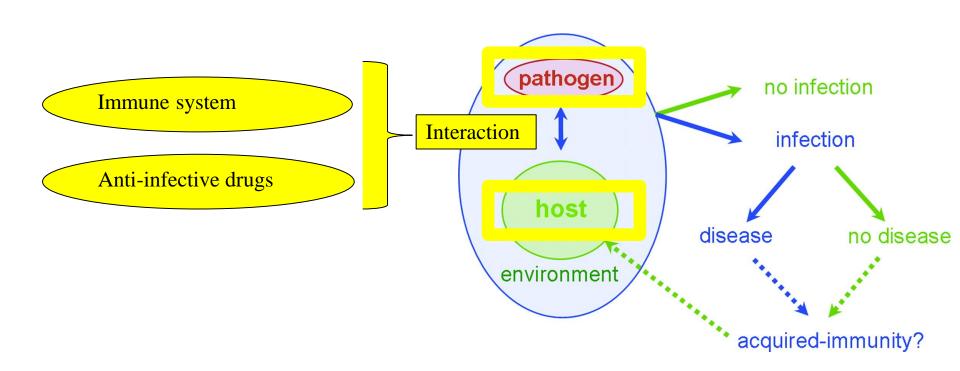
Schematic representation of host-pathogen interactions



IMMUNE SYSTEM

Actively acquired - When the body has already experienced an infection by that pathogen

Natural

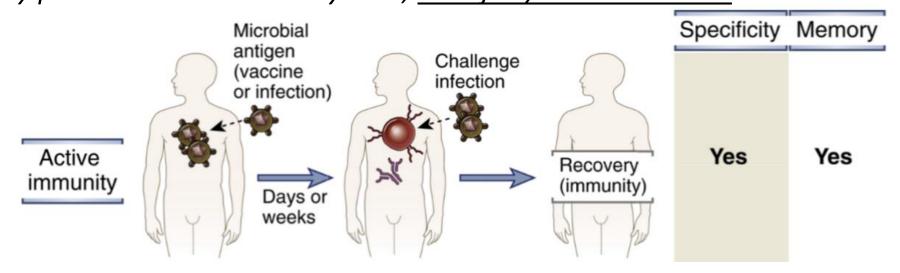
Passively acquired - Antibodies pass across placenta providing a newborn baby with immunity against disease. Antibodies are also present in breast milk.

Acquired immunity

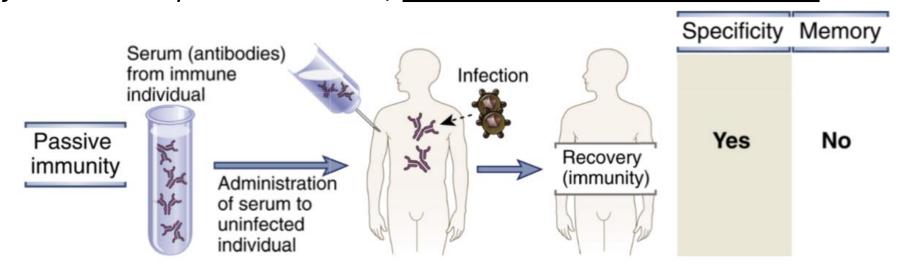
Non natural induced **Actively acquired** – ACTIVE IMMUNIZATION. This is by **vaccination** at a suitable time in the person's life.

Passively acquired – PASSIVE IMMUNIZATION. This is by administrating ready-made **antibodies** which are able to immediately neutralize the viruses

Active immunity (immunoprophylaxis) Protection conferred by person's own immune system, <u>lasts for years or decades</u>



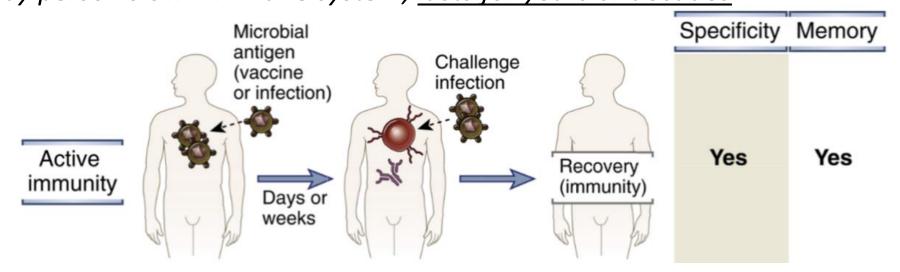
Passive Immunity (immunotherapy) Protection transferred from another person or animal, <u>disappears after two-three weeks</u>

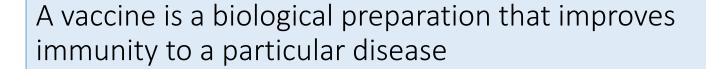


Box 1. Indications for blood-derived antibodies for infectious diseases with a current American or European Union market authorization^a

- Anthrax: treatment of inhaled anthrax.
- Botulism: treatment of botulinum.
- Clostridium botulinum: treatment of infant botulism caused by type A or B C botulinum in patients < 1 year.
- Cytomegalovirus: prophylaxis of cytomegalovirus disease associated with transplantation of kidney, lung, liver, pancreas and heart.
- Diphtheria: treatment of diphtheria and rarely as prophylactic of diphtheria in asymptomatic, non-immunized individuals who have been exposed.
- Hepatitis A: protection from hepatitis A in household and other close contacts.
- Hepatitis B: prevention of Hepatitis B recurrence following liver transplantation; treatment of acute exposure to Hepatitis B-containing blood, sexual exposure to infected persons, infants born to infected mothers and household exposure to persons with acute infection.
- Hepatitis C: Prevention of recurrent hepatitis C virus-induced liver disease in liver transplant recipients.
- Measles: postexposure prophylaxis for suspected measles in unvaccinated persons.
- Rabies: postexposure prophylaxis to rabies category III exposure (i.e. single or multiple transdermal bites or scratches, contamination of mucous membrane with saliva from licks; exposure to bat bites or scratches).
- Rubella: prophylaxis of rubella to exposed individuals in early pregnancy.
- Staphylococcus aureus: treatment of S aureus bacteraemia.
- Tetanus: immediate prophylaxis after tetanus prone injuries in patients not adequately vaccinated, with unknown immunization status, severe deficiency in antibody production or vaccinated patients with high risk wounds.
- Vaccinia: prevention or treatment of vaccinia/smallpox. Treatment and/or modification of conditions which are complications resulting from smallpox vaccination.
- Varicella: prophylaxis against varicella zoster virus infection in at-risk exposed patients.

Active immunity (immunoprophylaxis) Protection conferred by person's own immune system, <u>lasts for years or decades</u>

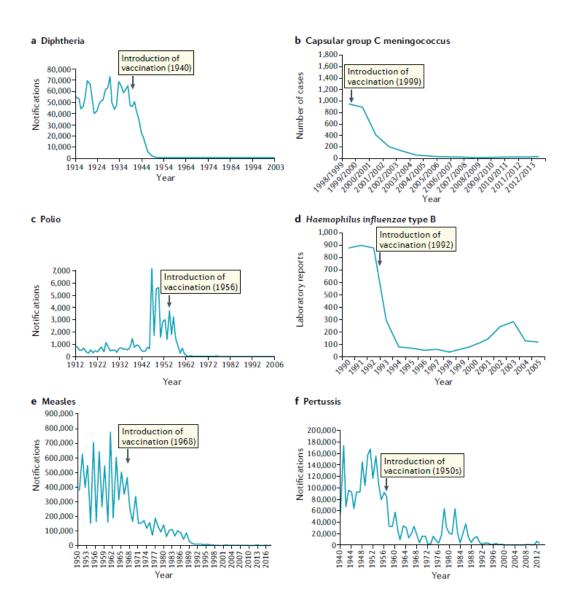






.....A vaccine typically contains an agent that resembles a disease-causing microorganism The agent stimulates the body's immune system to recognize the agent as foreign, "destroy it, and remember it", so that the immune system can more easily recognize and destroy any of these microorganisms that it later encounters......

Vaccines have proved to be one of the most powerful and effective ways of reducing disease and vaccination is considered the most cost-effective medical intervention ever introduced



Modificato da: Stanley Plotkin

PNAS | **August 26, 2014** | vol. 111 | no. 34

Outline of the development of human vaccines

Empiric development

Rational development

| Live attenuated | Killed whole organisms | Purified proteins or polysaccharides | Genetically engineered |
|-----------------------------------|---|---|--|
| 18th Century | | | |
| Smallpox (1798) | | | |
| 19th Century | | | |
| Rabies (1885) | Typhoid (1896) | | |
| | Cholera (1896) | | |
| | Plague (1897) | | |
| Early 20th Century, first half | | | |
| Tuberculosis (bacille | Pertussis (1926) | Diphtheria toxoid (1923) | |
| Calmette-Guérin) (1927) | | | |
| Yellow fever (1935) | Influenza (1936) | Tetanus toxoid (1926) | |
| | Rickettsia (1938) | | |
| 20th Century, second half | | | |
| Polio (oral) (1963) | Polio (injected) (1955) | Anthrax secreted proteins (1970) | Hepatitis B surface antigen recombinant (1986) |
| Measles (1963) | Rabies (cell culture) (1980) | Meningococcus polysaccharide (1974) | Lyme OspA (1998) |
| Mumps (1967) | Japanese encephalitis (mouse brain) (1992) | Pneumococcus polysaccharide (1977) | Cholera (recombinant toxin B) (1993) |
| Rubella (1969) | Tick-borne encephalitis (1981) | Haemophilus influenzae type B polysaccharide (1985) | |
| Adenovirus (1980) | Hepatitis A (1996) | H.influenzae type b conjugate (1987) | |
| Typhoid (Salmonella TY21a) (1989) | Cholera (WC-rBS) (1991) | Typhoid (Vi) polysaccharide (1994) | |
| Varicella (1995) | Meningococcal conjugate (group C) (1999) | Acellular pertussis (1996) | |
| Rotavirus reassortants (1999) | S-October Commission | Hepatitis B (plasma derived) (1981) | |
| Cholera (attenuated) (1994) | | | |
| Cold-adapted influenza (1999) | | | |
| 21st Century | | | |
| Rotavirus (attenuated and new | Japanese encephalitis (2009) | Pneumococcal conjugates* | Human papillomavirus |
| reassortants) (2006) | (Vero cell) | (heptavalent) (2000) | recombinant (quadrivalent) (200 |
| Zoster (2006) | Cholera (WC only) (2009) | Meningococcal conjugates* | Human papillomavirus |
| | | (quadrivalent) (2005) | recombinant (bivalent) (2009) |
| | | Pneumococcal conjugates* | Meningococcal group B proteins |
| | | (13-valent) (2010) | (2013) |

Rational development

1- ATTENUATION – attenuated vaccines

The idea of attenuation of virulent infections developed slowly over the course of centuries. It was based on the idea that an agent virulent for animals might be attenuated in humans. Serial propagation of a pathogen in inhabitual hosts may lead to "attenuation"

Old examples: Jenner's use of an animal poxvirus to prevent smallpox

<u>Cell cultures.</u> In 1948-1949, a revolution happened with the discovery that cells could be cultured in vitro and used as substrates for viral growth. **Passage in cell culture leads to adaptation to growth in that medium,** and the mutants best capable of growth have often lost or modified the genes that allow them to infect and spread within a human host.

More recent examples of attenuation" in vitro in cell culture: measles, rubella, mumps, grown in cell culture

Rational development

2- INACTIVATION inactivated vaccines

Another discovery toward the end of the 19th century was that immunogenicity could be retained if microorganisms were carefully killed by heat or chemical treatment

3- PROTEIN-BASED VACCINES "subunit" vaccines

Some vaccines used today are **purified from microorganism preparation** or generated by growing the viruses in vitro and then breaking up the whole virus with detergents. Then viral protein is purified to serve as the vaccine antigen. Several vaccines consist of partly or fully purified proteins.

Sometimes that vaccine is a "conjugated "vaccine. Conjugation is a procedure that combines a type of subunit vaccine (with a weak antigen) with a strong antigen (as a carrier) so that the immune system has a stronger response to the weak antigen.

Rational development

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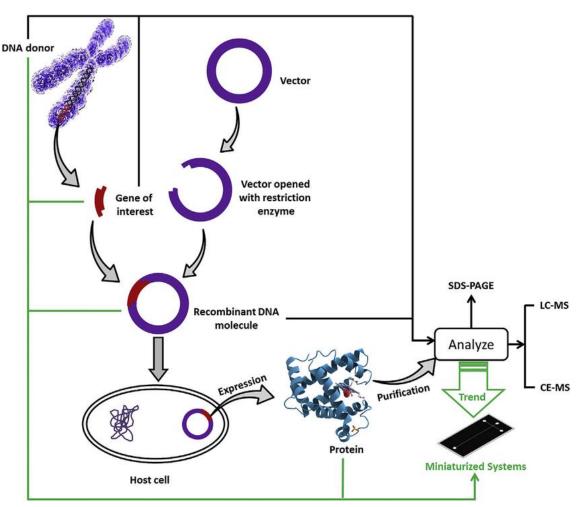
Sometimes that vaccine is a "conjugated "vaccine. Conjugation is a procedure that combines a type of subunit vaccine (with a weak antigen) with a strong antigen (as a carrier) so that the immune system has a stronger response to the weak antigen.

4- GENETIC ENGINEERING

The revolution of genetic engineering toward the end of the 20th century has greatly impacted vaccine development. The first fruit of that revolution was the vaccine against hepatitis B.

Vaccino - Proteine ricombinanti

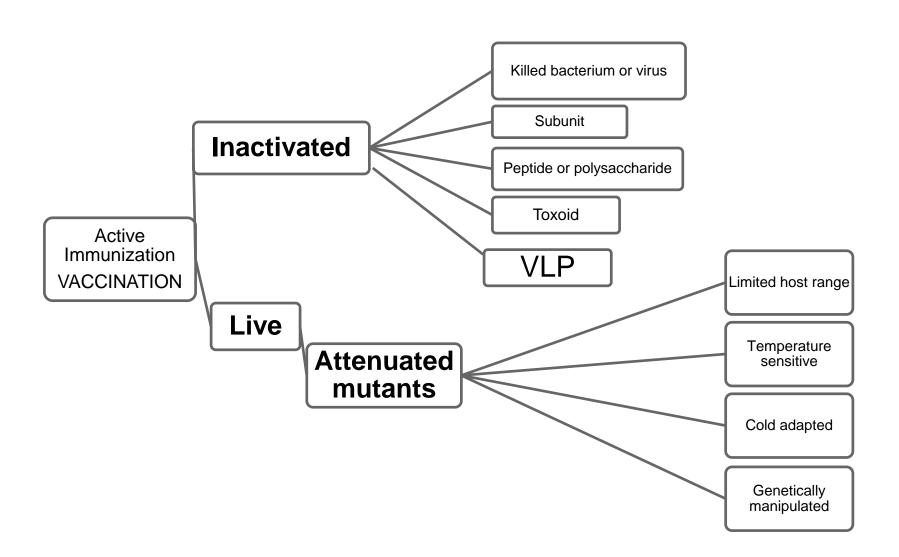
This technique is based on the possibility to allow the cells of the subject to be immunized to directly produce the antigens against which we want to induce an immune response. This is possible using a cDNA encoding the protein of interest carried by a plasmid

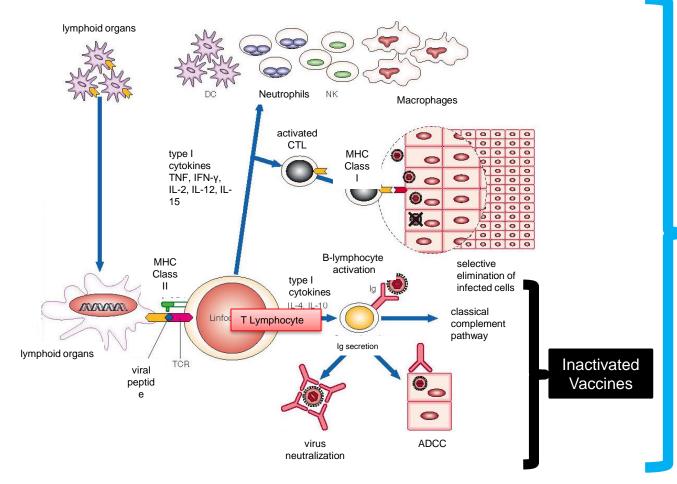


Gianini Morbioli G. eAnalltica Chimica Acta · September 2016

Strategies for the development of vaccines

| PROCEDURE | EXAMPLES |
|----------------------------------|--|
| | |
| attenuation, inactivation, grown | |
| and purification | many |
| no combinant DNA took note my | |
| recombinant DNA technology | HBV, HPV |
| | |
| conjugation technology | Haemophilus influenzae B |
| | Streptococcus pneumoniae (different serotypes) |
| | Neisseria meningitidis A |





General Rule

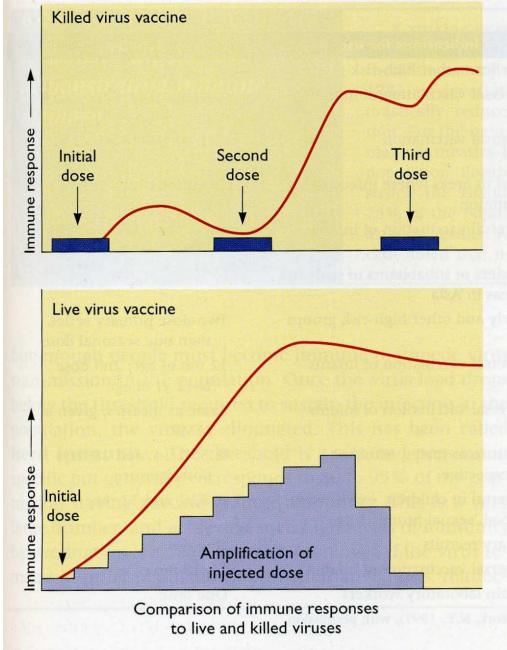


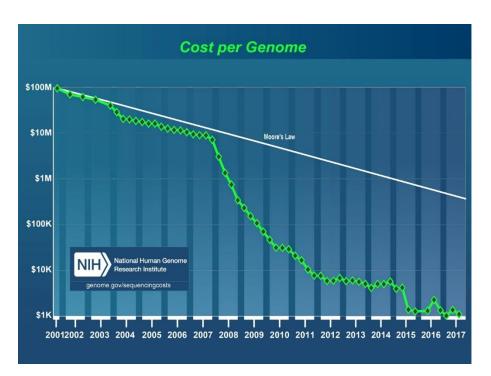
Figure 19.2 Comparison of the predicted immune responses to live and killed viruses used in vaccine protocols.

Further challenges for vaccinologists

| Challenges (examples) | Determinants (examples) | Microorganism (Examples) |
|--|---------------------------------------|--|
| | | |
| Severe infections | No availabilty of curative drugs, | HIV, HCV |
| | , , , , , , , , , , , , , , , , , , , | |
| Severe infections | Variability of the pathogen | Influenza |
| | | |
| | Incidence, frequency and | |
| Climate changes | distribution | vector-borne diseases, |
| | of several infection infections | ancient bacteria and viruses |
| | | |
| Demolation amountly and only a continu | Danislatian danaits | and a subject to a subject to the su |
| Population growth and urbanization | Population density | mosquito-trasmitted diseases, |
| in developing countries | | leptospirosis, rabies, etc. |
| | | |
| | | Methicillin-resistant staphyloccocus aureus |
| Antibiotic resistance | Hospital acquired infections | (MRSA) |
| | Misuse of antibiotics | Pseudomonas aeruginosa |
| | | Clostridium difficile |
| | | |
| | | |
| Emerging infections | The number of emerging viruses | Norovirus, SARS-CoV-1, MERS, SARS-CoV-2 |
| | is significantly increasing | |

| Year of first description | Name | Deaths | Comments |
|---------------------------|----------------------------------|---|--|
| 1918 | 'Spanish influenza' | In the range of about 50 million to 100 million | 1918: H1N1; other pandemics in 1957-1958 (H2N2), 1968 (H3N2) and 2009 (H1N1) |
| 1931 | Rift Valley Fever | Overall CFR < 1%; ~50% for hemorrhagic fever | Contact with blood or organs of infected animals and mosquito-borne; several outbreaks in 1977, 1997–1998, 2000–2016 |
| 1937 | West Nile fever | CFR~5% | Mosquito-borne; worldwide outbreaks (most recent 1999-2010, USA) |
| 1967 | Marburg hemorrhagic fever | ~470; very high CFR (24-88%, WHO) | Contact with African green monkey; numerous outbreaks in Africa 1969–2018 |
| 1969 | Lassa fever | ~5,000 deaths annually; CFR 1–2%; Nigerian CFR 25% | Contact with rodents or contaminated food or items; mostly in West Africa (Nigeria 2018) |
| 1969 | Acute hemorrhagic conjunctivitis | Rare | First identified in 1969; pandemic in 1981; frequent outbreaks worldwide |
| 1976-2020 | Ebola hemorrhagic fever | >15,000; CFR 75% | First identified in 1976; first major outbreak in 2013–2016 in West Africa and in 2018 in Democratic Republic of Congo; 29 regional epidemics in 2020 in West and Central Africa |
| 1981 | HIV/AIDS | ~37 million | Ongoing pandemic |
| 1996 | Avian flu | High CFR (60%) | H5N1 and H7N9 viruses from poultry; several outbreaks worldwide; last outbreak in China in 2018 |
| 1999 | Nipah fever | <1,000?; very high CFR | Outbreaks in Malaysia, Singapore, Bangladesh and India |
| 2002 | SARS | 813; CFR~10% | Contained—did not turn into pandemic |
| 2009 | H1N1; H7N9 'swine flu' | 284,000; CFR 2.9-9% | Pandemic |
| 2012 | MERS | 935; CFR 34.4% | Major outbreak in 2012–2019; ongoing (camels, humans); detected in 27 countries but mostly in Middle Eastern countries |
| 2014 | Chikungunya | Rare | Mosquito-borne |
| 2015 | Zika | Unknown | Mosquito-borne |
| 2019-ongoing | COVID-19 (SARS-CoV-2) | >2.3 million; CFR 2-10%; high in elderly and individuals with comorbidities | Pandemic—animal-to-animal, animal-to-human and human-to-human transmission |

Next generation sequencing and the metagenomic revolution



| Sequencing technology | | | |
|-----------------------|-------------|----------------------|--------------------|
| | 4: | | |
| year | time needed | cost | needs |
| 2000 | 10 years | 1 billion US dollars | world-wide teams |
| 2010 | 10 days | 50,000 US dollars | 1 laboratory group |
| now | 1 day | 1,000 US dollars | |

Strategies for the development of new vaccines

Using the computer to rationally design vaccines starting with information present in the genome, without the need to grow the specific microorganisms.

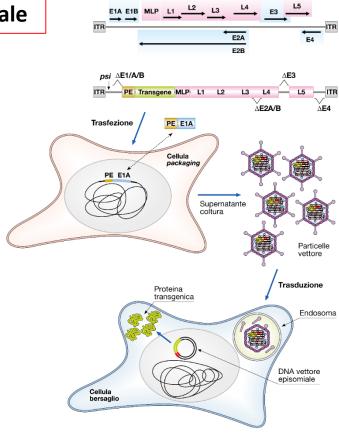
Strategies for the development of vaccines

| PROCEDURE | EXAMPLES |
|---------------------------------|--|
| | |
| growth, isolation, inactivation | many |
| | |
| recombinant DNA technology | HBV, HPV |
| | |
| conjugation technology | Haemophilus influenzae B |
| | Streptococcus pneumoniae (different serotypes) |
| | Neisseria meningitidis A |
| | |
| genomics; genome-based | |
| approach | Ebola virus, Influenza virus, SARS-CoV-2 |

Vaccini a DNA - vettore adenovirale

Questa tecnica si basa sulla possibilità di far produrre direttamente alle cellule dell'individuo da immunizzare gli antigeni contro i quali si vuol indurre una risposta immunitaria. Questo è possibile utilizzando un cDNA che codifica la proteina di interesse veicolata da un virus non citopatico o difettivo (vettore virale)

A cDNA encoding the protein of interest carried by a noncytopathic or defective virus (viral vector)



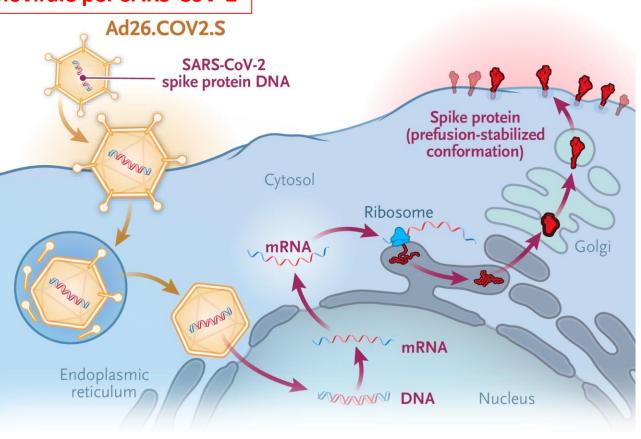
Pentoni / Core / Esoni

Ceppo parentale

Figura 70.3 Produzione e trasduzione con vettore adenovirale.

Vaccini a DNA/vettore adenovirale per SARS-CoV-2

Questa tecnica si basa sulla possibilità di far produrre direttamente alle cellule dell'individuo da immunizzare la proteina S del SARS-CoV-2 contro la quale si vuol indurre una risposta immunitaria. Questo è possibile utilizzando un cDNA che codifica la proteina di interesse veicolata da un adenovirus difettivo.



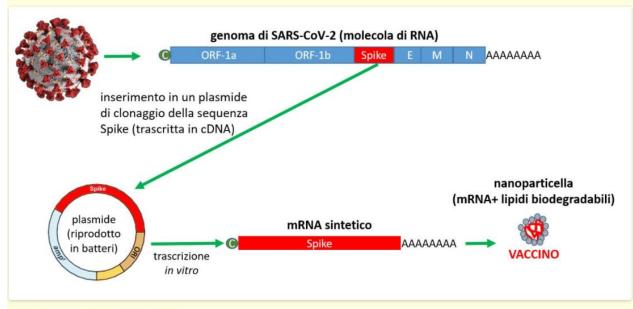
A cDNA encoding the S-protein of SARS_CoV-2 carried by a non-cytopathic or defective virus (viral vector)

Sadoff | et al. DOI: 10.1056/NEJMoa2101544

Vaccini a RNA – SARS-CoV-2 RNA messaggero

Questa tecnica si basa sulla possibilità di far produrre direttamente alle cellule del soggetto vaccinato gli antigeni contro i quali si vuol indurre una risposta immunitaria.

Questo è possibile utilizzando un RNA messaggero corrispondente alla proteina S di SARS-CoV-2 veicolata da un liposoma per far entrare l'RNA nella cellula.



[Disegno: Ferdinando Di Cunto]

l **liposomi** sono vescicole con un diametro fra i 25 nm e 1 μ m le cui pareti sono composte da colesterolo e lipidi e sono in grado di veicolare al loro *interno* diverse sostanze, ad esempio farmaci o vaccini.

I **virosomi** sono strutture assimilabili ai liposomi che contengono proteine presenti sulla superficie dei virus, sia incapsulate al loro interno sia al loro esterno.

a messenger RNA corresponding to the protein of interest carried by a virosome or liposome to let the RNA enter the cell.

SARS-CoV-2: Frontiere della ricerca



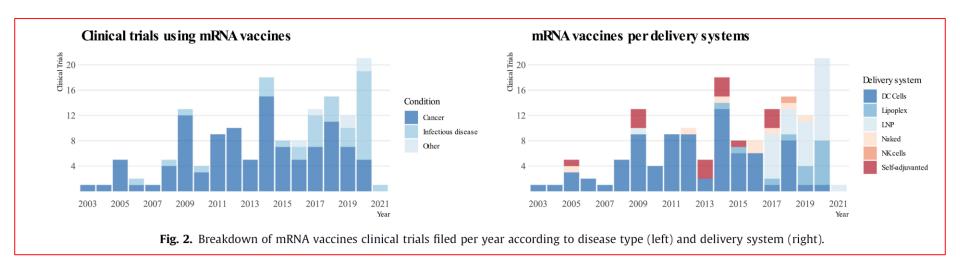


Table 4 | Leading mRNA vaccine developers: research focus, partners and therapeutic platforms

| Institution | mRNA technology | Partners | Indication (disease target) |
|-------------------------------|---|------------------------------|---|
| Argos Biotechnology | mRNA neoantigens (Arcelis platform) | NA | Individualized cancer vaccines, HIV-1 |
| BioNTech RNA | Nucleoside-modified mRNA (IVAC Mutanome, FixVAC) | Genentech/Roche | Individualized cancer vaccines |
| Pharmaceuticals GmbH | | Bayer AG | Veterinary vaccines |
| CureVac AG | Sequence-optimized, purified mRNA (RNActive, RNArt, RNAdjuvant) | Boehringer Ingelheim GmbH | Cancer vaccines (lung cancer) |
| | | Johnson & Johnson | Viral vaccines |
| | | Sanofi Pasteur | Infectious disease vaccines |
| | | BMGF | Infectious disease vaccines |
| | | IAVI | HIV vaccines |
| eTheRNA Immunotherapies | Purified mRNA (TriMix) | NA | Cancer (melanoma, breast), viral vaccines (HBV and/or HPV) |
| GlaxoSmithKline/ Novartis | Self-amplifying mRNA (SAM) (alphavirus replicon) | NA | Infectious disease vaccines |
| Moderna Therapeutics | Nucleoside-modified mRNA | Merck & Co. | Individualized cancer vaccines, viral vaccines |
| | | BMGF, DARPA, BARDA | Viral vaccines (influenza virus, CMV, HMPV, PIV, chikungunya virus, Zika virus) |
| University of Pennsylvania | Nucleoside-modified, purified mRNA | NA | Infectious disease vaccines |

BARDA, Biomedical Advanced Research and Development Authority; BMGF, Bill & Melinda Gates Foundation; CMV, cytomegalovirus; DARPA, Defense Advanced Research Projects Agency; HBV, hepatitis B virus; HMPV, human metapneumovirus; HPV, human papillomavirus; IAVI, International AIDS Vaccine Initiative; NA, not available; PIV, parainfluenza virus.

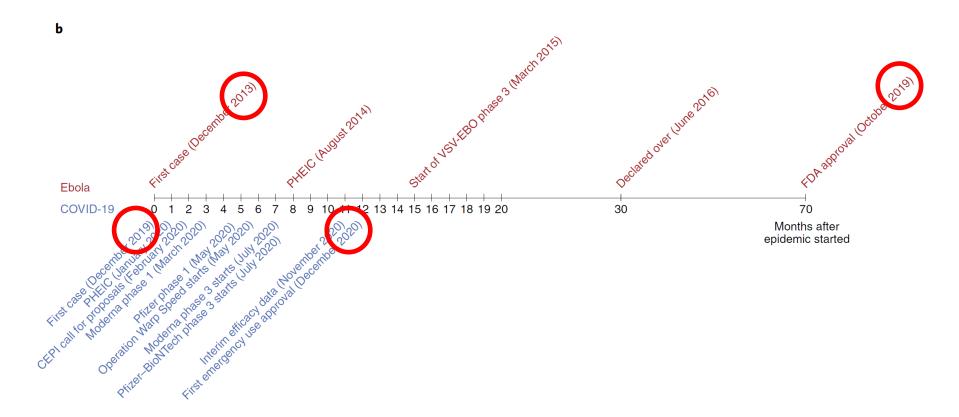
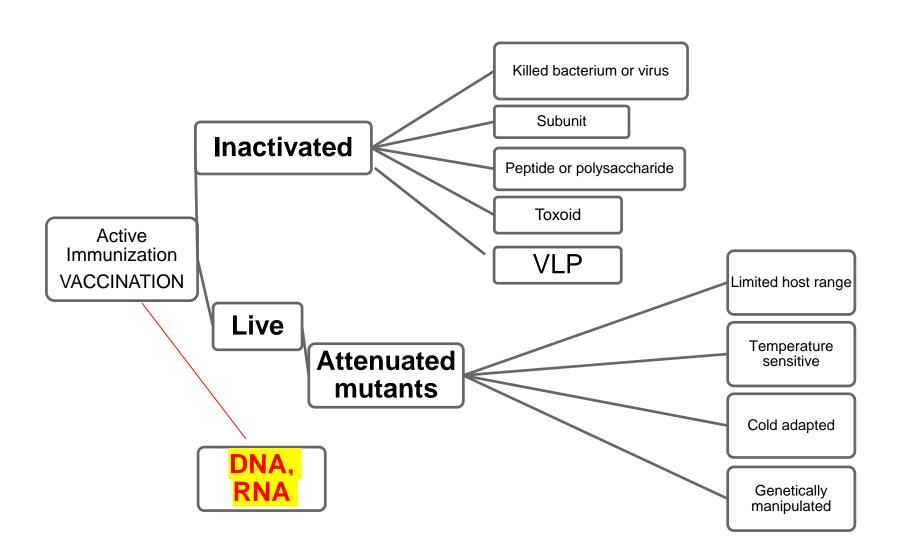
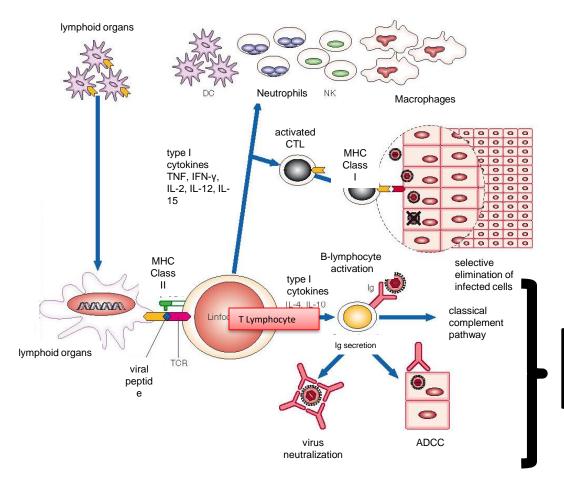


Table 2 | Examples of different vaccine platforms and vaccines currently developed or under development for emerging viral infectious diseases Vaccine platform Other specifications Under development or Developed for Shortcomings and advantages stopped^a for Live attenuated Influenza; yellow fever; COVID-19; RVF Biosafety level 3 manufacturing plant for poliomyelitis (veterinary and human handling dangerous viruses use) Lassa fever; chikungunya Whole inactivated With or without adjuvant Influenza; poliomyelitis; SARS^a; Zika; RVF Biosafety level 3 manufacturing plant for COVID-19 (veterinary use); dangerous viruses; needs adjuvant; HPB chikungunya regimens possible DNA Electroporation; adjuvant SARS^a; MERS; Zika; Lassa Poorly immunogenic; electroporation fever; COVID-19 requires device; difficult use for rollout; HPB regimens possible mRNA COVID-19 Lassa fever; disease X Rapidly adaptable to new emerging viruses; HPB regimens possible; ultracold chain currently unpractical for large-scale use in resource-limited settings Recombinant vectors Nonreplicating Ad5 COVID-19 Preexisting immunity to Ad5 ChAd3 Ebola Cell-line-produced; adaptable construct to emerging virus in MERS; RVF; Lassa fever; ChAdOx1 COVID-19 5-6 months; HPB regimens possible Nipah; Zika; chikungunya Ad26 Ebola; COVID-19 Live attenuated MVA Ebola **MERS** VSV Fbola COVID-19a; Lassa fever; Nipah Measles MERS: Lassa fever: Nipah; chikungunya; COVID-19ª Protein based Requires more time to adapt to new emerging viruses; likely needs adjuvant; With adjuvant COVID-19 COVID-19 Virus-like particle HPB regimens possible COVID-19: RFV: Nipah Monomer; dimer; With adjuvant trimer Molecular clamp With adjuvant Influenza; MERS; COVID-19^a

aVaccine development stopped.





Vaccines , Subunit, DNA, RNA

Inactivated

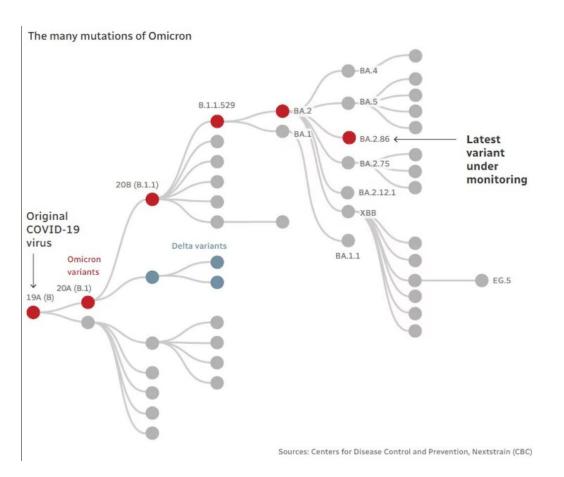
Viral Vaccines

| Virus | Vaccines Components | Who Should Receive Vaccinations |
|-----------------------|---|---|
| Polio, inactivated | Trivalent (Salk vaccine) | Children |
| Attenuated polio | Live (oral polio vaccine, Sabin vaccine) | Children |
| Measles | Attenuated | Children |
| Mumps | Attenuated | Children |
| Rubella | Attenuated | Children |
| Varicella-zoster | Attenuated | Children |
| Rotavirus | Human-bovine hybrids Attenuated | Infants |
| Human papilloma-virus | VLP | Girls aged 9-26 yr |
| Influenza | Inactivated Attenuated (nasal spray) | Children, adults, especially medical personnel, and the elderly 2-50 yr |
| Hepatitis B | Subunit (VLP) | Newborns, health care workers, high risk groups (e.g. sexually promiscuous, intravenous drug users) |
| Hepatitis A | Inactivated | Children, child care workers, travelers to endemic areas, Native Americans and Alaskans |
| Adenovirus | Attenuated | Military personnel |
| Yellow fever | Attenuated | Travelers at risk to exposure, military personnel |
| Rabies | Inactivated | Anyone exposed to virus Preexposure: veterinarians, animal handlers |
| Smallpox | Live vaccinia virus | Protection from bioterrorism, military |
| Japanese encephalitis | Inactivated | Travelers at risk to exposure |

Advantages and Disadvantages of Live versus Inactivated Vaccines

| Property | Live | Inactivated, "Subunit", DNA, RNA |
|-------------------------------|--------------------------|----------------------------------|
| Route of administration | Natural or injection | Injection |
| Dose of virus, cost | Low | High |
| Number of doses, amount | Single, low | Multiple, high |
| Need for adjuvant | No | Yes |
| Duration of immunity | Long-term | Short-term |
| Antibody response | IgG, IgA | IgG |
| Cell-mediated immune response | Good | Poor |
| Potential lability | Yes | No |
| Interference | Occasional | None |
| Side effects | Occasional mild symptoms | Rare |
| Reversion to virulence | Rarely | None |

Modified From Murray et al Medical Microbiology Elsevier 2016



BA.2.86 Variant- September 2023

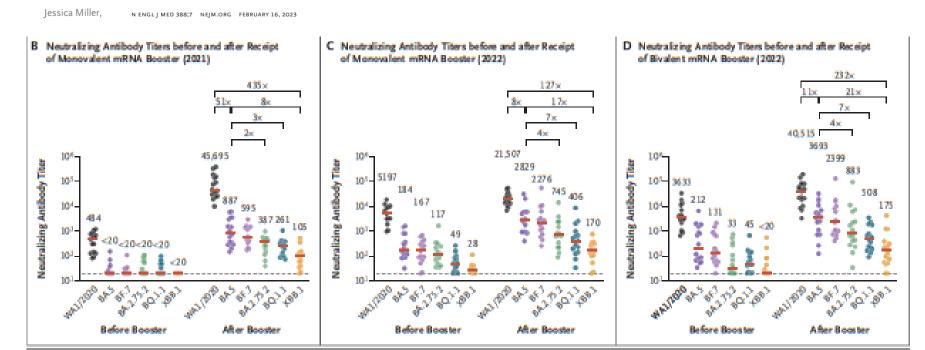
BA.2.86 Variant hyper-mutated variant has shown up in many places now. To date, the BA.2.86 variant has been detected in Israel, Denmark (3 individuals), the UK, the US (2 individuals, one coming back from Japan), and South Africa (2 individuals). It has also been detected in wastewater in 1 region in Switzerland (2% level), along with wastewater detection in Ohio and in Thailand. It's safe say that BA.2.86's presence is widespread across the world at this point.

BA.2.86 has probably been circulating in a region of the world with poor viral surveillance and

has now been repeatedly exported to other places in the world

The lineage seems to be descended from an Omicron subvariant called BA.2, which caused large case spikes in early 2022. However, the BA.2.86 spike protein carries 34 changes relative to BA.2. Large numbers of spike mutations have been observed in people with long-term SARS-CoV-2 infections, and it is likely that BA.2.86 emerged from one such chronic infection

Substantial Neutralization Escape by SARS-CoV-2 Omicron Variants BQ.1.1 and XBB.1



BQ.1.1 and XBB.1 variants escaped neutralizing antibodies substantially more effectively than the BA.5 variant by factors of 7 and 17, respectively, after monovalent mRNA boosting and by factors of 7 and 21, respectively, after bivalent mRNA boosting

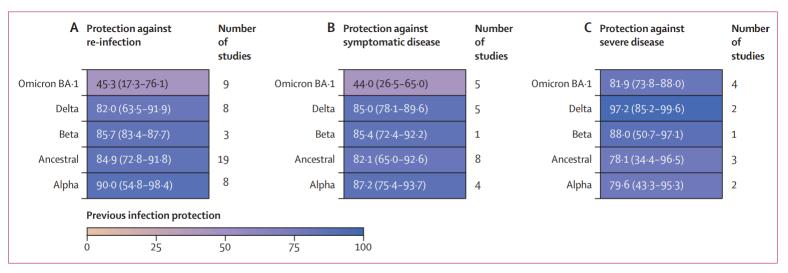
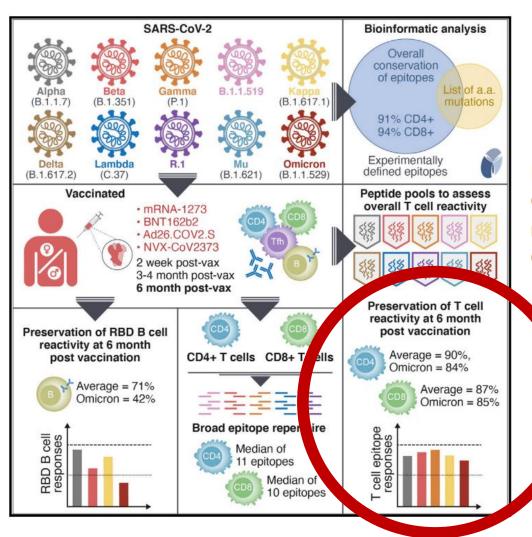


Figure 2: Pooled estimate of protection from past SARS-CoV-2 infection against re-infection, symptomatic disease, and severe disease by variant, and number of included studies in each meta-analysis estimate

Data are pooled estimate (95% uncertainty interval). Estimates of protection against re-infection (A), symptomatic disease (B), and severe disease (C).



- T cells of vaccinees recognize SARS-CoV-2 variants, including Omicron
- RBD memory B cells' recognition of Omicron is reduced

Human memory T cells induced by SARS-CoV-2 vaccines maintain the ability to recognize viral variants, including the Omicron variant.

CellPress

Tarke et al., 2022, Cell 185, 847–859 March 3, 2022 © 2022 Elsevier Inc. https://doi.org/10.1016/j.cell.2022.01.015

Vaccination against infectious diseases: challenges

- Are broadly neutralizing antibodies an absolute necessity?
- We do not know how to generate long-lasting protective antibodies at mucosal surfaces
- There are multiple viral serotypes/genotypes and antigenic variation requires constant updating of vaccine formulations;
- For some infectious diseases, no validated immunological correlates of immunity (protective or pathological) do exist

The 'best' vaccine

Any effort to rank the vaccines must take into account their reported effectiveness, but also:

- their ability to fend off emerging viral variants
- the durability of the protection they offer
- the logistics of deploying them
- the supply and cost issues