Channelopathies

- Long QT syndrome
- Skeletal muscle channelopathies
- ALS associated channelopathies
- Epileptic channelopathies

>340 human genes code for Ionic Channels:

muscular and nervous excitation

hormonal secretion

cell proliferation

signal transduction

learning and memory

Blood pressure

hydro-salt balance

cell death

Mutations in> 60 human genes that code for Ionic Channels have been associated with diseases

CHANNELOPATHIES = diseases resulting from the failure of ion channels (mutations in genes coding for ion channels or accessory regulatory subunits)

original sequence

codon	ACC	ATC	GGT	TAT	GGC
amino acid	T	I	G	Y	G

point mutation

codon	ACC	ATC	**A**GT	TAT	GGC
amino acid	T	I	**S**	Y	G

Nonsense mutation

codon	ACC	ATC	AGT	TA**G**	GGC
amino acid	T	I	G	*	

frameshift

codon	ACC	**G**AT	CGG	TTA	TGG	C
amino acid	T	**D**	**R**	**L**	**W**	

Mutations alter the structure and function of ion channels

Loss of function mutations:

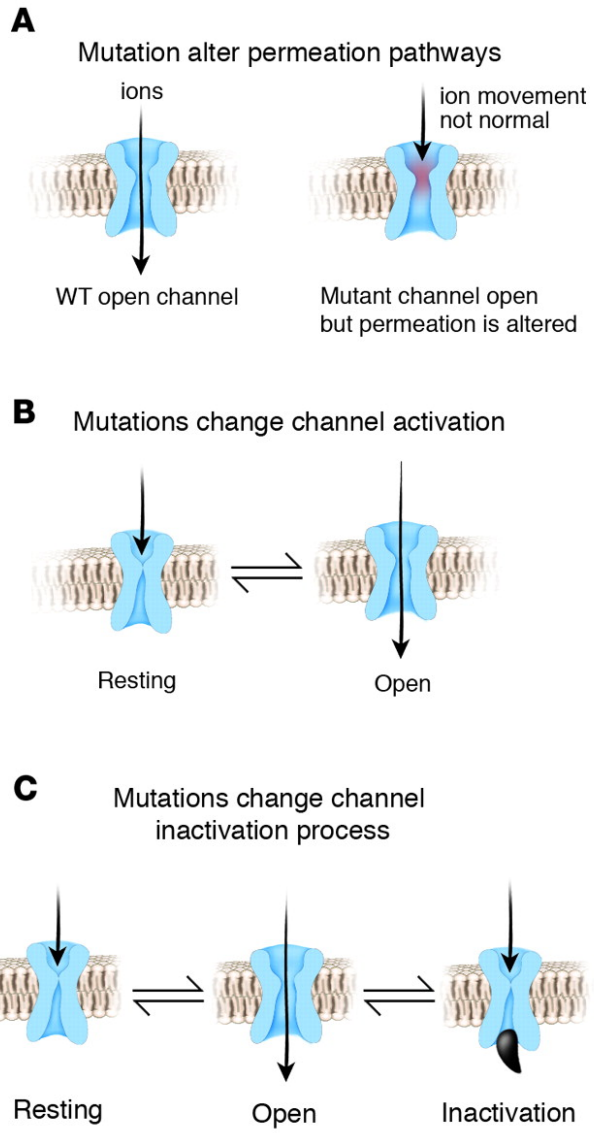
lack of protein synthesis

failed or incorrect insertion in the membrane

destruction of ligand binding sites

or proteins / accessory subunits ("modulation")

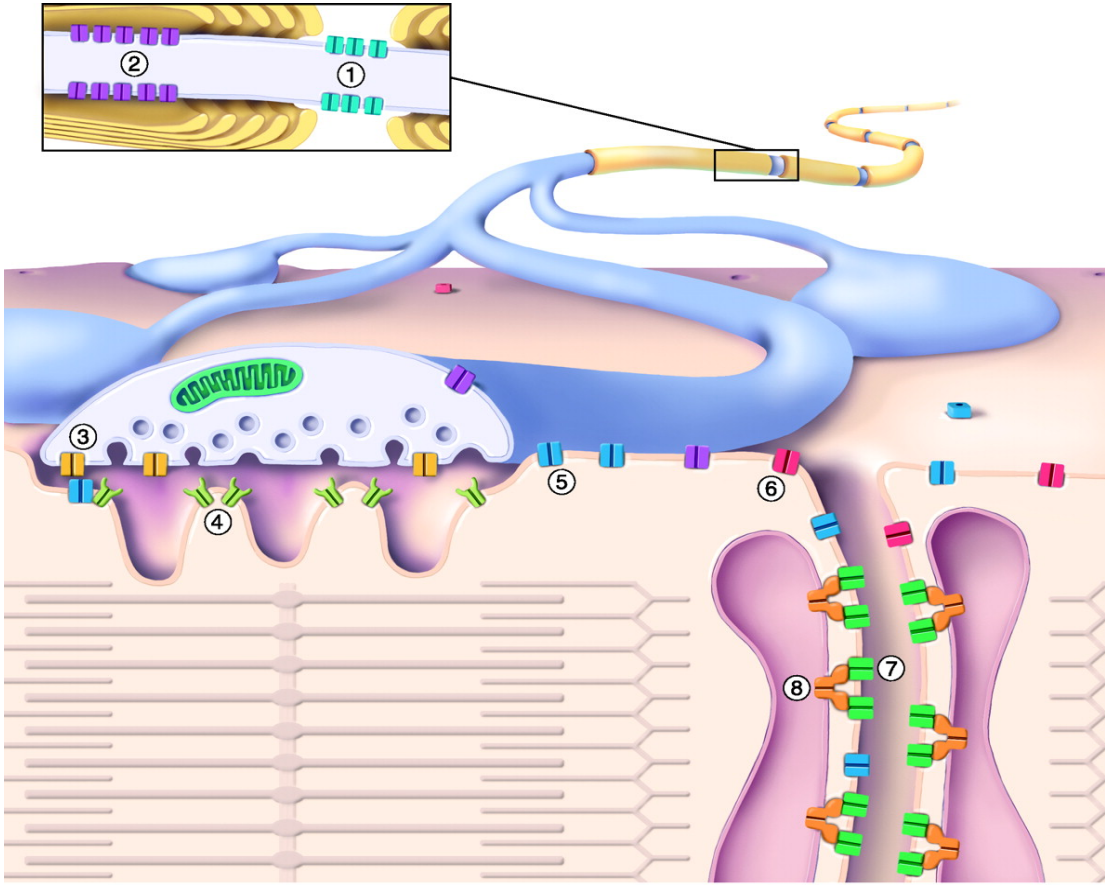
Gain of function mutations



Kass, R. S. J. Clin. Invest. 2005;115:1986-1989

Long QT syndrome

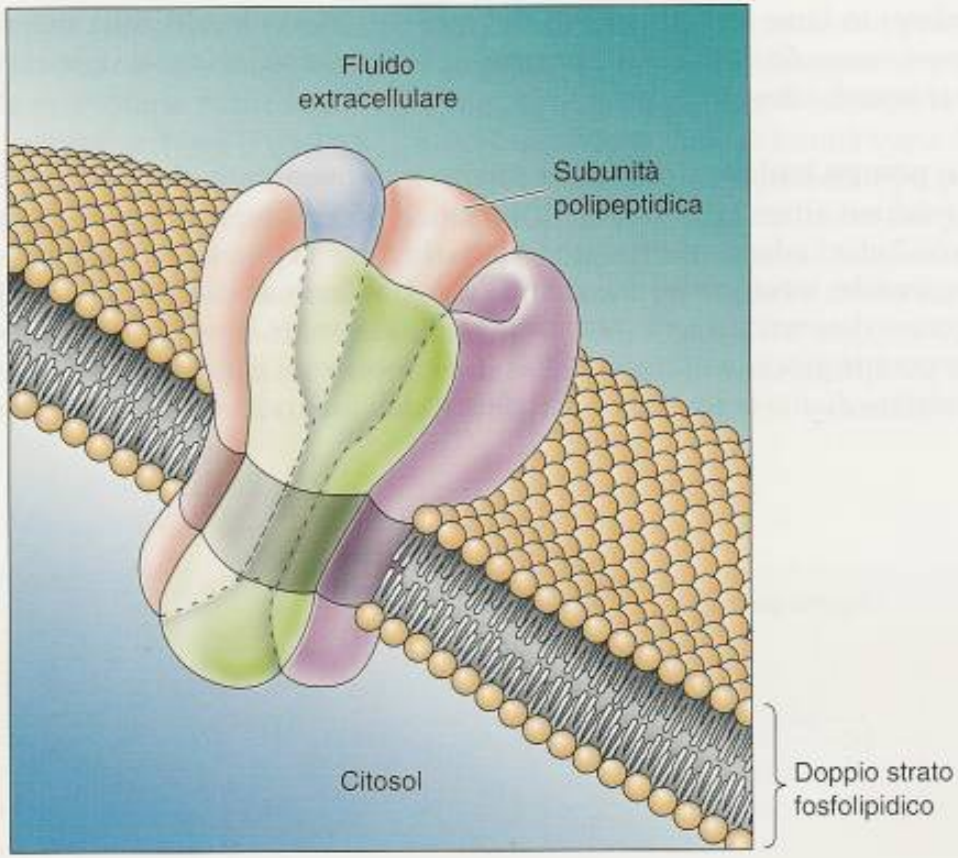
Pathologies of ion channels



① Nerve voltage-gated sodium channel
② KCNA voltage-gated potassium channel
③ Nerve voltage-gated calcium channel
④ Nicotinic acetylcholine receptor
⑤ Skeletal muscle voltage-gated sodium channel
⑥ Skeletal muscle voltage-gated chloride channel
⑦ Transverse tubule voltage-gated calcium channel
⑧ Sarcoplasmic reticulum calcium release channel

Cooper, Edward C. and Jan, Lily Yeh (1999) Proc. Natl. Acad. Sci. USA 96, 4759-4766

Figura 3.7
I canali ionici di membrana. I canali ionici sono costituiti da proteine che attraversano la membrana, unite tra loro a formare un poro. In questo esempio il canale proteico è costituito da cinque subunità polipeptidiche. Ciascuna subunità è dotata di una regione superficiale idrofobica (ombreggiata) che si unisce efficacemente al doppio strato fosfolipidico.



Properties of Ionic Channels.

- **Selectivity = ability to discriminate between one ionic species and another**

  dimension

  charge

- **Gating = transition process between an "open" and "closed" state**

  Ligand-gated channel

  Voltage-gated channel

  Temperature

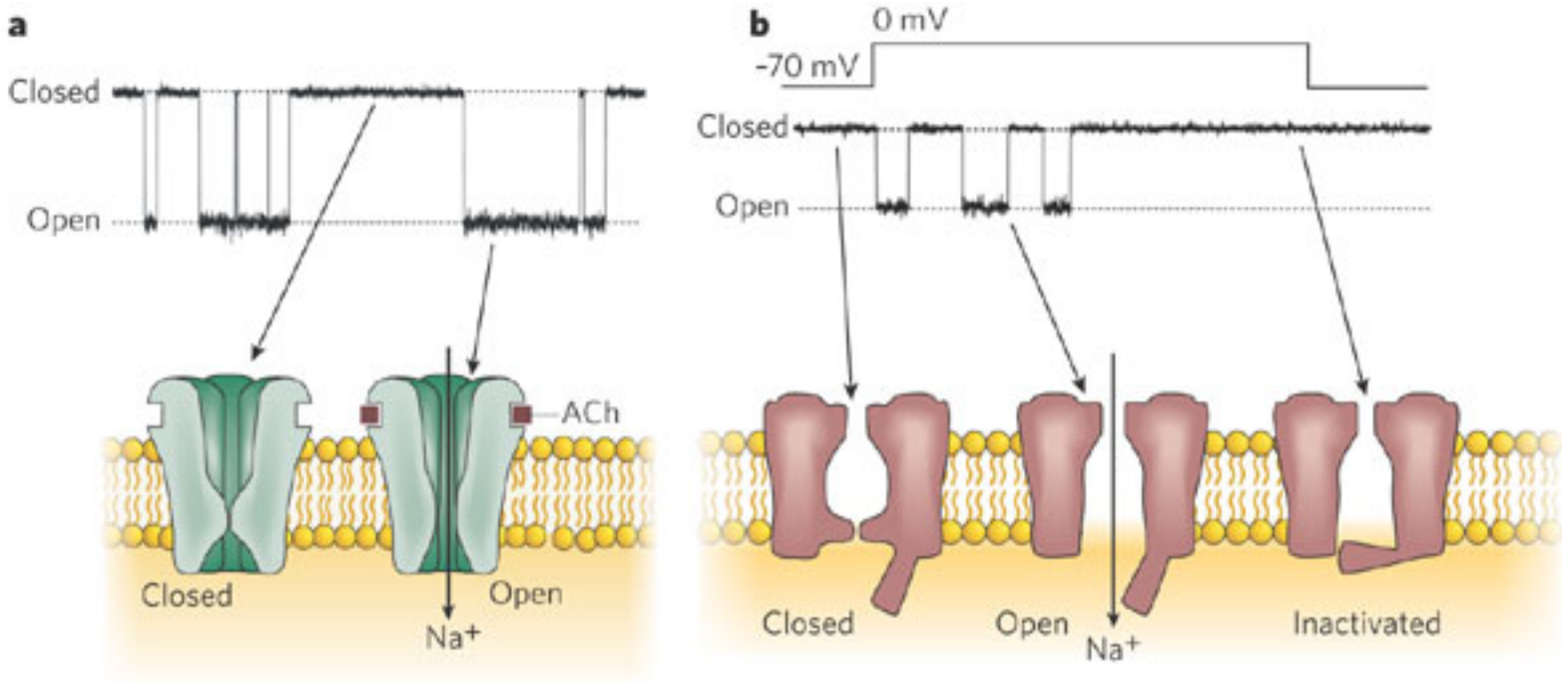
  Mechanical Stress

- **Modulation (phosphorylation)**

Gating = transition process between an "open" and "closed" state

Ligand-gated channel

Voltage-gated channel



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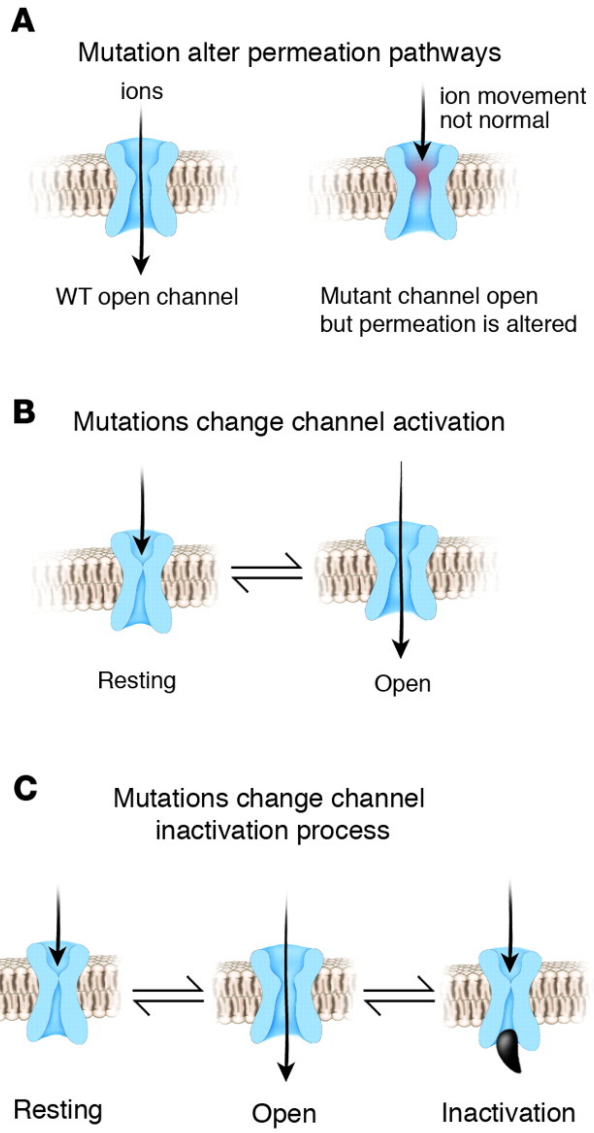


Kass, R. S. J. Clin. Invest. 2005;115:1986-1989

CARDIAC ACTION POTENTIAL

Different types of cells are distinguished: nodal (sinus-atrial node and atrioventricular node); conduction (His bundle and Purkinije fibers, common or working) The electrical and membrane mechanisms are similar to those already seen: we will focus mainly on the differences.

Heart cardiomyocytes: potential in 5 phases

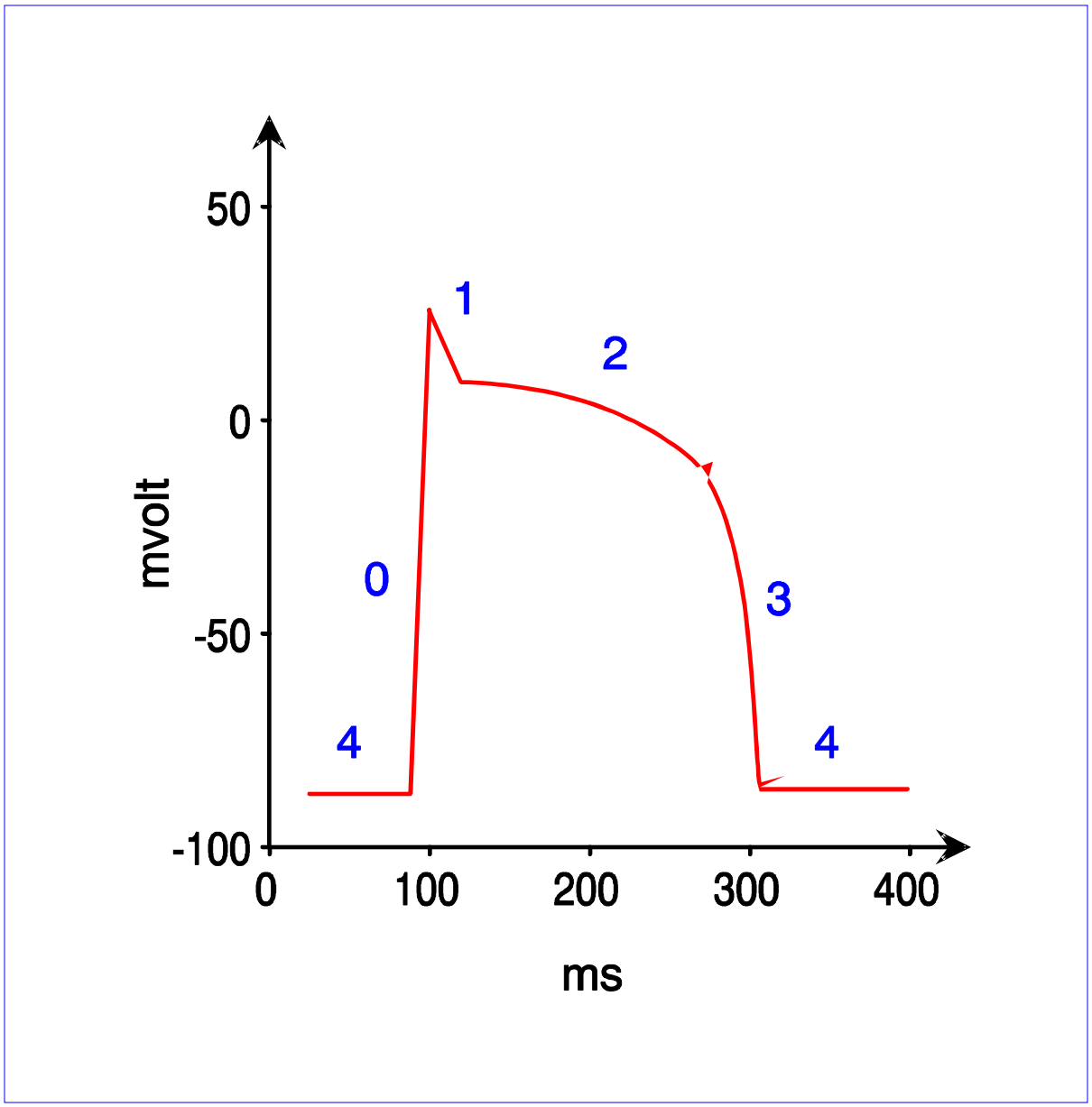
0 - rapid depolarization for opening of voltage-gated sodium channels

1 - partial short repolarization due to transient increase in chloride and potassium conductance

2 - plateau: stable potential on slightly positive values for about 0.2 s; due to the increase in calcium conductance (opening of "slow channels") and reduction of K conductance

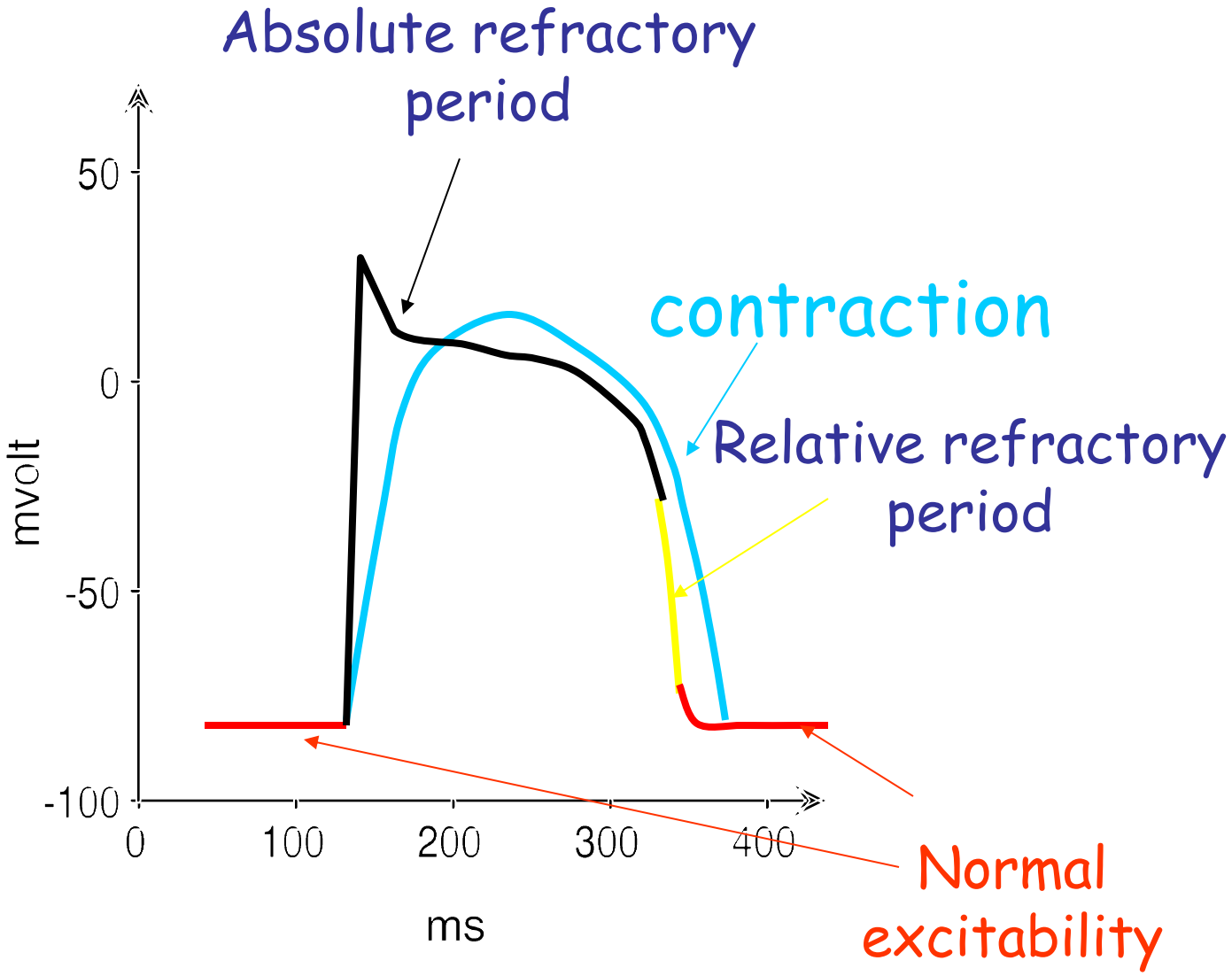
3 - repolarization due to progressive increase in potassium conductance and closure of slow channels;repolarization due to progressive increase in potassium conductance and closure of slow channels;

4 - resting potential, stable at -90 mV.


50
0
-50
-100
mvolt
0
100
200
300
400
ms
0
1
2
3
4
4

During the plateau a calcium current occurs, very important for the electromechanical coupling and for the regulation of contractility

Changes in excitability during action potential: refractory periods. The mechanical response appears during the potential and has approximately the same duration: the heart can not be tetanized


Absolute refractory period
contraction
Relative refractory period
Normal excitability
50
0
-50
-100
mvolt
0
100
200
300
400
ms

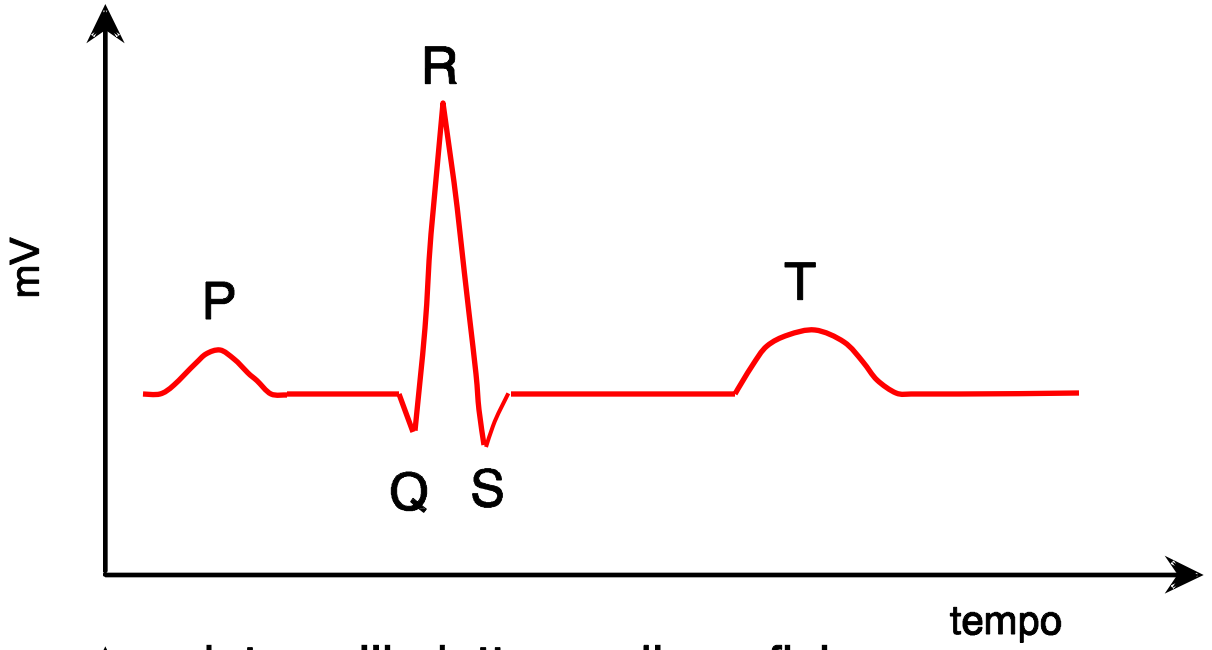
ELECTROCARDIOGRAM:

physiological bases; arrangementof the electrodes in the derivationsstandard;

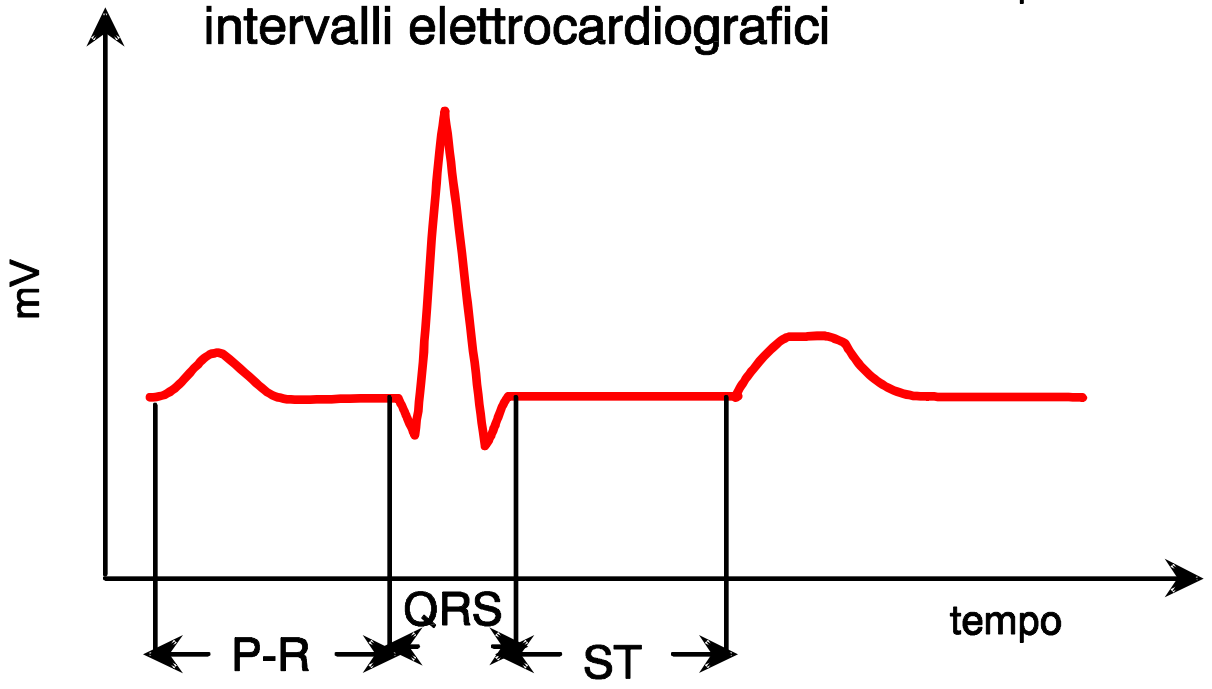
ECG waves.

What he says ewhat the ECG does not say

onde elettrocardiografiche

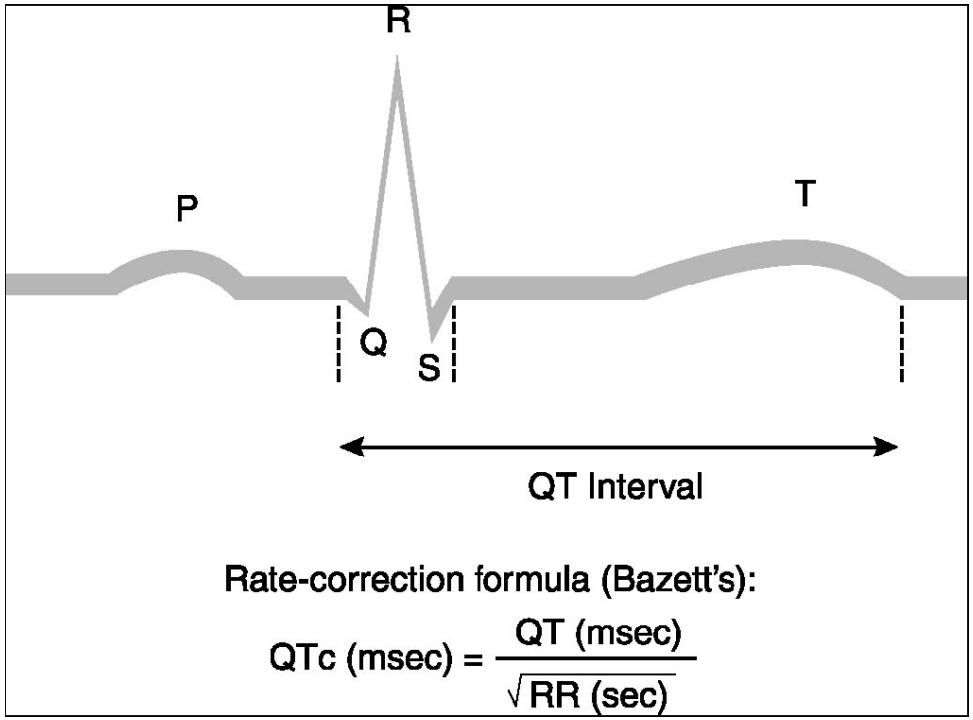


intervalli elettrocardiografici



Long QT syndrome (LQTS)

The congenital (idiopathic) form of long QT (LQTS) is mostly caused by gene mutationswhich encode proteins for cardiac ionic channel subunits



Among the various genotypes responsible for LQTS, the most common characteristic predisposing to arrhythmias is the lengthening of the ventricular action potential during cardiac repolarization, measured as the QT interval in the ECG.

LQT e morte improvvisa

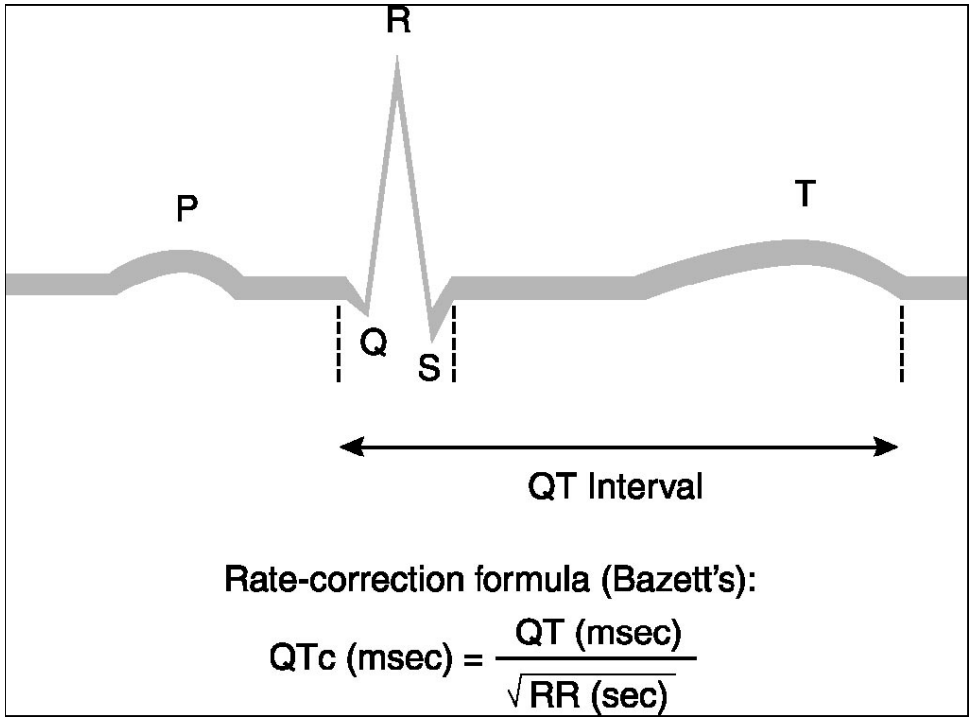
- La morte improvvisa e senza causa apparente nei neonati è stata a lungo spiegata come squilibrio funzionale del sistema nervoso autonomo.

- In realtà si tratta di *canalopatie*, disturbi della funzionalità dei canali ionici voltaggio dipendenti, che sottendono l'attività elettrica del cuore.

- Si ha un rallentamento della ripolarizzazione delle cellule cardiache dovuto a modificazioni geniche di alcuni canali ionici , quindi un prolungamento della durata del potenziale d'azione cardiaco (prolungamento dell'intervallo QT *long-QT syndrome*, LQTS) nell'elettrocardiogramma

La sindrome del QT lungo (LQTS)

può essere ereditaria o acquisita
i pazienti affetti sono predisposti ad improvvisa insorgenza di aritmie ventricolari pericolose (tipica è la torsione di punta) che si possono manifestare come episodi sincopali o, nei casi più gravi possono causare morte improvvisa da arresto cardiaco



La LQTS è una malattia genetica trasmessa come carattere autosomico dominante, dovuta a mutazioni su almeno sei geni, tutti codificanti per canali ionici responsabili del controllo dell'attività elettrica delle cellule cardiache.

Mutations

- ***Molecular genetic aspects: Channelopathy***
  - *7 genes have been linked to LQTS*
  - *mutations of the fast and slow K+ - channel (loss of function, reduces repolarizing K currents)*
  - *mutations of the Na+-channel (gain of function, increases inward INa currents)*

prolongation of the repolarization

- ***congenital QT-syndrome***
- ***acquired QT-syndrome (drug induced)***

Table 1
Genes and electrophysiology[a]

LQT subtype	Gene name	Locus	Configurations with 2 variant gene copies[b]	Encoded protein	Ion current affected	Effect of mutation	Common triggers [5]	Spectrum of ECG findings [6]
LQT1	KvLQT1, KCNQ1	11p15.5	Homozygous mutations cause Jervell and Lange-Nielsen syndrome (JLNS)[4][c]; compound heterozygous mutations described	Alpha subunits forming a tetramer	I_{Ks}	Loss-of-function; rare gain-of-function with short QT syndrome has been observed[d]	Exercise, esp. swimming; emotional stress	Normal appearing T-wave; broad based T-wave; late-onset normal-appearing T-wave
LQT2	HERG, KCNH2	7q35-36	Homozygous and compound heterozygous mutations described; homozygous may present with congenital AV block	Alpha subunits forming a tetramer	I_{Kr}	Loss-of-function; rare gain-of-function with short QT syndrome has been observed	Rest/sleep, auditory stimuli, emotional stress; postpartum state	Subtle, obvious, or widely split notched/bifid T-wave; low-amplitude T-wave
LQT3	SCN5A	3p21-24	Homozygous and compound heterozygous mutations described	Four-domain alpha subunit	I_{Na}	Gain-of-function; loss-of-function mutations lead to varied presentations (Brugada syndrome, conduction system disease)	Rest/sleep	Late-onset peaked, biphasic T-wave; asymmetrical peaked T-wave
LQT4	ANKB, ANK2	4q25-27		Membrane anchoring protein	Affects Na^+, K^+, Ca^{2+} exchange	Loss-of-function	Exercise, emotional stress (based on limited data)	Sinus bradycardia; inverted, bifid, or low-amplitude T-wave; inconsistent QT prolongation; prominent U-wave; frequent PVCs
LQT5	minK, IsK, KCNE1	21q22.1-2	Homozygous mutations can cause JLNS [4]; (2) compound heterozygous mutations described	Beta subunit to KCNQ1	I_{Ks}	Loss-of-function	(Insufficient data)	(Insufficient data)
LQT6	MiRP1, KCNE2	21q22.1		Beta subunit to HERG	I_{Kr}	Loss-of-function	(Insufficient data)	(Insufficient data)
LQT7	Kir2.1, KCNJ2	17q23		Kir2.1 subunits forming a tetramer	I_{K1}	Loss-of-function; rare gain-of-function with short QT syndrome has been observed	Accompanied by alterations in serum K^+ level in some cases	Prominent U-wave; prolonged terminal T downslope; modest QT prolongation; frequent PVCs; bigeminy; bidirectional ventricular tachycardia
LQT8	CACNA1C	12p13.3		Alpha subunits forming a tetramer	$I_{Ca,L}$	Gain-of-function	Hypoglycemia, sepsis (2 cases)	Severe QT prolongation (up to 650 ms); 2:1 atrioventricular block; overt T-wave alternans

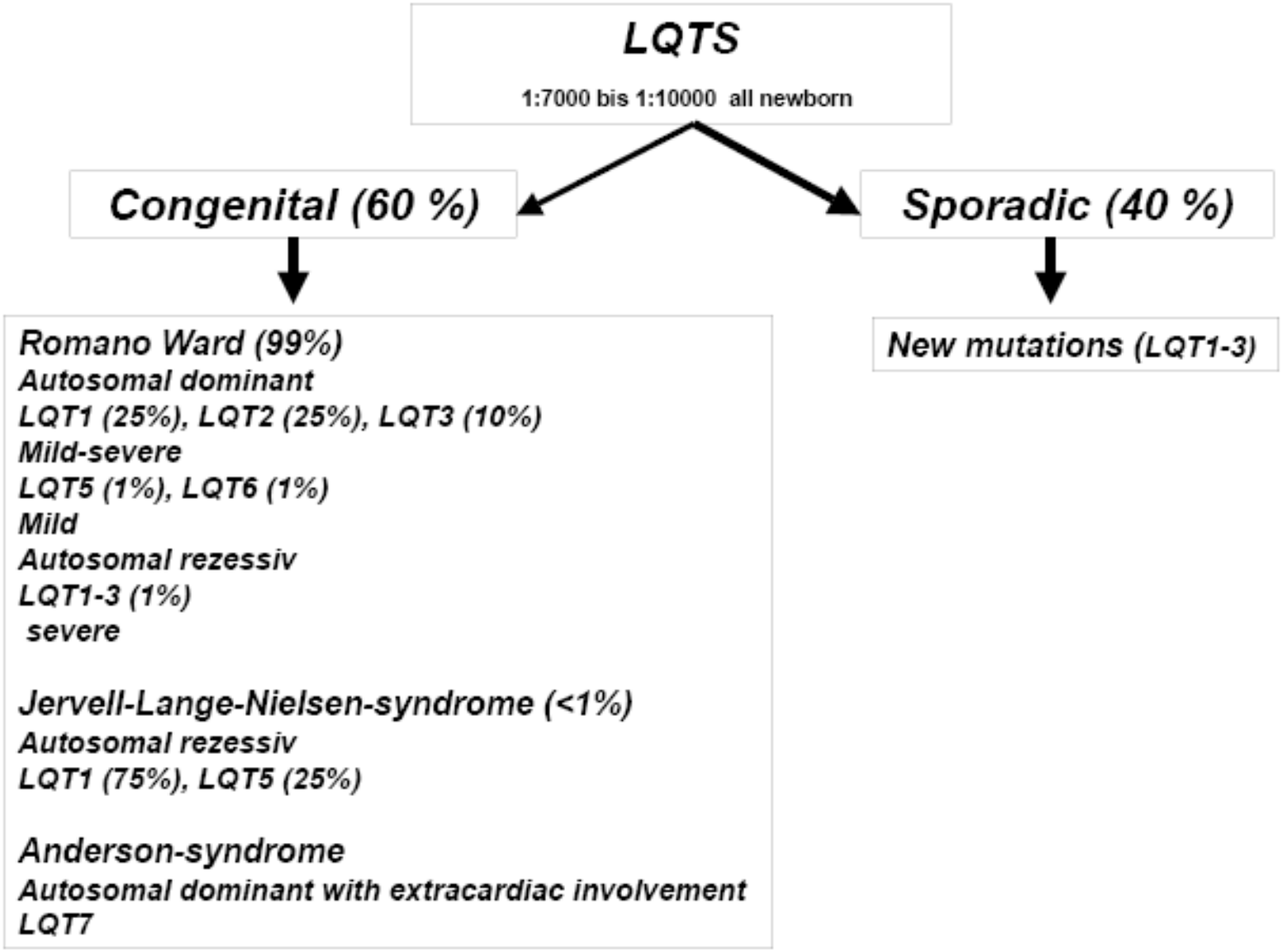
[a]Table completed 6/29/05.
[b]The normative LQTS gene configuration is heterozygous with one variant gene copy.
[c]Numbers with brackets refer to citations in Reference section.
[d]See Disease Associations section.

Genetics

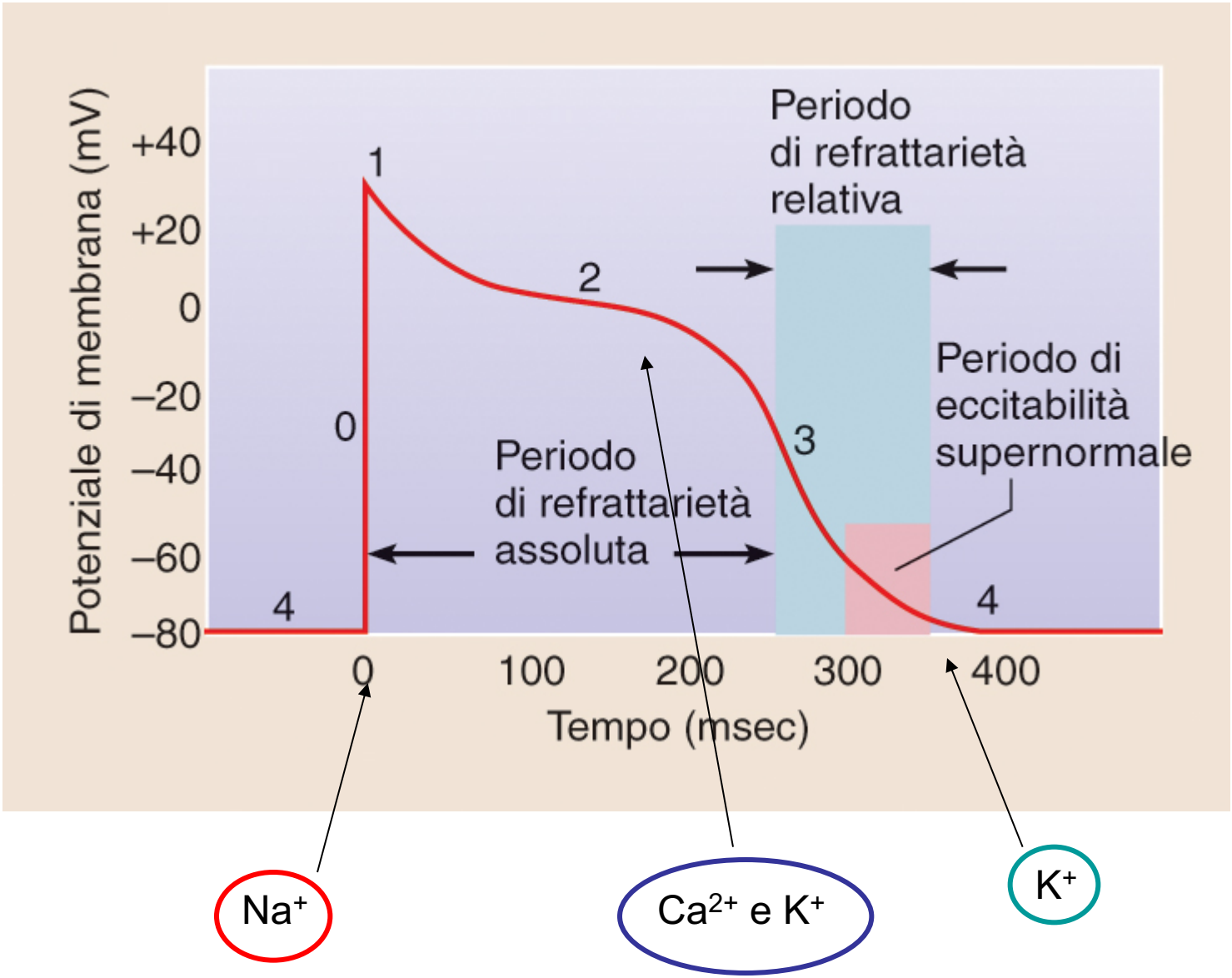
- 7 Genes on chromosoms 3, 4, 7, 11 and 21 identified
- Most frequent are mutations on KCNQ1-gene (LQT1 30%) and on KCNH2-gene (LQT2 30%).

LQT1-Syndrom:	KCNQ1, mutations slow K-channel
LQT2-Syndrom:	KCNQ2 mutations fast K-channel
LQT3-Syndrom:	SCN5A, mutations Na-channel

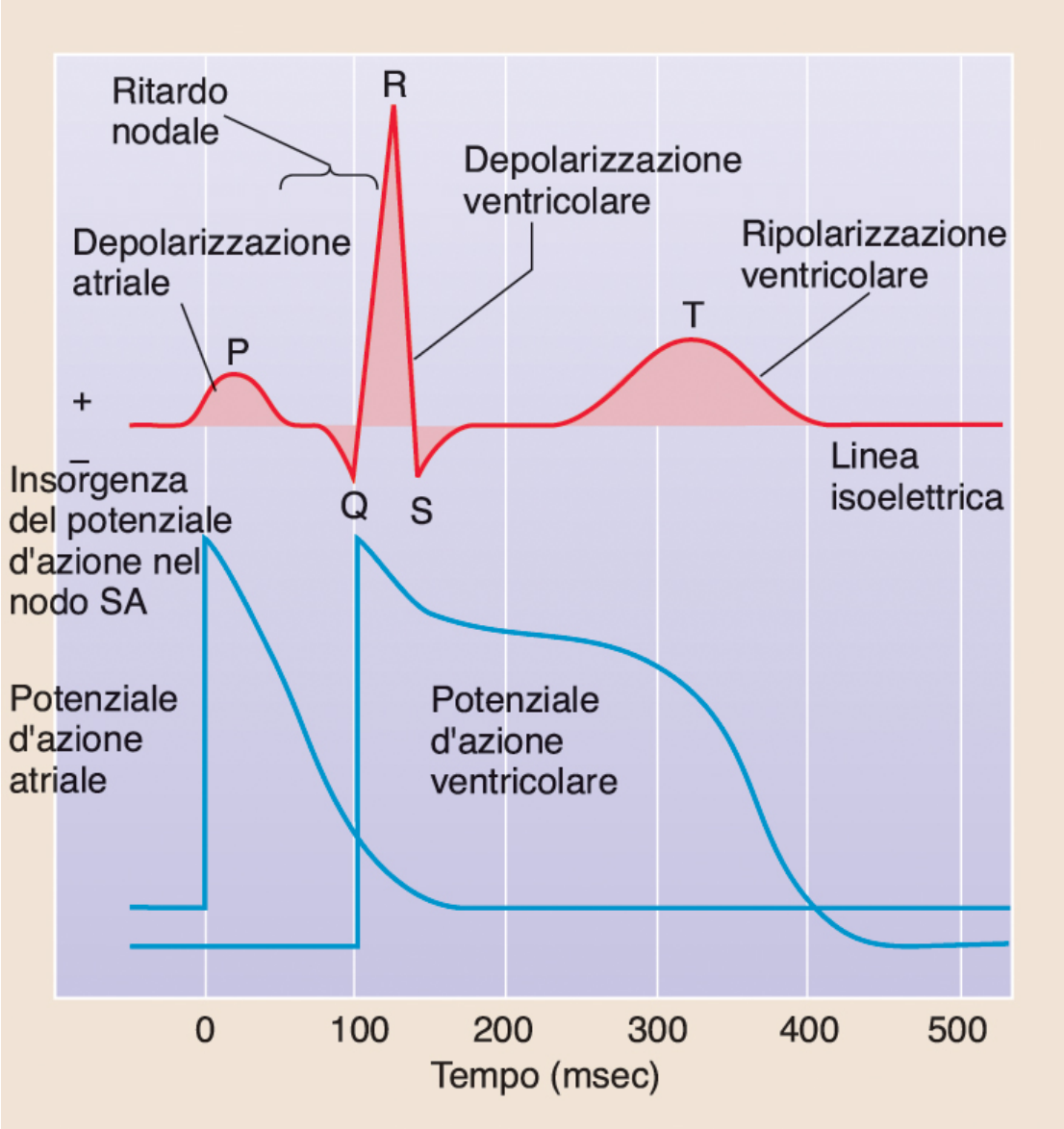
LQT4-Syndrom:	ankyrin B, mutations unknown
LQT5-Syndrom:	KCNQ1, mutations slow K-channel
LQT6-Syndrom:	KCNQ2, mutations fast K-channel


LQTS
1:7000 bis 1:10000 all newborn
Congenital (60 %)
Sporadic (40 %)
Romano Ward (99%)
Autosomal dominant
LQT1 (25%), LQT2 (25%), LQT3 (10%)
Mild-severe
LQT5 (1%), LQT6 (1%)
Mild
Autosomal rezessiv
LQT1-3 (1%)
severe
Jervell-Lange-Nielsen-syndrome (<1%)
Autosomal rezessiv
LQT1 (75%), LQT5 (25%)
Anderson-syndrome
Autosomal dominant with extracardiac involvement
LQT7
New mutations (LQT1-3)

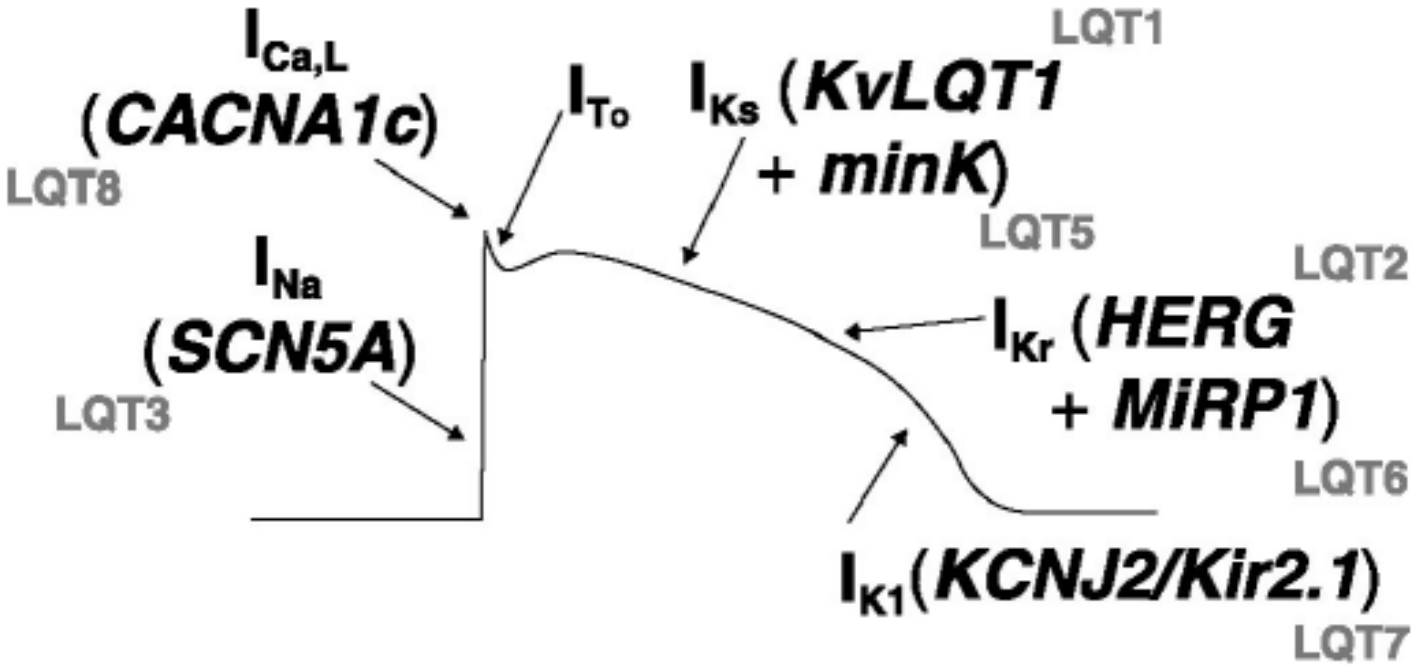
Ventricular action potential



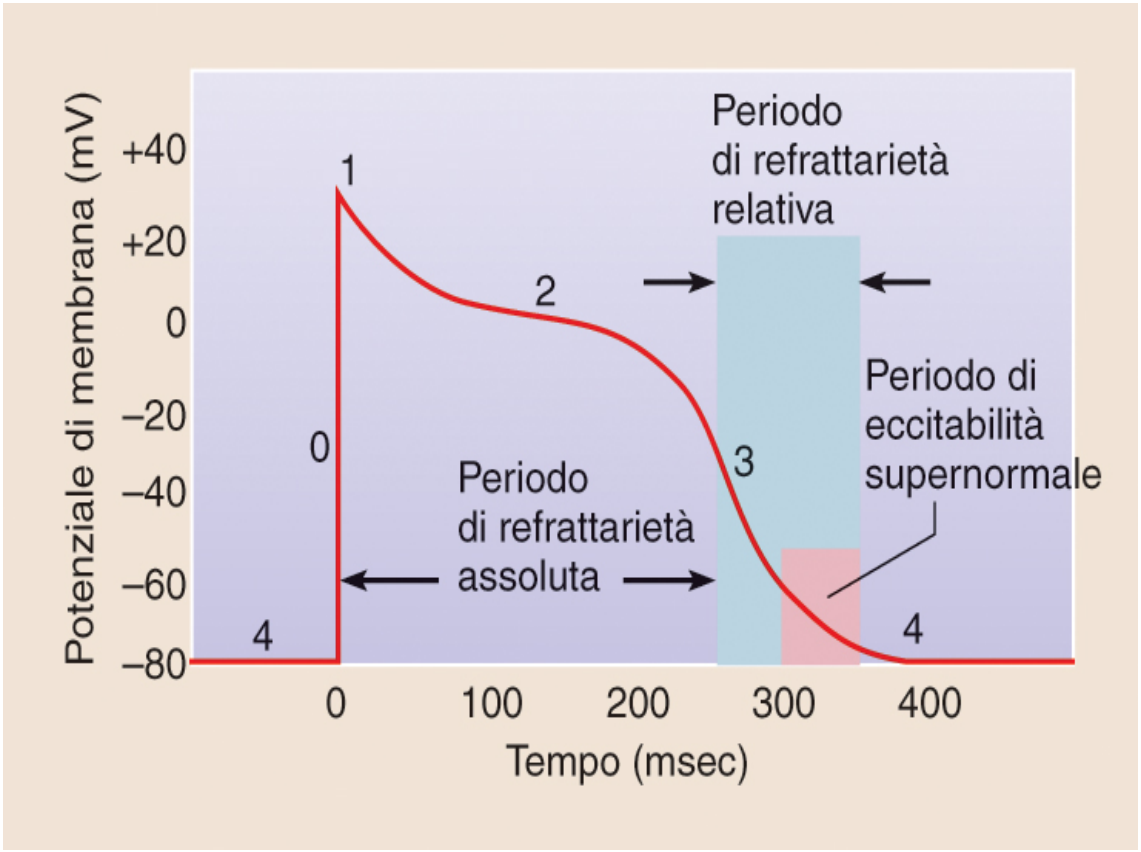
ECG


Ritardo nodale
R
Depolarizzazione ventricolare
Depolarizzazione atriale
Ripolarizzazione ventricolare
T
P
+
–
Linea isoelettrica
Insorgenza del potenziale d'azione nel nodo SA
Q
S
Potenziale d'azione atriale
Potenziale d'azione ventricolare
0
100
200
300
400
500
Tempo (msec)

Ventricular action potentials, genes responsible for LONG-QT and ionic currents

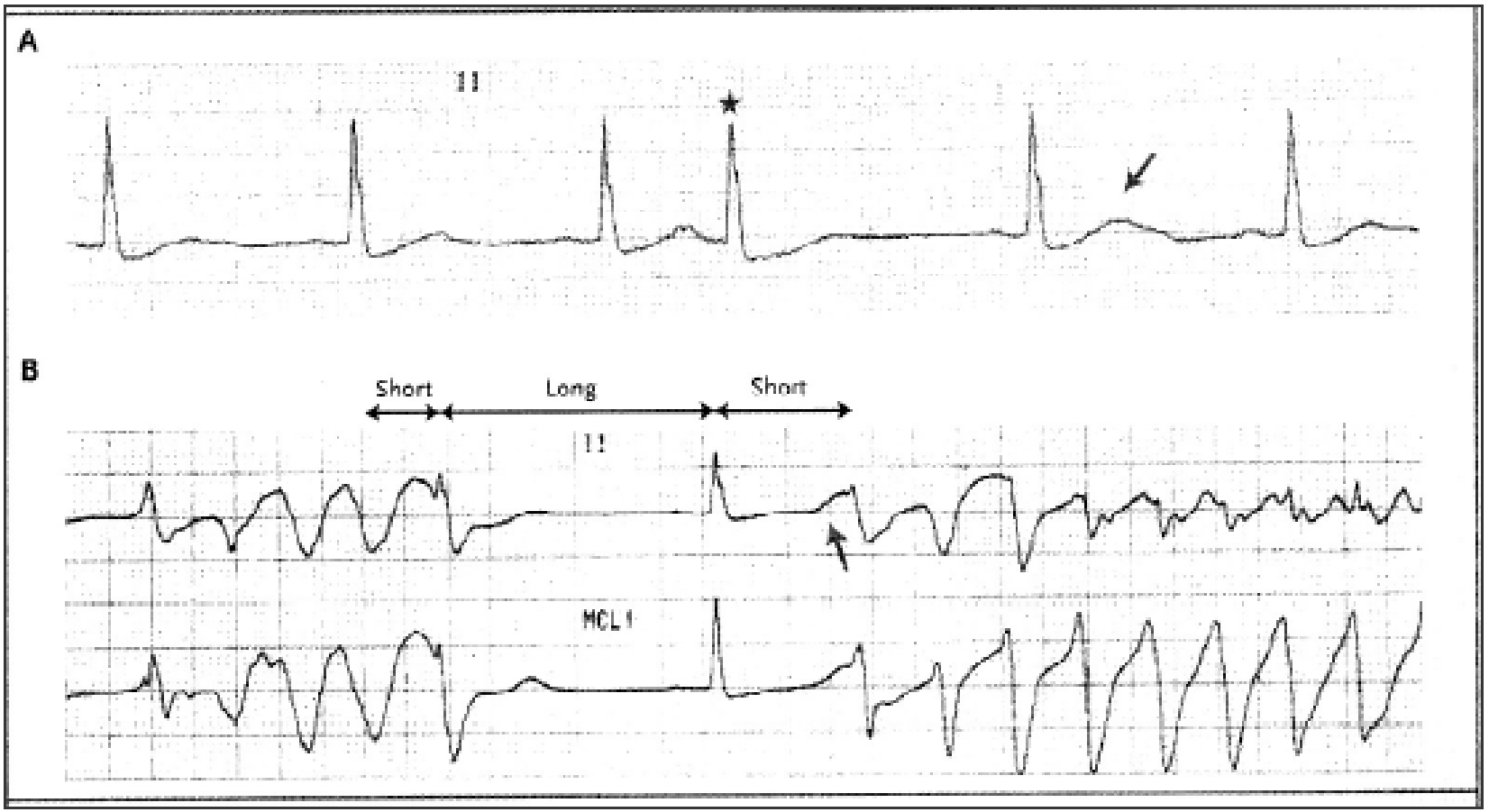


Fase 0: rapido influsso di Sodio (INa).
Fase 1 (picco): ripolarizzazione rapida transiente outward di potassio (Ito).
Fase 2 (plateau) : bilancio tra influsso di Na e Ca (L-type) (ICa,L), ed efflusso di K (IKs, IKr) e Cl.
Fase 3: ripolarizzazione ottenuta mediante l'aumento dela conduttanza rapida del K (IKr) con il contributo delle conduttanze lente del K (IKs) e delle rettificanti (IK1).
Fase 4: La cellula torna al suo potenziale di riposo



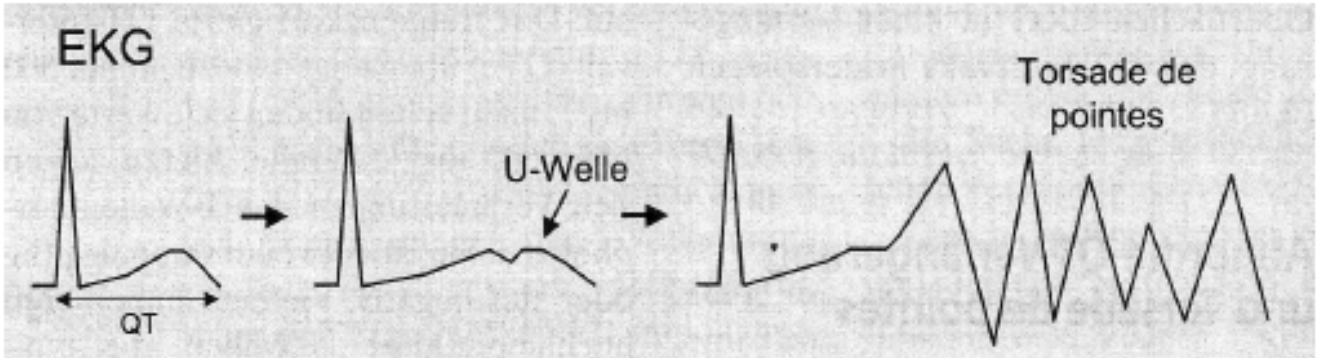
From a functional point of view, the prolongation of the QT interval corresponds,at the cellular level, a prolongation of the duration of the cardiac action potential.
The defects in the potassium channels are associated with a loss of function, which is followed by a "slowing down" of the repolarization phase of the cardiac myocytes and therefore a prolongation of the duration of the action potential.

ECG



ECG - LQT

- *QT-prolongation, QT-dispersion*
- *T wave changes, U - wave*
- *Bradycardia*
- *polymorphic VT/TDP*



- *QT interval > 500 ms is commonly regarded as conferring an increased risk*

Electrical consequences

Slowing of repolarization (phase 3)

The onset of an early action potential is possible (activation of L channels for Ca^{2+}) during the period of relative refractivness.

The longer the phase 3 is extended, the higher the risk of "Early post-depolarization”

It can start in a series of electrical complexes of variable conformation (torsades de pointes, TdP), which can spontaneously stop or turn into ventricular fibrillation.

In the first case there will be a syncope, in the second the risk of sudden death is very high.

Not all gene modifications that can cause sudden death induce TDP.

This form of tachyarrhythmia is frequent when gene modification involves a current at K + in phase 3 (LQT1, LQT2, LQT5, LQT6).LQT3, related to modifications of the gene that codes for the voltage-dependent Na + channel (SCN5A), manifests itself mainly as ventricular fibrillation

Are mutations all equivalent?

The site of the mutation determines the severity of the LQTS phenotype.

Patients with LQT2 mutation at the pore level appear to be at greater risk of cardiac events than mutations at sites outside the pore.

SINDROME	CROMOSOMA	NOME	PROTEINA	CORRENTE
LQT1	11	*KCNQ1* o *KVTLQT1*	subunità- α canale al K+	corrente *IKs*.
LQT2	7	*HERG (human ether-a-go-go related gene)*	subunità-α del canale al K+	corrente *IKr*
LQT3	3	*SCN5A*	subunità- α del canale al Na+	corrente *INa*.
LQT4	4	*ANK2*	proteina citoscheletrica ankirina B	
LQT5	21	*KCNE1* o *MinK*	subunità-β del canale al K+	corrente *IKs*
LQT6	21	*KCNE2* o *MiRP1*	subunità-β del canale al K+	*IKr*
LQT7	17	*KCNJ2*	subunità-β del canale al K+	subunità-β del canale al K+

LQT1 and LQT2 have a higher frequency of cardiac events, but LQT3 has the highest incidence of lethal cardiac episodes.

Potassium channel mutations are not just responsible for LQTS

Organ				
Cell type	Cardiac myocyte	Stria vascularis	Outer hair cells	Forebrain neurons
Channel subunit composition	KCNQ1 + KCNE1	KCNQ1 + KCNE1	KCNQ4 (+ KCNQ3)	KCNQ2 + KCNQ3
Current	Isk	Isk	I_M-like	I_M-like
Phenotype	Long QT syndrome	Congenital deafness	Childhood onset deafness	Epilepsy

LQT 3Mutation in SCN5A (Na + Channel, INa)

LQT III

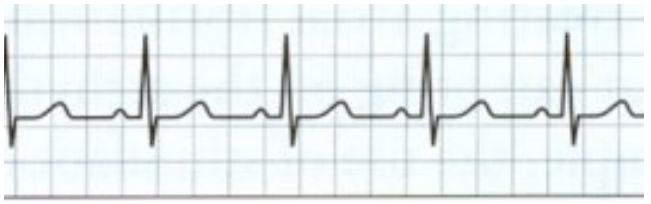
- Sodium channel mutated
- Incomplete inactivation of the channel

Symptoms

- Syncope
- Seizures
- Cardiac arrest
- Unexpected death

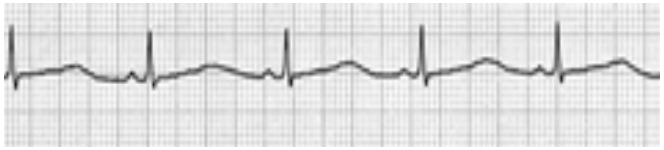
Diagnosis

- Analysis of the T wave of the ECG.



Normal ECG

- Genetic analysis on the SCN5A gene



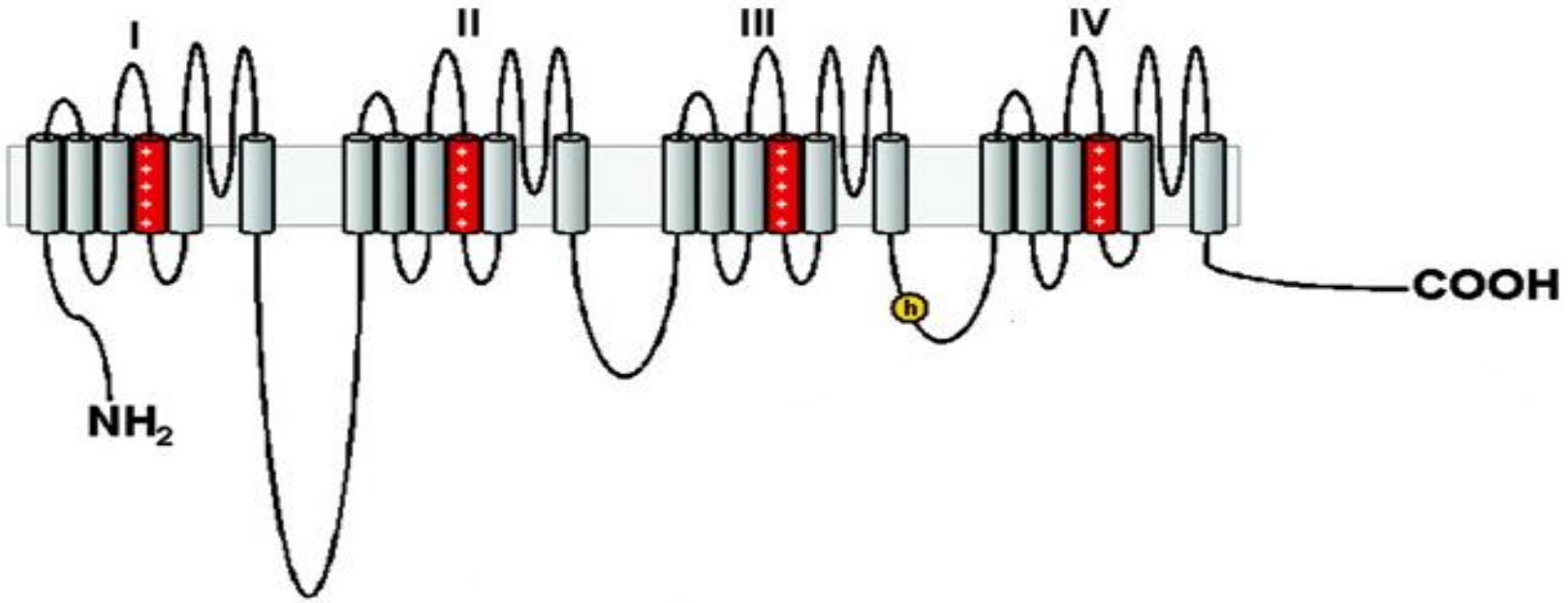
Long QT syndrome

Statistics

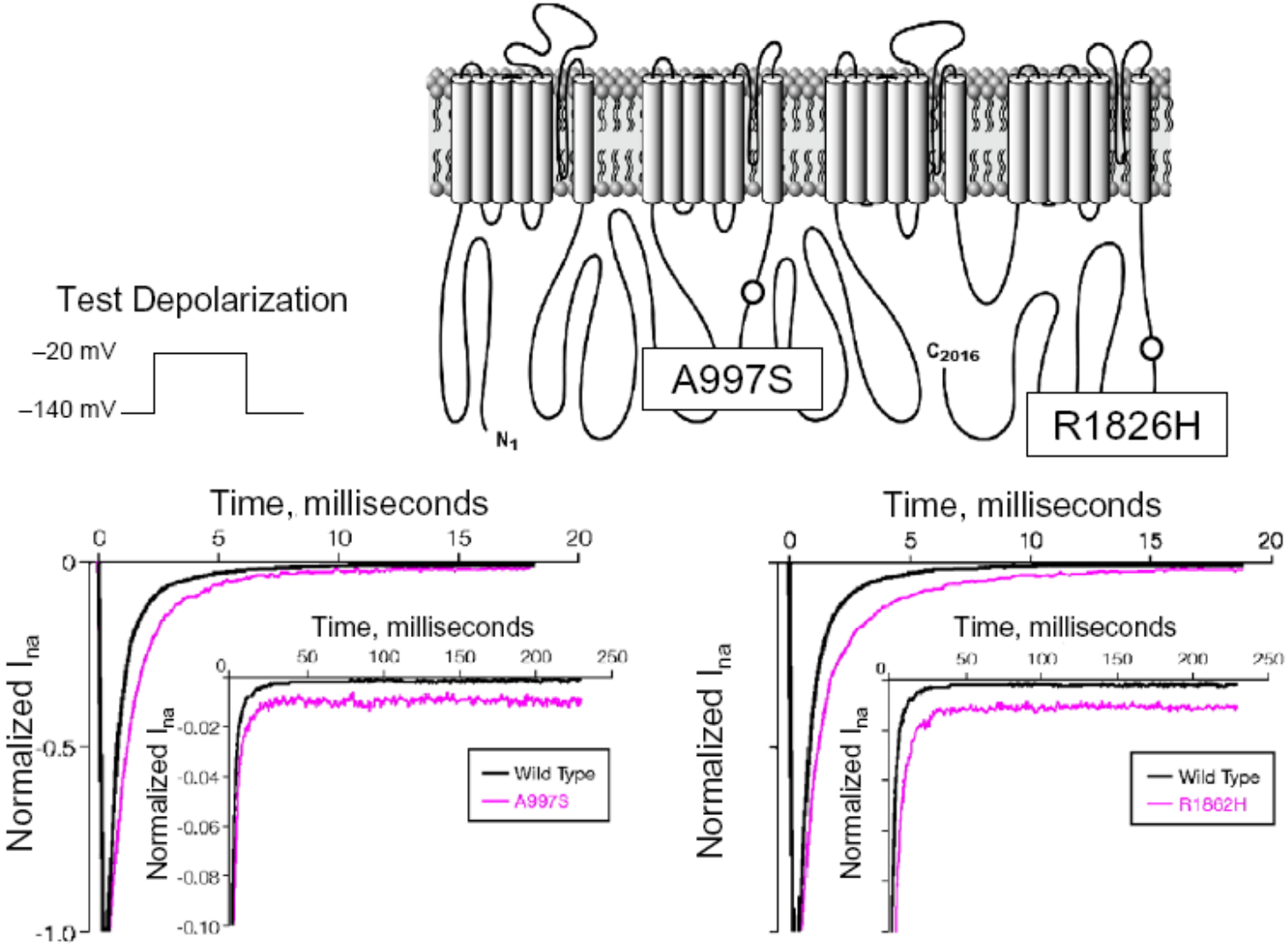
- 8% of patients with LQT are affected by the SCN5A mutation
- LQT-3 is one of the most deadly mutations
- Onset: 50% before 12 years; 90% before the age of 40

- Fatal arrhythmias - 39% at rest, 32% during physical exertion or emotional stress

Protein

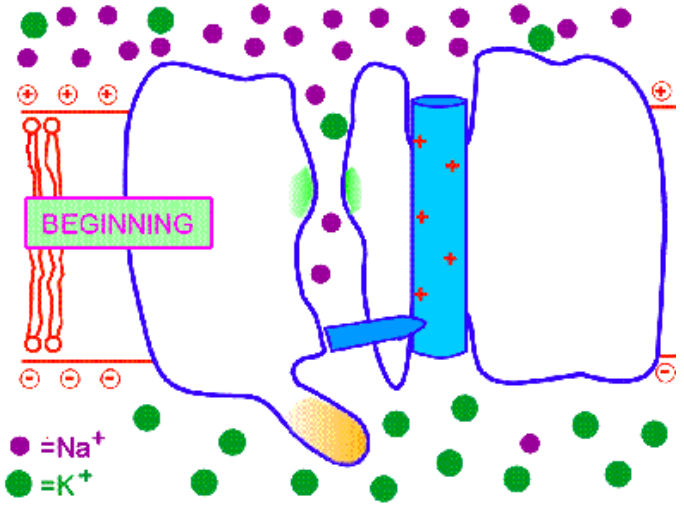


Mutation SCN5A



Function

- Selective channels to Na + voltage dependent
- Voltage sensors



- Responsible for the rapid ascent of the action potential

Introduction to LQT

Currently Recognized LQTS Disease Genes

Disease	Gene	Chromosome	Ion Channel
LQT1	KVLQT1	11p 15.5	I_{Ks} subunit
LQT2	HERG	7q35-6	I_{Kr}
LQT3	SCN5A	3q21-24	Na^+
LQT4	Unknown	4q25-27	Unknown
LQT5	Min K #	21	I_{Ks} Subunit

84

- Disorder caused by mutations in cardiac ion channels
- Most associated with K+ channels

Post-synaptic myasthenia syndromes

Characteristics of the disease:

myasthenic symptoms:

hyposthenia and muscular exhaustion of the skeletal and bulbous innervation muscles,
without involvement of the immune system (absence of autoantibodies)

prevalence less than 1: 500 000

early onset, usually immediately after birth.

There are several forms whose clinic depends on which molecule of the neuromuscular junction (GNM) is genetically altered: they can be pre-synaptic, synaptic or post-synaptic.

slow-channel syndrome

fast-channel syndrome

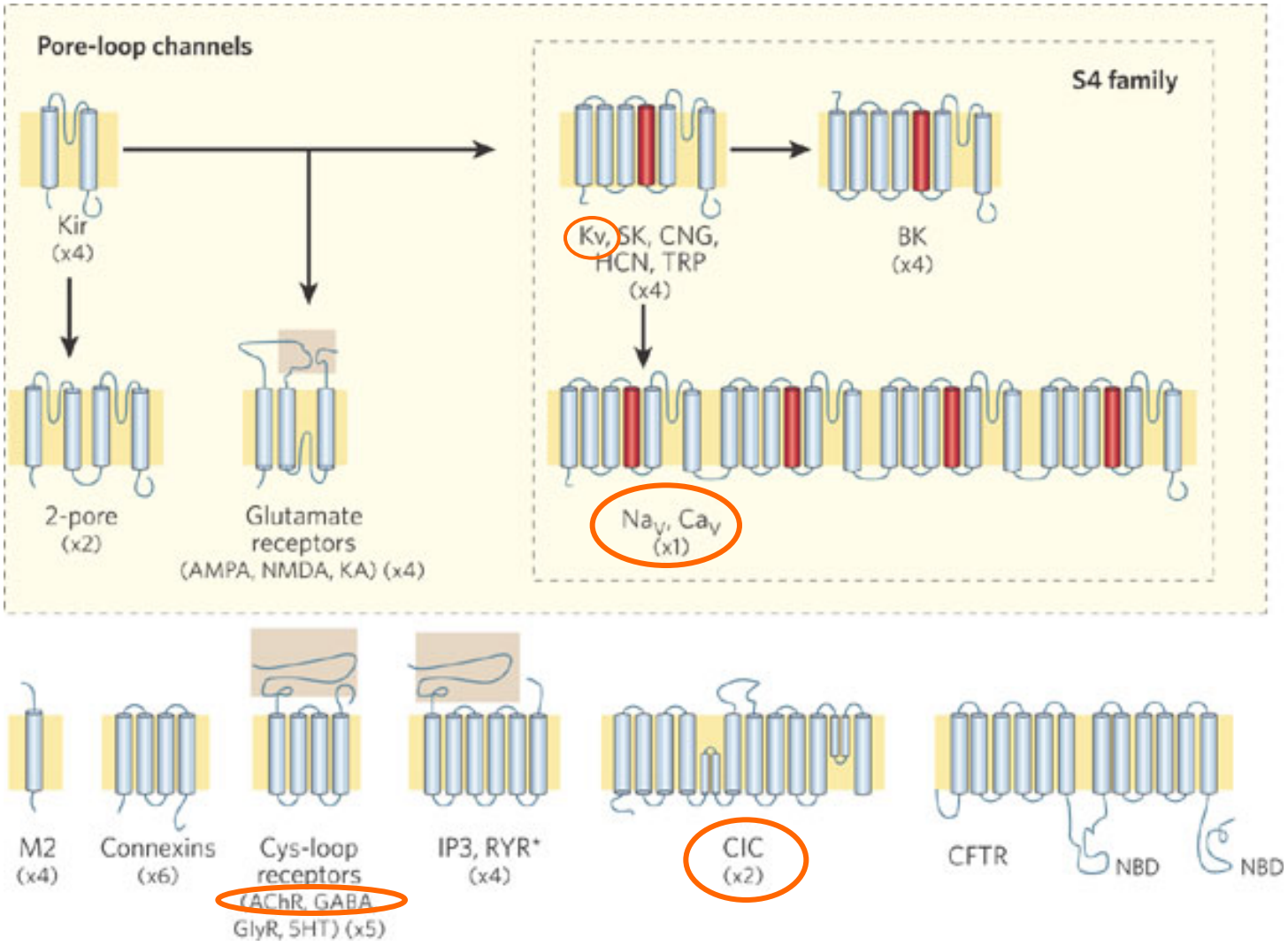
Acetylcholine receptor deficiency (AChR).

May arise during childhood (severe form) or later, up to the 7th decade (lighter form)

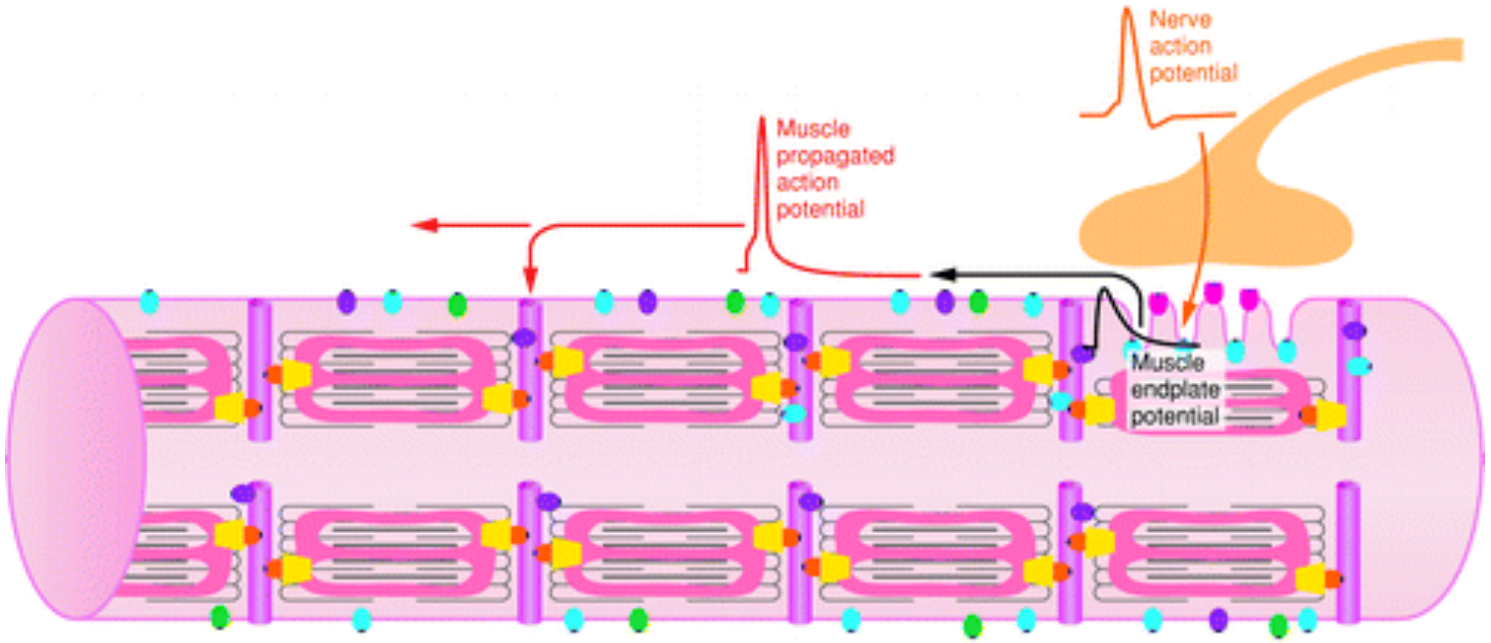
The extensor muscles of the forearm, the musculature of the neck and that of the scapula are selectively affected.

Skeletal Muscle Channelopaties

Ionic Channels



Neuromuscular junction and the propagation of excitation in skeletal muscle



Sarcoplasmic reticulum

Transverse tubule

Actin / myosin filaments

nAChR (congenital myasthenic syndrome)

NaV1.4 (periodic paralysis, myotonia)

ClC-1 (myotonia congenita)

Kir2.1 (Andersen-Tawil syndrome)

CaV1.1 - RyR1 (hypokalemic periodic paralysis)

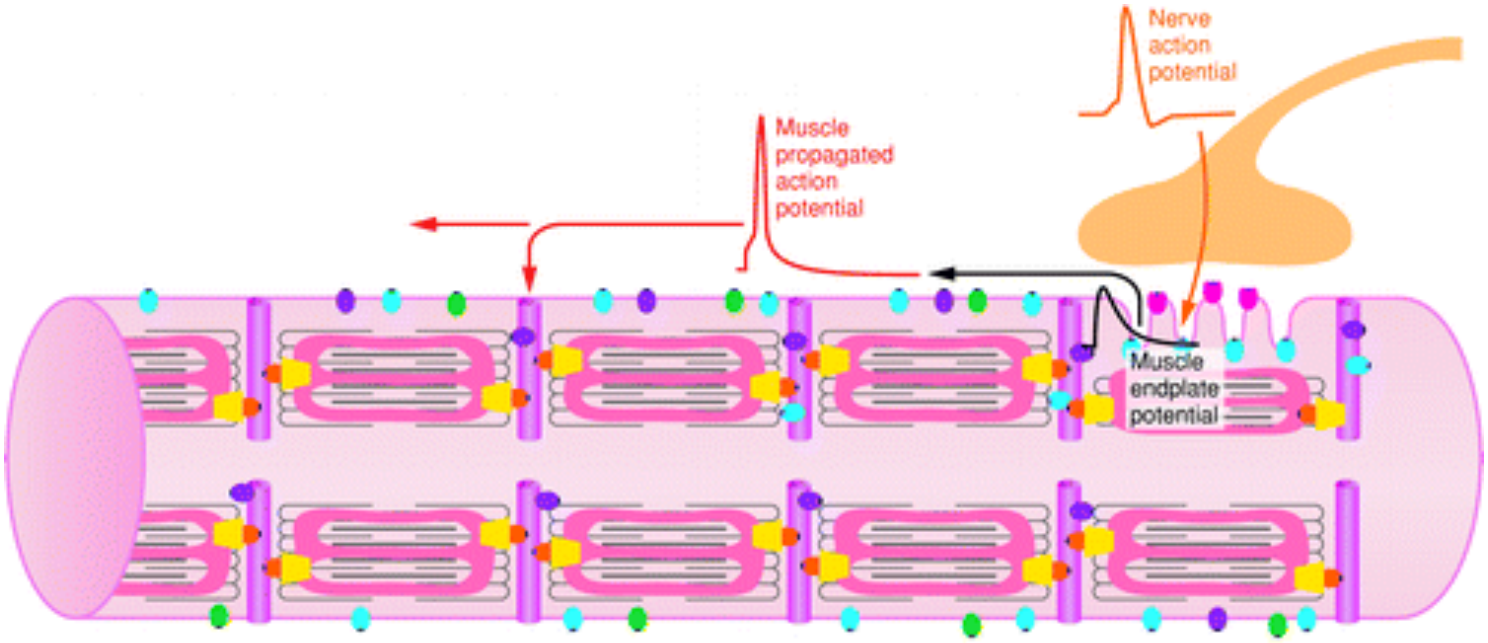
Cannon SC. 2006.
Annu. Rev. Neurosci. 29:387–415

Common features:

- **Symptoms often manifest as transient attacks separated by long periods of normal function**
- **Mostly autosomal dominant mutations**
- **Clinical phenotype generally limited to the involvement of a single organ**

Skeletal Muscle Channelopaties

1) Defects in neuromuscular transmission
2) Defects in the Sarcolemma excitability



Defects in neuromuscular transmission

Congenital Myasthenic Syndromes (CMS):

- **hereditary diseases in which neuromuscular transmission is damaged by one or more specific mechanisms: the Plaque Potential is insufficient to activate the NaV channels responsible for the propagation of the action potential.**
- **They occur at birth or early adolescence**
- **They involve the muscles of the eyes, the skull and the limbs**

Neuromuscolar Junction

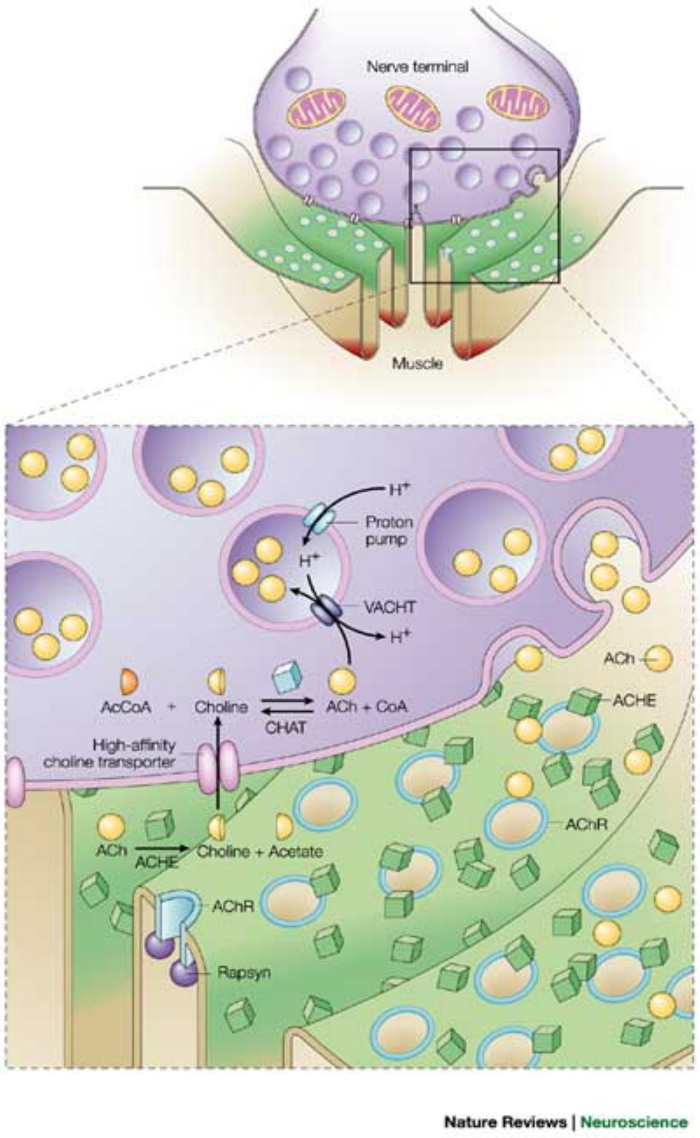
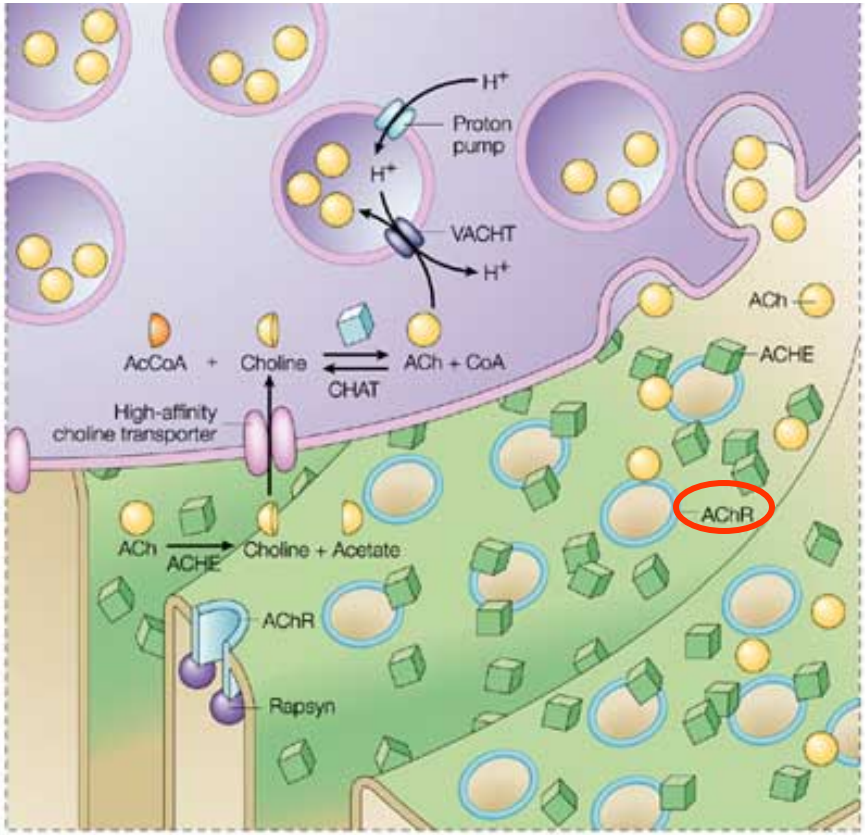


Table 1 | **Classification of CMSs**

Site of defect	Index cases
Presynaptic defects (7%)	
CHAT deficiency‡	6
Paucity of synaptic vesicles and reduced quantal release	1
Lambert-Eaton syndrome like	1
Other presynaptic defects	4
Synaptic basal lamina-associated defects (14%)	
Endplate ACHE deficiency‡	26
Postsynaptic defects (79%)	
Kinetic abnormality of AChR with/without AChR deficiency‡	45
AChR deficiency with/without minor kinetic abnormality‡	83
RAPSYN deficiency‡	17
Plectin deficiency	1
Total (100%)	***185***

*Classification based on cohort of congenital myasthenic syndrome patients investigated at the Mayo Clinic between 1988 and 2003. ‡Gene defects identified.
ACHE, acetylcholinesterase; AChR, acetylcholine receptor; CHAT, choline acetyltransferase; CMSs, congenital myasthenic syndromes.

CMS caused by defects in the Acetylcholine Receptor (AChR):



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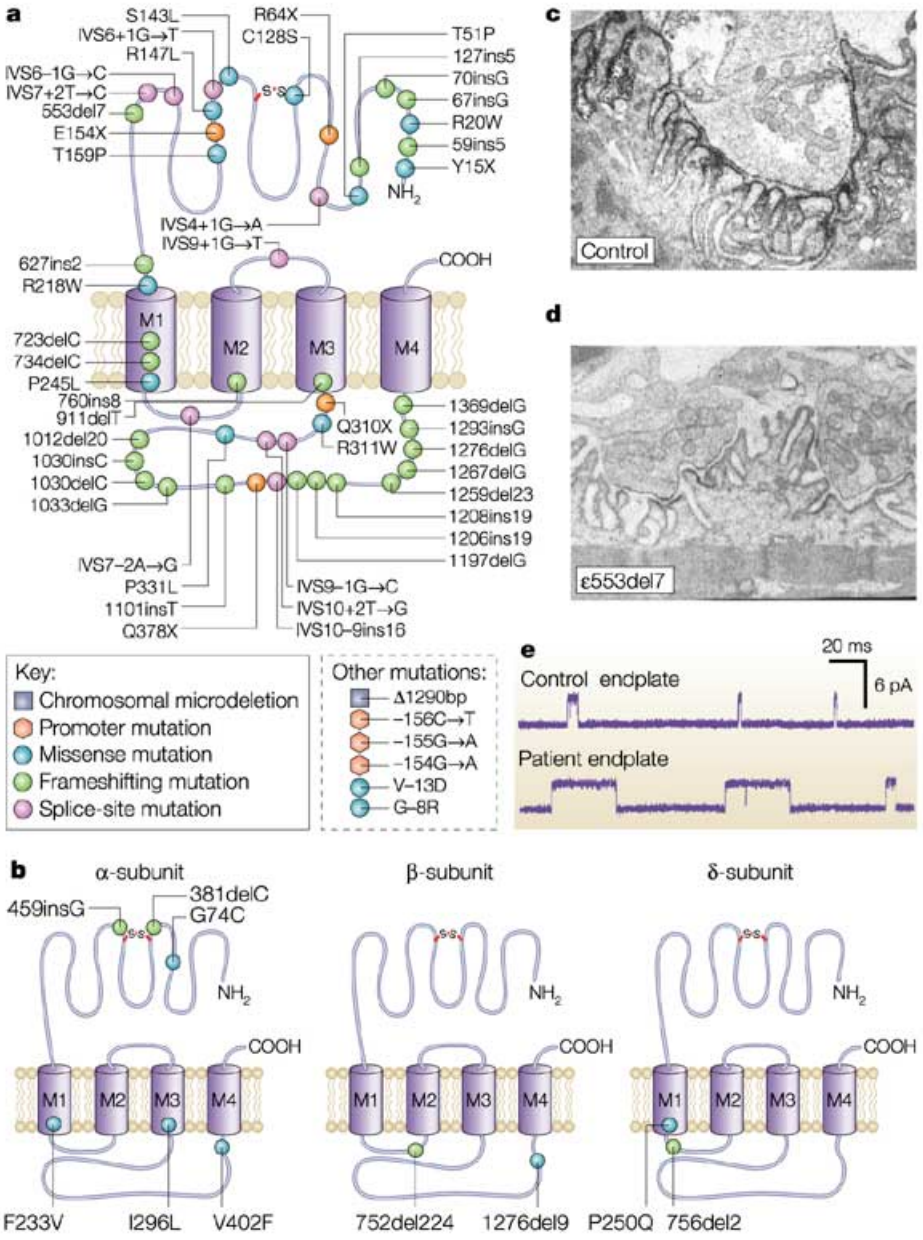
Mutations in different domains

of the different subunits:

Reduced receptor expression

Altered kinetic properties

("Slow & Fast Channel CMS")



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nAChR is a ligand-activated post-synaptic membrane receptorheteropentamer: 2 sub α, β, δ, ε (γ)

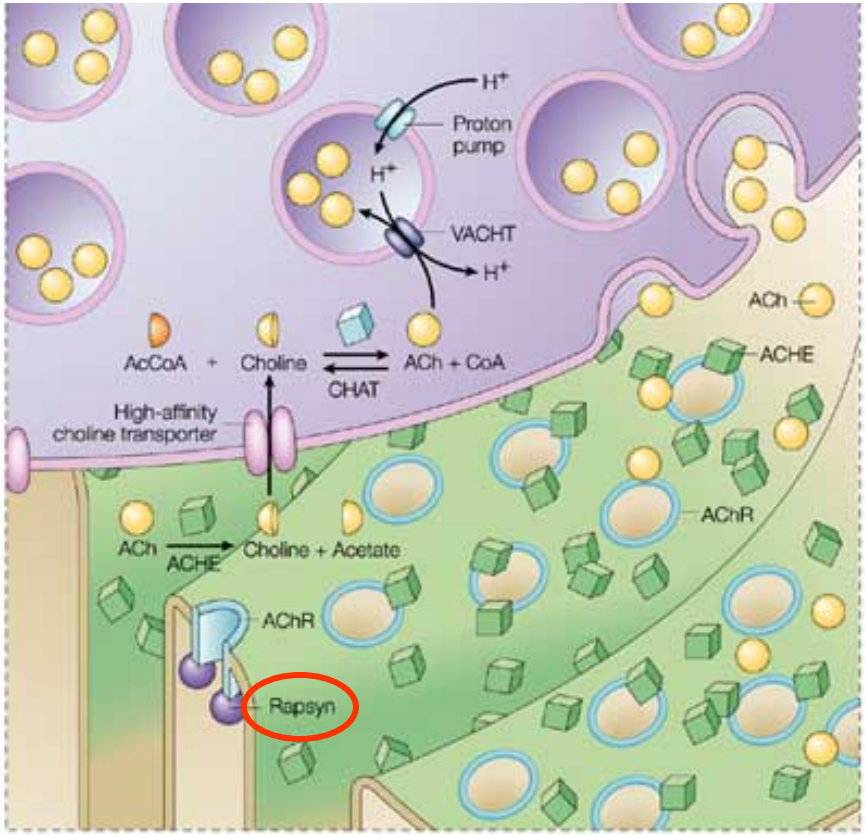
The part that protrudes in the junction has 2 binding domains for the Ach at the interface between the subunits

The transmembrane part forms the channel

The cytoplasmic extension contains structures that modulate the opening of the canal and anchor the cytoplasmic protein receptor

Mutations affect the maturation and expression in the receptor membrane

CMS caused by defects in the Rapsin protein:



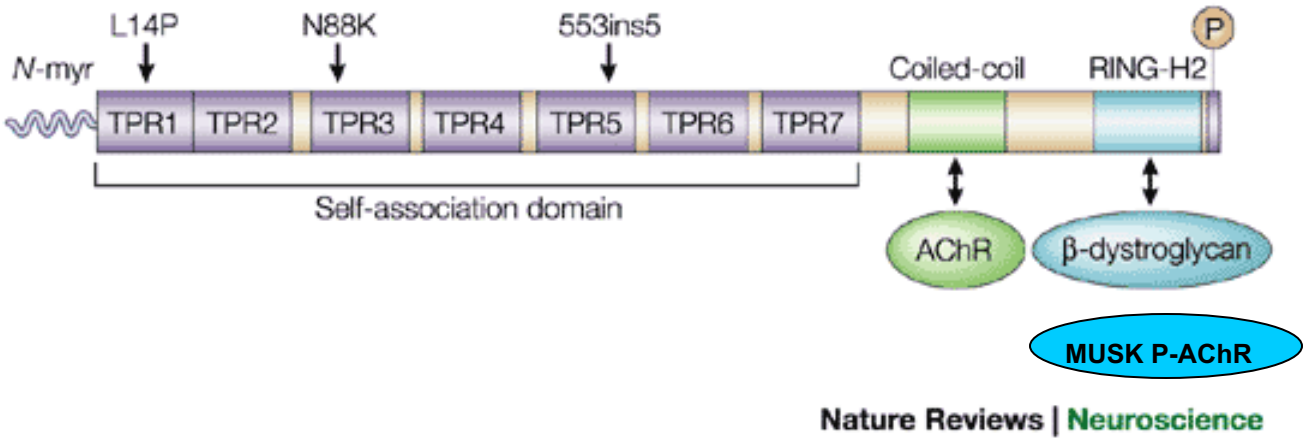
Nature Reviews | Neuroscience

Rapsin, together with other proteins such as Agrin and MUSK, regulates the membrane expression of AChR

It associates with itself and binds the cytoplasmic domains of AChR subunits

By binding to β-dystroglycan, it binds the receptor to the post-synapse cytoskeleton

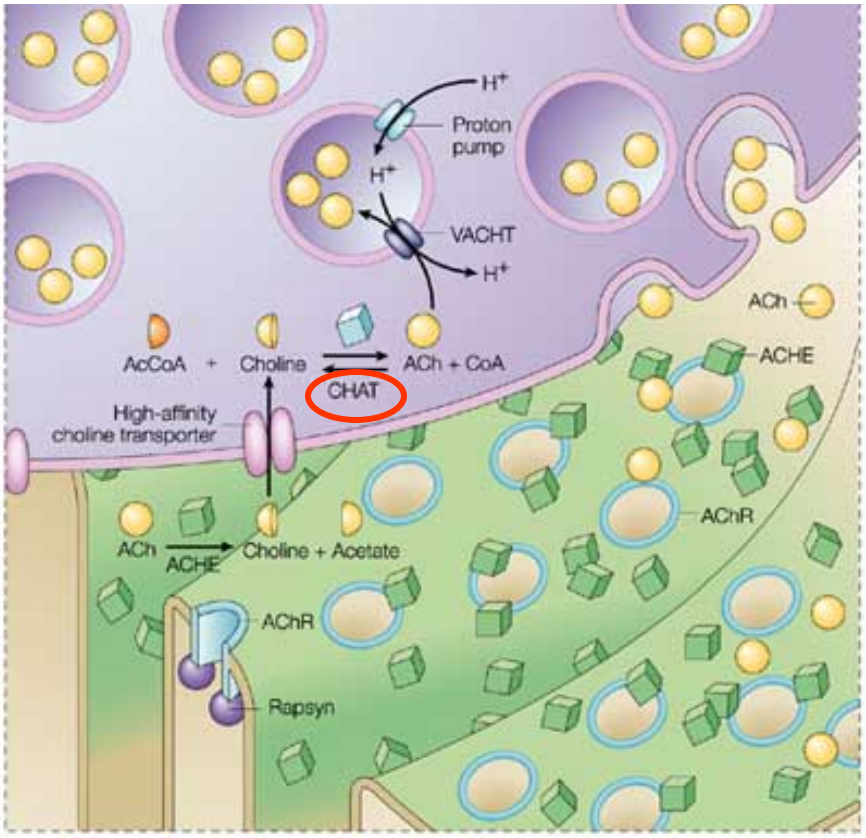
Mutations in Rapsin protein domains



Mutations compromise co-carriage of the AChR receptor with Rapsin

Post-synaptic regions are poorly developed

CMS caused by defects in Choline-acetyltransferase (CHAT):



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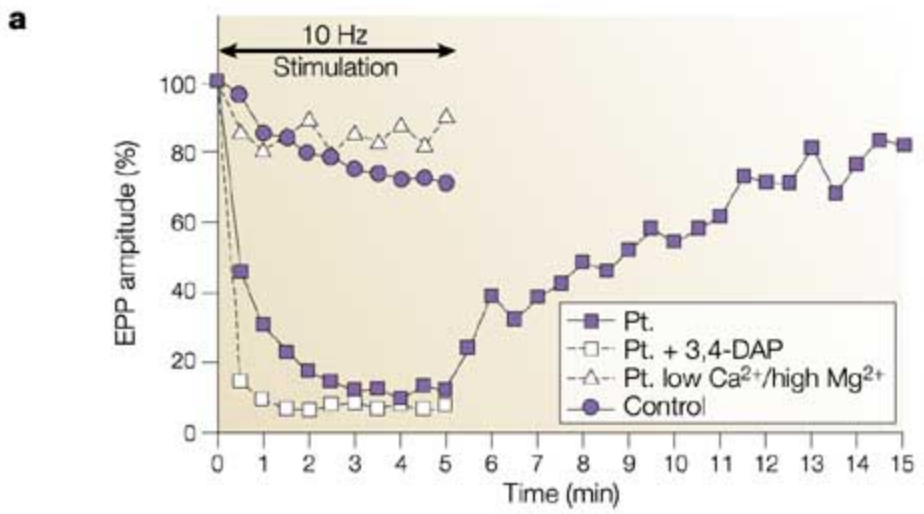
It manifests at birth or during childhood or early adolescence

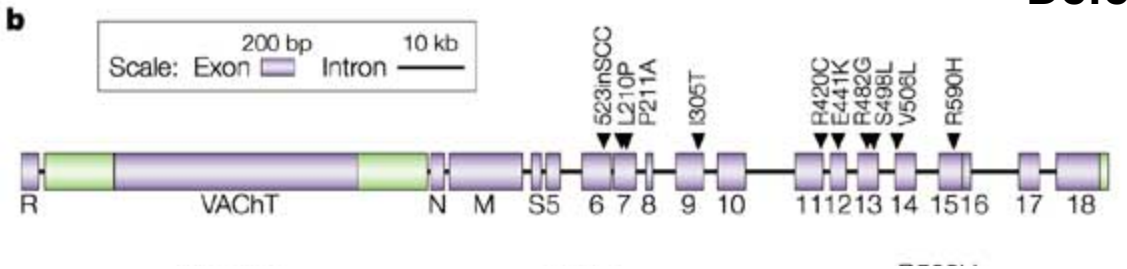
Respiratory problems and / or episodic apnea attacks

The concentration of AChR and the structure of the post-synapse is normal, but the synaptic vesicles are smaller than normal both in the resting muscle and after stimulation



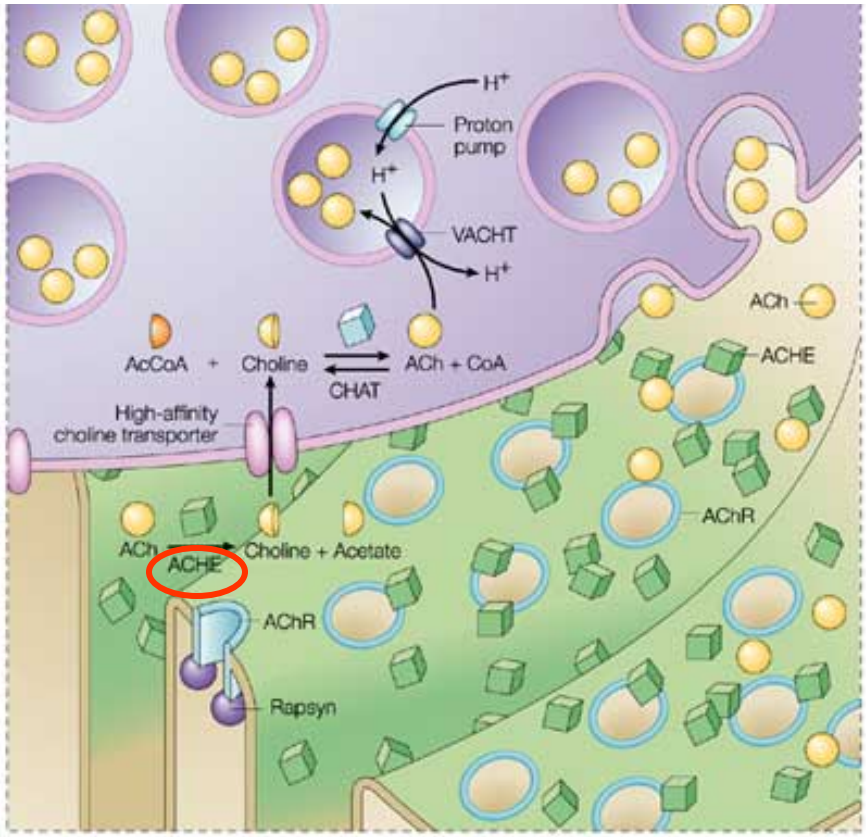
The synaptic response to the Ach, measured as the amplitude of the End Plate Potential is normal in the muscle at rest but decreases following a stimulation of 10 Hz for 5 min and recovers slowly

Defects in the synthesis of the Ach



Genetic analysis of patients → 10 mutations in the CHAT gene

CMS caused by defects in Acetylcholinesterase (ACHE):



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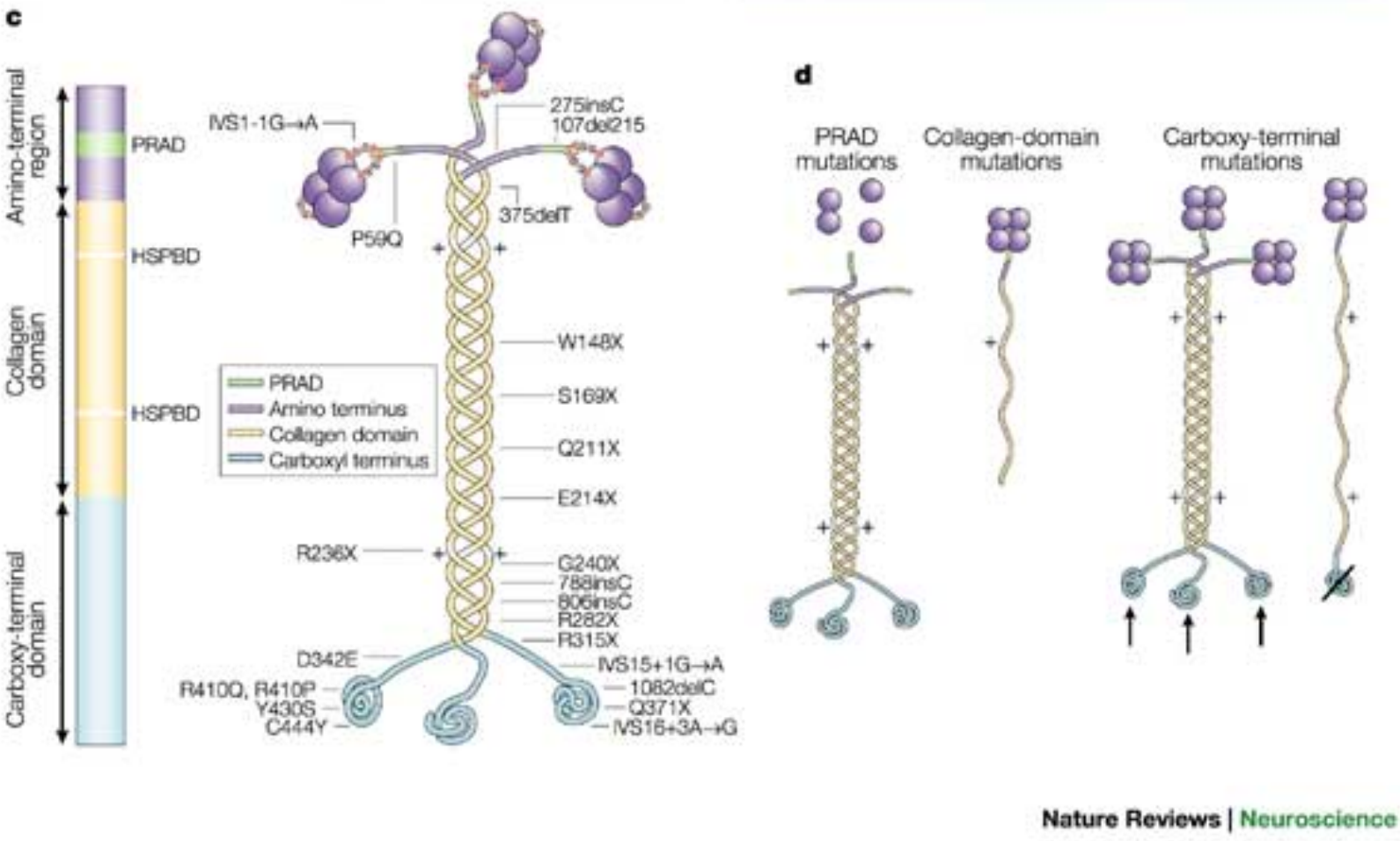
It manifests itself during early adolescence

ACHE is absent or non-functional in neuromuscular plaque

The presynaptic terminals are smaller than normal, the release of Ach is low and the synaptic response is prolonged beyond the refractory period.

Mutations in the ACHE protein domains



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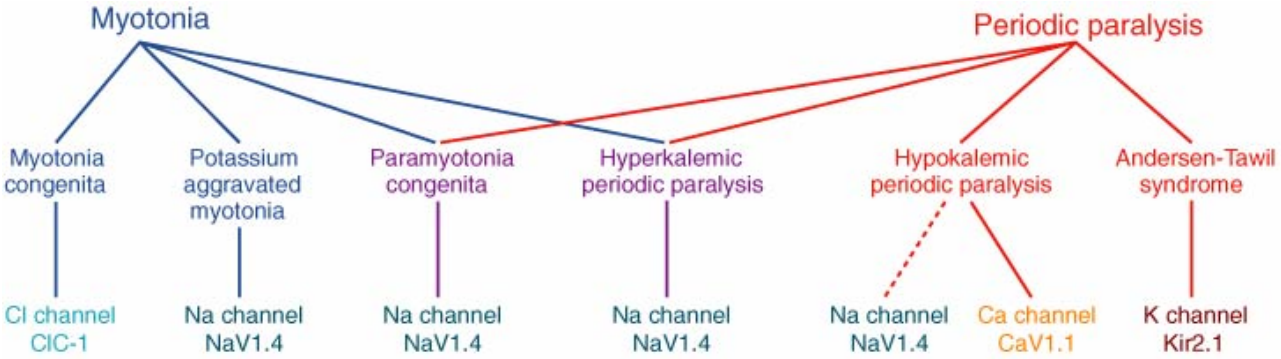
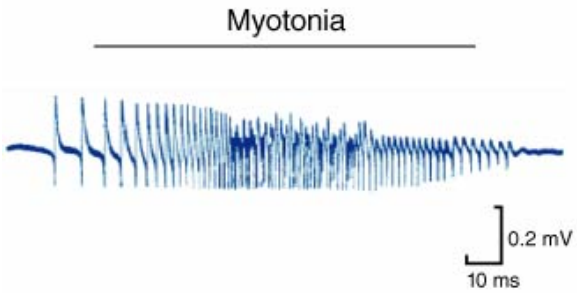
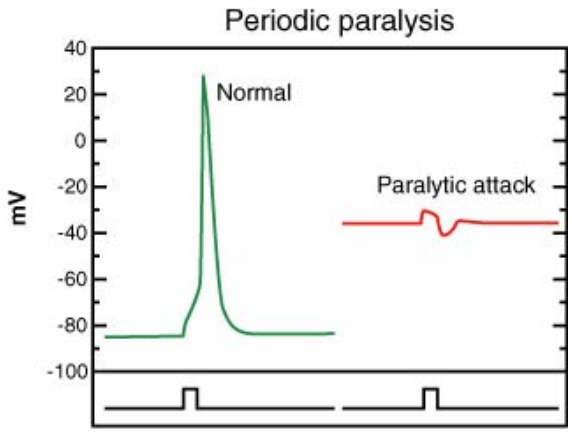
Defects in the excitability of Sarcolemma

Myotonia and Periodic Paralysis

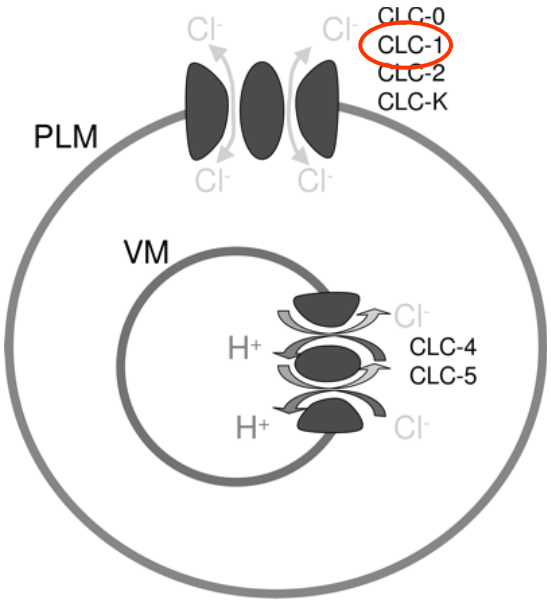


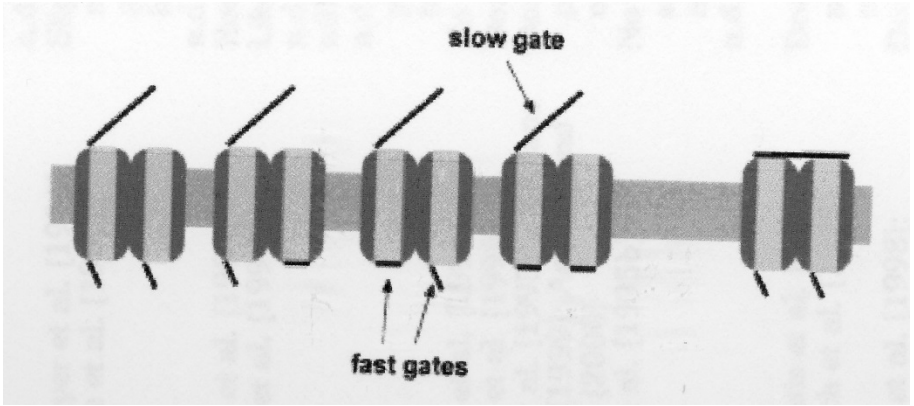




Cannon SC. 2006.
Annu. Rev. Neurosci. 29:387–415

Malfunction of the Cl- Channel





Congenital myotonia: Hyperexcitability of the plasma membrane of skeletal muscle, due to mutations in the CLCN1 gene that encodes the CLC-1 Chlorine channel, which reduce the conductance

Dystrophic myotonia: expansion of repeated nucleotides in a UTR of the CLCN1 gene which causes the transcript not to be matured

Dysfunction of the Na + Channel

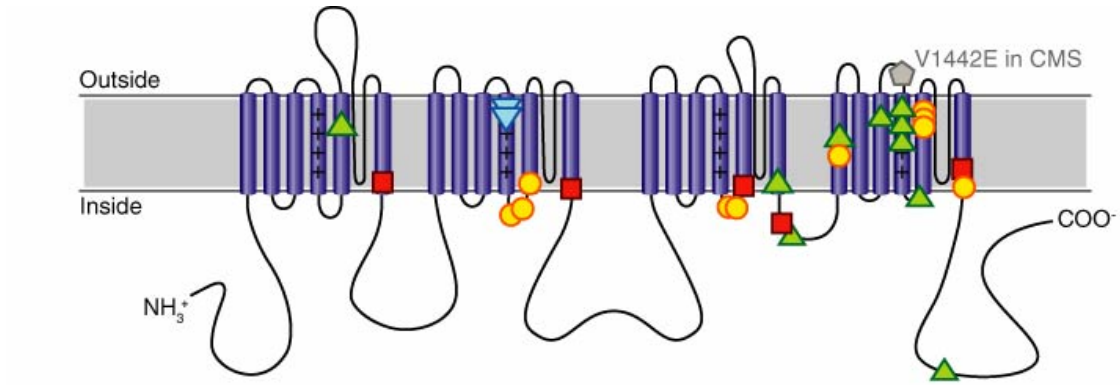
Mutations that increase NaV1.4 channel activation or destroy inactivation:

Myotonia aggravated by K

Paratyotonia

Mutations that increase channel inactivation:

Periodic paralysis



PAM			PMC			HyperPP			HypoPP		
Val	445	Met	Leu	266	Val	Leu	689	Ile	Arg	669	His
Ser	804	Phe	Val	1293	Ile	Ile	693	Thr	Arg	672	His / Gly / Ser
Ile	1160	Val	Thr	1313	Met / Ala	Thr	704	Met			
Gly	1306	Ala / Val / Glu	Met	1370	Val	Ala	1156	Thr			
Val	1589	Met	Leu	1433	Arg	Pro	1158	Ser			
			Arg	1448	Cys / His / Pro / Ser	Met	1360	Val			
			Gly	1456	Glu	Ile	1490	Leu / Met 1493 Ile			
			Val	1458	Phe	Ile	1495	Phe			
			Phe	1473	Ser	Met	1592	Val			
			Phe	1705	Ile						

Cannon SC. 2006.
Annu. Rev. Neurosci. 29:387–415

Ca2+ Channel dysfunction

Mutations in the α1 subunit of the Ca2 + channel in the sensitive region

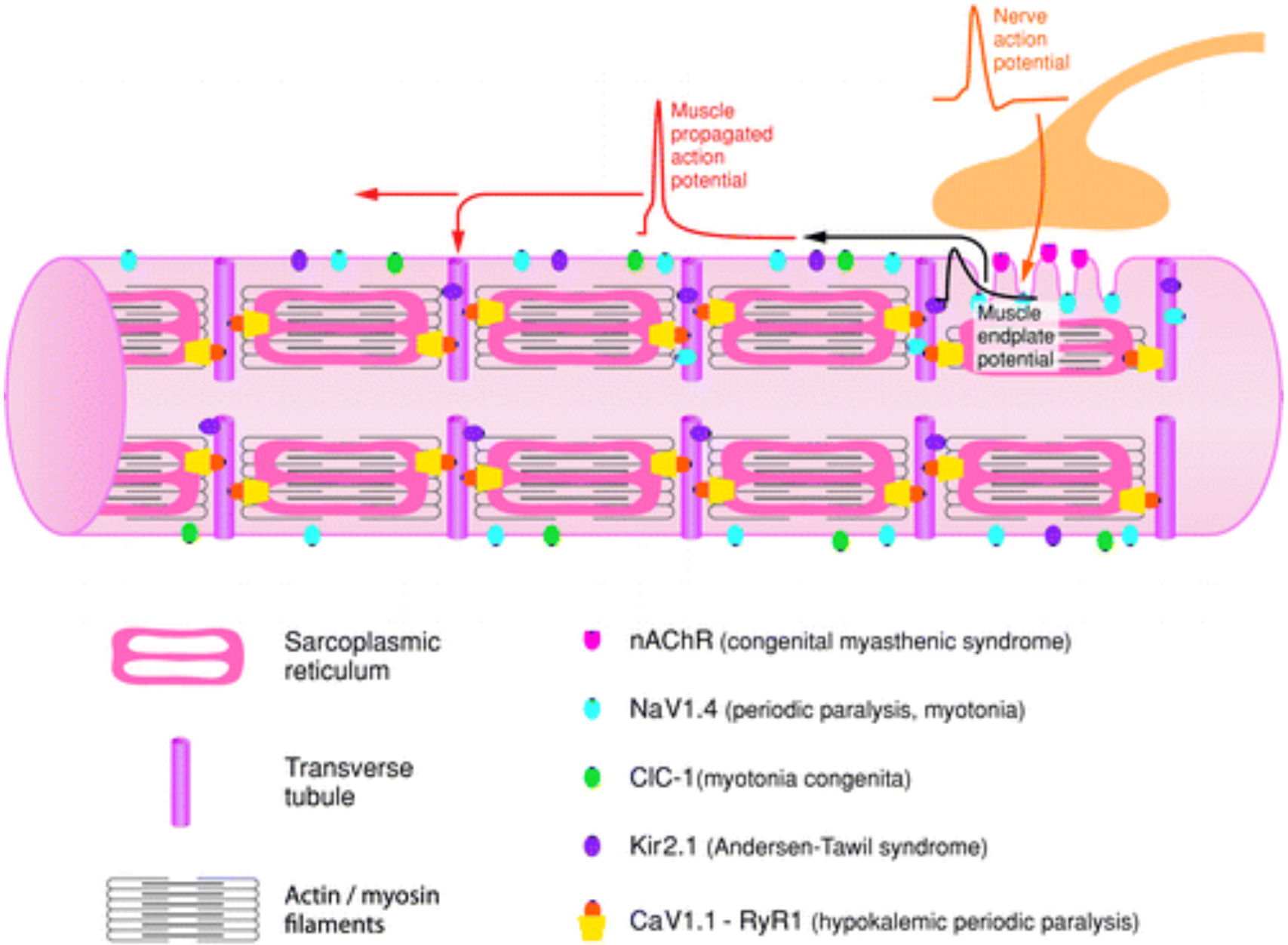
reduced ion current density and slowed activation kinetics:

Periodic paralysis

K+ Channel dysfunction

Mutations in the gene coding for the Kir2.1 subunit of the K + channel, expressed in skeletal muscle, in the heart and in the brain, for which the homotetrameric channel is not formed

Andersen-Tawil syndrome: periodic paralysis, ventricular arrhythmias, skeletal abnormalities



Cannon SC. 2006.
Annu. Rev. Neurosci. 29:387–415

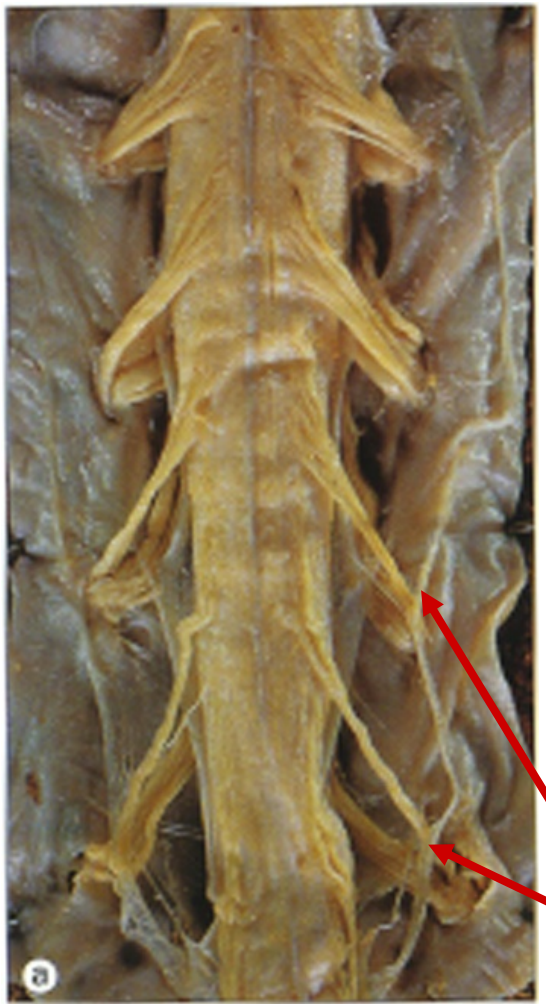
ALS and nicotinic receptor

Amyotrophic Lateral Sclerosis (ALS) or Motor Neuron Disease

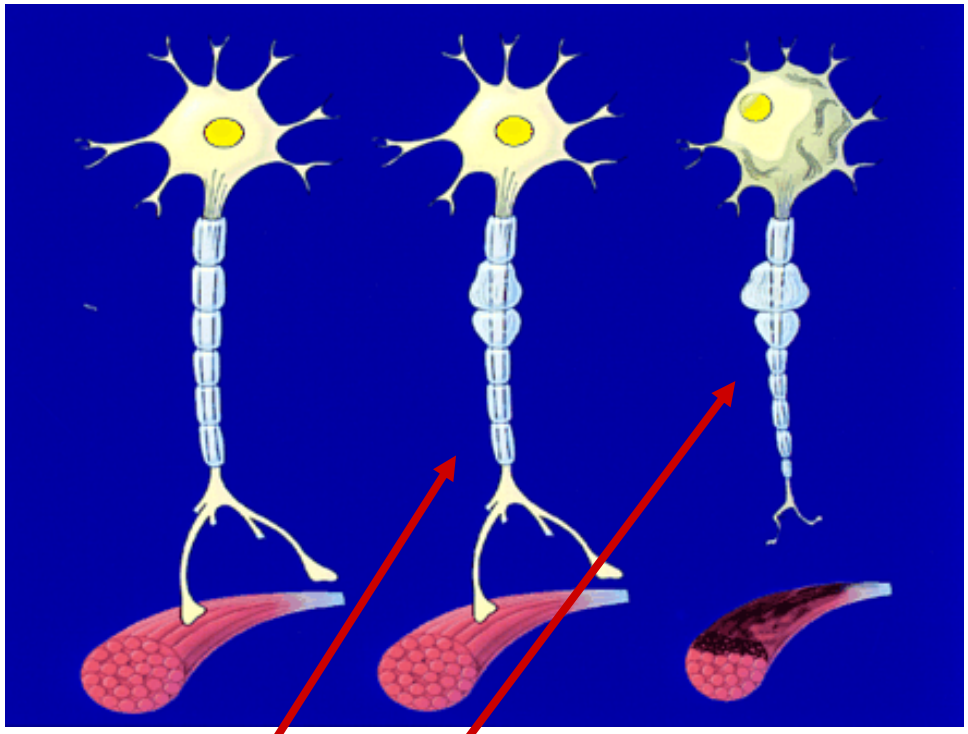
"Lou Gehrig's Disease" named for famous baseball player

- Progressive neuronal loss
- Upper and lower motoneuron
- Familiar (5%) or sporadic (95%)
- Different phenotypes

Death of motor neurons causes paralysis of muscles in ALS



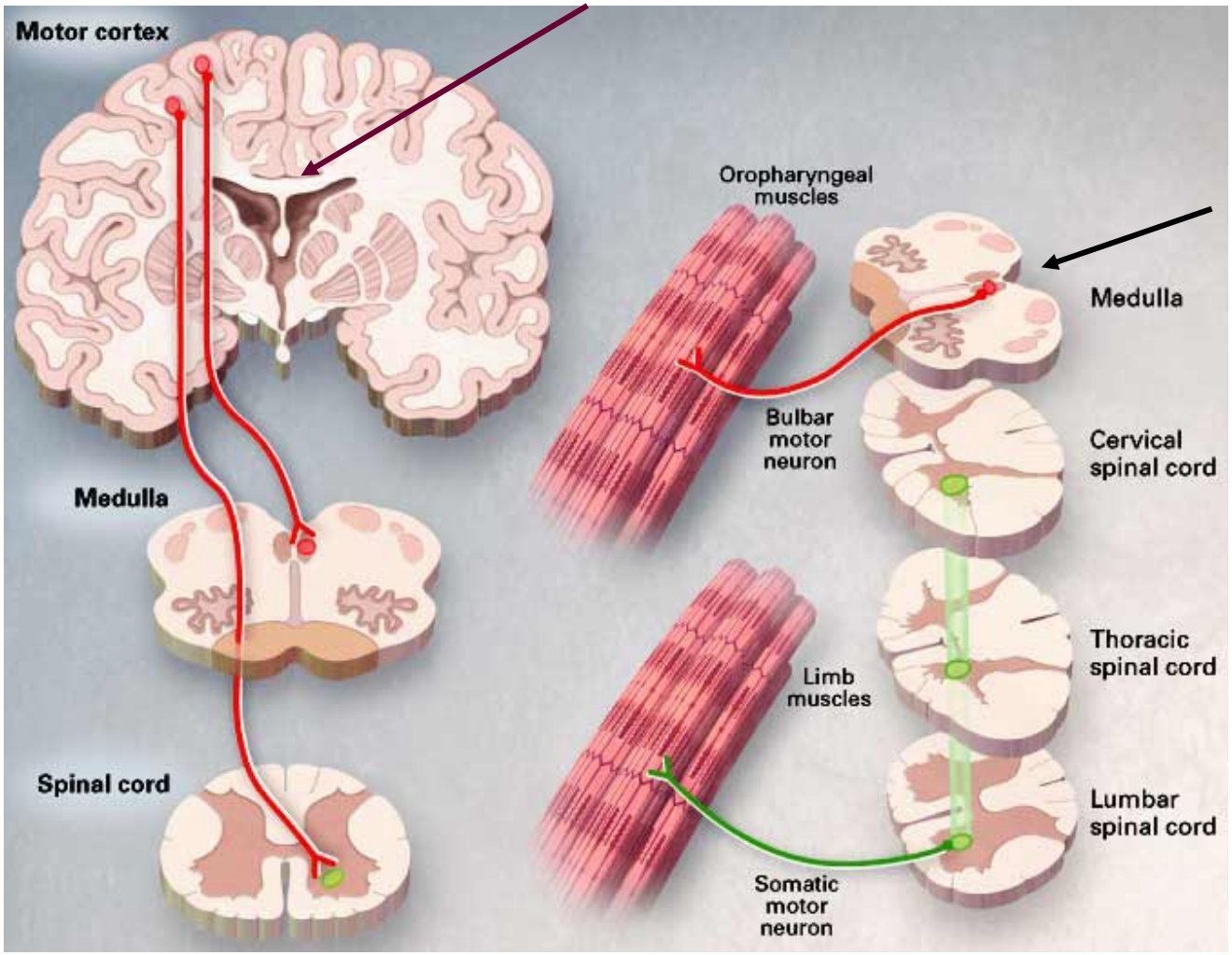
ALS spinal cord



Dying nerves

sALS phenothypes and loss of motoneurons

Loss of brain motor neurons (UMN) → spasticity



Loss of spinal motor neurons (LMN) → skeletal muscle denervation

Rowland and Schneider, NEJM 322:1688 (2001)

Causes of ALS

• In fALS mutation in the gene coding the cytosolic copper-zinc binding enzyme (SOD1 enzyme)- 1% of ALS cases

•Excitotoxicity:

excitatory neurotransmitters may participate to motoneurons death

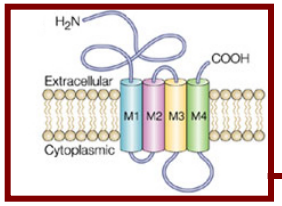
•Neuroinflammation

•Viruses, toxins, oxidative stress…

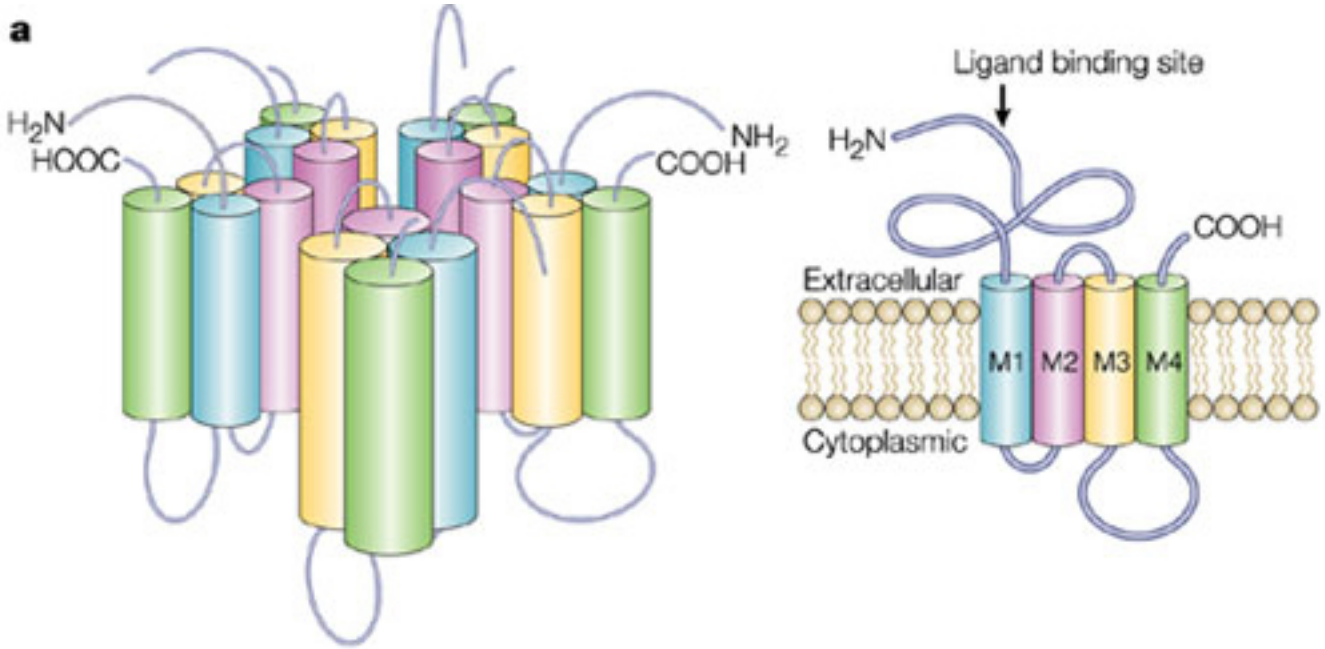
Why nAChRs?

nAChR controls Glu release

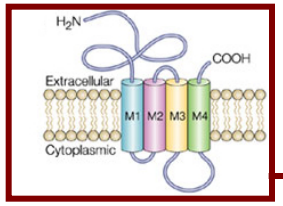
Ambigous role of smoke on neurodegenerative disease



Neuronal nicotinic receptors



- Ligand gated ion channels
- Prototypes of pentameric ionotropic receptors
- Role in synaptic transmission
- First discovered structure



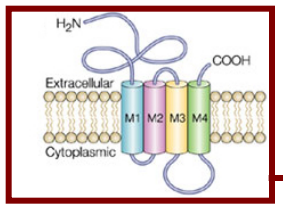
Mutation in nAChRs and diseases

monogenic familiar diseases

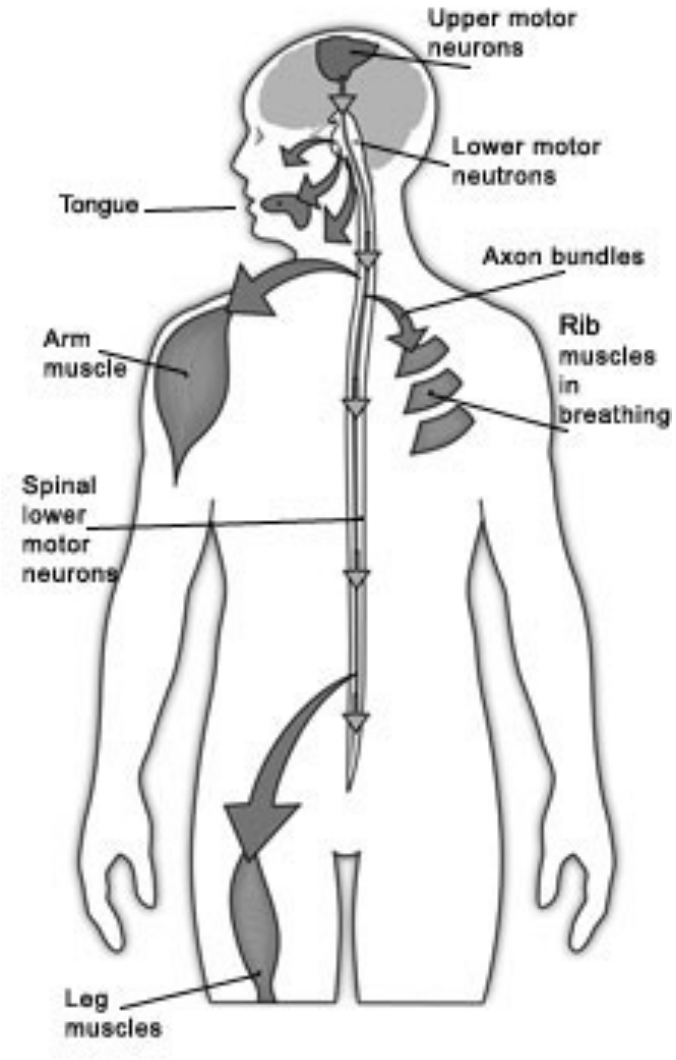
- Slow channel syndrome (muscle AChRs)
- Epilepsy (ADNFLE) neuronal nAChRs

susceptibilty factors in non familiar polygenic diseases

Mutations in the intracellular loop of nAChRs subunits have been found in patients affected by sALS



nAChRs distribution on spinal cord and upper motoneurons



Brainstem motoneuron

European Journal of Neuroscience, Vol. 22, pp. 2723–2734, 2005

Activation and desensitization of neuronal nicotinic receptors modulate glutamatergic transmission on neonatal rat hypoglossal motoneurons

Costanza Quitadamo, Elsa Fabbretti, Nerijus Lamanauskas and Andrea Nistri
Neurobiology Sector and CNR-INFM Unit, International School for Advanced Studies (SISSA), Via Beirut 4, 34014 Trieste, Italy

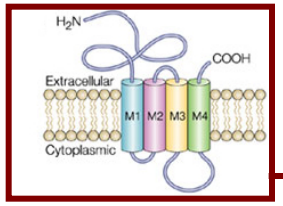
Renshaw cells and spinal motoneuron

J Neurophysiol
87: 3117–3125, 2002; 10.1152/jn.00745.2001.

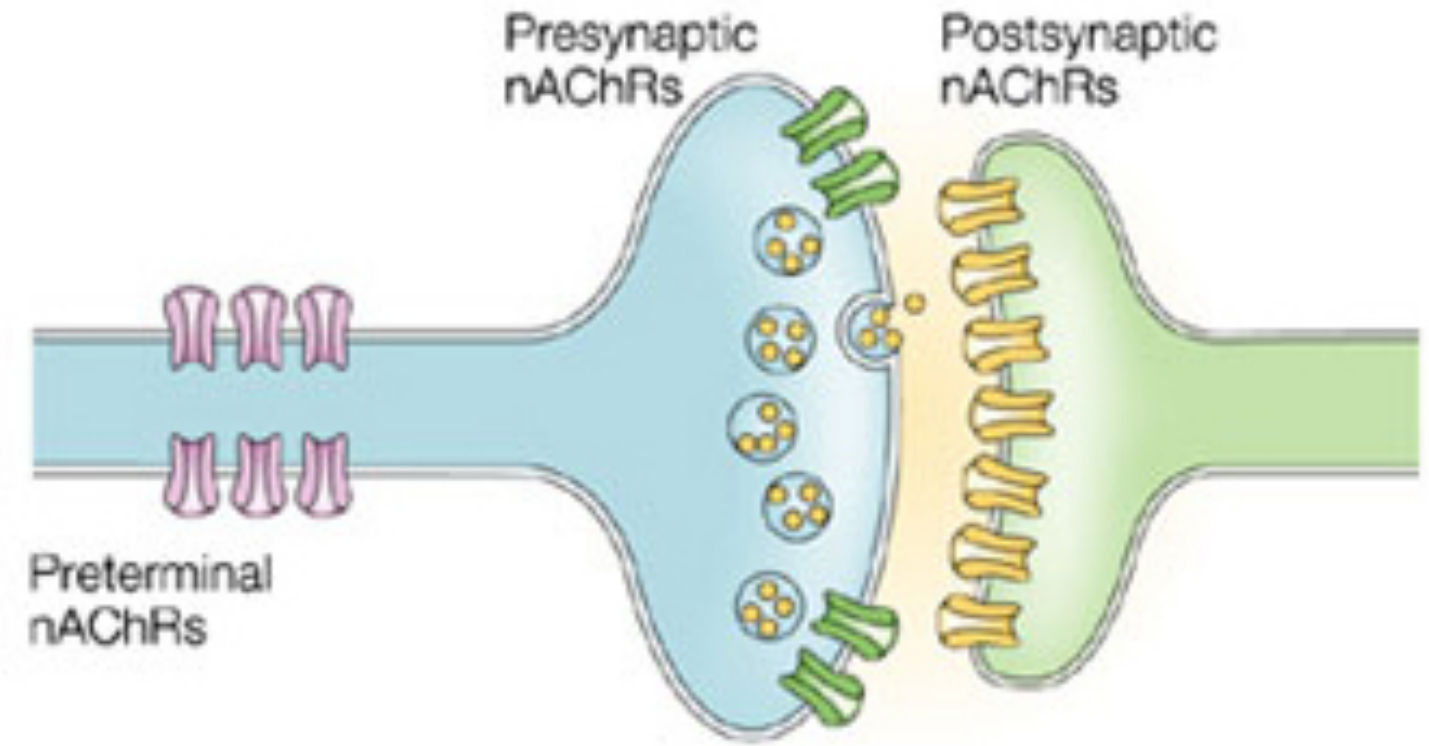
Properties of Nicotinic Receptors Underlying Renshaw Cell Excitation by α-Motor Neurons in Neonatal Rat Spinal Cord

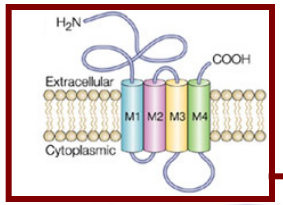
MICHELLE DOURADO AND PETER B. SARGENT
Departments of Stomatology and Physiology, University of California, San Francisco, California 94143

Received 4 September 2001; accepted in final form 29 January 2002

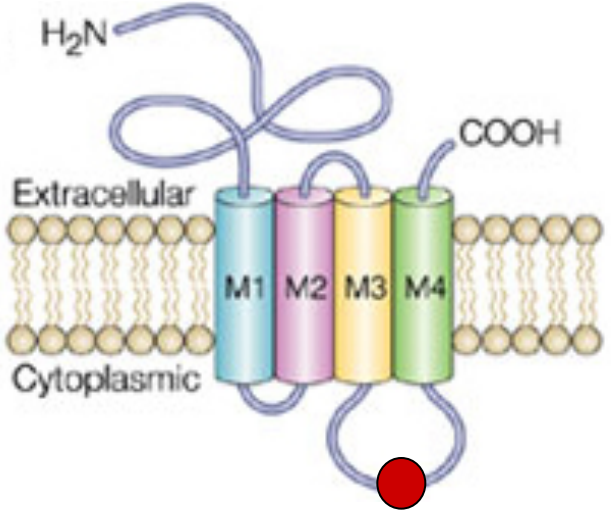


Pre and post synaptic role of nAchR





Mutations on nAchRs found in sALS patients



12 heterozygous different missense mutations

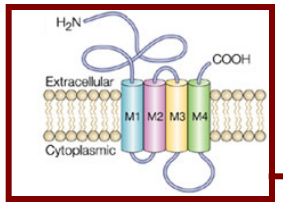
1 (A.C.)	α3	R385H
2 (S.L.)	α3	S388F
3 (M.F.)	α4	R345C
4 (P.P.)	α4	R495Q
5 (P.A.L.)	α4	Q572R
6 (M.L.)	α4	P451L
7 (TC.)	α4 β4	R487Q R349C
8 (P.G.)	α4	G454S
9 (S.A.)	α4	R336C
10 (R.M.)	β4	M456V
11 (I.M.)	β4	R349C
12 (Sb.A.)	β4	R349C
13(Se. A.)	β4	R349C
14 (C.M.)	β4	R349C
15 (M.A.) (trasmissione AD)	β2	Q397N

250 sALS patients analyzed

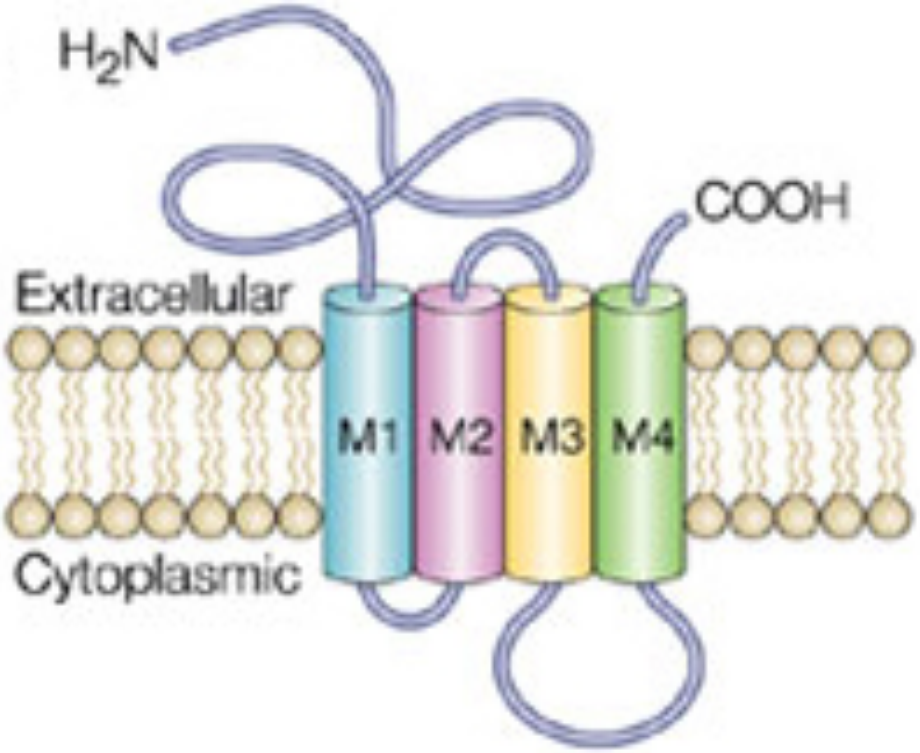
4.4% of sALS patients
Not found in 450 controls

Clinically:

- six patients had the predominant Upper Motor Neuron form



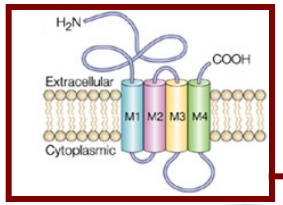
nAChRs subunit assemby



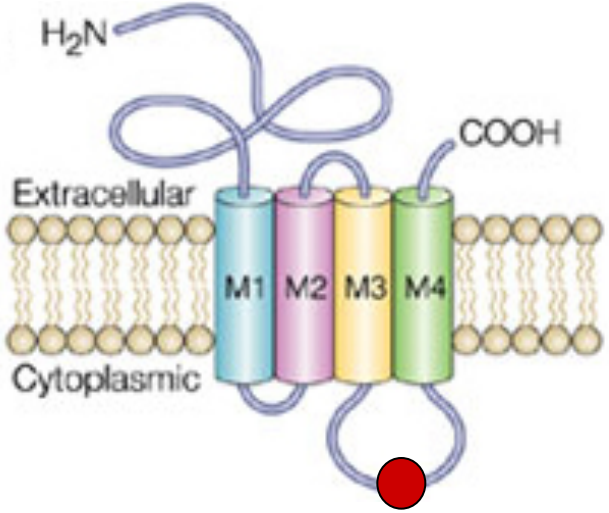
α3β4

α4β4

α4β2



Mutations on nAchRs found in sALS patients



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11 (I.M.)	β4	R349C
12 (Sb.A.)	β4	R349C
13(Se. A.)	β4	R349C
14 (C.M.)	β4	R349C
15 (M.A.) (trasmissione AD)	β2	Q397N

12 mutations in the M3-M4 cytosolic loop of α3 α4 β2 and β4 subunits

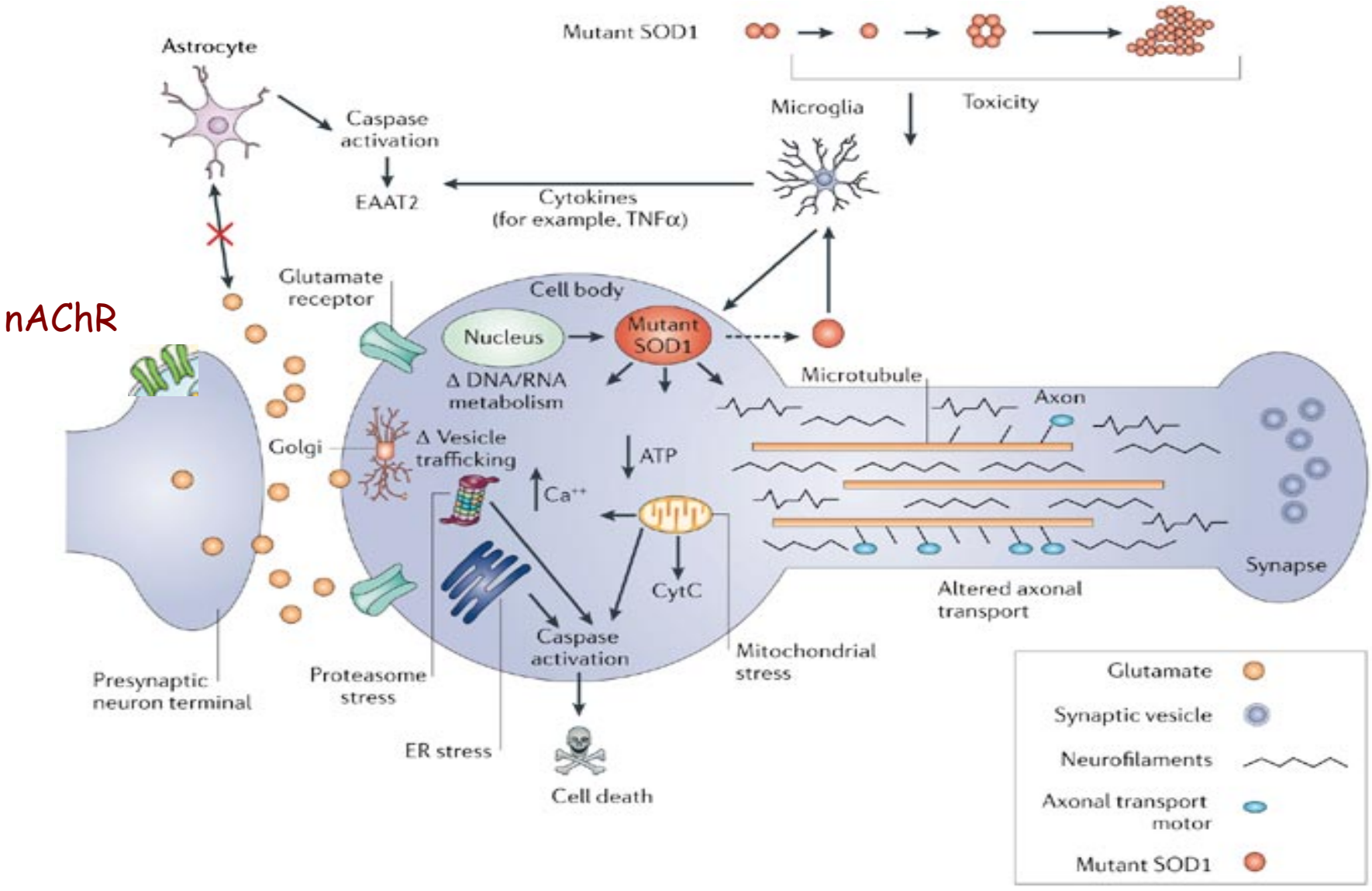
definite gain-of function of single mutant containing receptor

consistent with enhanced Ca2+ entry

the enhancement of Ca^{2+} entry may occur at pre- or postsynaptic sites

Reduced desensitization of presynaptic nAChRs at glutamatergic terminals might result in excessive excitatory glutamate release

potentiated activity of postsynaptic nAChRs might bring Ca2+
entry into neurons to neurotoxic levels.



Pasinelli and Brown *Nature Reviews Neuroscience* **7**, 710–723 (September 2006)

Idiopathic epilepsies: genetic alterations of ion channels

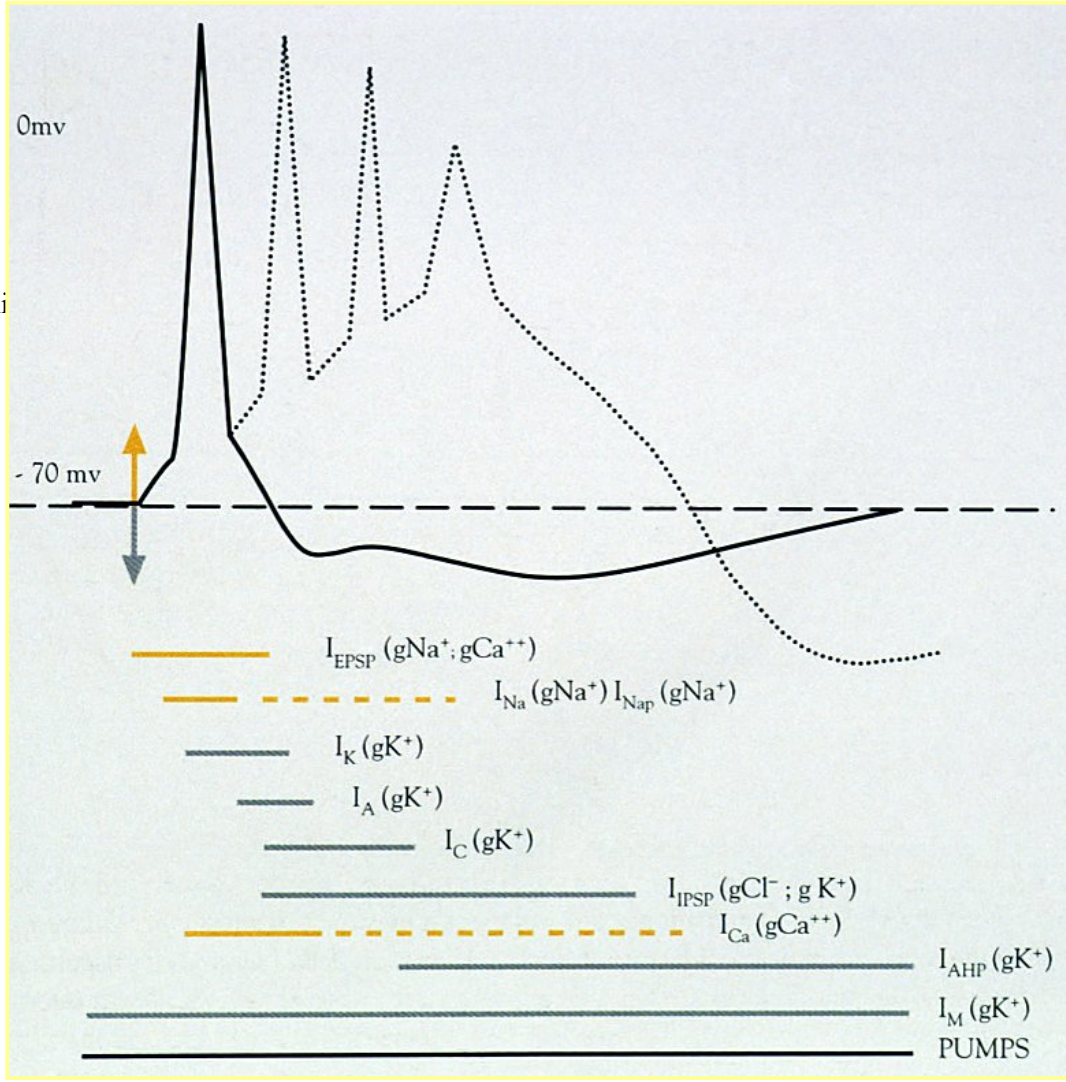
Idiopathic epilepsies

I. Reactive syndromes
- A. Neonatal seizures due to reversible causes
- B. Benign febrile convulsions
- C. “Low threshold” reactive seizures

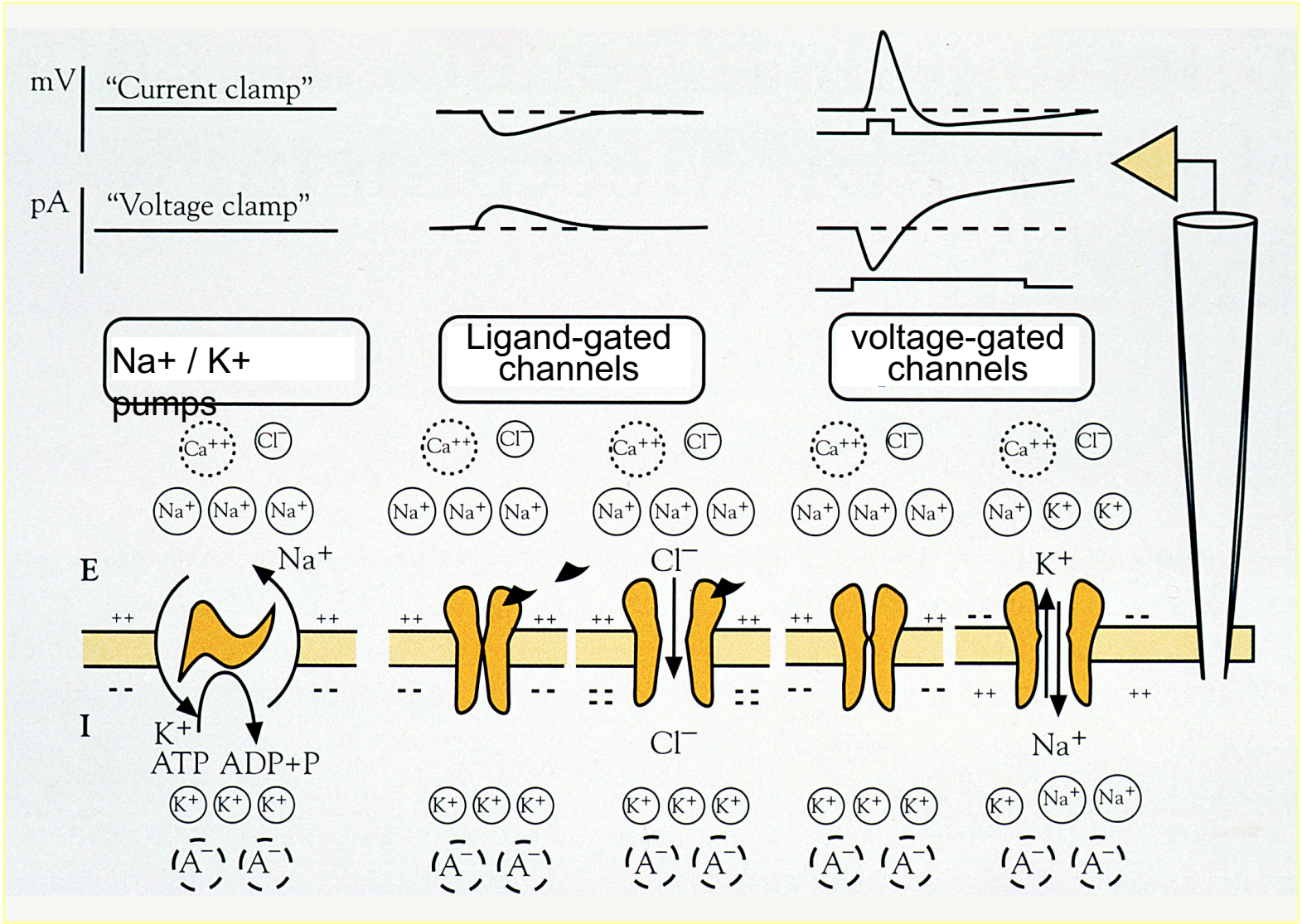
II. Idiopathic syndromes
- A. Benign Neonatal convulsions
  - I. Familial
  - 2. Non familial
- B. Partial syndromes
  - 1. Benign childhood epilepsy with centrotemporal spikes
  - 2. Childhood epilepsy with occipital paroxysms
- C. Generalized syndromes
  - 1. Childhood absence epilepsy (CAE)
  - 2. Juvenile absence epilepsy (JAE)
  - 3. Epilepsy with generalized tonic-clonic seizures on awakeni
  - 4. Juvenile absence epilepsy (JME)

III. Symptomatic syndromes
- A. Neonatal seizures due to irreversible causes
- B. Partial syndromes
  - 1. Epilepsia partialis continua (Kojewnikow’s syndrome)
    - a. Encephalopathic form (Rasmussen’s syndrome)
    - b. Focal form
  - 2. Temporal lobe epilepsy
- C. Generalized syndromes
  - 1. Early myoclonic encephalopathy
  - 2. Infantile spasms
  - 3. Lennaux-Gastaut syndrome

IV. Less well defined syndromes
- A. Severe myoclonic epilepsy of infancy
- B. Benign myoclonic epilepsy of infancy
- C. Epilepsy with myoclonic astatic seizures
- D. Epilepsy with myoclonic absences
- E. Acquired epileptic aphasia (Landau-Kleffner syndrome)
- F. Epilepsy with continuous spikes and waves during sleep (ESES)
- G. Reflex epilepsies



Epileptic Chanelopathies



Epileptic Chanelopathies

- Rare

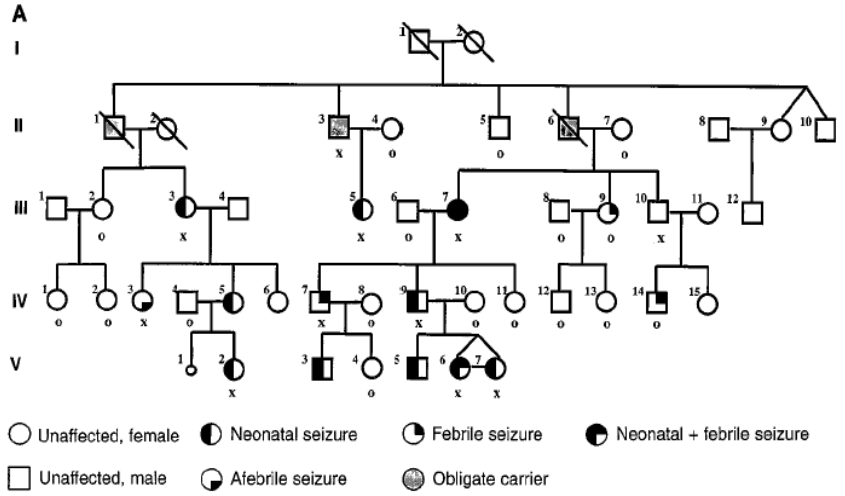
- Generalized and focal epilepsies
- Moderate phenotypic variability

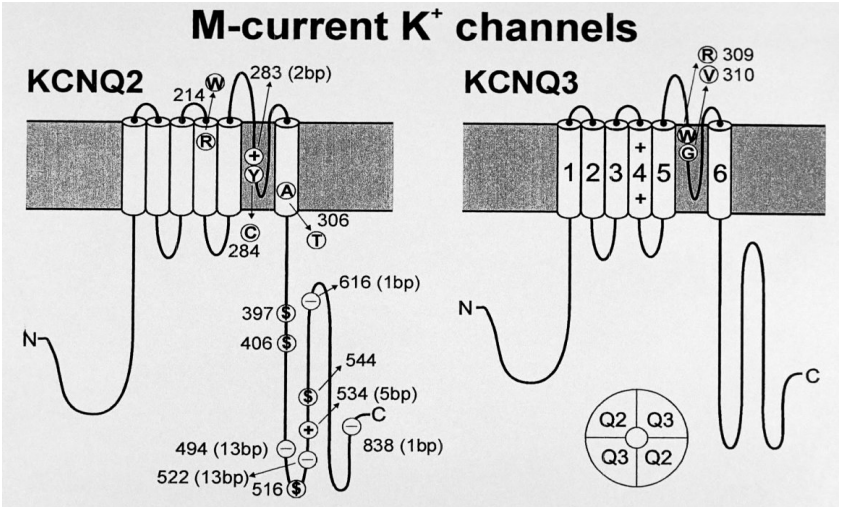
- Autosomal Dominant mode of inheritance
- Incomplete penetrance (~ 80%)

Voltage-Gated Potassium Channels

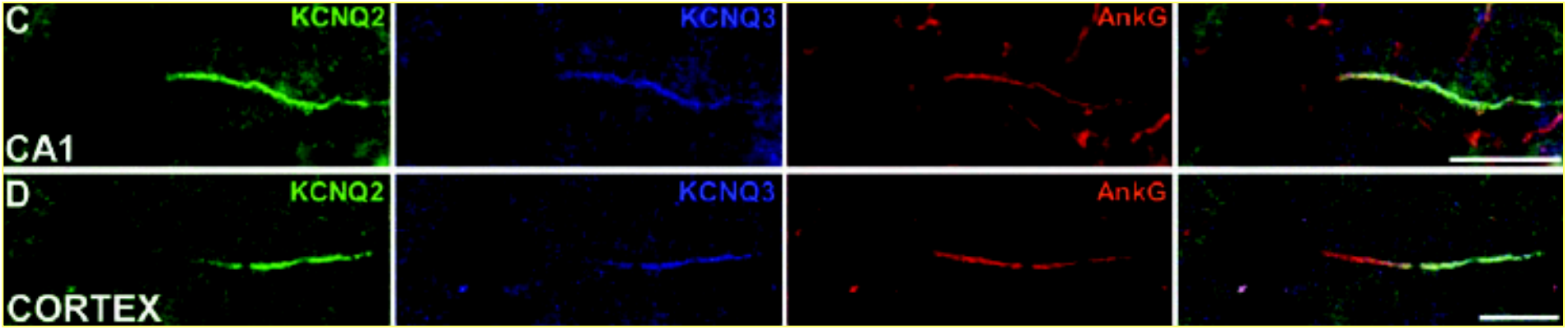
Mendelian idiopathic epilepsies

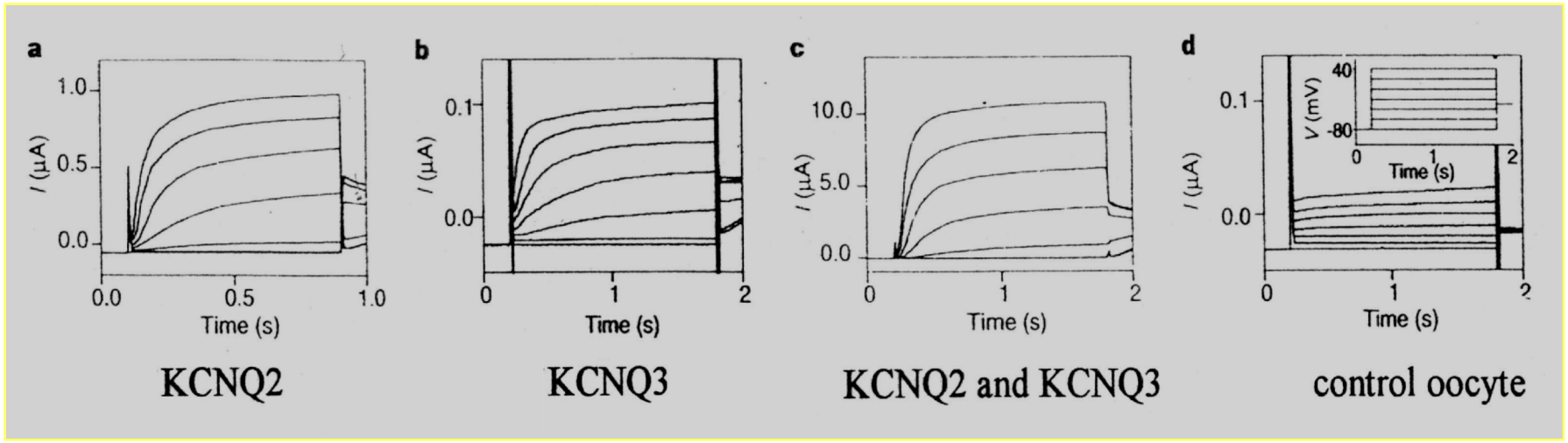
- Age of onset between 2 to 4 days of age
- Partial seizures
  - tonic posture progressing to clonic movements
  - ocular symptoms
  - apneic spells
- Normal interictal EEG
- Ictal EEG showing a sequence of generalized attenuation followed by slow waves, spikes and burst-suppression
- No psychomotor delay or brain lesions
- 10 % risk of febrile convulsions or epilepsy





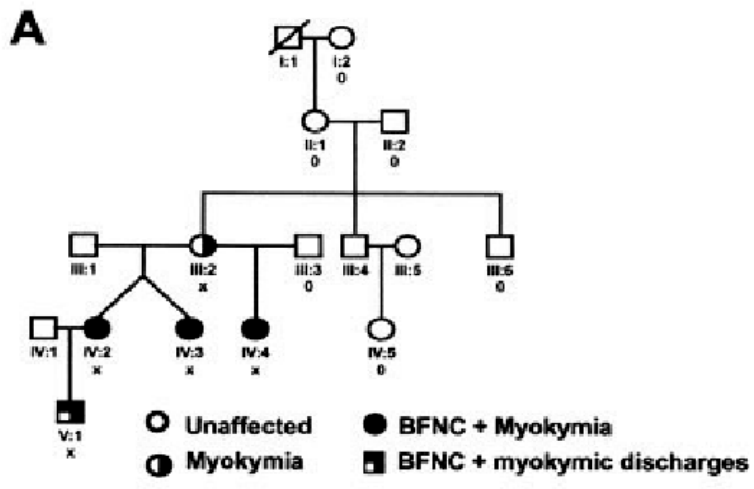
Distribution of KCNQ2 and KCNQ3 in the CNS

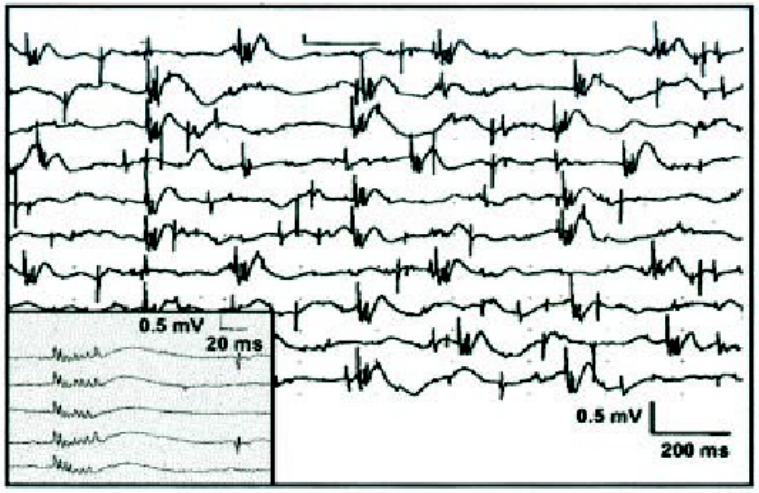




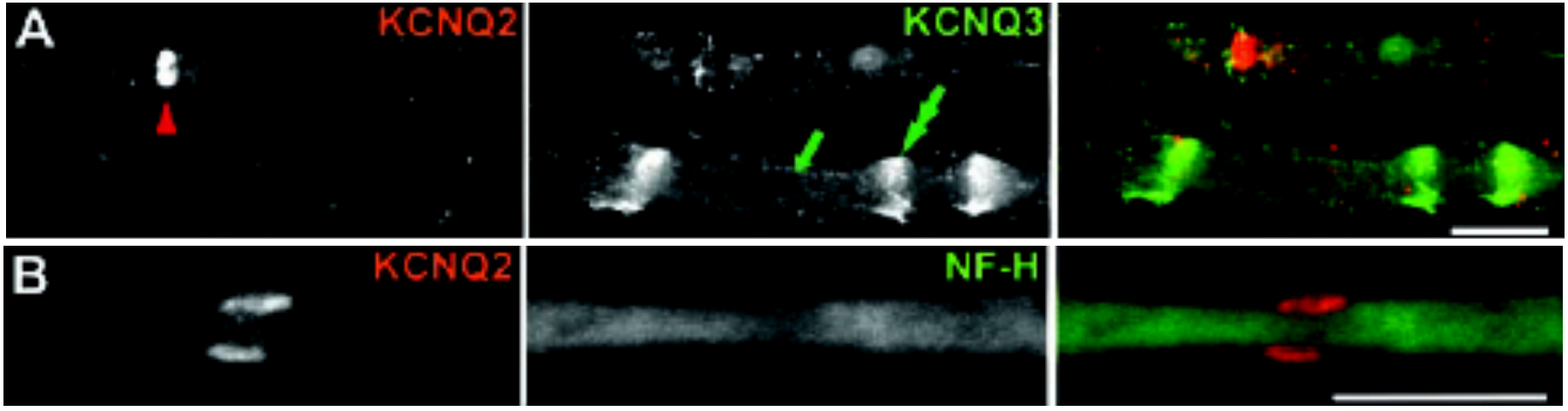
Myokymia and neonatal epilepsy caused by a mutation in the voltage sensor of the KCNQ2 K^+ channel

Karin Dedek*, Bernhard Kunath†, Colette Kananura‡, Ulrike Reuner†, Thomas J. Jentsch*§, and Ortrud K. Steinlein‡§

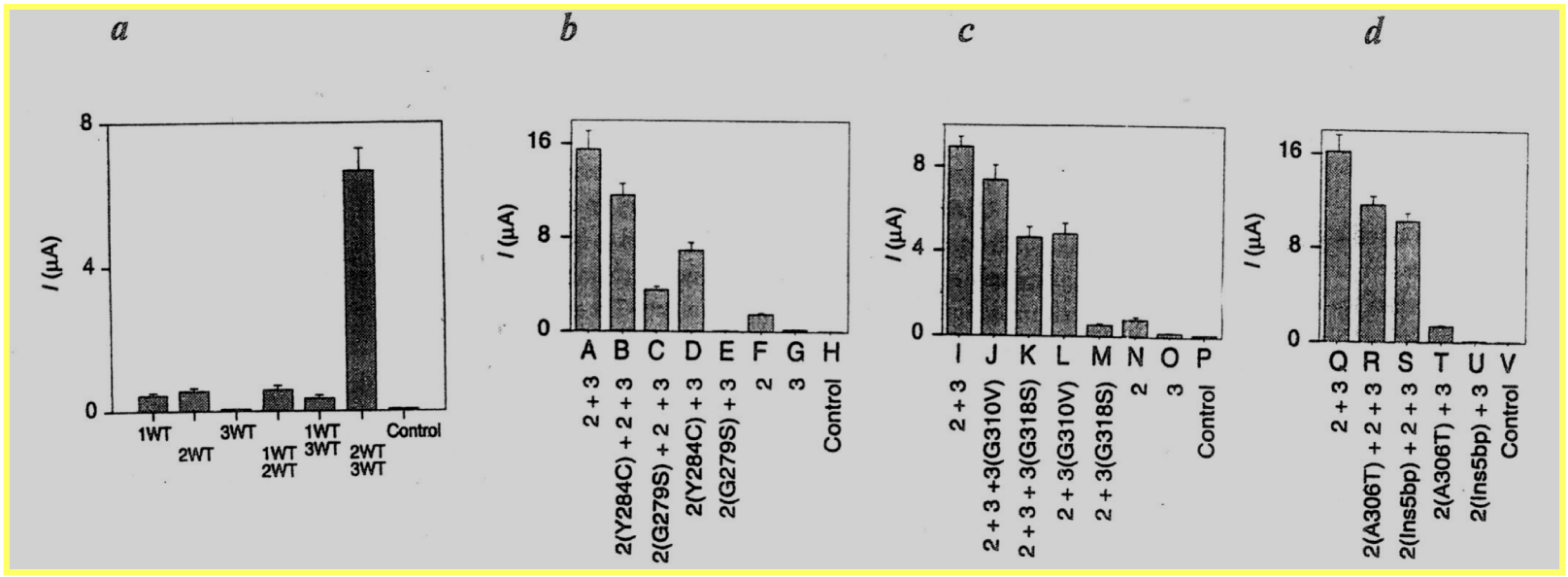




Distribution of KCNQ2 and KCNQ3 in the PNS



Functional analysis of voltage-gated K^+ channels



Mendelian idiopathic epilepsies

Variable age of onset (4-40 years, average ~20)

Simple Partial Seizures

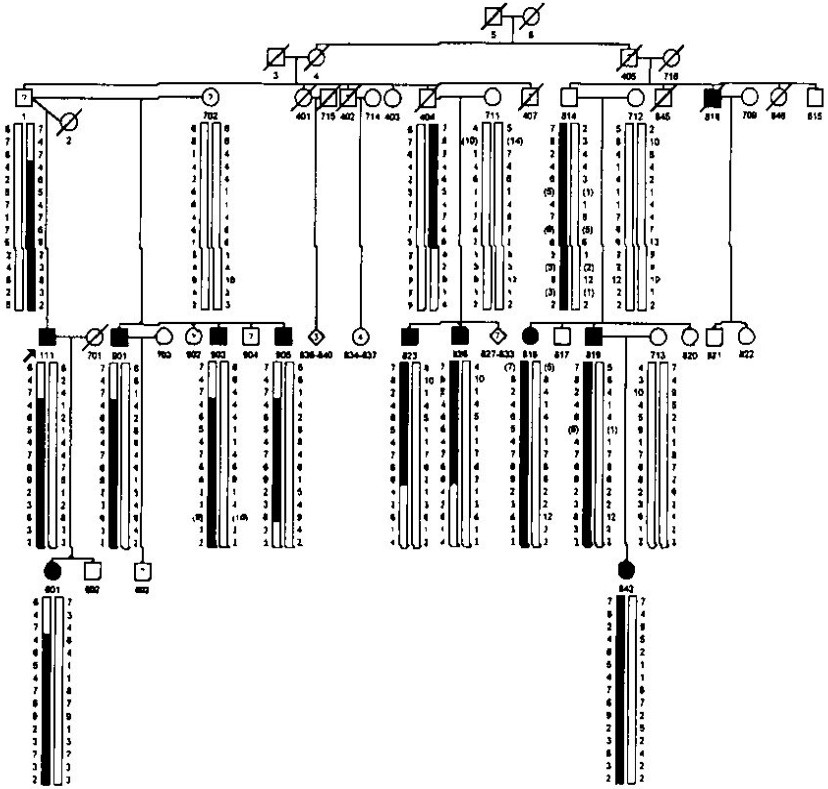
Auditory aurea (Visual symptoms, aphasia)

Secondarily Generalized Tonic-Clonic Seizures

Seizures sometimes pharmacoresistant

Normal interictal EEG or mild temporal abnormalities

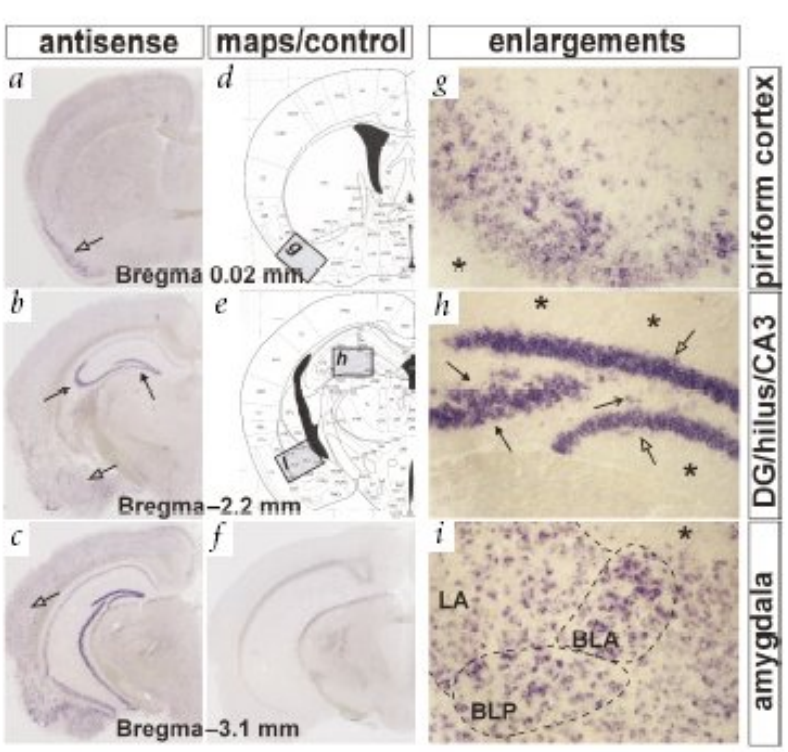
Absence of psychomotor delay or brain lesions

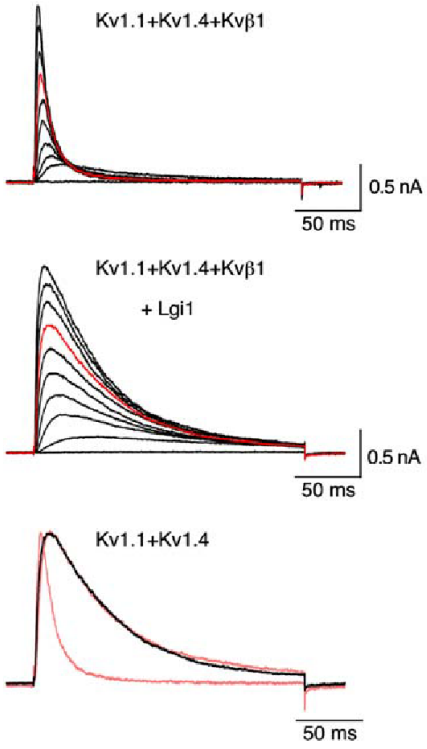


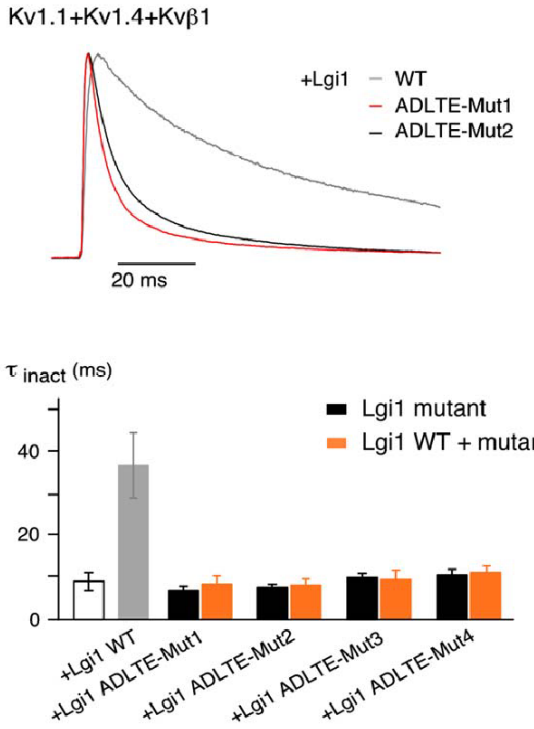
The Epilepsy-Linked Lgi1 Protein Assembles into Presynaptic Kv1 Channels and Inhibits Inactivation by Kvβ1

Uwe Schulte,[2] Jörg-Oliver Thumfart,[1] Nikolaj Klöcker,[1]
Claudia A. Sailer,[1,3] Wolfgang Bildl,[1]
Martin Biniossek,[5] Doris Dehn,[4] Thomas Deller,[4]
Silke Eble,[1] Karen Abbass,[2] Tanja Wangler,[2]
Hans-Günther Knaus,[3] and Bernd Fakler[1,*]

Neuron *49*, 697–706, March 2, 2006





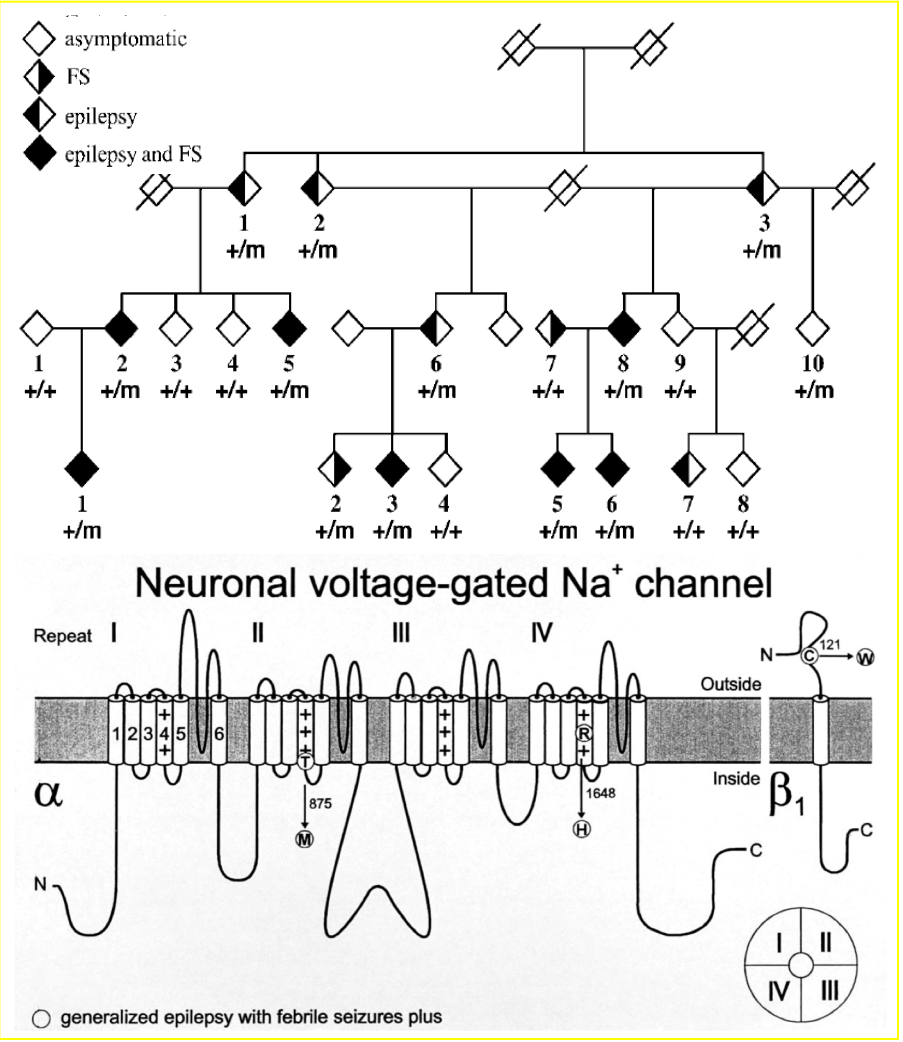


Voltage-Gated Sodium Channels

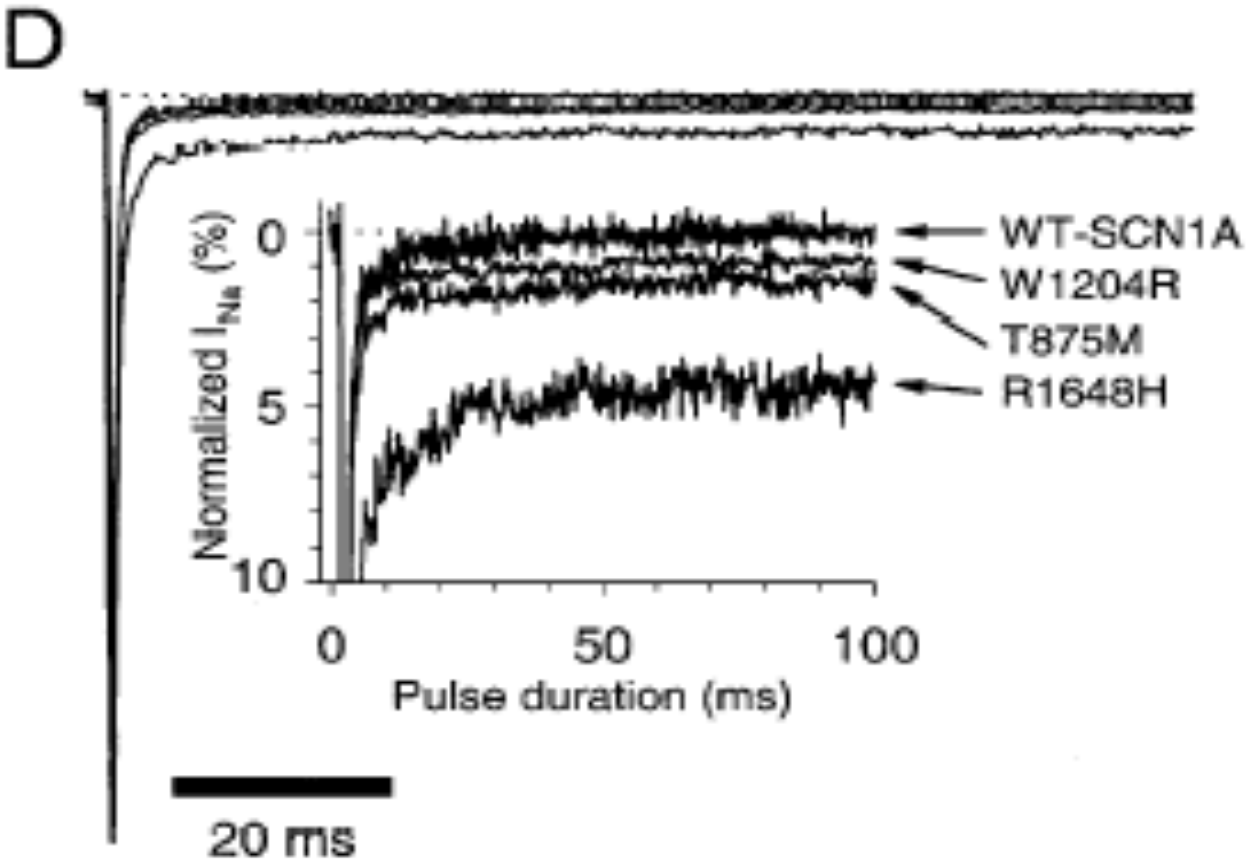
Generalized Epilepsy and Febrile Seizures plus

Clinical features

- Clinical Variability
- Febrile Seizures (< 6 y)
- Febrile Seizures "plus" (> 6y)
- Afebrile seizures, usually generalized (tonic-clonic, absence, myoclonic, atonic)
- Benign outcome
- No brain lesions or metabolic disorders



Functional analysis of voltage-gated Na^+ α1 subunit



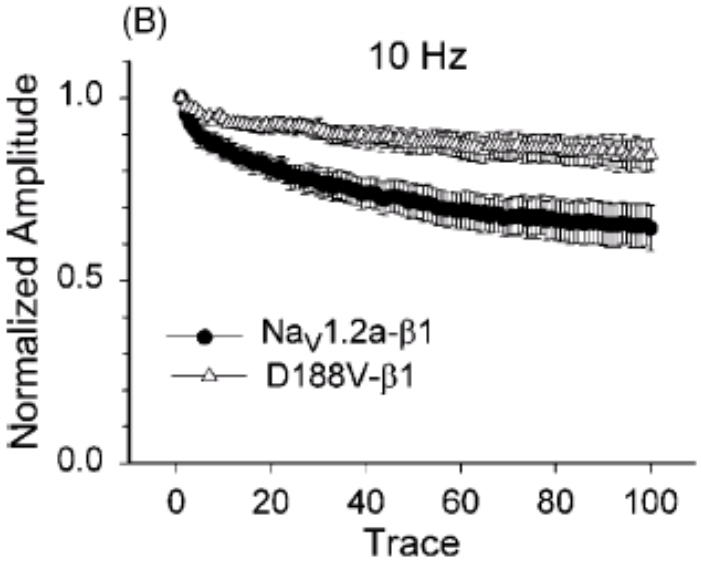
Epilepsy Research 53 (2003) 107–117

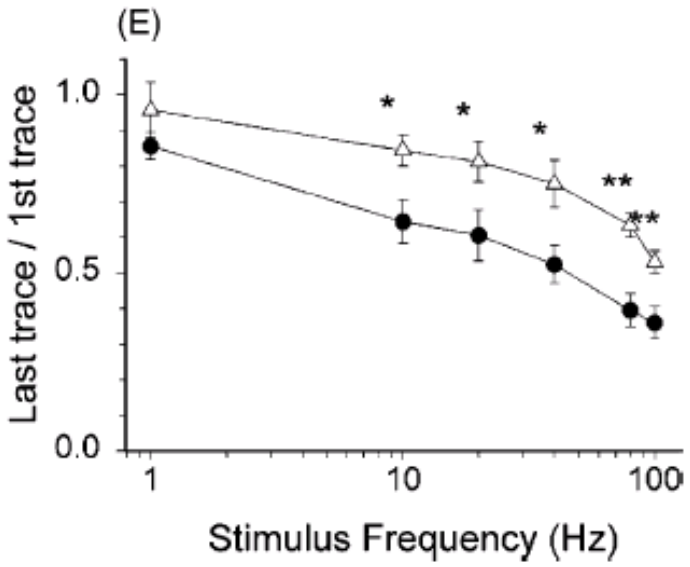
Functional characterization of the D188V mutation in neuronal voltage-gated sodium channel causing generalized epilepsy with febrile seizures plus (GEFS)

Patrick Cossette[a,1], Andrew Loukas[b,1], Ronald G. Lafrenière[a,c], Daniel Rochefort[a], Eric Harvey-Girard[a], David S. Ragsdale[b], Robert J. Dunn[a], Guy A. Rouleau[a,*]

Phenotype: GEFS+
Mutation: D188V

cDNA: rat SCN2A
Expression System: Human HEK





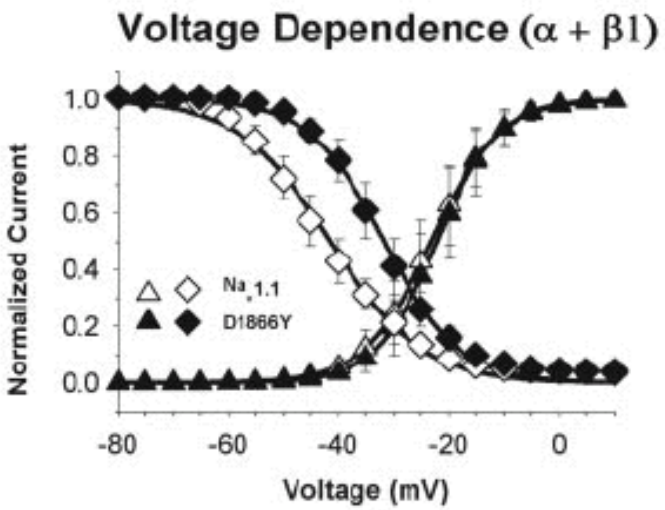
Resistance to cumulative inactivation during high frequency activation,

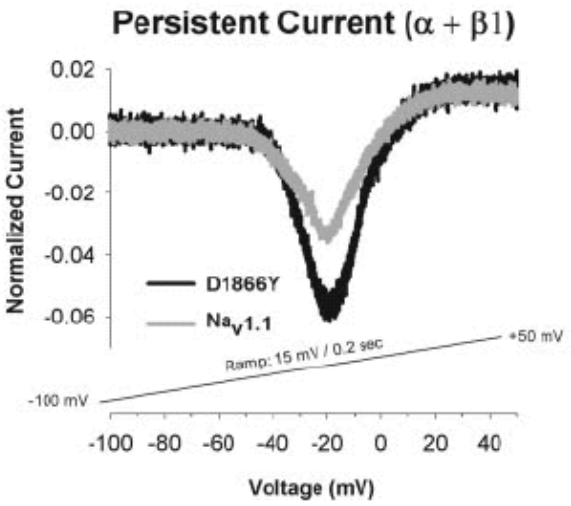
A Novel Epilepsy Mutation in the Sodium Channel *SCN1A* Identifies a Cytoplasmic Domain for β Subunit Interaction

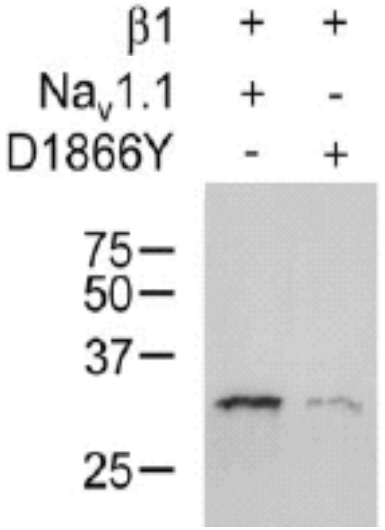
J. Spampanato,[1] J. A. Kearney,[3] G. de Haan,[3] D. P. McEwen,[4] A. Escayg,[3] I. Aradi,[2] B. T. MacDonald,[3] S. I. Levin,[3] I. Soltesz,[2] P. Benna,[5] E. Montalenti,[5] L. L. Isom,[4] A. L. Goldin,[1,2] and M. H. Meisler[3]

Departments of [1]Microbiology and Molecular Genetics and [2]Anatomy and Neurobiology, University of California, Irvine, Irvine, California 92697-4025, Departments of [3]Human Genetics and [4]Pharmacology, University of Michigan, Ann Arbor, Michigan 48109-0618, and [5]Department of Neurosciences, University of Torino, 10126 Torino, Italy

10022 • The Journal of Neuroscience, November 3, 2004 • 24(44):10022–10034







Am. J. Hum. Genet. 68:1327–1332, 2001

De Novo Mutations in the Sodium-Channel Gene *SCN1A* Cause Severe Myoclonic Epilepsy of Infancy

Lieve Claes,[1] Jurgen Del-Favero,[1] Berten Ceulemans,[2,3] Lieven Lagae,[3,4] Christine Van Broeckhoven,[1] and Peter De Jonghe[1,2]

[1]Department of Molecular Genetics, Flanders Interuniversity Institute for Biotechnology (VIB), University of Antwerp, and [2]Department of Neurology, University Hospital Antwerp, Antwerp; [3]Epilepsy Center for Children and Youth, Pulderbos, Belgium; and [4]Department of Child Neurology, University Hospital Gasthuisberg, Leuven, Belgium

DOI: 10.1093/brain/awg053 *Brain* (2003), **126,** 531–546

Mutations of sodium channel α subunit type 1 (SCN1A) in intractable childhood epilepsies with frequent generalized tonic–clonic seizures

Tateki Fujiwara,[1] Takashi Sugawara,[2] Emi Mazaki-Miyazaki,[2] Yukitoshi Takahashi,[3] Katsuyuki Fukushima,[1] Masako Watanabe,[1] Keita Hara,[1] Tateki Morikawa,[1] Kazuichi Yagi,[1] Kazuhiro Yamakawa[2] and Yushi Inoue[1]

$Na_V1.1$ channels with mutations of severe myoclonic epilepsy in infancy display attenuated currents

Takashi Sugawara [a], Yuji Tsurubuchi [b], Tateki Fujiwara [c], Emi Mazaki-Miyazaki [a], Keiichi Nagata [b], Mauricio Montal [d], Yushi Inoue [c], Kazuhiro Yamakawa [a,*]

Epilepsy Research 54 (2003) 201–207

[a] *Laboratory for Neurogenetics, Brain Science Institute, RIKEN, 2-1 Hirosawa, Wako, Saitama 351-0198, Japan*
[b] *Laboratory for Memory and Learning, Brain Science Institute, RIKEN, 2-1 Hirosawa, Wako, Saitama 351-0198, Japan*
[c] *National Epilepsy Center, Shizuoka Medical Institute of Neurological Disorders, 886 Urushiyama, Shizuoka 420-8688, Japan*
[d] *Section of Neurobiology, Division of Biological Sciences, University of California, San Diego, La Jolla, CA 92093, USA*

Received 28 January 2003; received in revised form 10 April 2003; accepted 14 April 2003

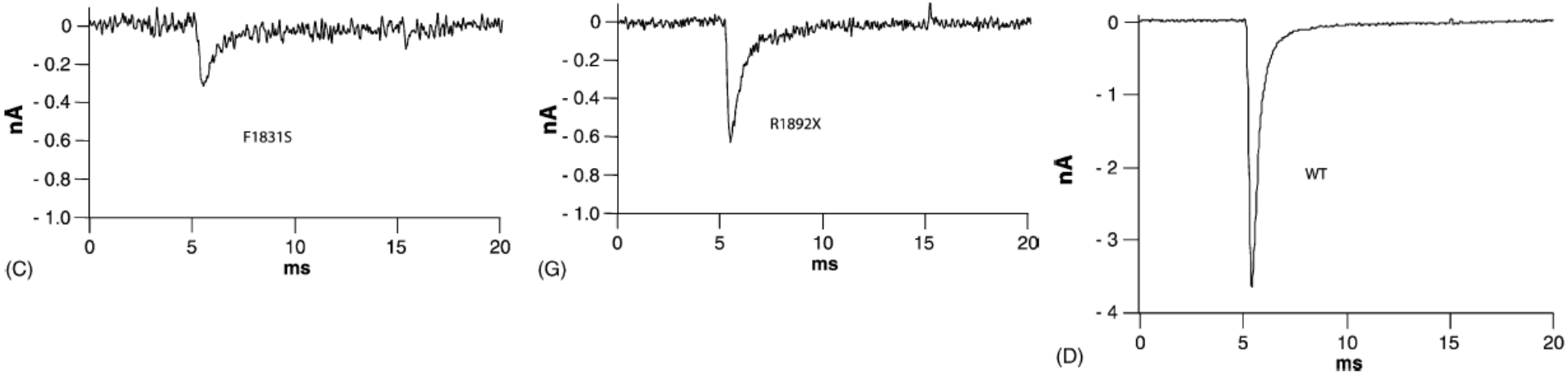


Fig. 2. Representative whole-cell current recordings in HEK293 cells expressing WT human $Na_V1.1$ channel, and those with SMEI-associated mutations. Currents were evoked from a holding potential of −120 to 0 mV. More than 10 fluorescent-active cells were recorded for each mutant channel, and maximal sodium currents were shown in the figure. (A–C): Mutant channels bearing missense mutations (G979R, N985I, and F1831S). (D): WT control. (E–G): Mutant channels bearing nonsense mutations (R712X, R1407X, and R1892X).

Reduced sodium current in GABAergic interneurons in a mouse model of severe myoclonic epilepsy in infancy

Frank H Yu[1], Massimo Mantegazza[1,4], Ruth E Westenbroek[1], Carol A Robbins[2,3], Franck Kalume[1], Kimberly A Burton[1], William J Spain[3], G Stanley McKnight[1], Todd Scheuer[1] & William A Catterall[1]

VOLUME 9 | NUMBER 9 | SEPTEMBER 2006 NATURE NEUROSCIENCE

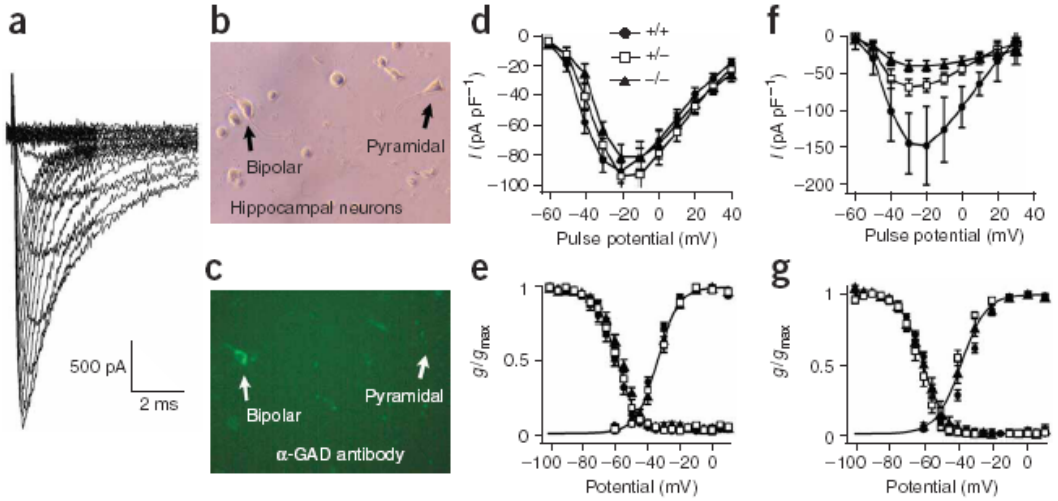


Figure 4 Sodium currents from hippocampal neurons in wild-type and heterozygous and null $Na_V1.1$ mice. (**a**) A representative set of sodium current traces from hippocampal pyramidal cells after subtraction of traces recorded in the presence of 1 μM tetrodotoxin, which were elicited by depolarizing steps from –60 to –15 mV in 5-mV increments from a holding potential of –100 mV. (**b**) Representative bright-field view of hippocampal neurons that were acutely dissociated from P14 wild-type mice. The pyramidal-shaped and bipolar-shaped neurons are indicated with arrows. (**c**) Same hippocampal neurons as in **b** but after immunocytochemical processing and staining with anti-GAD. The bipolar-shaped cells, but not the pyramidal-shaped cells, were strongly labeled, which indicates that they are GABAergic inhibitory interneurons. (**d**,**f**) Current density–voltage relationships of whole-cell sodium currents from hippocampal pyramidal (**d**) and bipolar (**f**) neurons for wild-type, heterozygous and homozygous mice. Currents of heterozygous and homozygous bipolar neurons were significantly smaller than those of wild-type neurons ($P < 0.05$). (**e**,**g**) Voltage dependence of activation (right curves) and steady-state inactivation (left curves) of sodium currents from hippocampal pyramidal (**e**) and bipolar (**g**) neurons. Same symbols for mouse genotypes as in **d**. Error bars are s.e.m.

Reduced sodium current in GABAergic interneurons in a mouse model of severe myoclonic epilepsy in infancy

Frank H Yu[1], Massimo Mantegazza[1,4], Ruth E Westenbroek[1], Carol A Robbins[2,3], Franck Kalume[1], Kimberly A Burton[1], William J Spain[3], G Stanley McKnight[1], Todd Scheuer[1] & William A Catterall[1]

VOLUME 9 | NUMBER 9 | SEPTEMBER 2006 NATURE NEUROSCIENCE

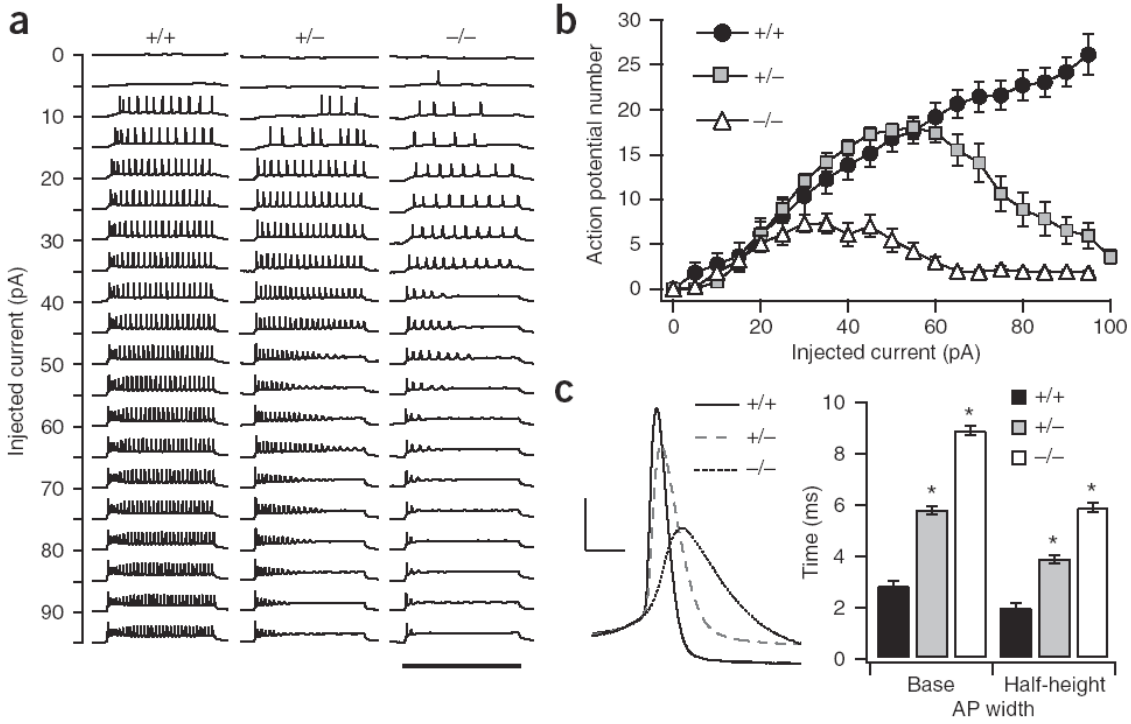


Figure 5 Depolarization-evoked firing activity of hippocampal interneurons from $Na_V1.1$ mutant mice. (**a**) Representative sets of action potential traces recorded from wild-type (+/+), heterozygous (+/–) and homozygous (–/–) interneurons during application of 800-ms injections of depolarizing current in +10-pA increments, from a holding potential of –80 mV. (**b**) Input-output relationship of the number of action potentials elicited versus the injected current. (**c**) Left panel, representative single action potentials recorded for each genotype of interneurons elicited by the same injected current amplitude (35 pA). Right panel, summary bar graph of action potential width and half-width, which are respectively defined as the width at the base and half-maximum amplitude of the action potential. Action potential parameters are listed in **Supplementary Table 1**. Error bars are s.e.m. $^*P < 0.01$.

Mendelian idiopathic epilepsies

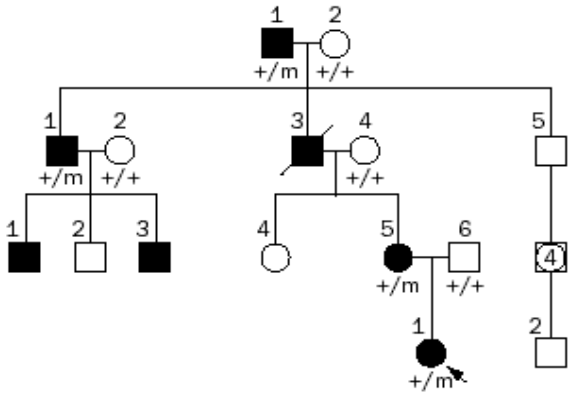
- Age of onset between 2 days and 6months of age
- Partial seizures, usually in cluster

  psychomotor arrest

  slow deviation of the head and eyes to one side

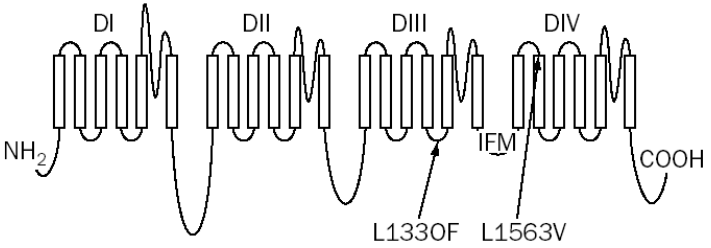
  asynchromous limb jerks

- Normal interictal EEG
- Ictal EEG showing a recruiting rhythm with central-occipital region outset and secondary generalization
- Absence of pshycomotor delay or brain lesions



SCN2A

Voltage-gated sodium channel α2 subunit



Effects in Neocortical Neurons of Mutations of the $Na_v1.2$ Na^+ Channel causing Benign Familial Neonatal-Infantile Seizures

The Journal of Neuroscience, October 4, 2006 • 26(40):10100–10109

Paolo Scalmani,[1] Raffaella Rusconi,[1] Elena Armatura,[1] Federico Zara,[2] Giuliano Avanzini,[1] Silvana Franceschetti,[1] and Massimo Mantegazza[1]

[1]Department of Neurophysiopathology, Istituto Neurologico C. Besta, 20133 Milan, Italy, and [2]Laboratory of Neurogenetics, Unit of Muscular and Neurodegenerative Disease, Istituto G. Gaslini, University of Genova, 16147 Genova, Italy

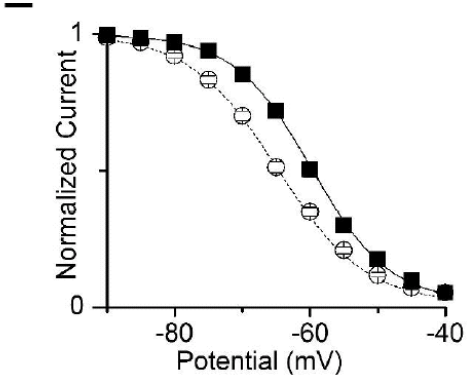


Figure 5. Functional effects of the mutation L1330F. E, mean voltage dependence of inactivation (solid for L1330F, dashed for wild type Nav1.2).

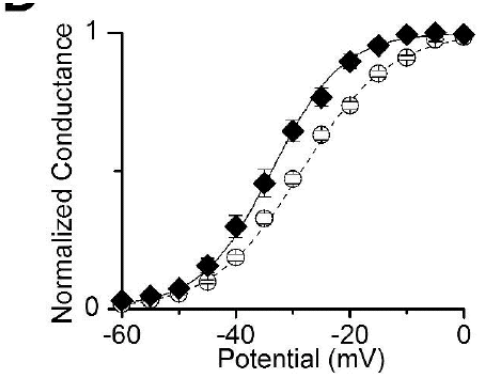
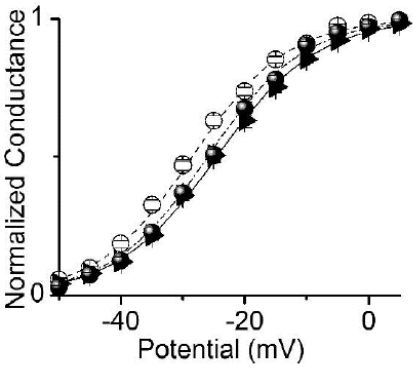


Figure 6. Functional effects of the mutation L1563V. D, mean voltage dependence of activation (solid for L1563V, dashed for wild type Nav1.2.



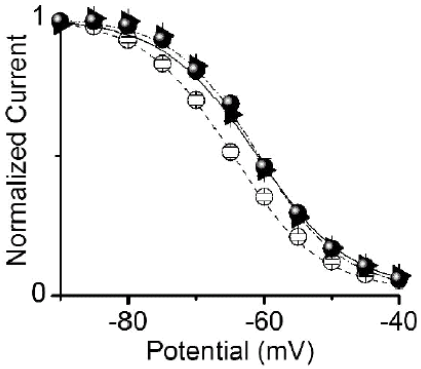


Figure 7. Functional effects of the mutations R223Q and R1319Q. D, mean voltage dependence of activation E, mean voltage dependence of inactivation (solid for R223Q, dashed-dotted for R1319Q and dashed for wild type Nav1.2).

Effects in Neocortical Neurons of Mutations of the $Na_v1.2$ Na^+ Channel causing Benign Familial Neonatal-Infantile Seizures

The Journal of Neuroscience, October 4, 2006 • 26(40):10100–10109

Paolo Scalmani,[1] Raffaella Rusconi,[1] Elena Armatura,[1] Federico Zara,[2] Giuliano Avanzini,[1] Silvana Franceschetti,[1] and Massimo Mantegazza[1]

[1]Department of Neurophysiopathology, Istituto Neurologico C. Besta, 20133 Milan, Italy, and [2]Laboratory of Neurogenetics, Unit of Muscular and Neurodegenerative Disease, Istituto G. Gaslini, University of Genova, 16147 Genova, Italy

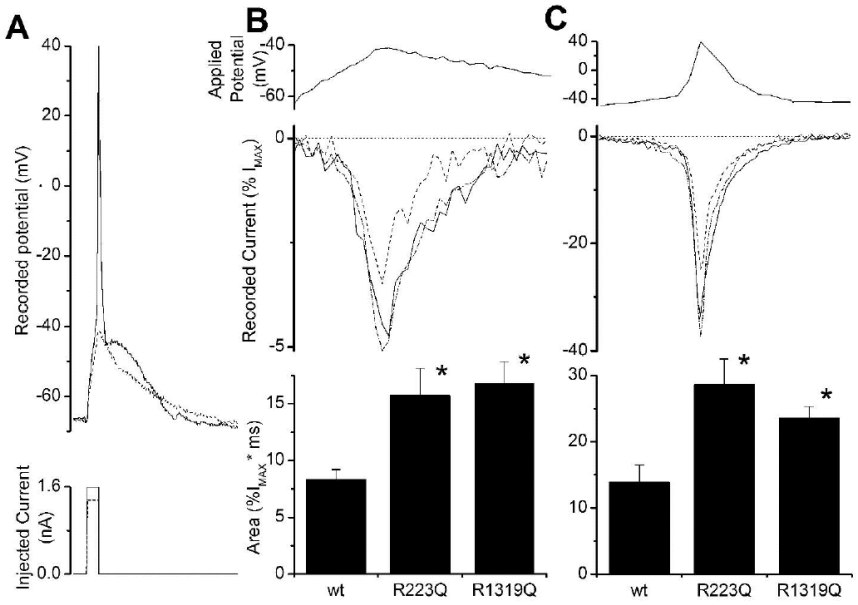
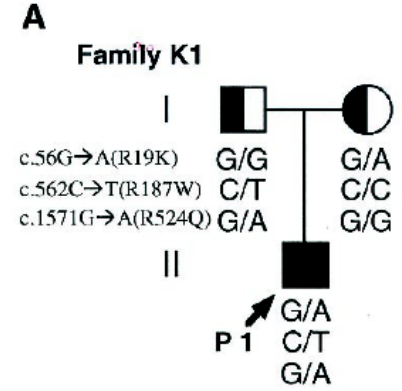


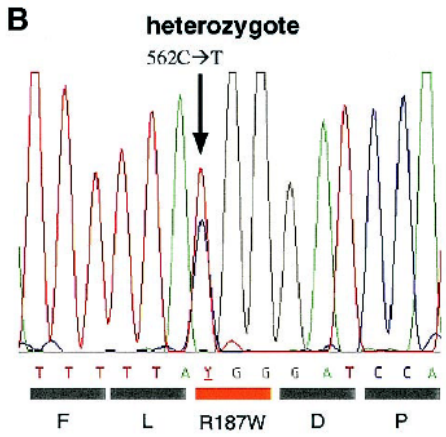
Figure 8. Functional effects of the mutations R223Q and R1319Q studied with physiological voltage stimuli. A, action potential and subthreshold response recorded with sharp microelectrodes in a Layer V neuron in neocortical slices, the lower panel is the injected depolarizing current pulse; scale bar 10ms. B, currents elicited in transfected neurons by the subthreshold response shown in A; the upper panel shows the subthreshold response used as voltage stimulus; the middle panel shows the recorded subthreshold currents (solid line for R223Q, dashed-dotted line for R1319Q and dashed line for wild type Nav1.2), scale bar 1ms; the bar-graph in the lower panel shows the comparison between the area subtended by the subthreshold currents (see also table 2). C, currents elicited in transfected neurons by the action potential shown in A; the upper panel shows the action potential used as voltage stimulus; the middle panel shows the recorded action currents (solid line for R223Q, dashed-dotted line for R1319Q and dashed line for wild type Nav1.2), scale bar 1ms; the bar-graph in lower panel shows the comparison between the area subtended by the action currents recorded in the three conditions (see also table2).

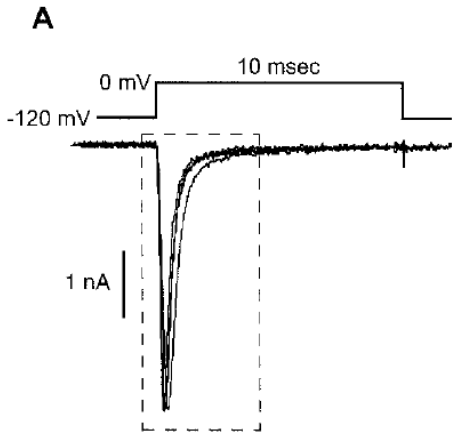
A missense mutation of the Na^+ channel α_{II} subunit gene $Na_v1.2$ in a patient with febrile and afebrile seizures causes channel dysfunction

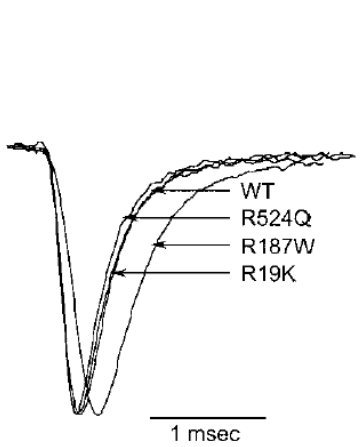
Takashi Sugawara*, Yuji Tsurubuchi†, Kishan Lal Agarwala*, Masatoshi Ito‡, Goryu Fukuma§, Emi Mazaki-Miyazaki*, Hiroshi Nagafuji¶, Masaharu Noda‖, Keiji Imoto**, Kazumaru Wada††, Akihisa Mitsudome§, Sunao Kaneko††, Mauricio Montal‡‡, Keiichi Nagata†, Shinichi Hirose§,§§, and Kazuhiro Yamakawa*,§§

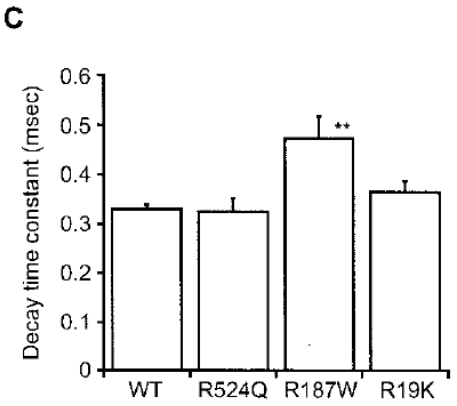
6384–6389 | PNAS | May 22, 2001 | vol. 98 | no. 11







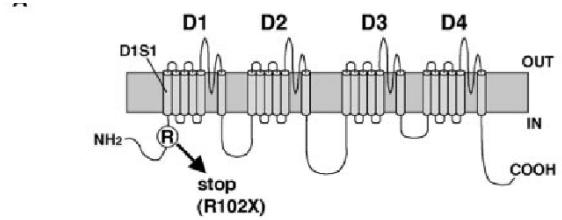


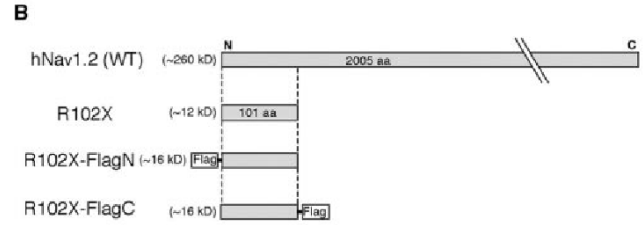


A Nonsense Mutation of the Sodium Channel Gene *SCN2A* in a Patient with Intractable Epilepsy and Mental Decline

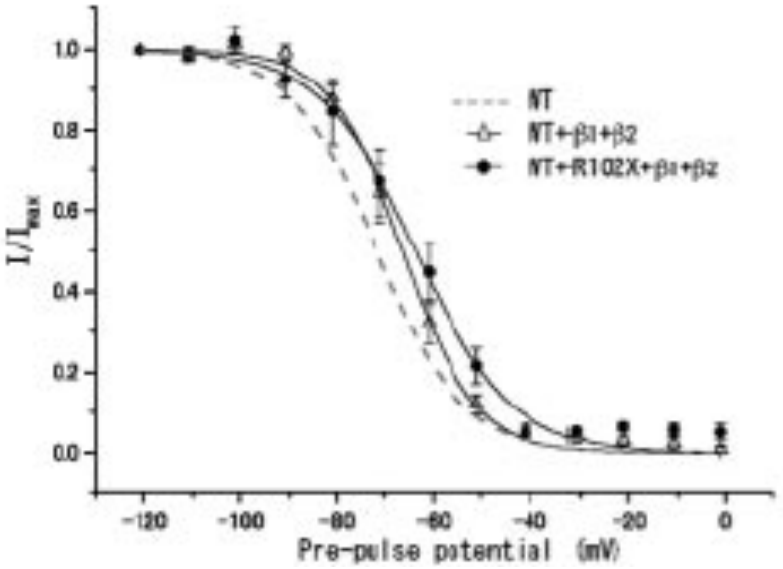
Kazusaku Kamiya,[1] Makoto Kaneda,[2] Takashi Sugawara,[1] Emi Mazaki,[1] Nami Okamura,[1] Mauricio Montal,[3] Naomasa Makita,[4] Masaki Tanaka,[5] Katsuyuki Fukushima,[5] Tateki Fujiwara,[5] Yushi Inoue,[5] and Kazuhiro Yamakawa[1]

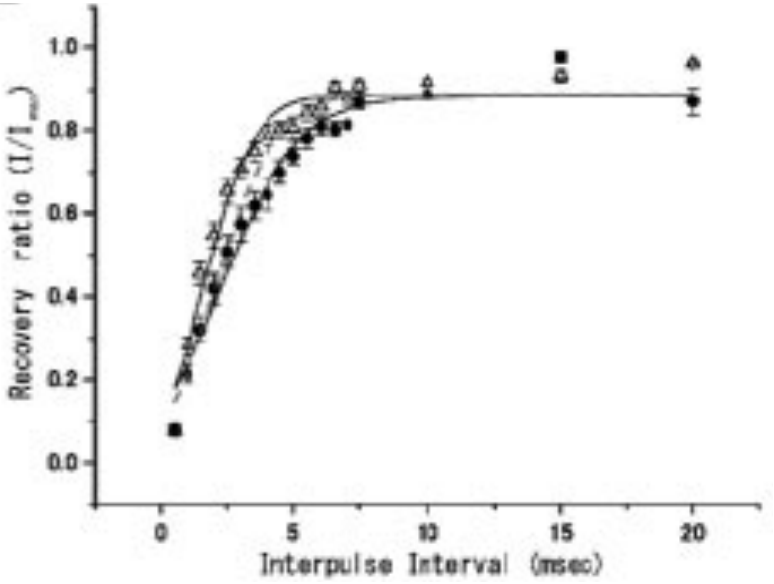
2690 • The Journal of Neuroscience, March 17, 2004 • 24(11):2690–2698





$\alpha + \beta$ subunit





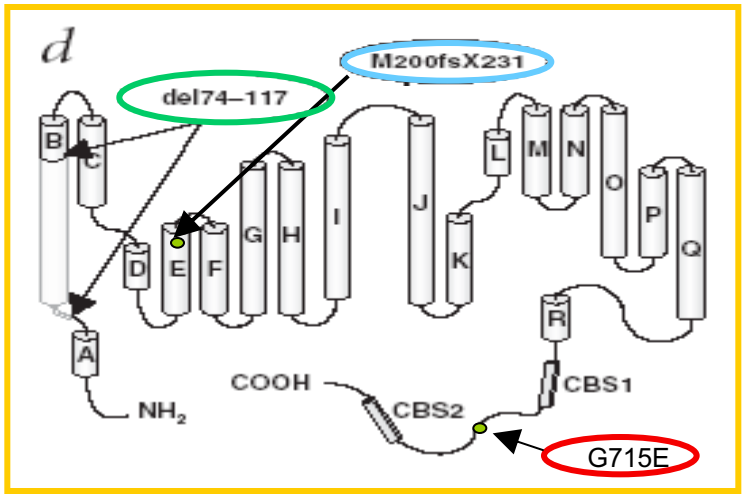
Voltage-Gated Chloride Channels

I. Reactive syndromes
- A. Neonatal seizures due to reversible causes
- B. Benign febrile convulsions
- C. “Low threshold” reactive seizures

II. Idiopathic syndromes
- A. Benign Neonatal convulsions
  - I. Familial
  - 2. Non familial
- B. Partial syndromes
  - 1. Benign childhood epilepsy with centrotemporal spikes
  - 2. Childhood epilepsy with occipital paroxysms
- C. Generalized syndromes
  - 1. Childhood absence epilepsy (CAE)
  - 2. Juvenile absence epilepsy (JAE)
  - 3. Epilepsy with generalized tonic-clonic seizures on awakening (EGMA)
  - 4. Juvenile absence epilepsy (JME)

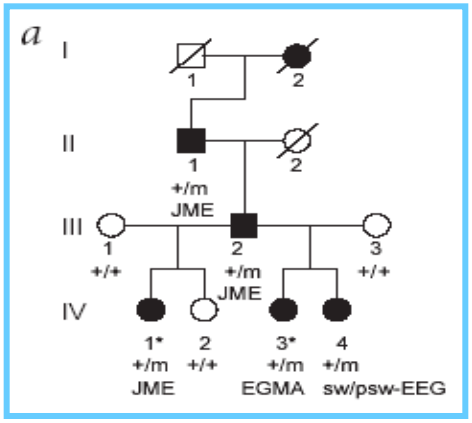
III. Symptomatic syndromes
- A. Neonatal seizures due to irreversible causes
- B. Partial syndromes
  - 1. Epilepsia partialis continua (Kojewnikow’s syndrome)
    - a. Encephalopathic form (Rasmussen’s syndrome)
    - b. Focal form
  - 2. Temporal lobe epilepsy
- C. Generalized syndromes
  - 1. Early myoclonic encephalopathy
  - 2. Infantile spasms
  - 3. Lennaux-Gastaut syndrome

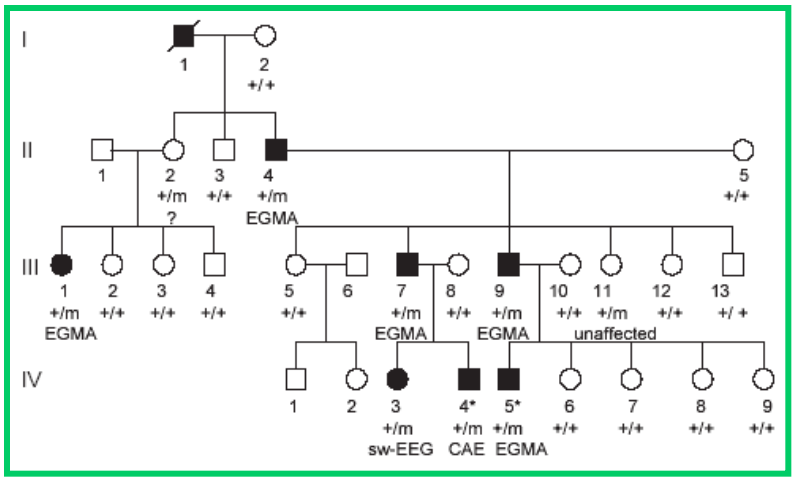
IV. Less well defined syndromes
- A. Severe myoclonic epilepsy of infancy
- B. Benign myoclonic epilepsy of infancy
- C. Epilepsy with myoclonic astatic seizures
- D. Epilepsy with myoclonic absences
- E. Acquired epileptic aphasia (Landau-Kleffner syndrome)
- F. Epilepsy with continuous spikes and waves during sleep (ESES)
- G. Reflex epilepsies

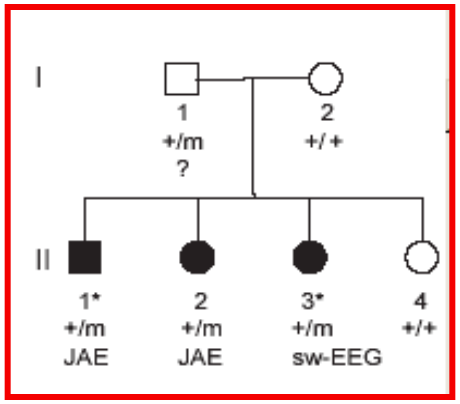


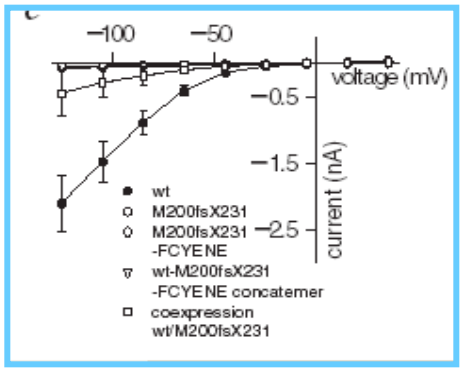
Voltage-gated chloride channel type 2 (CLCN2)

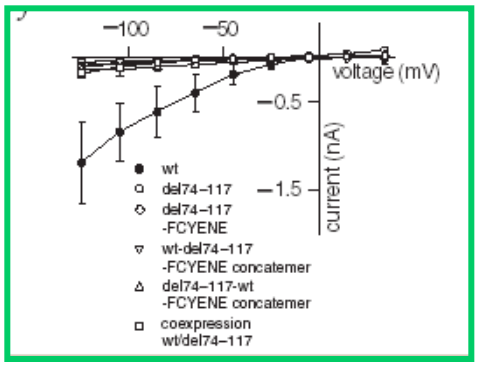
Functional analysis of CLCN2

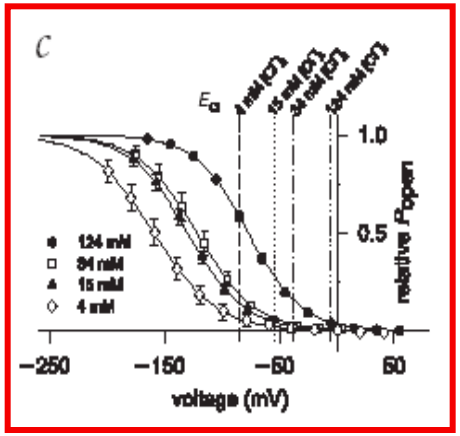


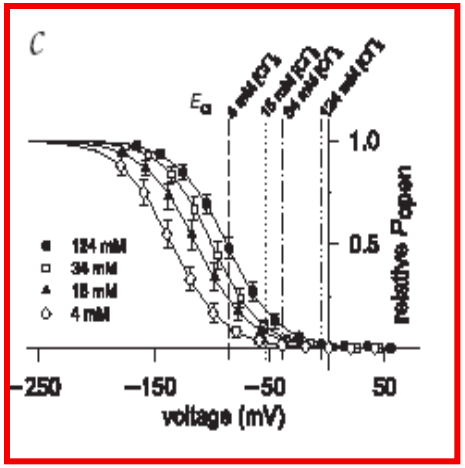










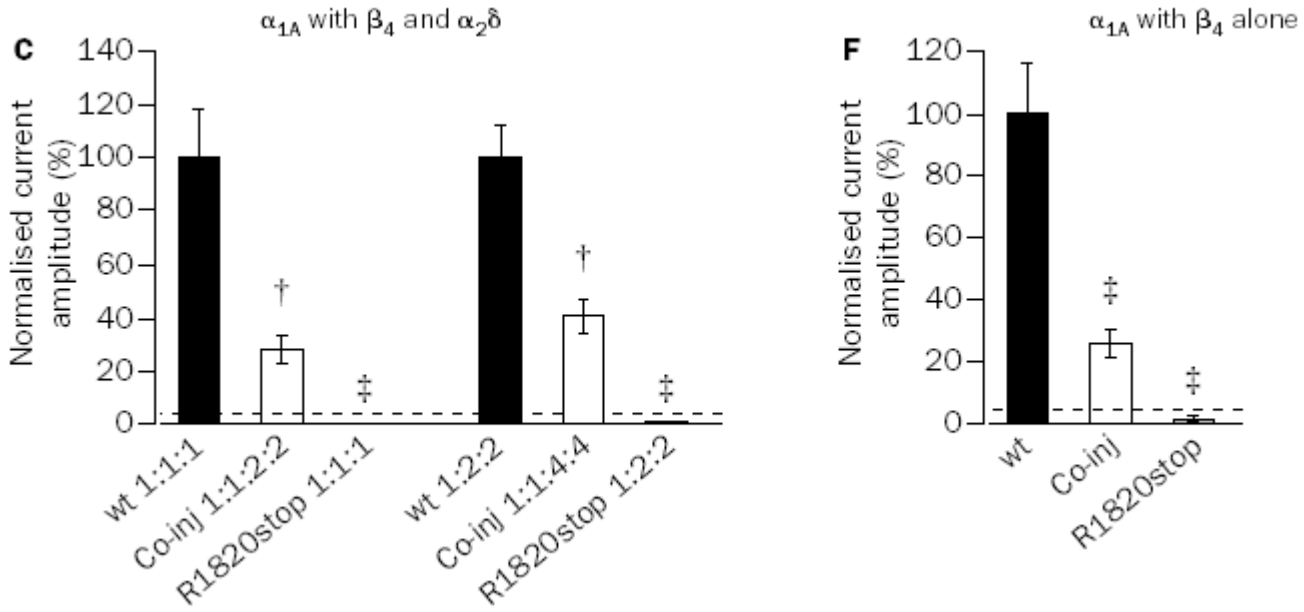


Voltage-Gated Calcium Channels

Human epilepsy associated with dysfunction of the brain P/Q-type calcium channel

Lancet 2001; **358:** 801–07

Anne Jouvenceau, Louise H Eunson, Alexander Spauschus, Venkataswaran Ramesh, Sameer M Zuberi, Dimitri M Kullmann, Michael G Hanna



Am. J. Hum. Genet. 66:1531–1539, 2000

Coding and Noncoding Variation of the Human Calcium-Channel β_4-Subunit Gene *CACNB4* in Patients with Idiopathic Generalized Epilepsy and Episodic Ataxia

Andrew Escayg,[1] Michel De Waard,[2] David D. Lee,[1] Delphine Bichet,[2] Peter Wolf,[3] Thomas Mayer,[3] Janine Johnston,[4] Robert Baloh,[5] Thomas Sander,[6] and Miriam H. Meisler[1]

[1]Department of Human Genetics, University of Michigan, Ann Arbor; [2]INSERM U464, Institut Federatif Jean Roche, Faculté de Medecine Nord, Marseille; [3]Clinic Mara, Epilepsy Center Bethel, Bielefeld, Germany; [4]Department of Neurology, University of Manitoba, Winnipeg; [5]Department of Neurology, University of California School of Medicine, Reed Neurological Research Center, Los Angeles; and [6]Department of Neurology, University Hospital Charite, Campus Virchow Clinic, Humboldt University of Berlin, Berlin

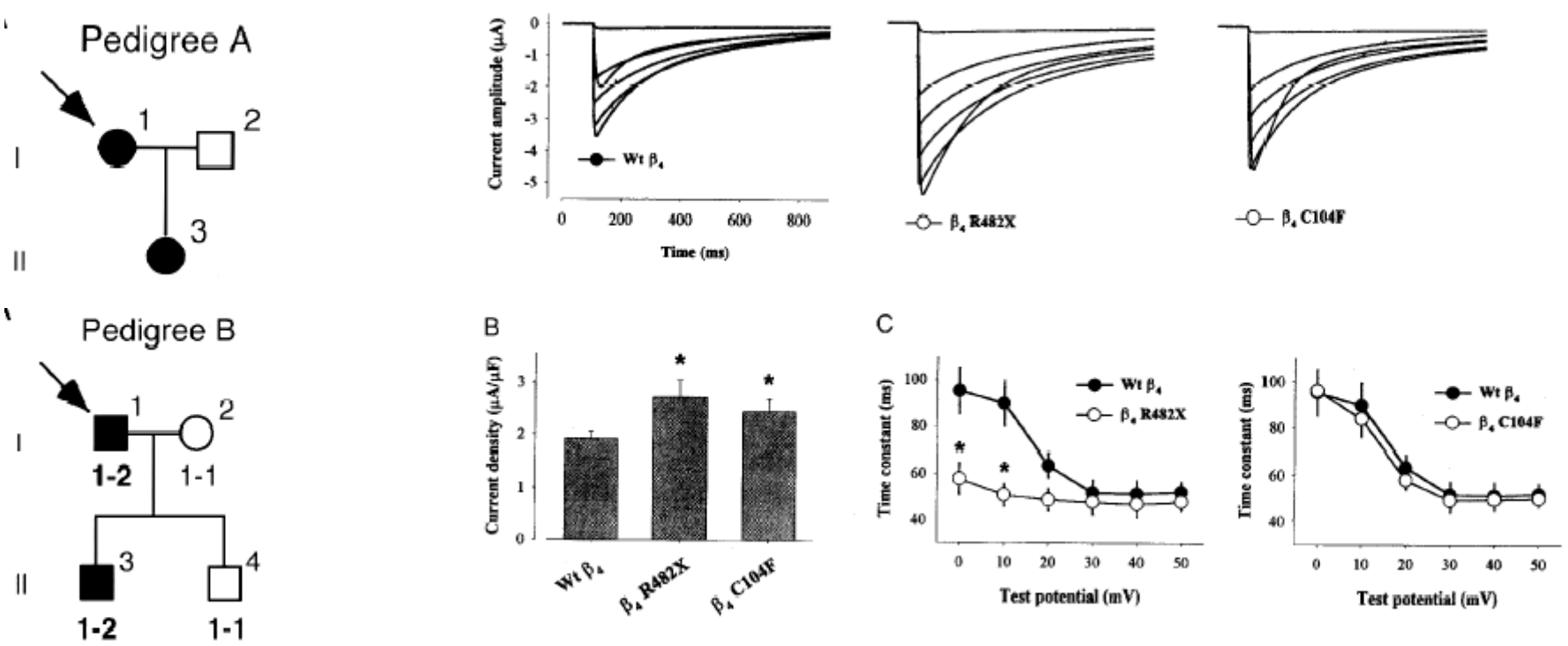


Figure 4 Electrophysiological analysis of mutant β_4 subunits. *Xenopus* oocytes were injected with mRNA from either the wild-type or mutant β_4 subunit, in combination with the *CACNA1A* α_1 subunit. *A*, Sets of current traces at test potentials of −20, −10, 0, 10, 20, and 30

Absence epilepsy and voltage-gated calcium channels

Animal models

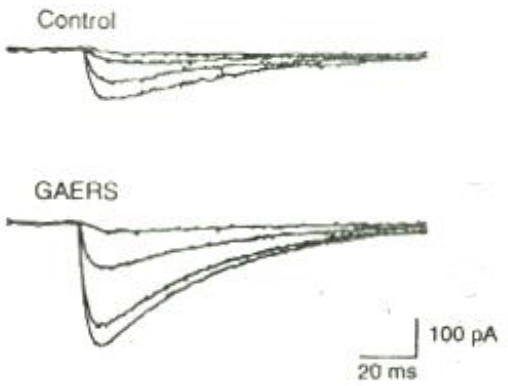
Tottering	Voltage-gated Ca+ a1 subunit *(Cacna1a)*
Lethargic	Voltage-gated Ca+ a1 subunit *(Cacnb4)*
Stargazer	Voltage-gated Ca+ a1 subunit *(Cacng2)*

Functional studies

Selective Increase in T-Type Calcium Conductance of Reticular Thalamic Neurons in a Rat Model of Absence Epilepsy

Evdoxia Tsakiridou,[1] Laura Bertollini,[2] Marco de Curtis,[2] Giuliano Avanzini,[2] and Hans-Christian Pape[1,3]

[1]Abteilung für Neurophysiologie, Medizinische Fakultät, Ruhr-Universität, D-44780 Bochum, Germany, [2]Dipartimento di Neurofisiologia, Istituto Nazionale Neurologico "Carlo Besta," I-20133 Milano, Italy, and [3]Institut für Physiologie, Medizinische Fakultät, Otto-von-Guericke-Universität, D-39120 Magdeburg, Germany



Acetylcholine Receptors

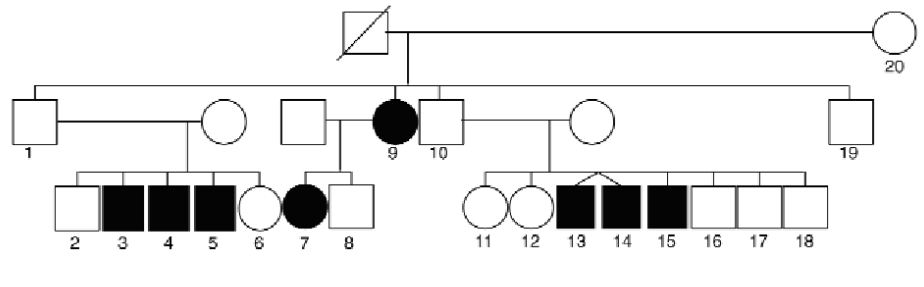
Mendelian idiopathic epilepsies

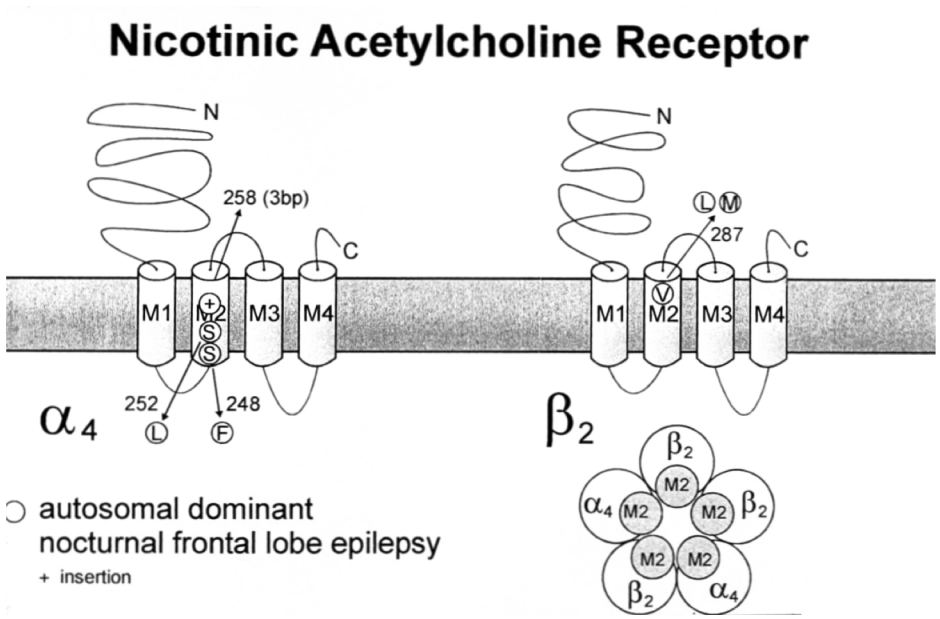
- Age of onset: 6 months - 55 years
- Partial seizures

  Nocturnal motor seizures in clusters

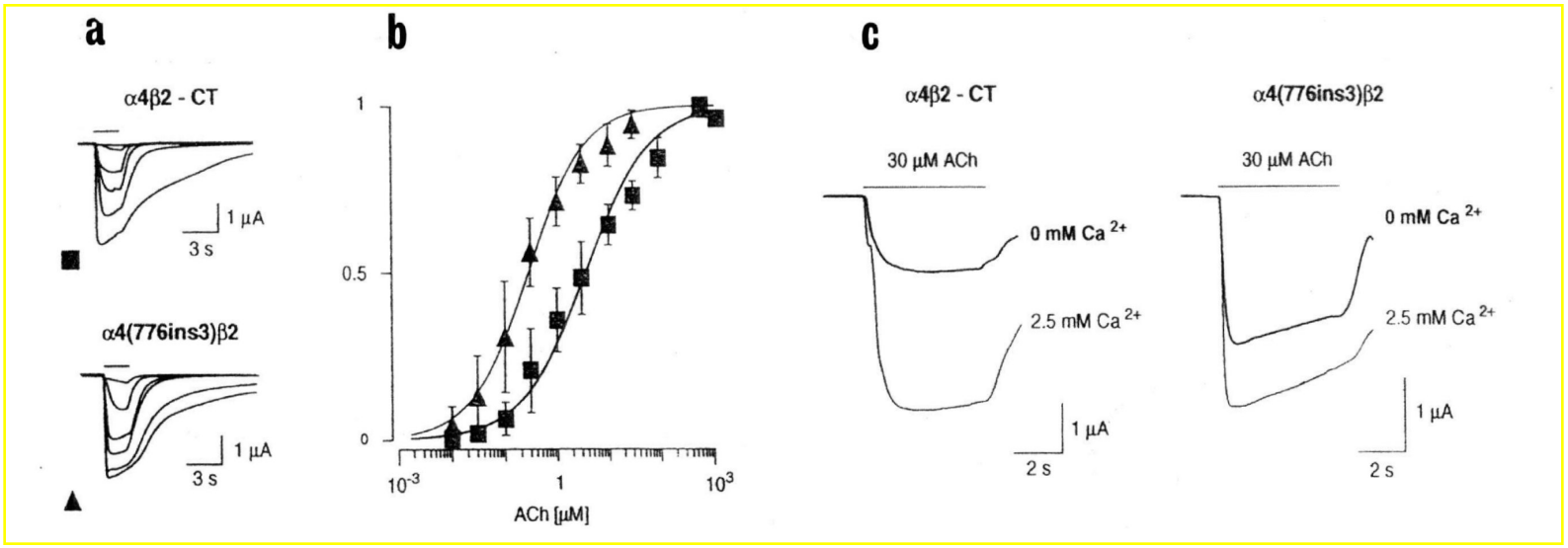
  Aura (daytime)

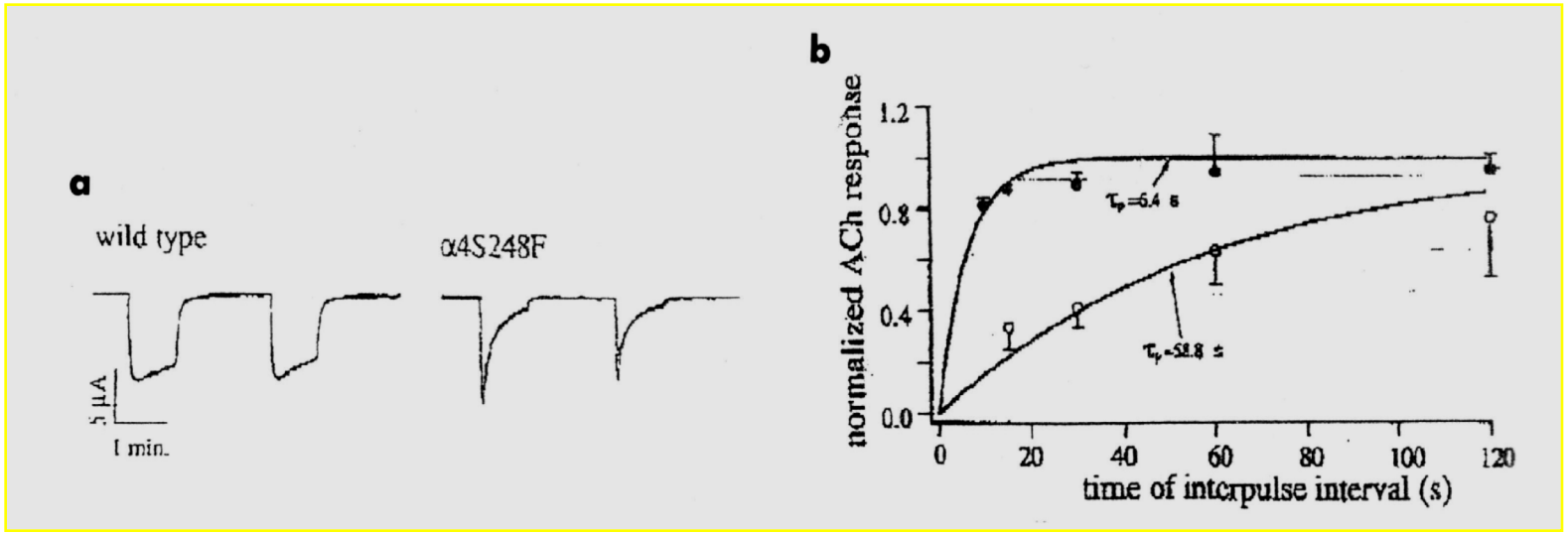
- Normal interictal EEG
- Ictal EEG showing generalized high-voltage slow and sharp activity followed by a fast bi-frontal rhythm.
- Absence of pshycomotor delay or brain lesions





Functional analysis of Neuronal AChRs

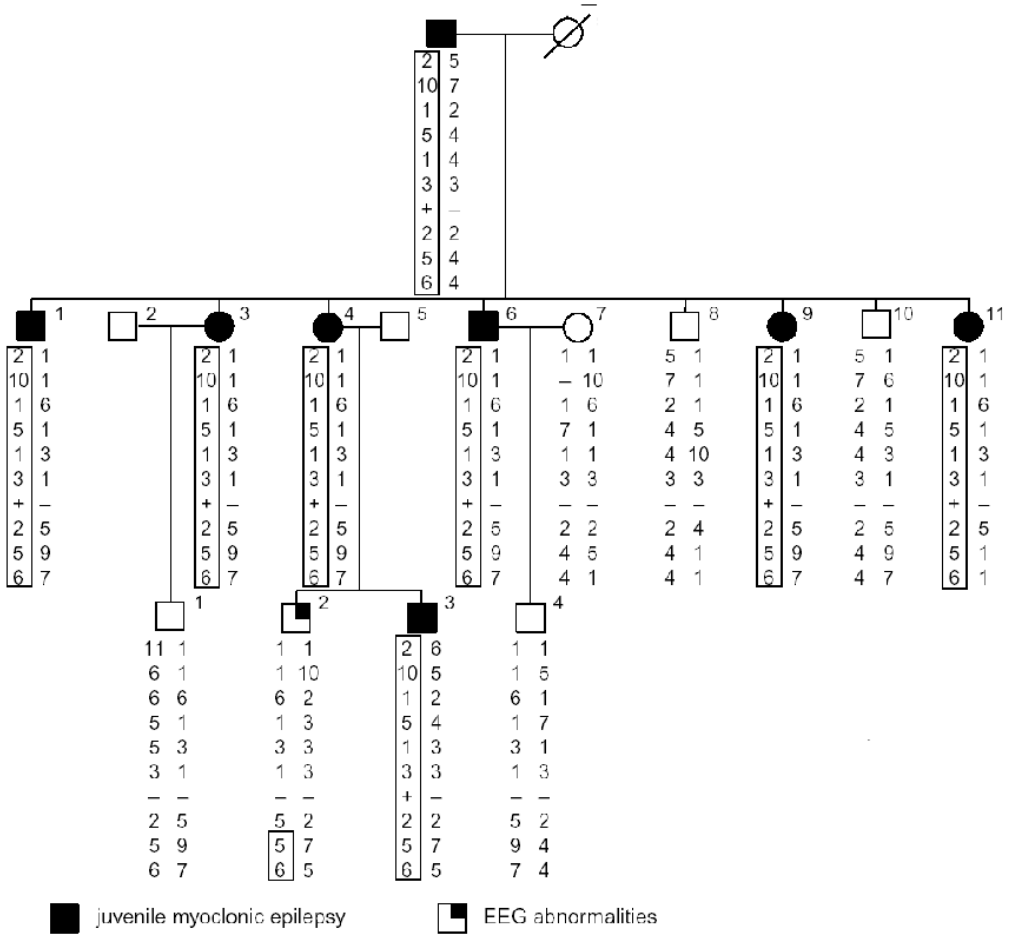


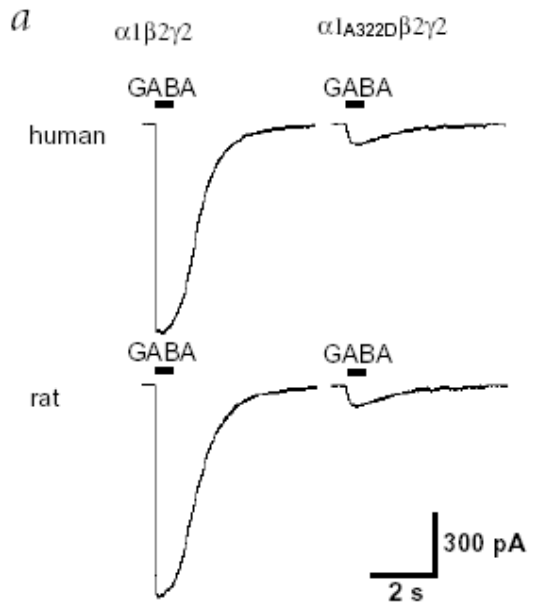


$GABA_A$ Receptors

Mutation of *GABRA1* in an autosomal dominant form of juvenile myoclonic epilepsy

Patrick Cossette[1,5], Lidong Liu[2], Katéri Brisebois[1], Haiheng Dong[3], Anne Lortie[4], Michel Vanasse[4], Jean-Marc Saint-Hilaire[5], Lionel Carmant[4,5], Andrei Verner[6], Wei-Yang Lu[3], Yu Tian Wang[2] & Guy A. Rouleau[1]

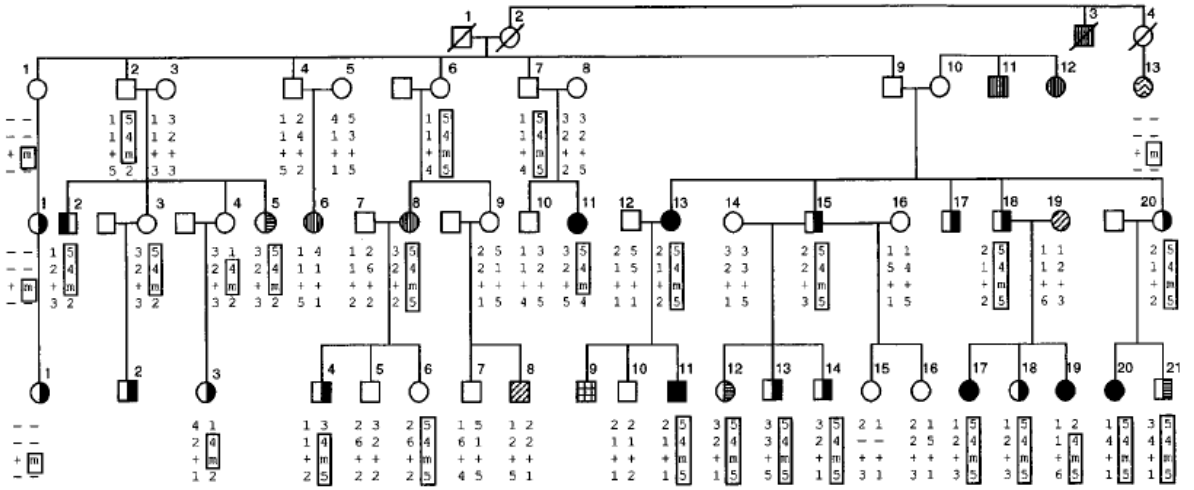




Generalized epilepsy with febrile convulsions plus

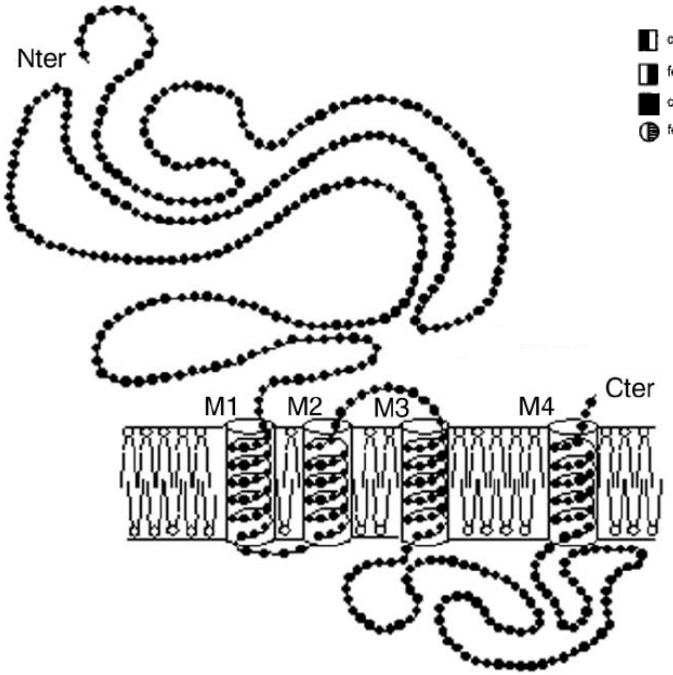
GABRG2

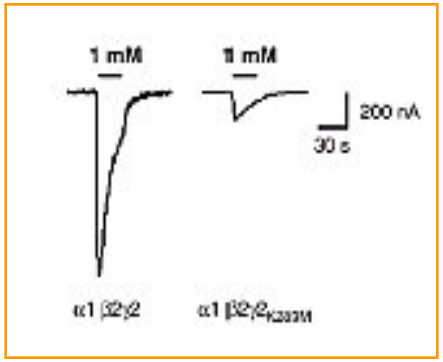
$GABA_A$ receptor γ_2 subunit

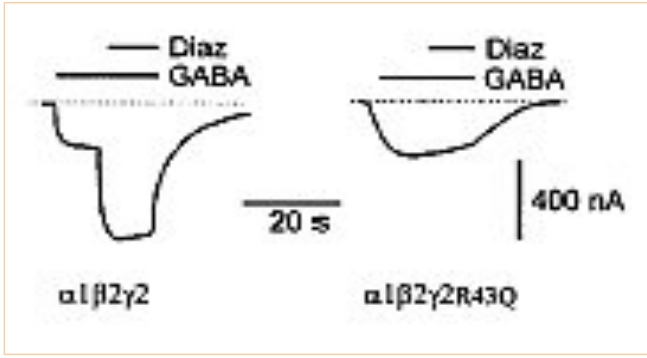


childhood absence epilepsy
febrile seizures
childhood absence epilepsy and febrile seizures
febrile seizures plus
myoclonic-astatic epilepsy
temporal lobe epilepsy
unclassified epilepsy
EEG abnormalities

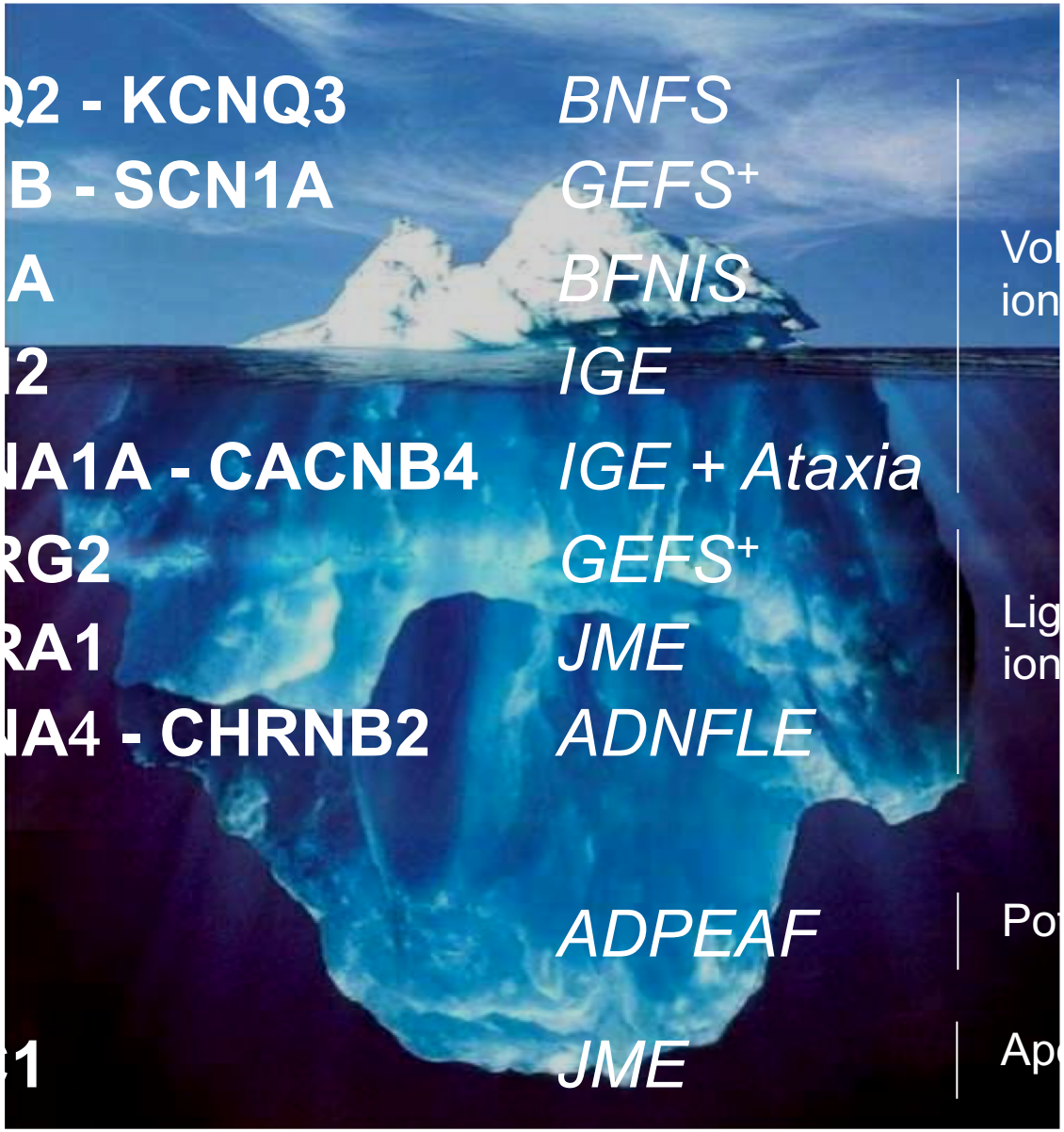
Marker	Zmax	Theta
D5S2112	2.36	0.1
D5S422	6.06	0.0
GABRG2c.245G>A	8.83	0.0
D5S2040	2.51	0.1



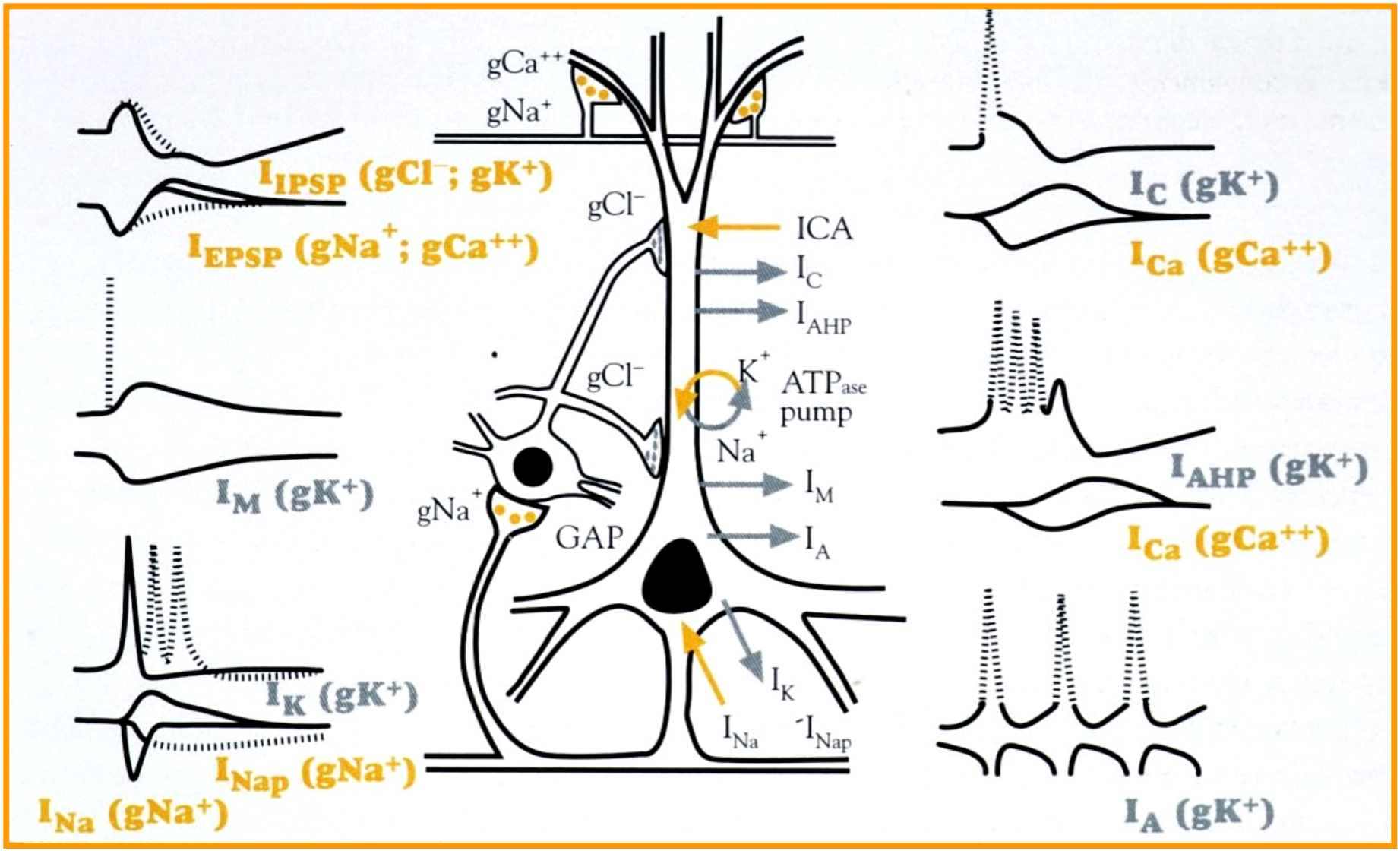


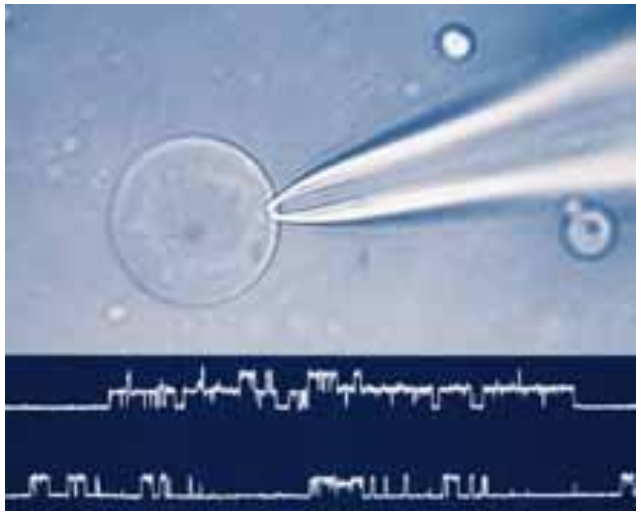


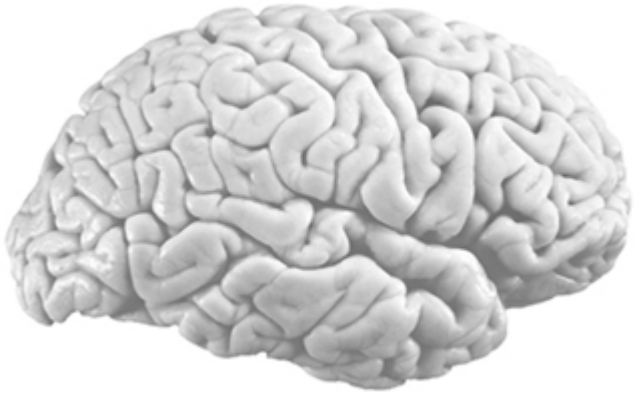
Genes involved in human idiopathic epilepsy

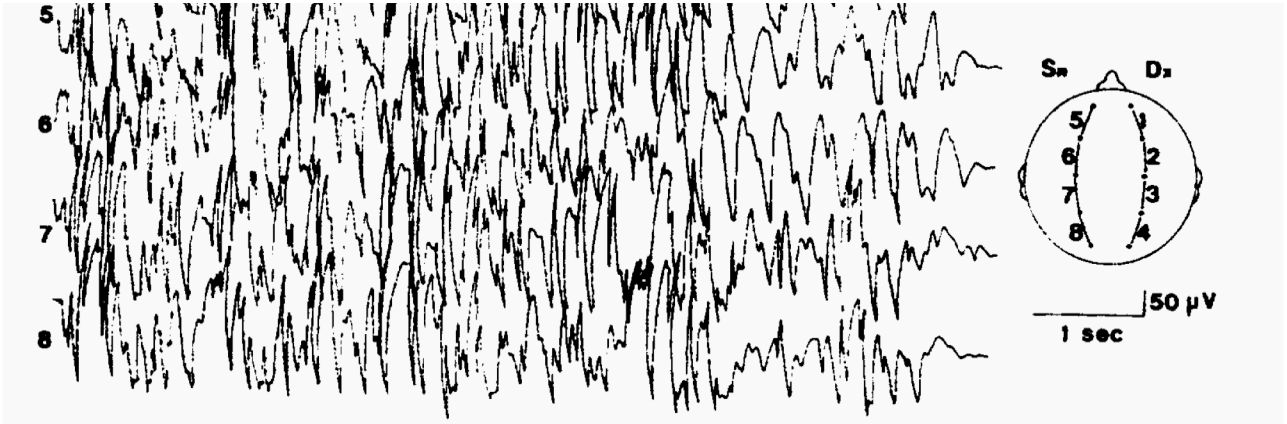


Epileptic Chanelopathies





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