DATING OF THE HOMINID (*Homo neanderthalensis*) REMAINS ACCUMULATION FROM EL SIDRÓN CAVE (PILÓÑA, ASTURIAS, NORTH SPAIN): AN EXAMPLE OF A MULTI-METHODOLOGICAL APPROACH TO THE DATING OF UPPER PLEISTOCENE SITES

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The age of Neanderthal remains and associated sediments from El Sidrón cave has been obtained through different dating methods (14CAMS, U/TH, OSL, ESR and AAR) and samples (charcoal debris, bone, tooth dentine, stalagmitic flowstone, carbonate-rich sediments, sedimentary quartz grains, tooth enamel and land snail shells). Detrital Th contamination rendered Th/U dating analyses of flowstone unreliable. Recent 14C contamination produced spurious age-values from charcoal samples as well as from inadequately pretreated tooth samples. Most consistent 14C dates are grouped into two series: one between 35 and 40 ka and the other between 48 and 49 ka. Most ESR and AAR samples yielded concordant ages, ranging between 39 and 45 ka; OSL dating results permitted adequate bracketing of the sedimentary layer that contained the human remains. Our results emphasize the value of multi-dating approaches for the establishment of reliable chronologies of human remains.

KEYWORDS: SPAIN, NEANDERTHAL, DATING, 14C, U/TH, ESR, OSL, AAR

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INTRODUCTION

The El Sidrón cave site constitutes a paradigm of paleoanthropological findings. In 1994, members of a speleological group discovered the first human remains on the floor of a gallery of the cave. These remains ended up in the neighbouring Guardia Civil (a police corps) presidio which sent the bones to the National Forensic Institution, where the experts immediately realized that they were not of modern humans but much older ones. This initial mistake can be understood in historical terms, because during the Spanish Civil War (1936–1939), the cave was used as a guerilla refuge. In fact, at the entrance of the cave there is a tomb in the shadow of an ancient chestnut tree. Today flowers decorate the grave, which has become the focus of local pilgrimage.

J. L. Prieto from the National Forensic Institute sent a bone sample to the University of Arizona for dating, but not enough collagen was found for AMS dating. The bones were subsequently studied by B. Vandermeersch and M. D. Garralda who ascribed these remains to Homo sapiens with very archaic characteristics (published in local press). However, Rosas and Aguirre (1999) attributed the remains to Homo neanderthalensis, a view which was reconfirmed by others (Egocheaga et al. 2000; Prieto et al. 2001). Since 2000, fieldwork, including archaeological excavation, prospecting and geophysics, has been conducted by J. Fortea and M. de la Rasilla in order to obtain more palaeoanthropological and archaeological material and to discover their autochthonous position (Fortea et al. 2003; Rosas et al. 2006).

The aim of this paper is not only focused on the dating of El Sidrón cave hominid remains, but also on a discussion of the value of results obtained through the use of different dating techniques which were applied on fossil remains and sediment samples.

GEOGRAPHICAL AND GEOLOGICAL SETTINGS

El Sidrón cave (5° 19′ 34″ W; 43° 23′ 07″ N; elevation 167 m) is located in the vicinity of Borines, a small village in Asturias (northern Spain) (Fig. 1). The wider area consists of Jurassic and Cretaceous limestones and Tertiary outcrops. In the north and south, border quartzites, black shales and limestones of Palaeozoic age appear (Gervilla et al. 1978; Martínez 1989). The area was affected by alpine compression generating NNE–SSW and NW–SE fault systems, which favoured the installation of a palaeofluvial drainage network. During the Pleistocene a distensive phase produced an E–W fault system.

El Sidrón cave is developed in Tertiary carbonate conglomerates alternating with fine- to medium-grained sandstones, dipping 20–30° to the north. Dissolution processes preferentially affect conglomerates. The dip of strata and the fault systems explain both the geometry of the karst system and the morphology of individual galleries. The cave axis (Fig. 2) consists of a 600 m long NNW–SSE phreatic tube with an E–W oriented central segment of 200 m length, following the strike of the bedding plane. Faults are visible on the ceiling of the galleries. Less developed galleries orthogonal to the main gallery are also conditioned by the fault system. One of them, the Galería del Osario (GO, Ossuary Gallery), houses the archaeological and palaeontological site. Formerly, the GO was presumably open air, connected through a sink hole, but nowadays this entrance is sealed with collapsed blocks and finer grained sediments.

Sedimentology and stratigraphy

The excavations performed in 2000 started in section 3, squares E, F, G and H-8 (Fig. 2), with the aim of obtaining the widest stratigraphical section possible between the GO walls E and W.
Figure 1  Geographical setting of El Sidrón cave.

Figure 2  Map of the excavated areas in El Sidrón cave (Ossuary Gallery – GO).
During the following years, the excavation works continued by means of 33 cm wide strips towards N and S. Drawings and photographs of every new section obtained were taken, and projection points were determined (as x, y, z) for archaeological and anthropological material, as well as mineral or organic elements, to be dated in each section. This gave something like a stratigraphical tomography with more than 14 sections separated every 33 cm, containing all the relevant information properly documented. There is no room here to include all these sections; Figure 3 is shown as an example. Complementarily, Figure 4 shows the horizontal position of some of the elements that have been dated.

In short, four major sedimentary units have been recognized in the infill of the central part of the GO – sections 2, 3 and 5 (Fig. 2). They can be summarized, from bottom to top, as follows (Fig. 5):

0: Unit of massive mud located at the bottom of the column. No clear sedimentary structures can be distinguished. Provisionally they appear to be sediments deposited through a low energy outflow.
I. Unit of laminated fine sands and mud, with cross-stratification. It includes low-intensity fluvial-karstic material with a relative increase in energy at the top.
II: Unit of poorly sorted gravels, sands and mud. Until now, it has represented the lower limit of the fossiliferous units. The fluvial-karstic materials originated from a high energy event and are clearly erosive in nature.

Figure 3  Stratigraphic section in E-H/8, with the localizations of SID-22 (OSL 1) and SID 23 (OSL 5) samples (the latter respect the coordinates X and Z, but Y is located 105 cm to the south in the stratigraphic section δ, between Units III/IV – see also Figs 4 and 5). Lithic and bone remains are projected in the section.
III: Unit of massive clays with dispersed levels of gravels, sands and silts. Interbedded silts and fine sands showing water-scape structures are common. At the base, this unit is very similar to the previous one and, in general, the grain decreases upward. In the western part of the gallery, the grain size of the unit is also coarser, with a predominance of pebble and gravel deposits. A prominent feature is the existence of calcareous crusts (IIIc) on the top of the unit.

IV: Unit of massive mud with some interbedded sands. These sediments formed in a very low energy fluvial-karstic environment, and correspond to the final infill episode in the gallery which can be regarded as still in progress.

In general, the unit that holds most of the bone and lithic remains (III) consists of a massive deposit (debris flow) composed of a chaotic mixture of gravels, mud and water. Therefore, the arrival and the accumulation of the remains (bones) must have happened immediately after the entry of the previously mentioned gravel deposit (Unit II), and would have been associated with the same high-energy event. The data accumulated so far indicate that they were probably dragged down from a higher level, where very little or no activity had taken place until that time due to the position above the water table as a result of the confinement of the external fluvial network channels.
A mass of loose material situated in that higher cavity/gallery went downstream into the cave in a single occurrence of mud seepage due to a collapse and/or a storm event. The outcome was the arrival of a mass of pebbles, sands, bones and clays that would accumulate over a substratum (rock or previous sediments) and/or remain wedged in the subvertical fissures that are characteristic of the western part of the gallery. The particular layout of the remains and of the unit containing them suggest the possibility that they entered the gallery through one of the shafts located in the vertical section of bore I (squares G/H-9/10) and currently blocked up by a filler of clays (Sánchez-Moral et al. 2007).

Next, an intermittent entry of archaeological and anthropological remains occurred while the sequence upper part was being formed (Unit IV). In addition, displacement towards the main gallery and to the lower phreatic stratum took place in the case of some materials previously deposited in lines D and E (next to the east wall) due, on the one hand, to a fluvial–karstic flow coming from the south of the Ossuary Gallery and, on the other, to a stream sink located in squares E–F/9.

The palaeoanthropological record

The Neanderthal human group displays a set of well-defined derived features, e.g., apomorphies (Wolpoff 1980; Trinkaus 1988; Tattersall and Schwartz 1998; Rosas 2001; Conde 2003;
among others). Thus, the human species represented at El Sidrón cave correspond to *Homo neanderthalensis* (Rosas and Aguirre 1999; Rosas *et al.* 2006, 2008). The El Sidrón teeth are large, with crenulated enamel and accessory cusps. Neanderthal lineage incisor features (Bermúdez de Castro 1993) observed in the sample include shovel-shaping, marked labial convexity, and strongly developed lingual tubercles. On the premolars (Bailey 2002), an asymmetric lingual contour, strong transverse crests and accessory lingual cusps are present (e.g., SD-763). The posterior dentition shows some cases of a noticeable taurodontism (e.g., SD-763). The mandibular body tends to be high and thick, and Neanderthal lineage features include mental foramen below M1, deep pterygoid fossa, and inclined mylohyoid line (Fig. 6).

On the other hand, the anatomy of cranial remains corresponds to the set of features detected in Late Pleistocene Neanderths. The SD-436 frontal shows a marked anterior projection with the development of a supraglabellar fossa, and the supratoral sulcus is well defined; the degree of pneumatization is elevated. The temporal bones (SD-315 and SD-359) are still covered by concretions, but several diagnostic features can be distinguished, including a low projection of the mastoid process, flattened glenoid fossa, and an inclined anterior wall of this fossa. The

occipital SD-1219 is large, with a marked nuchal torus and open sutures connecting with a well-preserved temporal pyramid. A large suprainiac fossa is present. SD-1149 is smaller and partially covered by thin breccia. Right transverse sinuses are observed in both cases (Rosas et al. 2008).

The postcranial skeleton shows clear apomorphic features, in spite of its degree of fragmentation. The postcranial skeleton is principally represented by hand and foot metapodials and phalanges, the latter being the most abundant bones in the assemblage. Size and robustness of the first metacarpal and the enlarged distal tuberosity in the distal hand phalanges are among the diagnostic features (as defined by Musgrave 1973). Also, some Neanderthal dentine and enamel volumetric studies have been carried out by means of high-resolution imaging techniques (microCT) (Olejniczak et al. 2008).

Mitochondrial DNA (mtDNA) sequences have been retrieved so far from two El Sidrón samples (Lalueza-Fox et al. 2005, 2006): a right I2 (SD-441) and a femur fragment (SD-1252). The first sample yielded a 48-bp sequence (between positions 16 230 and 16 278 of the mtDNA reference sequence), and the second yielded an almost complete mtDNA hypervariable region 1 (302-nt sequence, between positions 16 076 and 16 378). Subsequently, nuclear DNA was studied, in particular specific genes FOX P2, related to oral ability (Krause et al. 2007), and MC1R, responsible for skin and hair pigmentation (Lalueza-Fox et al. 2007). During fieldwork, a non-contamination protocol for bone extraction in situ was followed (Fortea et al. 2008). These results have made it possible for El Sidrón to be included in the ‘Neanderthal Genome’ project of the Max Planck Institute, Leipzig (Germany).

The archaeological evidence
A total of 415 lithic artefacts have been recovered, including 13 cores, 352 flakes, 20 blades, 14 bladelets and 50 tools (Bordes 1961; Boëda 1993, 1994) comprising side scrapers, denticulates, a hand axe, and several Levallois points, flakes and blades (Fig. 7). The raw material comes from the immediate cave environment, and it is mostly chert (84%), and quartzite (16%).

The lithic assemblage is largely flake based, with a few laminar products and some Levallois supports. Up until now, 67 artefacts have been refitted, 18 of them belong to the same core; reflecting knapping at the primary location of the assemblage. This industry matches well with the Middle Palaeolithic flake industries that began in Europe in the middle part of the Middle Pleistocene, remaining until the lower half of the Upper Pleistocene (Santamaría et al., in press).

DATING
The dating process consisted of determining the terminus ante quem and post quem of the sedimentary unit containing human remains; obtaining the chronology of organic or inorganic material contextually related to the Neanderthal fossils in the same sedimentary unit; dating the fossils themselves. For these purposes, the methods below have been used:

- optically stimulated luminescence (OSL): analysis of sediments bracketing the layer containing the human remains;
- uranium-series disequilibrium dating on flowstones interbedded with the sediments containing the human remains;
- amino acid racemization (AAR) dating of land snails associated with the human remains;
- radiocarbon dating (14C AMS) of charcoal associated samples;
- electron spin resonance (ESR) dating of a human tooth;
- AAR dating of human teeth;
- 14C AMS dating of human bones and teeth.

For a more accurate and instructive analysis of the obtained age, it is interesting to establish a ‘calendar of events’.

A bone sample was sent in 1996 to the University of Arizona but not enough collagen was found for $^{14}$C AMS dating (SID-0/BS01, see Table 1). This negative result (due to bad collagen preservation or age of the sample) and the chronological amplitude of the *Homo neanderthalensis* suggested using other dating methods, without ruling out $^{14}$C.

Figure 7  *Lithic devices from El Sidrón cave (Photo J. Fortea).*

Table 1  Ages obtained from the samples recovered from the Ossuary Gallery (GO) section; U/Th samples are not included. (AA – Arizona University; GX – Geochron; GIfA – Gif sur Yvette; Beta – Beta Analytic Laboratory; ANU – Australia National University; BSL – Biomolecular Stratigraphy Laboratory; OSL – Gif sur Yvette)

<table>
<thead>
<tr>
<th>Sample number</th>
<th>Invent. No.</th>
<th>Lab. ref.</th>
<th>Dating method</th>
<th>Type</th>
<th>Age BP</th>
</tr>
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<tr>
<td>SID-0/BS01</td>
<td>IAF</td>
<td>AA20283</td>
<td>14C AMS</td>
<td>Hominid bone</td>
<td>Insuf. collagen</td>
</tr>
<tr>
<td>SID-00A</td>
<td>B/G</td>
<td>GifA99167</td>
<td>14C AMS</td>
<td>Hominid bone</td>
<td>48 500 ± 2600</td>
</tr>
<tr>
<td>SID-00B</td>
<td>B/G</td>
<td>GifA99704</td>
<td>14C AMS</td>
<td>Hominid bone</td>
<td>49 200 ± 2500</td>
</tr>
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<td>SID-2A</td>
<td>441</td>
<td>GX-30241</td>
<td>14C AMS</td>
<td>Hominid bone</td>
<td>11 289 ± 79</td>
</tr>
<tr>
<td>SID-3A</td>
<td>531</td>
<td>GX-30242</td>
<td>14C AMS</td>
<td>Hominid molar</td>
<td>10 340 ± 70</td>
</tr>
<tr>
<td>SID-8B</td>
<td>312</td>
<td>Beta-89644</td>
<td>14C AMS</td>
<td>Hominid tooth</td>
<td>34 940 ± 680</td>
</tr>
<tr>
<td>SID-13</td>
<td>566</td>
<td>Beta-189645</td>
<td>14C AMS</td>
<td>Hominid tooth</td>
<td>Insuf. collagen</td>
</tr>
<tr>
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<td>736</td>
<td>Beta-189646</td>
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<td>Insuf. collagen</td>
</tr>
<tr>
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<td>SID-18</td>
<td>50</td>
<td>Beta-192064</td>
<td>14C AMS</td>
<td>Hominid tooth</td>
<td>Insuf. collagen</td>
</tr>
<tr>
<td>SID-19</td>
<td>500</td>
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<td>Hominid tooth</td>
<td>40 840 ± 1200</td>
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<td>SID-20</td>
<td>509a</td>
<td>Beta-192066</td>
<td>14C AMS</td>
<td>Hominid bone</td>
<td>37 300 ± 830</td>
</tr>
<tr>
<td>SID-21</td>
<td>763a</td>
<td>Beta-192067</td>
<td>14C AMS</td>
<td>Hominid tooth</td>
<td>38 240 ± 890</td>
</tr>
<tr>
<td>SID-1</td>
<td>9</td>
<td>GX-28272</td>
<td>14C AMS Quercus charcoal</td>
<td>6070 ± 40</td>
<td></td>
</tr>
<tr>
<td>SID-4</td>
<td>3</td>
<td>GX-30243</td>
<td>14C AMS Quercus charcoal</td>
<td>2230 ± 50</td>
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<td>SID-5</td>
<td>7</td>
<td>GX-30244</td>
<td>14C AMS Quercus charcoal</td>
<td>5990 ± 60</td>
<td></td>
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<tr>
<td>SID-6</td>
<td>12</td>
<td>GX-30245</td>
<td>14C AMS Quercus charcoal</td>
<td>6200 ± 60</td>
<td></td>
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<tr>
<td>SID-2C</td>
<td>441</td>
<td>ANU</td>
<td>ESR</td>
<td>Hominid tooth</td>
<td>EU: 37 000 ± 3000</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>LU: 40 000 ± 3000</td>
</tr>
<tr>
<td>SID-2B, 3B, 8A, 9</td>
<td>441, 531, 312, 313</td>
<td>BSL-4069, 4374, 4068, 4063</td>
<td>AARD</td>
<td>Hominid teeth</td>
<td>44 400 ± 8500 (average)</td>
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<td>311</td>
<td>BSL-4068</td>
<td>AARD</td>
<td>Hominid tooth</td>
<td>Insuf. collagen</td>
</tr>
<tr>
<td>SID-10, 11, 12</td>
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<td>BSL-4179, 4180, 4181</td>
<td>AARD</td>
<td>Gastropod (Cepaea nemoralis)</td>
<td>39 400 ± 7100 (average)</td>
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<td>SID-22</td>
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<td>OSL-1</td>
<td>OSL</td>
<td>Sediment</td>
<td>46 900 ± 5200</td>
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<tr>
<td>SID-23</td>
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<td>OSL-5</td>
<td>OSL</td>
<td>Sediment</td>
<td>30 400 ± 2700</td>
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<tr>
<td>SID-24</td>
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<td>OSL-7</td>
<td>OSL</td>
<td>Sediment</td>
<td>28 000 ± 2500</td>
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</table>
After the commencement of the systematic excavations in 2000 by two of the authors (J.F. and M.R.), a new attempt was made to date the human remains. In 2001, five human teeth (SID-2B, SID-3B, SID-7, SID-8A and SID-9) were dated using the AAR method (AARD) at the Biomolecular Stratigraphy Laboratory of the Madrid School of Mines. The mean age, calculated using a four-point calibrated model, was reasonably old (Fortea et al. 2003), around 32 ± 11 ka. Likewise, three land snail (SID-10, SID-11, SID-12) samples, which were recovered within the horizons containing the human remains, returned ages around 40 ka BP.

In 2001 an enamel sample (SID-2C) was analysed using ESR in the Research School of Earth Sciences of the Australian National University (Canberra, Australia). The obtained age of about 40 ka BP matched the data obtained through the amino acid racemization analysis.

Samples SID-2A, SID-2B and SID-2C (inventory number 441) were taken from the same specimen being $^{14}$C, AARD and ESR dated. Samples SID-3A and SID-3B (inventory number 531) also from the same specimen were $^{14}$C and AARD dated (see Table 1).

In 2002 and 2003, a large set of flowstone samples was collected for Th/U dating. Some samples were picked from a flowstone interbedded in the human remain-bearing section. Other samples were taken for comparison from different flowstone deposits throughout the cave. Drip water was also sampled and analysed. All the samples were strongly contaminated with detrital thorium and almost all the ages obtained were out of the method range.

Human bone (SID-2A) and tooth (SID-3A) samples were sent to the Geochron Laboratories (Cambridge, MA) in 2003, and the results showed that the samples were astonishingly young, around 11 ka BP ‘in uncalibrated radiocarbon years’. Charcoal samples (SID-1, SID-4, SID-5, SID-6) gave younger dating (2–7 ka BP).

Taking into account the AAR and ESR ages obtained in 2003, a set of human bones and teeth (SID-8B, SID-13, SID-14, SID-15, SID-16, SID-17, SID-18, SID-19, SID-20, SID-21) was sent to Beta Analytic Laboratories (Florida, USA) in 2004. The results of around 35 and 40 ka BP were consistent with those obtained earlier with AAR and ESR.

In 2005, a more complete calibration algorithm for dating dentine collagen was calculated at the Biomolecular Stratigraphy Laboratory based on new $^{14}$C, U/Th and ESR dating results from many Iberian localities. The El Sidrón age was re-calculated and showed a better agreement with the aforementioned ages.

In 2006, OSL dating results on sediment samples (SID-22, SID-23 and SID-24) recovered in 2003 and 2005 from the GO section became available, again providing results consistent with the independent age estimates. In spite of many samples being signal-saturated due to the high radionuclide content of the sediments, the OSL age from the stratigraphical Unit II, underlying the bone-bearing Unit III, as well as the top of Unit III were consistently dated at 47 and 30 ka BP, respectively.

In 2006, the $^{14}$C AMS results for the human femur sent by Vandermeersch and Garralda to the LSCE and dated in 1998 (unpublished, now included in this paper) became available. They yielded c. 49 ka BP (SID-00A and SID-00B in Table 1).

**MATERIAL AND METHODS**

**Radiocarbon dating**

The first set of samples (SID-2A, SID-3A, SID-1, SID-4, SID-5, SID-6) was sent to Geochron Laboratories. The charcoal fragments were separated for any foreign matter, including rootlets and treated with 1N HCl to remove carbonates, NaOH to remove humic acids and other organic
contaminants, and followed by a further HCl treatment. Afterwards, the sample was combusted to recover CO₂ for analysis. The human teeth samples were crushed and reacted with 1N HCl to dissolve the mineral fraction. The residue was filtered and washed and later boiled for 8 h in slightly acid distilled water. The broth was filtered through fiberglass. This process allowed the removal of rootlets, humic acids and other contaminants. The recovered collagen was combusted and the CO₂ was recovered for analysis. All samples (charcoal and teeth) were dated using AMS. As complementary information, this laboratory only reported the δ¹³C value, which for the sample SID-2 was −19.9‰ and for SID-3 was −20.6‰ (Table 1).

Later, a wider set of bone and teeth samples was sent to Beta Analytic Laboratories (SID-8B, SID-13, SID-14, SID-15, SID-16, SID-17, SID-18, SID-19, SID-20, and SID-21). The samples were treated with alkali (NaOH) to remove humic acids. Later, diluted HCl was applied until apatite was dissolved. For ¹⁴C AMS analysis, the carbon sample was transformed into graphite, together with standards and blanks, and sent to an AMS facility. Some teeth sent to Beta Analytical were selected according to their collagen content, which was estimated according to their aspartic acid abundance during AAR analysis. From a total of 10 samples (human bone and teeth), only four had enough collagen for ¹⁴C AMS dating (Table 1). As complementary information, this laboratory only reported the δ¹³C/δ¹⁵N ratio, which for SID-19, SID-20 and SID-21 was −18.1‰, −18.1‰ and −18.4‰, respectively.

Two samples (SID-00A and SID-00B) from the same human diaphysis fragment were dated at the LSCE. Elemental analysis (%C, %N, C/N) performed on two subsamples yielded nitrogen and carbon content of 0.69 and 0.47% (for N) and 2.95 and 1.78% (for C), respectively, with C/N ratios of 4.25 and 3.77 which suggest a rather good state of preservation of bone collagen.

The sample preparation was based on the specific reaction between collagen amino acids and ninhydrin (Nelson 1991). Each dating required c. 2 g of cortical bone which was crushed after being sandblasted with carbon-free alumina. After pretreatment with 0.5N HCl to remove the mineral fraction, the carbonate-free sample was treated with ninhydrin to eliminate any ‘free’ amino acids introduced from the archaeological sediment. After the collagen had been hydrolysed with 6N HCl, a second treatment with ninhydrin allowed the extraction of CO₂ from the carboxylic groups of amino acids (Tisnérat-Laborde et al. 2003). The extracted CO₂ was reduced to graphite (Arnold et al. 1987), which was submitted to the ‘Tandetron’ AMS Facility (UMS2004) (Table 1). Bone ‘blank’ specimens were prepared and measured alongside the archaeological samples. The δ¹³C/δ¹⁵N ratios measured during the AMS dating fall in the range of values obtained for bone; no other measurements were done on the mass spectrometer.

Electron spin resonance dating

ESR (and U-analysis) was carried out on an isolated incisor (SID-2C, Table 1). Because of the value of human material, we decided to follow semi-non-destructive analytical procedures (Grün 1995, 2006; Grün et al. 2003). A small piece of tooth enamel was removed along pre-existing breaks. During this process, the enamel separated completely from the dentine and was not further cleaned.

The enamel fragment was mounted in a Bruker ER 218PG1 programmable goniometer and measured at each dose step at 10° angle intervals for 420° (the spectra past 360° were used to check for short-term fading effects). ESR measurements were carried out on a Bruker 106 spectrometer with a 15 kG magnet and a rectangular 4102 ST cavity. The samples were recorded with the measurement parameters routinely applied in the laboratory of the Canberra Research School of Earth Sciences: accumulation of between 1200 (natural sample) and 200 scans (for the
higher dosed samples) with 1.015 Gpp modulation amplitude, 10.24 ms conversion factor, 20.48 ms time constant, 2048 bit spectrum resolution (resulting in a total sweep time of 20.972 s), 120 G sweep width and 2 mW microwave power. The enamel piece was successively irradiated with the following cumulative doses: 0, 11.9, 23.5, 45.2, 68.4, 113, 152 and 197 Gy. The total machine measuring time was 37 days. ESR intensity values were obtained by natural spectrum fitting (Grü n 2002), dose values were obtained by applying a single saturating function with linear conversion, and errors were obtained by Monte Carlo simulation (Grü n and Brumby 1994). Figure 8 shows the angular dose measurements, which yielded an average value of 32.3 ± 1.1 Gy.

The beta dose rate was determined from large sediment samples from Unit III in which the tooth was found. The sediment samples were collected in watertight plastic bags and analysed. The neutron activation results were 3.0 ± 0.2 ppm U, 11.5 ± 0.2 ppm Th and 1.00 ± 0.08% K. The water content was assessed to be 23%. The gamma dose rate was measured in situ with a Digidart portable CsI gamma spectrometer on both sides on the trench where Unit III was exposed, resulting in two measurements of 501 ± 25 and 540 ± 27 mGy/a. Because of the discrepancy between the gamma spectrometric measurements and the NAA results (the latter would result in a dose rate of about 850 mGy/a), the isotopes of the U-decay chain were analysed with a high-resolution Ge detector system (Simpson and Grü n 1998). The following isotope ratios were measured: $^{230}$Th/$^{238}$U = 1.06 ± 0.08, $^{226}$Ra/$^{238}$U = 1.06 ± 0.03, $^{214}$Bi/$^{238}$U = 1.01 ± 0.01, $^{214}$Pb/$^{238}$U = 1.00 ± 0.01 and $^{210}$Pb/$^{238}$U = 0.99 ± 0.02. This means that the U-decay chain is in equilibrium and that the discrepancy between the NAA and in-situ measurements is more likely caused by the fine sedimentary layering.

Uranium in the enamel and dentine was determined with laser ablation (for more details, see Eggins et al. 2003, 2005). The enamel was scanned twice, both scans resulting in closely similar U distributions (Fig. 9). We obtained U concentrations of 0.36 ± 0.05 (enamel) and
5.82 ± 0.31 ppm (dentine) and assumed a $^{234}$U/$^{238}$U ratio of 1.2 ± 0.2 in enamel and dentine as well as 15 ± 7% water in dentine.

**U-series disequilibrium dating of flowstones**

U-series dating of stalagmitic flowstones has been successfully used to establish the chronological framework of many archaeological sites in the Iberian Peninsula, such as Abric Romaní (Bischoff et al. 1994), El Castillo (Rink et al. 1997), L’Arbreda (Bischoff et al. 1994), Banyoles (Julià and Bischoff 1991) and Atapuerca (Bischoff et al. 1997, 2003).

Twenty-two samples were recovered in El Sidrón cave for U/Th dating purposes (Table 2). Twenty-one samples were recovered from calcitic flowstones and one sample consisted of dripping water. The isotopic composition of the samples was determined by alpha spectroscopy.

Ten samples were taken on a single calcarenitic bed correlated with the horizon where the Neanderthal remains appear in the GO section. Eleven samples were recovered from speleothem throughout the cave, with uncertain relation with the Neanderthal remains and underlying –overlying sediments attesting the complex infill history of the cave.

For the principles of the U-series dating technique, the reader is referred to Ivanovich and Harmon (1992). Chemical separation followed Bischoff and Fitzpatrick (1991): the sample was totally dissolved in nitric acid and a radioisotope with known activity was incorporated in order to determine the efficiency of the isotope separation process. The U and Th isotopes were isolated
Table 2  Alpha spectroscopy analyses of U-series isotopes and derived dates from calcarenite hominid bone-bearing bed (A), speleothems not stratigraphically related to the Neanderthal remains bearing section (B) and dripping water (C)

<table>
<thead>
<tr>
<th>Ref.</th>
<th>U ppm</th>
<th>Th ppm</th>
<th>234U/238U</th>
<th>230Th/234U</th>
<th>238U/232Th</th>
<th>234U/232Th</th>
<th>238U/232Th</th>
<th>Nominal date</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>603-T</td>
<td>0.2</td>
<td>0.42</td>
<td>1.12 ± 0.02</td>
<td>0.46 ± 0.01</td>
<td>0.766 ± 0.022</td>
<td>1.652 ± 0.049</td>
<td>1.474 ± 0.045</td>
<td>66746 ± 2916/2843</td>
</tr>
<tr>
<td>1104-L</td>
<td>0.15</td>
<td>1.56</td>
<td>1.02 ± 0.03</td>
<td>2.89 ± 0.08</td>
<td>0.860 ± 0.010</td>
<td>0.298 ± 0.008</td>
<td>0.292 ± 0.008</td>
<td>*</td>
</tr>
<tr>
<td>1604-R</td>
<td>1.45</td>
<td>7.62</td>
<td>0.95 ± 0.06</td>
<td>1.07 ± 0.06</td>
<td>0.598 ± 0.021</td>
<td>0.561 ± 0.029</td>
<td>0.590 ± 0.030</td>
<td>*</td>
</tr>
<tr>
<td>1204-T</td>
<td>0.34</td>
<td>1.88</td>
<td>0.90 ± 0.05</td>
<td>1.51 ± 0.13</td>
<td>0.756 ± 0.030</td>
<td>0.500 ± 0.041</td>
<td>0.554 ± 0.045</td>
<td>*</td>
</tr>
<tr>
<td>61012-T</td>
<td>0.14</td>
<td>0.85</td>
<td>0.96 ± 0.02</td>
<td>1.57 ± 0.03</td>
<td>0.772 ± 0.008</td>
<td>0.491 ± 0.010</td>
<td>0.512 ± 0.010</td>
<td>*</td>
</tr>
<tr>
<td>1304-L</td>
<td>0.16</td>
<td>1.00</td>
<td>1.04 ± 0.02</td>
<td>1.75 ± 0.10</td>
<td>0.882 ± 0.045</td>
<td>0.504 ± 0.029</td>
<td>0.485 ± 0.028</td>
<td>*</td>
</tr>
<tr>
<td>1704-R</td>
<td>3.97</td>
<td>21.22</td>
<td>1.01 ± 0.04</td>
<td>0.96 ± 0.04</td>
<td>0.565 ± 0.015</td>
<td>0.589 ± 0.024</td>
<td>0.581 ± 0.023</td>
<td>*</td>
</tr>
<tr>
<td>1404-L</td>
<td>0.33</td>
<td>3.68</td>
<td>1.01 ± 0.03</td>
<td>2.43 ± 0.08</td>
<td>0.688 ± 0.008</td>
<td>0.283 ± 0.009</td>
<td>0.280 ± 0.009</td>
<td>*</td>
</tr>
<tr>
<td>1504-L</td>
<td>0.24</td>
<td>2.24</td>
<td>1.04 ± 0.04</td>
<td>2.41 ± 0.12</td>
<td>0.821 ± 0.023</td>
<td>0.341 ± 0.016</td>
<td>0.328 ± 0.016</td>
<td>*</td>
</tr>
<tr>
<td>2204-R</td>
<td>1.6</td>
<td>8.15</td>
<td>1.08 ± 0.07</td>
<td>0.97 ± 0.06</td>
<td>0.639 ± 0.025</td>
<td>0.658 ± 0.039</td>
<td>0.609 ± 0.037</td>
<td>*</td>
</tr>
<tr>
<td>B</td>
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<td></td>
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</tr>
<tr>
<td>6201-T</td>
<td>0.11</td>
<td>0.54</td>
<td>0.94 ± 0.02</td>
<td>0.94 ± 0.03</td>
<td>0.54 ± 0.011</td>
<td>0.573 ± 0.016</td>
<td>0.607 ± 0.016</td>
<td>*</td>
</tr>
<tr>
<td>6301-T</td>
<td>0.09</td>
<td>0.46</td>
<td>1.01 ± 0.04</td>
<td>1.19 ± 0.05</td>
<td>0.684 ± 0.007</td>
<td>0.577 ± 0.022</td>
<td>0.573 ± 0.022</td>
<td>*</td>
</tr>
<tr>
<td>6401-T</td>
<td>0.15</td>
<td>0.83</td>
<td>0.97 ± 0.03</td>
<td>1.08 ± 0.04</td>
<td>0.589 ± 0.011</td>
<td>0.546 ± 0.017</td>
<td>0.564 ± 0.017</td>
<td>*</td>
</tr>
<tr>
<td>303-T</td>
<td>0.42</td>
<td>1.66</td>
<td>1 ± 0.02</td>
<td>0.83 ± 0.07</td>
<td>0.645 ± 0.04</td>
<td>0.774 ± 0.062</td>
<td>0.777 ± 0.063</td>
<td>194 899 +/−39 270</td>
</tr>
<tr>
<td>403-T</td>
<td>0.55</td>
<td>2.35</td>
<td>1.02 ± 0.02</td>
<td>0.75 ± 0.02</td>
<td>0.558 ± 0.007</td>
<td>0.741 ± 0.019</td>
<td>0.727 ± 0.019</td>
<td>150 682 +/−8325</td>
</tr>
<tr>
<td>503-L</td>
<td>0.14</td>
<td>0.51</td>
<td>1.11 ± 0.04</td>
<td>0.84 ± 0.03</td>
<td>0.789 ± 0.017</td>
<td>0.943 ± 0.033</td>
<td>0.851 ± 0.030</td>
<td>185 143 +/− 15 071</td>
</tr>
<tr>
<td>703-R</td>
<td>3.19</td>
<td>8.80</td>
<td>0.91 ± 0.02</td>
<td>0.71 ± 0.04</td>
<td>0.726 ± 0.038</td>
<td>1.027 ± 0.053</td>
<td>1.125 ± 0.058</td>
<td>137 647 +/− 14 850</td>
</tr>
<tr>
<td>3002-T</td>
<td>0.09</td>
<td>0.61</td>
<td>0.99 ± 0.04</td>
<td>2.38 ± 0.09</td>
<td>1.073 ± 0.021</td>
<td>0.450 ± 0.016</td>
<td>0.455 ± 0.016</td>
<td>*</td>
</tr>
<tr>
<td>3102-T</td>
<td>0.18</td>
<td>0.77</td>
<td>0.98 ± 0.03</td>
<td>0.98 ± 0.04</td>
<td>0.691 ± 0.017</td>
<td>0.709 ± 0.027</td>
<td>0.720 ± 0.027</td>
<td>*</td>
</tr>
<tr>
<td>203-T</td>
<td>0.14</td>
<td>0.71</td>
<td>0.97 ± 0.04</td>
<td>1.03 ± 0.08</td>
<td>0.617 ± 0.043</td>
<td>0.597 ± 0.041</td>
<td>0.619 ± 0.043</td>
<td>*</td>
</tr>
<tr>
<td>3202-T</td>
<td>3.97</td>
<td>12.41</td>
<td>0.99 ± 0.01</td>
<td>0.45 ± 0.03</td>
<td>0.445 ± 0.010</td>
<td>0.987 ± 0.056</td>
<td>0.993 ± 0.056</td>
<td>65 086 +/− 5305/− 5010</td>
</tr>
<tr>
<td>C</td>
<td></td>
<td></td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3302</td>
<td>1.76 10⁻³</td>
<td>1.22 10⁻³</td>
<td>1.22 ± 0.07</td>
<td>0.26 ± 0.02</td>
<td>1.428 ± 0.115</td>
<td>5.473 ± 0.407</td>
<td>4.495 ± 0.344</td>
<td>32 508 +/− 2515/− 2462</td>
</tr>
</tbody>
</table>

T = total sample dissolution, L = HNO₃, leachable fraction, R = HNO₃, insoluble fraction.

*No date could be obtained due to detrital Th contamination.
by ion-exchange chromatography and later analysed in an alpha spectrometer. For age calculation, the UDATE program from Rosenbauer (1991) was used.

The method’s reliability depends on the chemical purity of the samples. Only six samples provided nominal dates younger than the method limit of about 350 ka (Table 2).

**Optically stimulated luminescence dating**

Nine sediment samples were collected by hammer-driven 20 cm long aluminium tubes in the sections. The exposed surface had been scraped to avoid contamination by grains bleached during the excavation. The current gamma dose rates were measured *in situ* by inserting a Nanoprobe portable spectrometer equipped with a 1.5 inch Na detector into the sampling holes. The recorded dose rates ranged from 562 to 856 μGy/a; the lower dose value is similar to the *in-situ* dose value obtained by Grün (this paper) using a different portable gamma spectrometer (501–540 μGy/a). The observed variation in the dose rates (a factor 1.5) may be due to mineralogical heterogeneity in the sampled beds.

The samples were treated to extract middle-sized quartz grains (80–100 μm) by sieving and acid leaching (hydrochloric, hydrofluoric and hexafluorosilicic acids) to dissolve carbonate and feldspar grains. Hydrogen peroxide was used to eliminate organics.

A large number of aliquots were prepared from each sample, placing small amounts of quartz grains on stainless-steel discs; silicon oil was used as an adhesive. It can be estimated that each aliquot contained about 100 grains. The luminescence measurements were performed using a TL/OSL-DA-15 Risø reader equipped with a ⁹⁰Sr/⁹⁰Y beta source. Optical stimulation was by blue light-emitting diodes (LEDs) with peak emission at 470 nm (Bøtter-Jensen *et al.* 2000). OSL signals were detected with an EMI 9235QA photomultiplier tube through three 3 mm Hoya U-340 filters.

The determination of the ED value of each aliquot followed the single-aliquot regenerative (SAR) dose protocol (Murray and Wintle 2000). The method allows the construction of a sensitivity-corrected regenerative OSL dose–response curve. The ED can then be calculated via interpolation of the corrected natural OSL signal on a saturating exponential curve fitted towards the corrected regenerative dose signals (Fig. 10). A first series of measurements was carried out for the determination of the temperature to which the aliquots have to be preheated for removing unstable signals prior to the luminescence measurements (Fig. 11). This temperature was 250°C for 10 s. Afterwards the SAR protocol was validated on six of the nine samples by performing recovery dose tests. In this second set of experiments, quartz grains were bleached with the LEDs and the aliquots irradiated with the beta dose of 60 Gy. For each sample, the SAR protocol was applied to eight aliquots. The mean measured doses ranged from 59.4 ± 2.4 to 60.6 ± 3.0 Gy, thus validating the measurement conditions.

**Amino acid racemization method**

Three gastropod shells (*Cepaea nemoralis* Linnaeus) as well as five *Homo neanderthalensis* teeth (three incisors, one premolar and one molar) were analysed (Table 1). The gastropod remains came from the same stratigraphical unit where the *Homo neanderthalensis* remains were found. The gastropod shells were sonicated and cleaned with water to remove sediment. Peripheral parts, approximately 20–30%, were eliminated after chemical cleaning of the sample with 2N HCl. The weight of material used for amino acid racemization analysis was 70–90 mg.

The Biomolecular Stratigraphy Laboratory (BSL) uses dentine for amino acid racemization dating of vertebrates. The use of bones is rejected because they are more prone to diagenetic interference (Masters 1986). Dentine collagen samples were obtained by drilling the root of the teeth with a dental diamond drill. A hole 2–3 mm in diameter was drilled near the tooth neck, trying to reach the part of the dentine, which is most protected by the crown. Between 5.5 and 16 mg of dentine were obtained. The outermost part of the root (mostly cementum) was rejected. The samples were pretreated to eliminate foreign and indigenous free amino acids (Marzin 1990; Torres et al. 1999, 2000) through a first acid (HCl) treatment at room temperature and a posterior dialysis step (Spectra/Por mnco 3500 D membrane) to eliminate dissolved mineral fraction and free amino acids. The sample preparation and analysis protocols are described in Goodfriend (1991) and Goodfriend and Meyer (1991) for mollusc-derived proteins.

Figure 10  Sensitivity-corrected regenerative OSL dose–response curve of an aliquot of sample OSL-1. The sensitivity changes which may affect the quartz are corrected in measuring after each regenerative dose (here 20, 50, 87, 132, 0 and 20 Gy); the OSL signal is induced by a constant dose (a test dose preheated at 160°C). The efficiency of this correction is evaluated by measuring the signals induced by a same dose – here 20 Gy – given at the beginning and end of the measurement cycle. The ED was calculated by interpolation of the corrected natural OSL signal onto a saturating exponential curve fitted towards the corrected regenerative dose signals.

Figure 11  EDs of 24 aliquots of sample OSL-1 measured with different preheat temperatures (from 160 to 300°C in steps of 20°). This test allows us to choose a preheat temperature of 250°C for 10 s which was systematically applied to the regenerative doses in all the following measurements.
RESULTS AND DISCUSSION

Radiocarbon dating

The $^{14}$C ages obtained at the Beta Analytic Laboratory (four dates) and the two obtained at the LSCE range from c. 35 000 to 49 000 years BP; the results obtained in each laboratory are internally coherent. However, the Beta Analytic Laboratory ages are younger than those of the LSCE. Such a wide age range can be explained by assuming that the site saw several occupation periods between 35 000 and 50 000 years BP, but the sedimentary infill and the bone and lithic refittings are at odds with this explanation. Another possibility is that the site was only occupied during part of this period, but some of the $^{14}$C ages are possibly underestimated due to contamination by modern carbon.

One has to keep in mind that for the time period in question even a small amount of contamination by modern carbon has an important impact on the results. For instance, the presence of only 0.5% of modern carbon in a 50 000-year-old sample results in an age underestimate of c. 9000 years (Delibrias 1985; Bronk-Ramsey et al. 2004). Compared to the Beta Analytic Laboratory protocol which uses the whole collagen for the dating, the ninhydrin protocol is more selective as it allows extraction of the CO$_2$ from the carboxylic groups of amino acids. This protocol is more effective in eliminating carbonaceous contamination and isolating the intrinsic carbon of the bone. Hence, the oldest ages are probably more reliable so that the Sidrón human remains can be considered to be older than 45 000 years BP.

In contrast, the two c. 11 000 years $^{14}$C yr BP dates (SID-2A, SID-3A) obtained at the Geochron Lab. on the hominid bone (SID-2A) and molar (SID-3A) are much younger than the ESR and AARD results (between 37 000 and 44 000 years) obtained on the same samples (Table 1). It is also obvious that the dated Quercus charcoals (SID-1, SID-4, SID-5 and SID-6) between 2000 and 6000 years are much younger than the Palaeolithic occupation. Consequently one cannot exclude the possibility that the samples dated at the Geochron Lab. were severely contaminated by modern carbon and that the radiocarbon ages are underestimated. One can also suggest that the Quercus trees grew during the Holocene period and that they were introduced late into the Palaeolithic layer.

ESR and U-analysis

For age calculation, we used an updated version of the ESR-DATA program which uses the Monte Carlo beta attenuation values of Marsh (1999) and an alpha efficiency of 0.13 ± 0.02 (Grün and Katzenberger-Apel 1994). The resulting ESR age estimates are 37 ± 3 ka for early uranium uptake and 40 ± 3 ka for linear U-uptake (for more details on U-migration and modeling, see Grün and McDermott 1994). At present, we are not able to further constrain the U-uptake history, therefore the best age estimate for the tooth lies between 34 and 43 ka.

Uranium/thorium from flowstones

The results of radiometric age calculation appear in Table 2. It is noticeable that there is a very high $^{230}$Th contamination. In all samples the value of the $^{230}$Th/$^{232}$Th ratio is above the threshold value (20) established by many authors (Schwarz 1980; Ford and Schwarz 1981; Julià and Bischoff 1991; Bischoff et al. 1994; Horvatincic et al. 2000; Auler and Smart 2001). This extremely high Th contamination, the dripping water has a nominal age older than 30 ka, makes
it unrealistic to try to use the isochron method proposed by Bischoff and Fitzpatrick (1991) and Ku and Liang (1984) to remove discrete $^{232}$Th contamination. In any case, the radionuclide concentrations along the same bed were also extremely variable, confirming the unreliability of these methods for these materials.

The uranium-series disequilibrium method did not work adequately in El Sidrón cave sediment samples.

**Optically stimulated luminescence dating**

Because the OSL was saturated in most of the aliquots of six samples, it was not possible to calculate their EDs. This probably indicated that the quartz grains were scarcely exposed to sunlight in the open air environment before their transportation into the cave; in any case, any reworking process inside the ossuary gallery took place in total darkness. For the remaining aliquots, the scattering of the EDs was very large (see histogram of Fig. 12). Only three sediment samples (SID-22, SID-23 and SID-24) produced good results – see an example in Figure 13. For these three samples, the mean ED was estimated by considering the highest probability of each histogram, i.e., the highest peak was interpreted to be the main component of the sediment.

The obtained values for samples SID-22, SID-23 and SID-24 were $53.5 \pm 3.6$, $43.3 \pm 0.6$ and $51.0 \pm 0.8$ Gy, respectively (Table 3).

The OSL ages were $46.9 \pm 5.2$ ka, $30.4 \pm 2.7$ ka and $28.0 \pm 2.5$ ka for samples SID-22, SID-23 and SID-24 (Table 3). Samples SID-22 and SID-23 were located in archaeological layers IVB and top IIIA, respectively (sedimentary units I and top III; see above: Sedimentology and stratigraphy), thus bracketing the human-bearing beds. Sample SID-24 was recovered from a higher stratigraphical layer (Figs 3 and 5).

**Amino acid racemization**

The amino acid racemization/epimerization ratios obtained in the gastropod and dentine collagen samples are detailed in Tables 4 and 5. For the dentine collagen, the age calculation was made...
from aspartic acid (Asp) racemization ratios because of its higher racemization rate. The aspartic acid d/l ratios of *Homo neanderthalensis* dentine collagen samples were introduced in the age calculation algorithm established for dentine collagen samples of bears (*Ursus deningeri* and *Ursus spelaeus*) from the Iberian Peninsula by Torres *et al.* (2001, 2002). The age calculation algorithms employed for *Cepaea nemoralis* samples are those established by Torres *et al.* (1997).

It is noteworthy that the d/l Asp values reported here differ from a single value (0.13) published in Lalueza-Fox *et al.* (2005), which did not significantly differ from the ‘magic threshold’ (0.10) for DNA preservation (Poinar *et al.* 1996). We think that this value was obtained from the total amino acid content, where the contribution of free amino acid fraction to the Asp racemization ratio may bias the final result due to its higher racemization rate (e.g., Ohtani and Yamamoto 1991).

For the age-calculation algorithm, we used the racemization values from eight cave localities dated through different radiometrical methods. In any case we have calculated the mean age from the individual values obtained from each sample and the age uncertainty is the standard deviation. The mean age for the dentine collagen samples from the Neanderthal teeth was 44.4±8.5 ka BP (Fig. 14).
In the case of gastropod samples we used the calculated ages from the racemization ratios of seven different amino acids but the final age was calculated as the mean value of the ages obtained from each amino acid from the three samples. For gastropods the average age was $39.4 \pm 7.1$ ka BP (Fig. 15).

**CONCLUSIONS**

Most of the dates based on radiometric methods applied to samples of different nature which come from the Palaeolithic levels (bones or teeth dated by C-14 BP uncalibrated, ESR and AARD) or on the geological context (quartz dated by OSL) fall between 38 000 and 50 000 years BP, showing that the Sidrón human remains might be contemporaneous with the last millennia of the Middle Palaeolithic. Actually the most reliable C-14 results are those obtained at the LSCE because the bone preparation used in this laboratory is more effective in eliminating modern contamination.

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**Table 4** Amino acid racemization ratios obtained in Cepaea nemoralis shells from El Sidrón cave. The abundance of each amino acid ($D + L$ forms) is also presented (peak areas are corrected for 100 mg sample weight and 4 $\mu l$ injected volume).

<table>
<thead>
<tr>
<th>Sample</th>
<th>BSL-4179</th>
<th>BSL-4180</th>
<th>BSL-4181</th>
</tr>
</thead>
<tbody>
<tr>
<td>Weight (mg)</td>
<td>80</td>
<td>91.5</td>
<td>67</td>
</tr>
<tr>
<td>Alanine</td>
<td>6871</td>
<td>9157</td>
<td>3613</td>
</tr>
<tr>
<td>d/l Ala</td>
<td>0.280</td>
<td>0.275</td>
<td>0.360</td>
</tr>
<tr>
<td>Valine</td>
<td>12.428</td>
<td>–</td>
<td>5966</td>
</tr>
<tr>
<td>d/l Val</td>
<td>0.351</td>
<td>–</td>
<td>0.285</td>
</tr>
<tr>
<td>Leucine</td>
<td>1272</td>
<td>1650</td>
<td>729</td>
</tr>
<tr>
<td>d/l Leu</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Aspartic acid</td>
<td>10.285</td>
<td>14.738</td>
<td>4876</td>
</tr>
<tr>
<td>d/l Asp</td>
<td>0.382</td>
<td>0.340</td>
<td>0.404</td>
</tr>
<tr>
<td>Glutamic acid</td>
<td>12.008</td>
<td>16.433</td>
<td>3023</td>
</tr>
<tr>
<td>d/l Glu</td>
<td>0.175</td>
<td>0.148</td>
<td>0.170</td>
</tr>
<tr>
<td>Phenylalanine</td>
<td>2844</td>
<td>3683</td>
<td>1275</td>
</tr>
<tr>
<td>d/l Phe</td>
<td>0.282</td>
<td>0.234</td>
<td>0.271</td>
</tr>
<tr>
<td>Isoleucine</td>
<td>3068</td>
<td>3794</td>
<td>1115</td>
</tr>
<tr>
<td>d/-aIle/l-Ile</td>
<td>0.183</td>
<td>0.153</td>
<td>0.158</td>
</tr>
</tbody>
</table>

**Table 5** Aspartic acid ratios obtained in Homo neanderthalensis (Fuhlrott and Schaaffhausen) dentine collagen samples from El Sidrón cave. The abundance of the aspartic acid ($D + L$ forms) is also presented (peak areas are corrected for 100 mg sample weight and 4 $\mu l$ injected volume). The results for other amino acids are not used for age calculation because $D$-forms were not detected in the analysis.

<table>
<thead>
<tr>
<th>Sample</th>
<th>BSL-4063</th>
<th>BSL-4064</th>
<th>BSL-4068</th>
<th>BSL-4069</th>
<th>BSL-4374</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tooth</td>
<td>I¹</td>
<td>P⁴</td>
<td>I¹</td>
<td>I²</td>
<td>M</td>
</tr>
<tr>
<td>Weight (mg)</td>
<td>12</td>
<td>16</td>
<td>8.5</td>
<td>5.5</td>
<td>14</td>
</tr>
<tr>
<td>Aspartic acid</td>
<td>1 393 917</td>
<td>27 337 438</td>
<td>–</td>
<td>89 933 455</td>
<td>463 814</td>
</tr>
<tr>
<td>d/l Asp</td>
<td>0.062</td>
<td>0.084</td>
<td>–</td>
<td>0.057</td>
<td>0.067</td>
</tr>
</tbody>
</table>
carbon contamination present in the sample which would otherwise yield an age underestimate. The uncertainty ranges of the other methods used are large enough for the results to be compatible with the oldest radiocarbon dates. In any case, some fossil bone samples are to be dated soon with the ultrafiltration method (Brown et al. 1988; Higham et al. 2006), which has also proved to be adequate for dating old and contaminated bone remains.

On the right-hand side of Figure 5, there is a synthetic section of the GO stratigraphy with the different age values obtained by the different dating methods. This allows us to confidently date the *Homo neanderthalensis* remains from El Sidrón cave more than a few millennia before 40 ka,
during the OIS 3, before *Homo sapiens* arrived in Europe. These results place the Sidrón archaeological record in the interval between the GIS 10 and 14, at the latest phase of the Middle Palaeolithic, as at some other Cantabrian sites, such as La Viña (Asturias, Spain), El Mirón, El Castillo (Cantabria, Spain), Arrilior, Axlor or Kurtzaleza (Basque Country, Spain) (Hoyos et al. 1999; Fortea 2001; Jöris et al. 2003; Straus and González Morales 2003; Bernaldo de Quirós and Stuart 2006).

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**REFERENCES**


