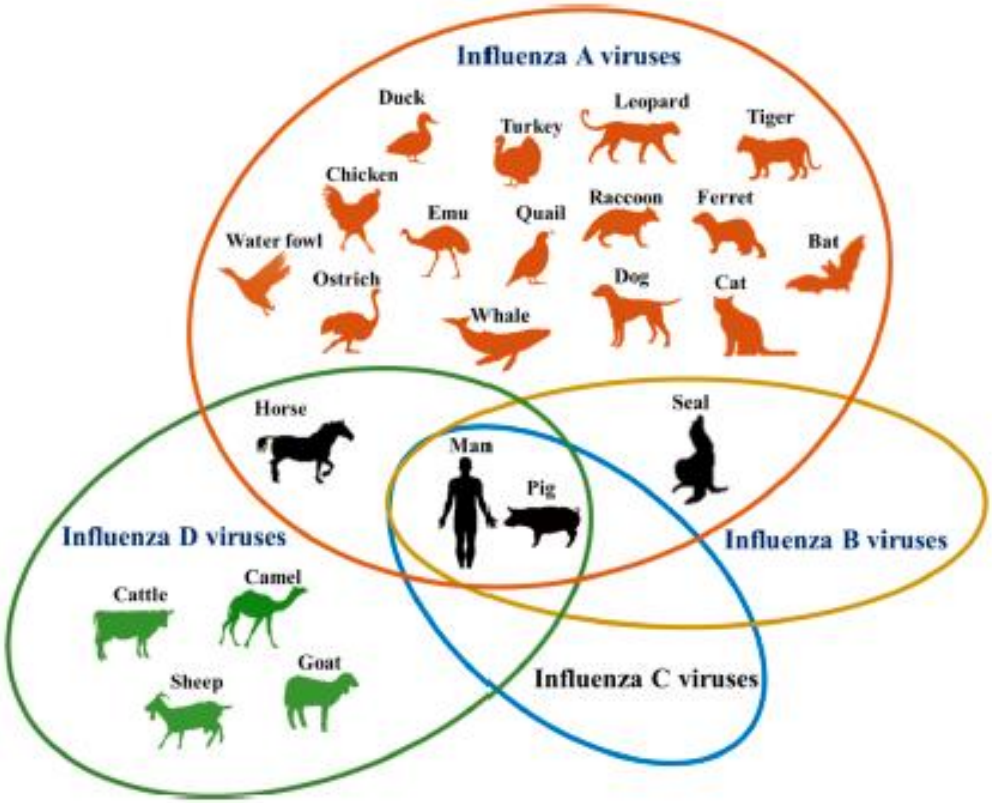
Orthomyxoviridae

Table 1. Revised classification of *Orthomyxoviridae* (ICTV 2017).

Genus	Species	Genomic Segments
Alphainfluenzavirus	Influenza A virus	8
Betainfluenzavirus	Influenza B virus	8
Deltainfluenzavirus	Influenza D virus	7
Gammainfluenzavirus	Influenza C virus	7
Isavirus	Salmon isavirus	8
Quaranjavirus	Johnston Atoll quaranjavirus	6
	Quaranfil quaranjavirus	
Thogotovirus	Dhori thogotovirus	6
	Thogoto thogotovirus	

ssRNA-

Host range of influenza viruses by species



Common hosts of more than one species are encompassed in overlapping ovals. Of the numerous hosts which support influenza virus infection, only four (horse, seal, man and pig) are known to be susceptible to more than one species.

Vet. Sci.2018,5, 71; doi:10.3390/vetsci5030071 www.mdpi.com/journal/vetsci

Influenza virus host range

•**Influenza A viruses** infect a wide variety of mammals **and** birds. IAV is the main human pathogen, associated with epidemics and pandemics. Many different subtypes exist according to the type of haemagglutinin (HA) and neuraminidase (NA) expressed. Pigs and birds are believed to be particularly important reservoirs, generating pools of genetically/antigenically diverse viruses, which can be transferred back to the human population via close contact between humans and animals.

•**Influenza B viruses** infect mammals only and cause disease. Unlike IAV, influenza B viruses do not have distinguishable subtypes.

•**Influenza C viruses** infect mammals only, but rarely cause disease.

•**Influenza D viruses** The virus was first isolated as an influenza C-like virus from pigs with respiratory illness in Oklahoma, USA, in 2011. The virus was subsequently classified as influenza D virus (IDV). While the precise role of IDVs in clinical disease in animals is not yet fully investigated, their role in causing respiratory infections in cattle has been implied.

Influenza virus Transmission

- AEROSOL
  - 100,000 - 1,000,000 VIRIONS/DROPPLET
- INCUBATION 18-72 HOURS

RECOVERY

- INTERFERON – SIDE EFFECTS INCLUDE:
  - FEVER, MYALGIA, MALAISE, FATIGUE
- CELL-MEDIATED IMMUNE RESPONSE
- TISSUE REPAIR:
  - CAN TAKE LONGER

Genome architecture of RNA viruses

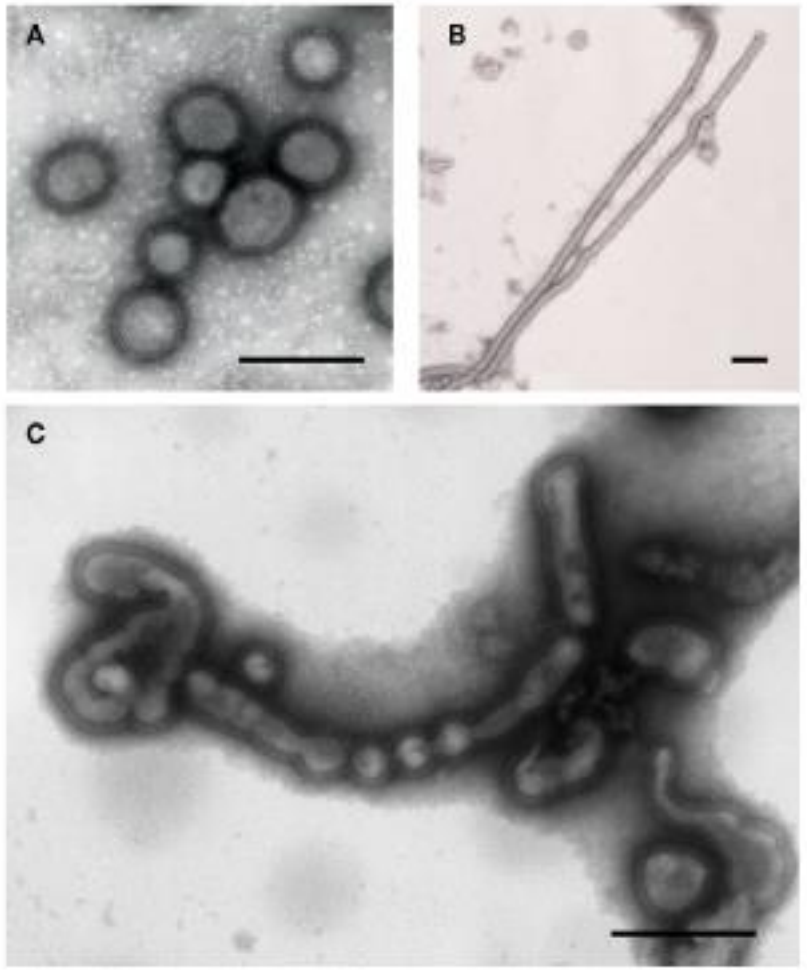
Viruses with negative-sense RNA genomes: that is, genomes with polarity opposite to that of messenger RNA

Some families have a **monopartite genome,** such as those belonging to the order **Mononegavirales**

others have a **segmented genome: Orthomyxoviridae with 8 or 7 segments,** Arenaviridae with 2 segments, and Bunyaviridae with 3 segments.

Morphology of influenza virions

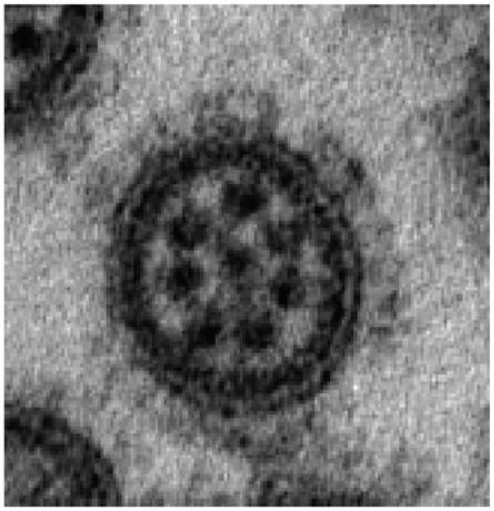
virions show spherical or filamentous shapes of about 100nm in diameter and occasionally irregular morphology, which exemplifies the pleomorphic nature of these virions.

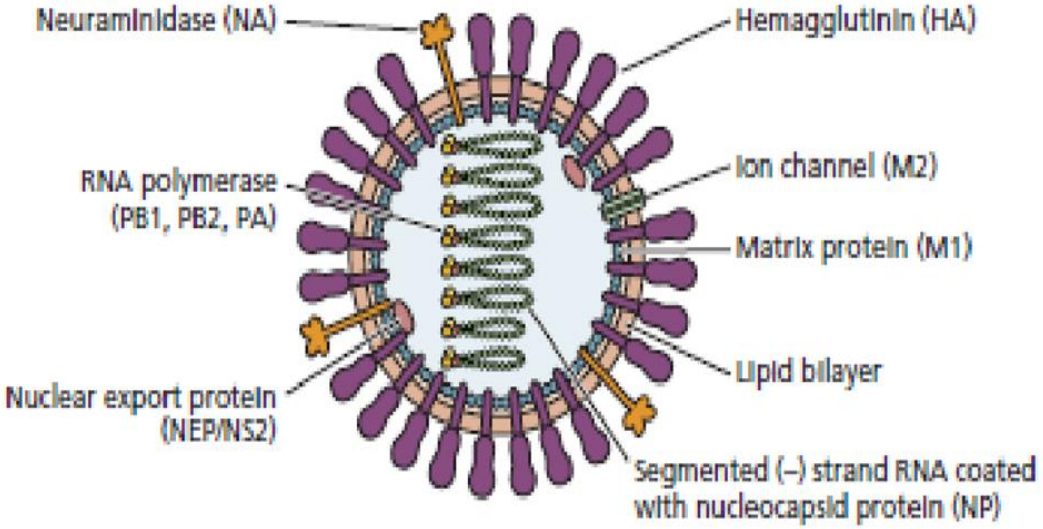


Segmented ssRNA(-): Orthomyxoviridae
Influenza A virus (IAV)

Influenza A has 8 genomic segments, each coated with nucleoprotein and displaying helical capsid symmetry. The segments share common nucleotide sequences at the 5’ and 3’ ends that are important for replication: they are complementary to each other, and within the virion the genomic segments are folded thanks to base pairing occurring at their ends

A





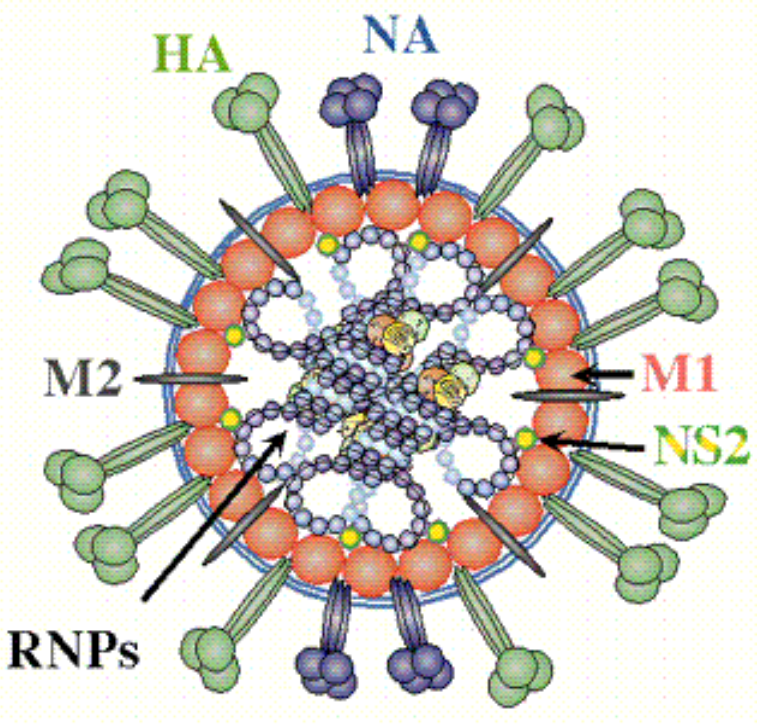
Structure and components

The outer layer of the lipid envelope is spiked with multiple copies of **Hemagglutinin** (**HA**) trimers and **Neuraminidase (NA)** tetramers.

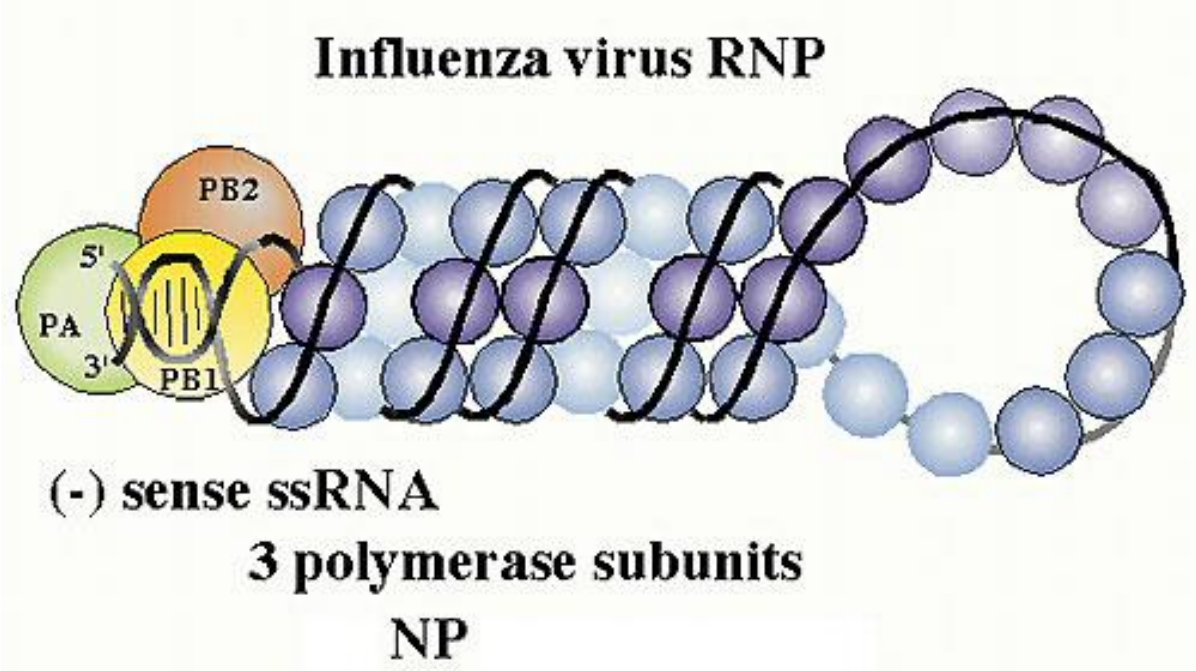
A small number of **M2** proteins forms the ion channels across the envelope

The **M1** (**Matrix protein**) molecules keep **vRNPs** attached to the inner layer

NS2, nuclear export protein (NEP)

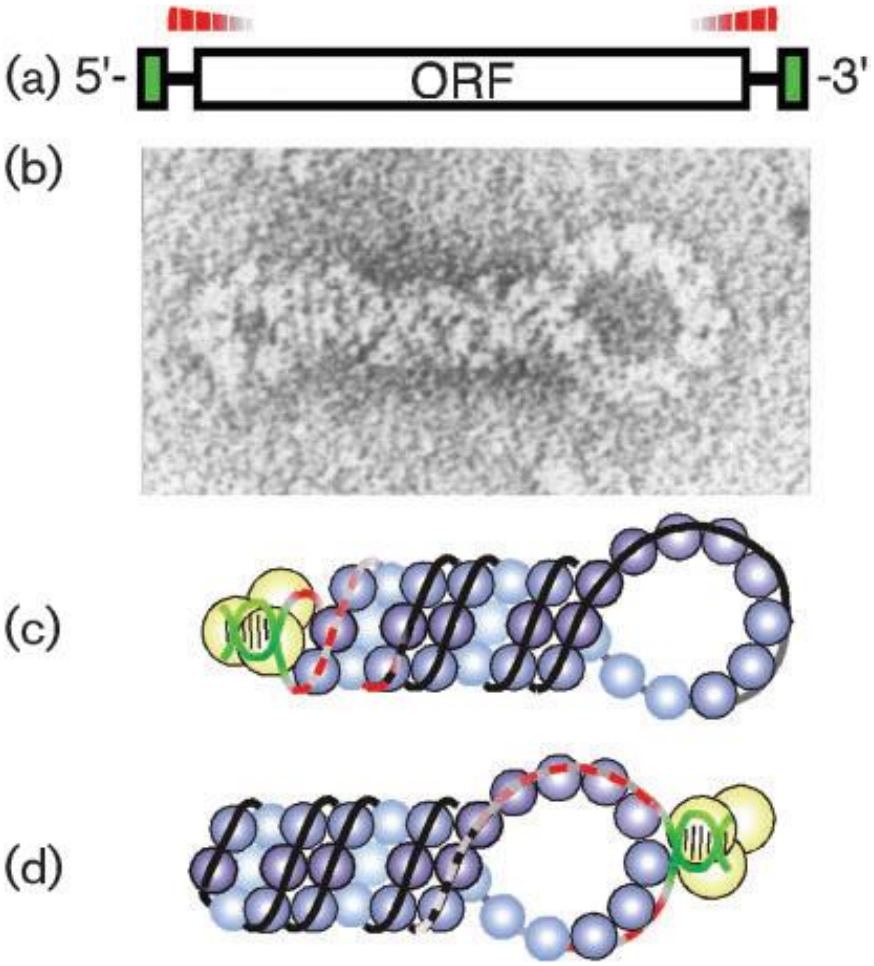


GENOME (-) ssRNA – 8 SEGMENTS



v-**RNP:** RNA + nucleoprotein (**NP).** The components of the **RNA-dependent RNA polymerase complex (PB1, PB2 and PA)** associates to each segment

GENOME (-) ssRNA – 8 SEGMENTS

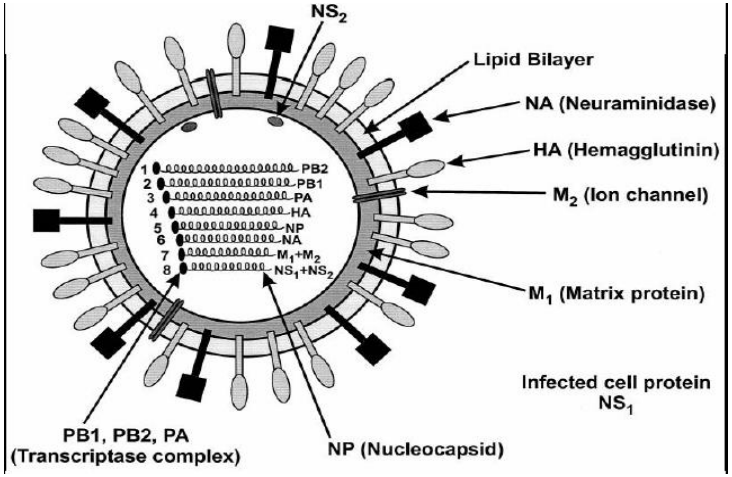


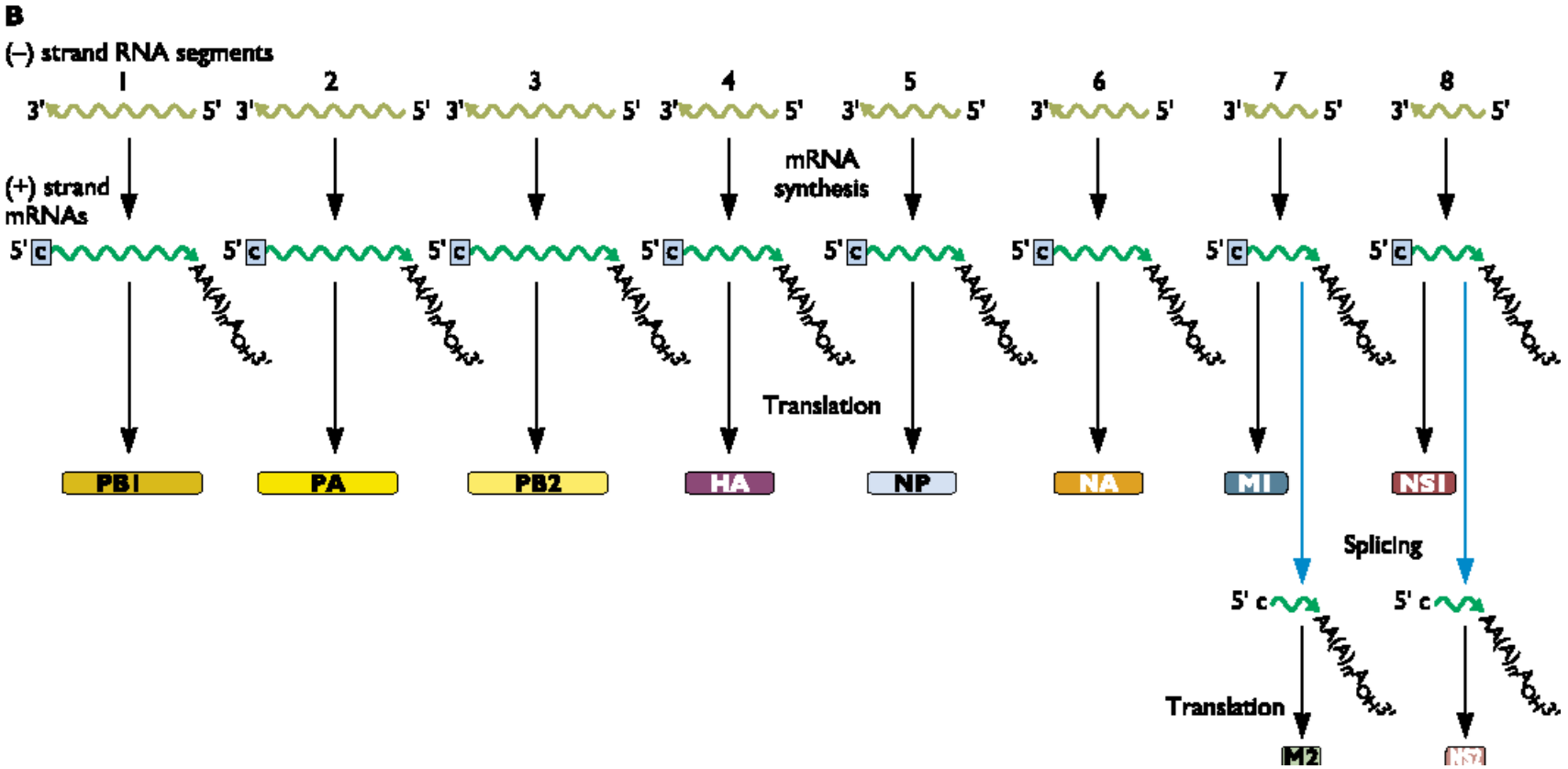
A large ORF (open box) is flanked by short UTRs containing terminal promoter sequences (green boxes) that form the polymerase binding site, they are essentially identical in all segments and show partial complementarity.
Specific packaging signals (red wedges) overlap the UTRs and terminal coding regions and are apparently discontinuous.

Journal of General Virology (2010), 91, 313–328

IAV genome segments

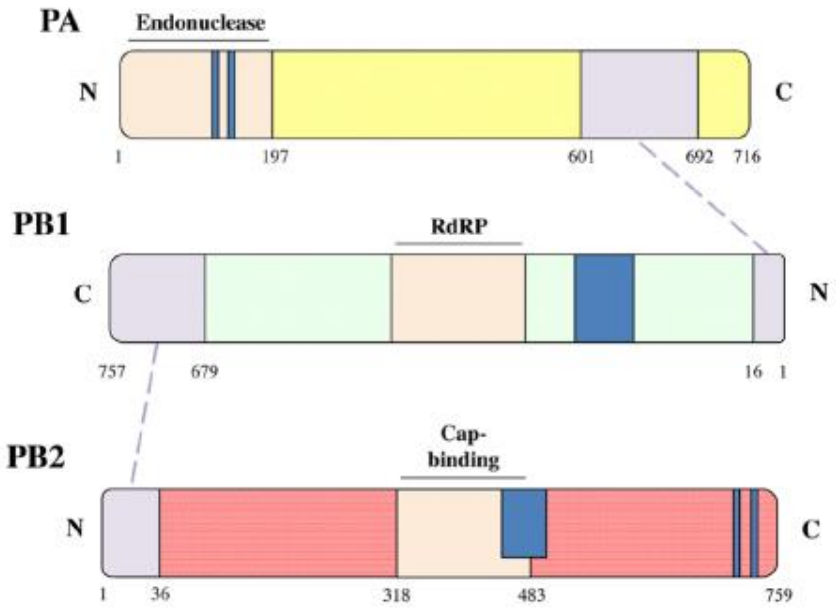
Segment	Size (nt)	Protein	Function
1	2341	PB2	Cap-binding (transcription)
2	2341	PB1	RNA-dependent RNA polymerase (transcription and replication)
3	2233	PA	Endonuclease
4	1778	HA	Hemagglutinin, receptor binding, fusion
5	1565	NP	Nucleoprotein, cytoplasm-nucleus translocation of v-RNP
6	1413	NA	Neuraminidase: virion release (main)
7	1027	M1 e M2	M1= matrix M2= ion channel
8	890	NS1 e NS2	NS1 = host function interference, absent inside the virion NS2 or NEP = v-RNP nuclear export





From Flint et al. *Principles of Virology* (2015), ASM Press

IAV polymerase complex



The interaction domains between proteins, as well as the functional domains are indicated. NLS of each protein are depicted in blue

Virus Research 209 (2015) 23–38

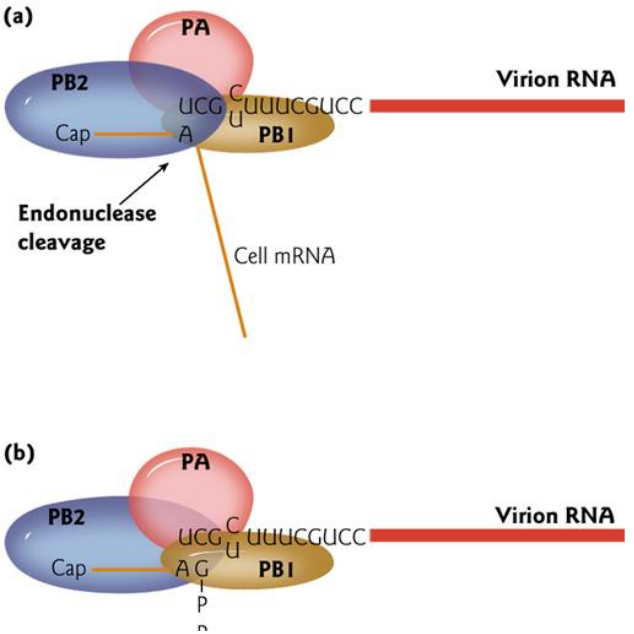
Influenza A Virus: transcription

Nigel J. Dimmock, Andrew J. Easton, Keith N. Leppard
Introduzione alla Virologia moderna *Casa Editrice Ambrosiana. 2017*

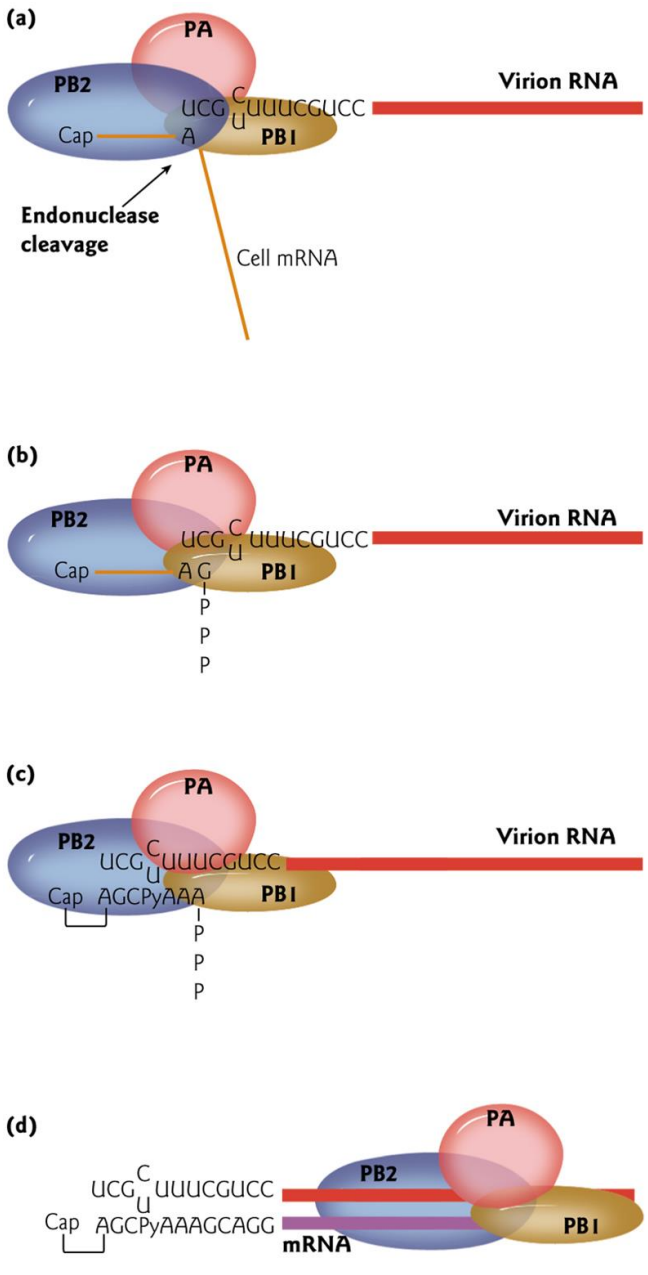
The polymerase complex transcribes in a primer-dependent manner and does not possess capping or methylation activity.

Transcription initiation occurs through **cap-snatching**:(i) the polymerase complex, together with the associated vRNA, binds the 5′ cap of a cellular mRNA via PB2;(ii) PA cleaves the mRNA 10–13 nucleotides downstream of the cap;(iii) the cleaved fragment is used as a primer and provides the cap for the nascent viral mRNAs.

Polymerase complex:
PB2: cap binding
PB1: initiation and elongation
PA: endonuclease activity



influenza A Virus: transcription



This is followed by the elongation phase of the messenger RNA until the termination signal is reached.Similarly to other class V viruses, the polymerase encounters a stretch of U residues (17–22 nucleotides from the 5′ end), which are copied repeatedly to generate the poly(A) tail through a stuttering-like mechanism.

The polymerase complex transcribes in a primer-dependent manner, whereas replication occurs in a primer-independent manner.

Nigel J. Dimmock, Andrew J. Easton, Keith N. Leppard
Introduzione alla Virologia moderna *Casa Editrice Ambrosiana. 2017*

RNAs in infected cells

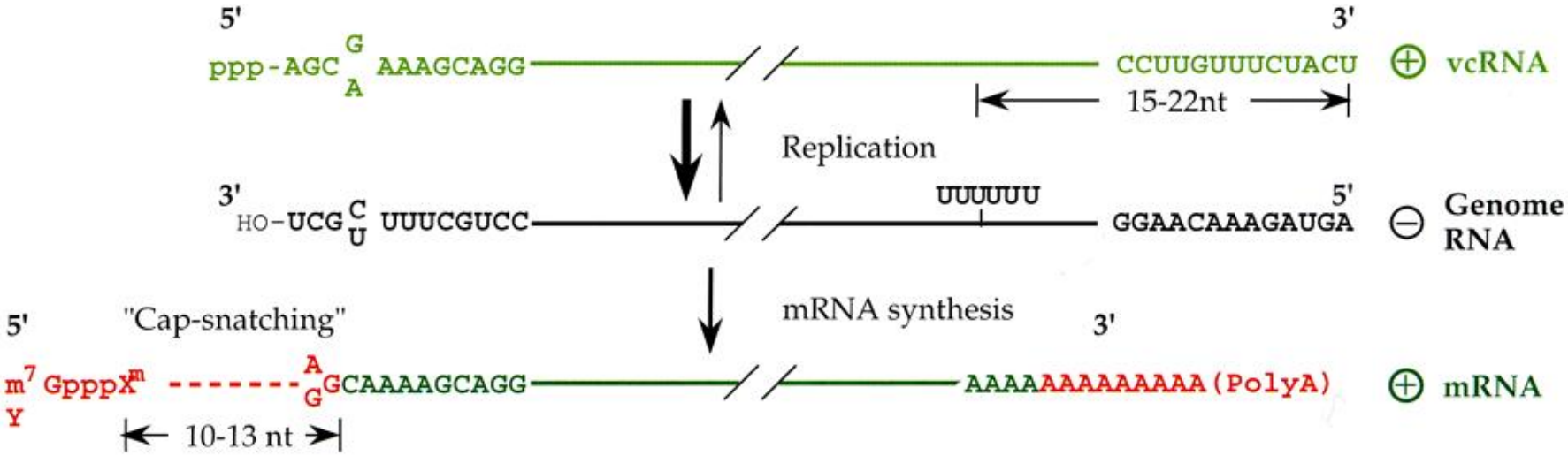


FIGURE 4.13 Relationship between genome RNAs, mRNAs, and vcRNAs of influenza virus. Synthesis of mRNAs in the cell nucleus requires a primer of 10–13 nucleotides derived from cellular pre-mRNAs by "cap-snatching," and mRNAs terminate with a poly(A) tail. Those portions of the mRNA that are not complementary to the genome RNA are shown in red. In contrast, vcRNAs are exact complements of the genomic minus strands. [Adapted from Strauss and Strauss (1997).]

IAV genome segments: accessory proteins

Segment	Size (nt)	Protein	Function
1	2341	PB2	Cap-binding (transcription)
2	2341	PB1 PB1-F2 N40	RNA-dependent RNA polymerase (transcription and replication) Apoptosis, inflammation, enhances polymerase activity Unknown
3	2233	PA PA-X	Endonuclease Shutoff of host protein expression at late stage of infection
4	1778	HA	Hemagglutinin, receptor binding, fusion
5	1565	NP	Nucleoprotein, cytoplasm-nucleus translocation of v-RNP
6	1413	NA	Neuraminidase: virion release (main)
7	1027	M1 e M2	M1= matrix M2= ion channel
8	890	NS1 e NS2	NS1 = host function interference, absent inside the virion NS2 or NEP = v-RNP nuclear export

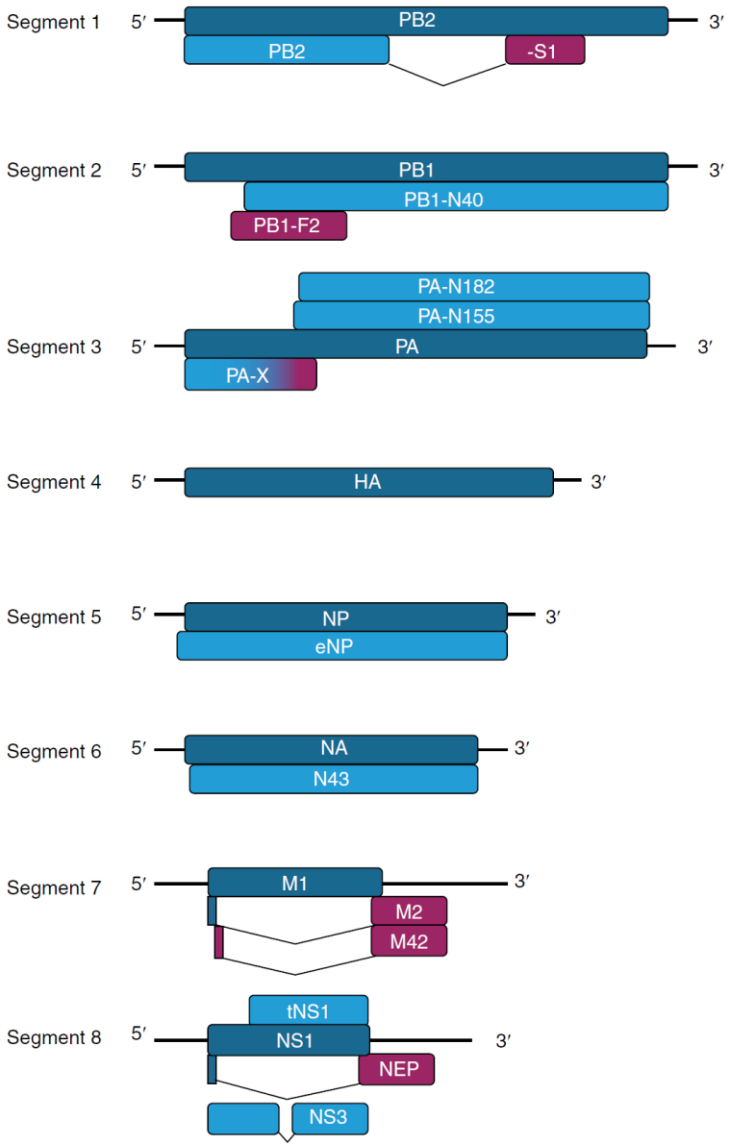


Figure 2. Schematic of the IAV coding strategy. mRNAs from the eight segments are symbolized by horizontal black lines. Coding regions (only approximately to scale) are represented by boxes with colors defining different reading frames (dark blue: primary products, frame 1; light blue: secondary products, frame 1; magenta: secondary products, frame 2). mRNA splicing in segments 1, 7, and 8 is denoted by deflected lines connecting coding regions.

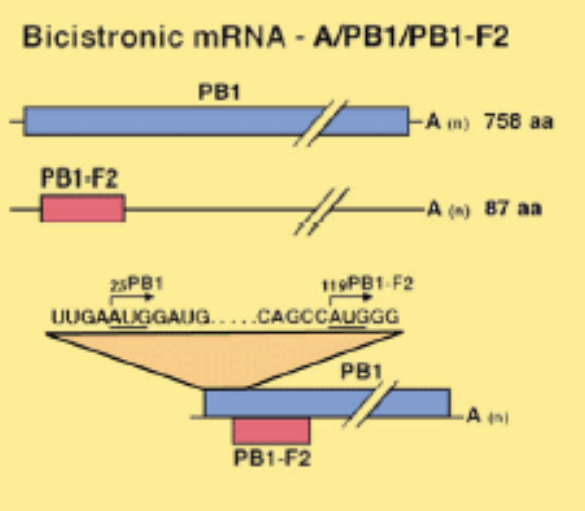
Table 1. Influenza A virus (IAV) gene products, expression mechanisms, and functions

Segment	Segment length (nt)	Protein products	Expression mechanism	Length (aa)	Molecular weight (kDa)	Main function(s)
1	2341	PB2	Canonical translation initiation	759	85.7	Component of the heterotrimeric viral RdRp; binds mRNA cap structures to promote cap snatching
		PB2-S1	mRNA splicing	508	55	Inhibitor of the RIG-I signaling pathway?
2	2341	PB1	Canonical translation initiation	757	86.6	Component of the viral RdRp; nucleotide polymerase
		PB1-N40	Alternative AUG initiation (translation reinitiation)	718	82.4	Unknown; interacts with the viral polymerase
		PB1-F2	Alternative AUG initiation (leaky scanning)	87	10.5	IFN antagonist; pro-apoptotic activity
3	2233	PA	Canonical translation initiation	716	84.2	Component of the viral RdRp; viral mRNA endonuclease responsible for cap snatching
		PA-X	Ribosomal frameshift	252	29	Exhibits endonuclease activity; contributes to host cell shutoff
		PA-N155	Alternative AUG initiation	562	62	Unknown
		PA-N182	Alternative AUG initiation	535	60	
4	1778	HA	Canonical translation initiation	566	61.5	Homotrimeric surface glycoprotein; mediates receptor binding and virus entry
5	1565	NP	Canonical translation initiation	498	56.1	Encapsidation of vRNA (to form RNP complexes); required for transcription activity and vRNP nuclear import
		eNP	Upstream AUG	504	56.8	Virulence factor for H1N1 IAVs
6	1413	NA	Canonical translation initiation	454	50.1	Homotetrameric surface glycoprotein; exhibits sialidase activity allowing release of virus progeny from the cell surface
		NA43	Alternative AUG initiation (leaky scanning?)	440	48.6	Uncertain, plasma-membrane-resident NA variant
7	1027	M1	Canonical translation initiation	252	27.8	Matrix protein; known to interact with RNPs and viral membrane proteins; involved in RNP nuclear export, virus assembly, and budding
		M2	mRNA splicing	97	11	Acts as proton channel; required during virus entry (uncoating) and for membrane scission at the budding stages of infection
		M42	mRNA splicing and alternative AUG initiation	99	13	Variant of M2 with common functions
8	890	NS1	Canonical translation initiation	230	26.8	Broad-spectrum IFN antagonist
		NEP/NS2	mRNA splicing	121	14.2	Involved in nuclear export of vRNPs
		NS3	mRNA splicing	187	21	Unknown
		NSP	unknown	167	18.1	Unknown; protein expression bioinformatically proposed, but yet not experimentally detected
		tNS1	Alternative AUG initiation	150/152	17	Involved in inhibition of IRF3

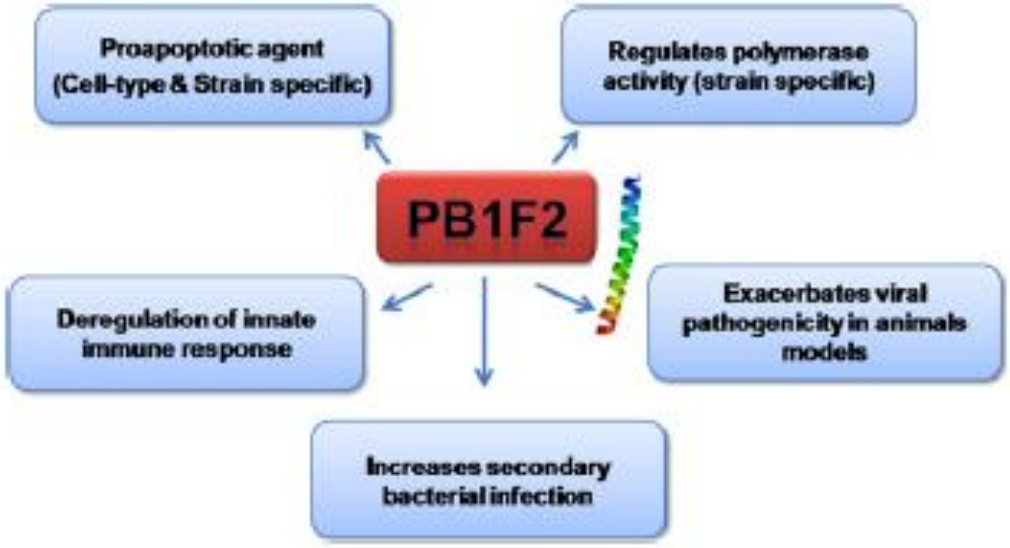
Segment and protein sizes based on A/Puerto Rico/8/1934 (H1N1).

(RdRP) RNA-dependent PNA polymerase, (mRNA) messenger RNA, (IFN) interferon, (vRNA) virion RNA, (IAV) influenza A virus, (RNP) ribonucleoprotein.

Genome segment 2 PB1-F2

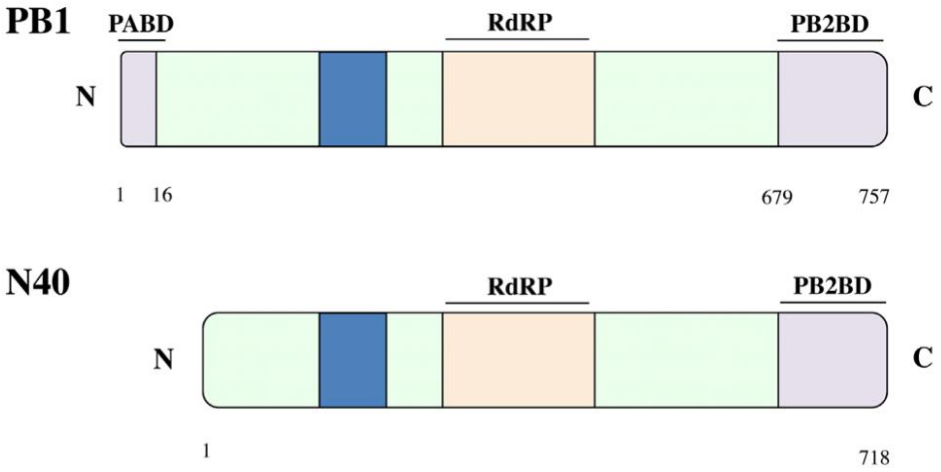


Here we see the two open reading frames of the PB1 gene segment of Influenza A Virus.
The red segment corresponds to the alternate reading frame that encodes the PB1-F2 protein whose start site is 120 bp downstream of the PB1 polymerase gene.



Translational regulation: alternative frame truncated protein

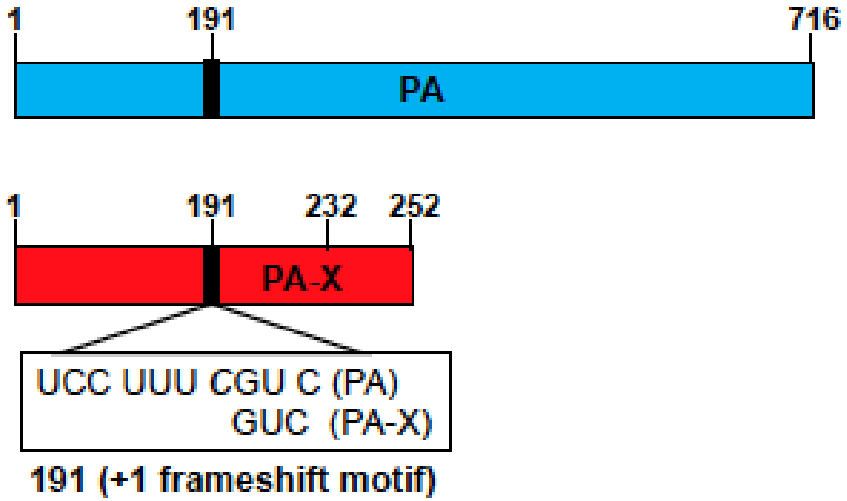
Genome segment 2 N40

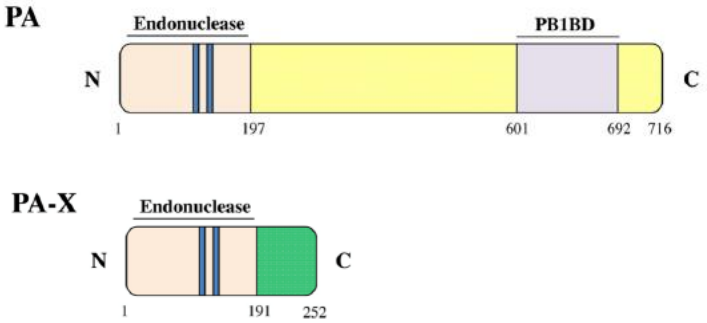


A leaky ribosomal scanning process is involved in N40 expression due to the presence of a strong Kozak translation initiation context in the 5th in frame AUG codon of PB1 gene. This initiation codon is located 115 nucleotides downstream from the first AUG in PB1 mRNA and thus, the N40 protein is an amino-terminally truncated version of the PB1 protein that lacks the first 39 amino acids, residues where the PA binding domain is located.

Virus Research 209 (2015) 23–38

Genome segment 3 PA-X



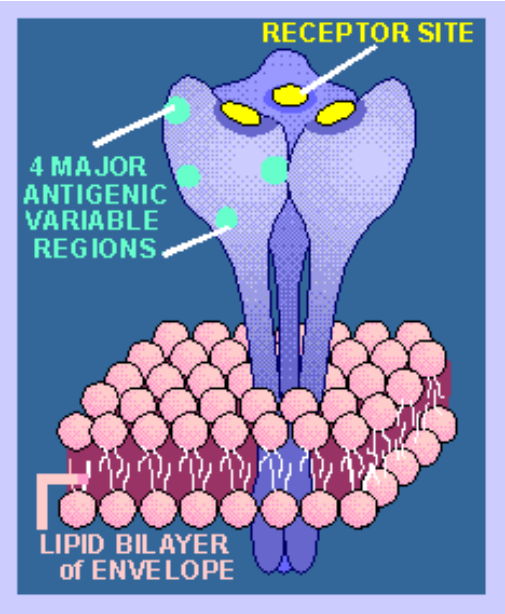


A schematic representation of the IAV PA viral segment and the PA and PA-X open reading frames (ORFs). Blue and red boxes indicate the ORF for PA and PA-X, respectively. The +1 frameshift motif (UCC UUU CGU C) at position 191 is indicated.
Bold and italics in the frameshift motif (C nucleotide) indicate that the nucleotide C is not read during PA-X translation.
PA-X proteins containing 232 or 252 amino acids if the C-terminal region has a 41 or 61 amino acid extension, respectively, are indicated. PA-X selectively degrades host RNA polymerase II (Pol II)-transcribed mRNAs and non-coding RNAs in the nucleus of infected cells, while sparing the products of polymerases I and III.

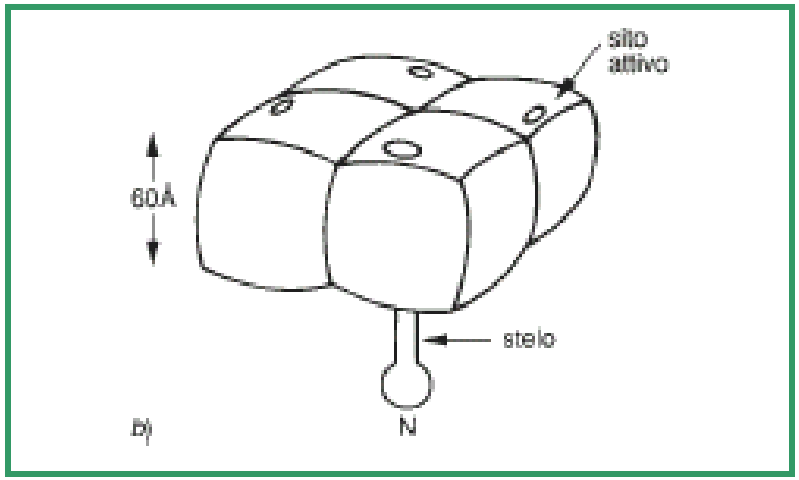
Viruses 2018, 10, 708

Virus Research 209 (2015) 23–38

Genome segment 4 and 6



On the HA head, the major antigenic variable regions localize close to the receptor binding site

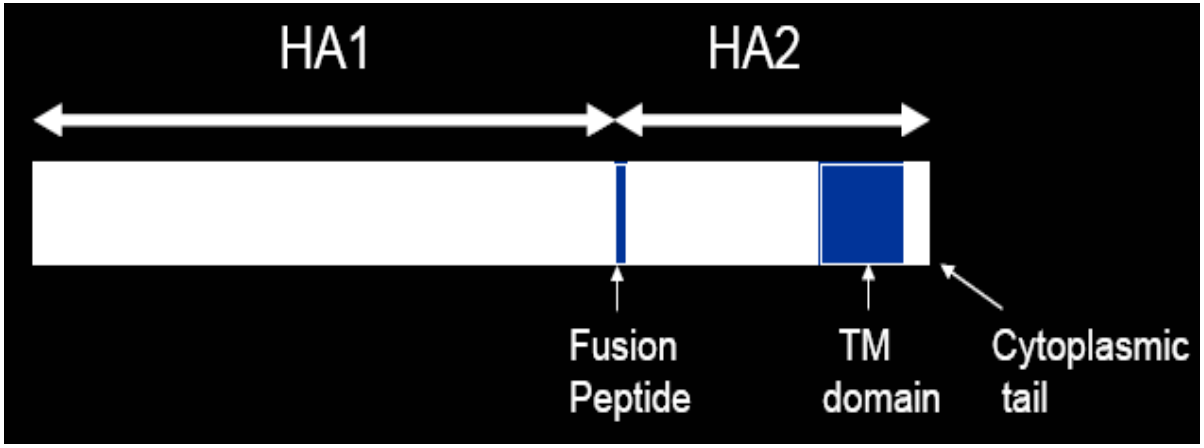


Antigenic variable epitopes characterize also neuraminidase (NA).

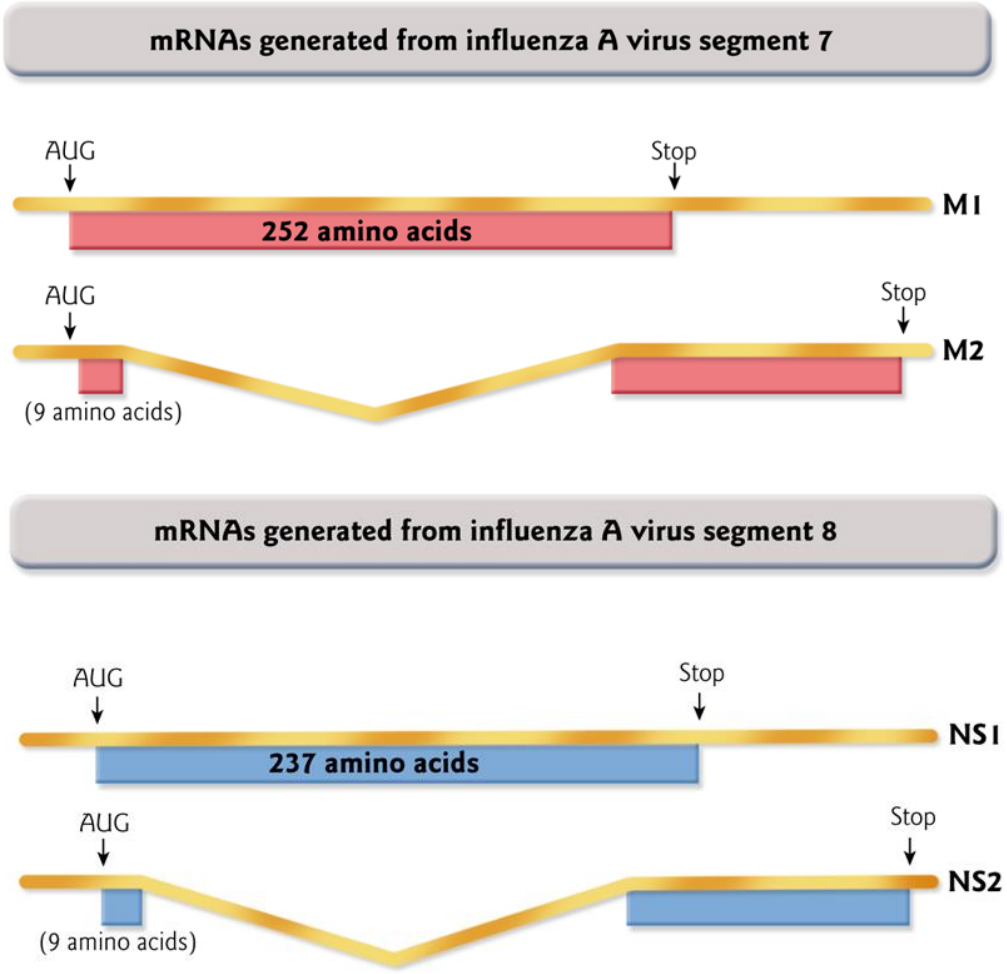
Post-translational regulation of HA

The proteolytic cleavage by **cellular proteases** is necessary for its function

HA is synthesized as a precursor (HA0), which is cleaved (trypsin-like enzymes or furin) into HA1 (responsible for interaction with the receptor) and HA2, which includes the trans-membrane domain and the fusion peptide (responsible for envelope fusion with endosomial membrane). The HA1 and HA2 fragments are held together by disulfide bridges.



Splicing of segment 7 and 8



Genome segment 7

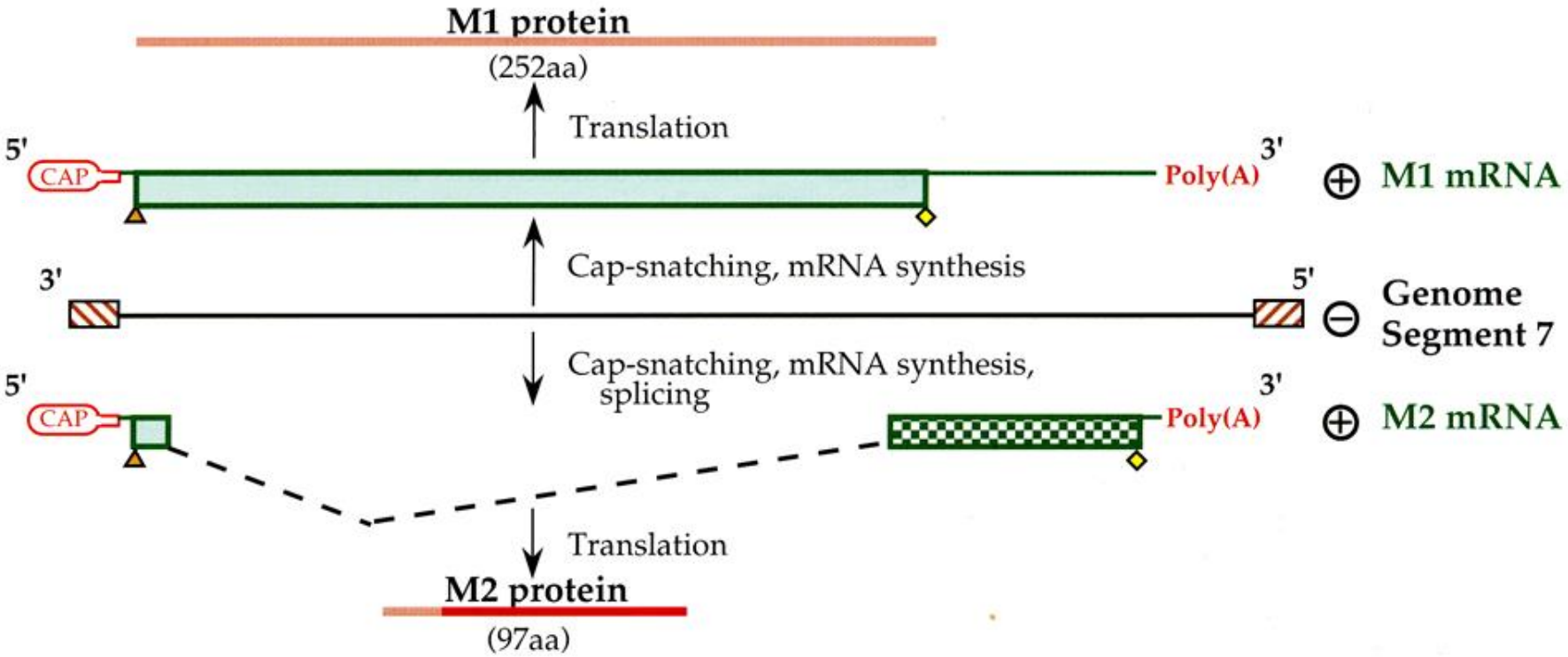
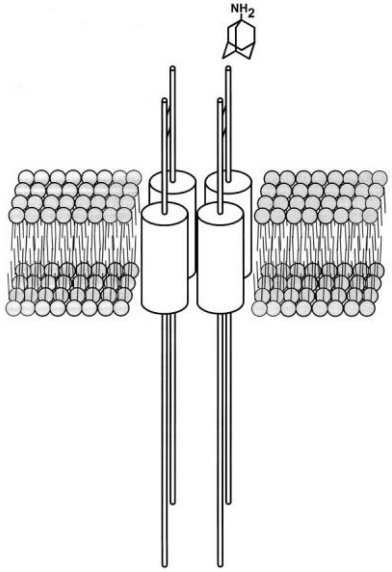


FIGURE 4.14 Synthesis of two mRNAs for the M1 and M2 proteins from gene segment 7 of influenza A. M1 RNA is translated from ORF1 (open box). M2 RNA starts identically, but after the splice it is translated in ORF2 (checked box). Both proteins are found in infected cells. The AUG initiation codon is shown as a triangle; termination codons are shown as filled diamonds. Patterned boxes at the end of the genome RNA are self-complementary sequences not present in the mRNAs that could form panhandles.

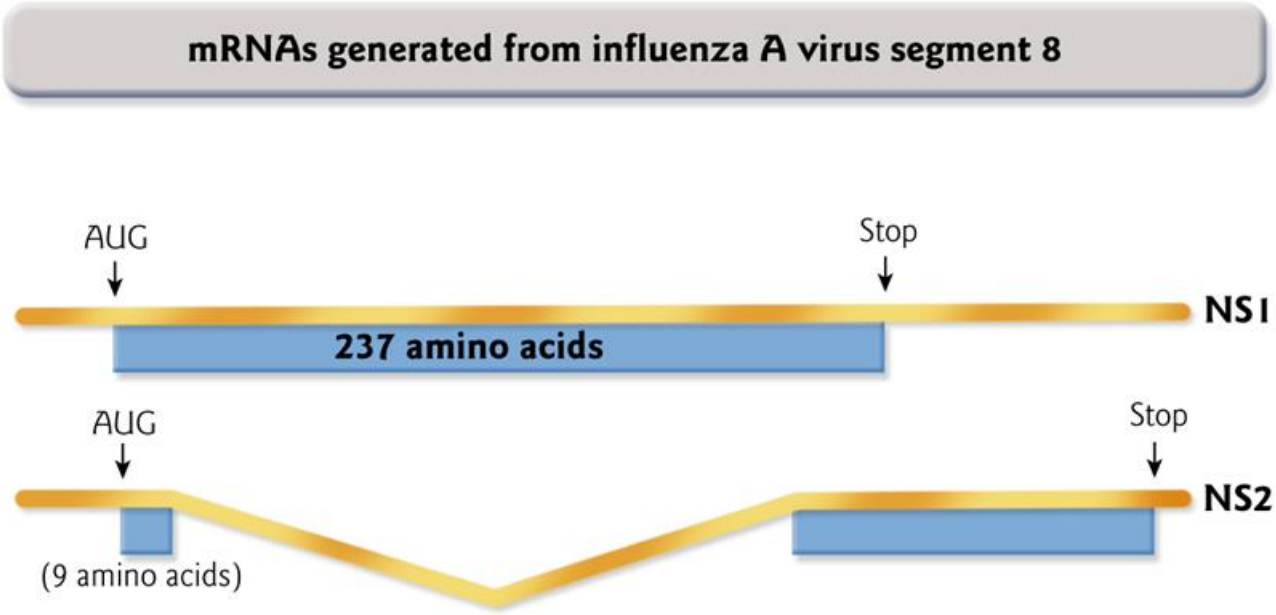
The M1 and M2 proteins

- The M1 matrix protein is located inside the lipid envelope and mediates the assembly and budding of the virus particle.
- The M2 protein forms active ion channels for proton transport. Ion channel activity is essential during the virus uncoating.
- These channels are specifically blocked by the antiviral drug amantadine hydrochloride.



The M2 protein forms homotetramers held together by disulfide bridges

Genome segment 8



The NS gene

NS1: anti-IFN activity, pre-mRNA accumulation in the nucleus

- The NS1 protein of the influenza A virus i) binds to dsRNA, ii) prevents activation of PKR and OAS mediated by dsRNA, iii) prevents the synthesis of Interferon. All these activities have been mapped to NS1 N-terminal domain.
- Through the interaction of the C-terminal portion with cellular proteins NS1 inhibits the polyadenylation of cellular mRNAs and their accumulation in the cytoplasm, increasing the pre-mRNA concentration in the nucleus and their availability for viral functions

NS2 (NEP for Nuclear Export Protein): involved in the nuclear export of the vRNPs

Splicing alternativo Orf alternativa Ribosomal frameshift Leaky scanning

Segment	Size (nt)	Protein	Function
1	2341	PB2	Cap-binding (transcription)
2	2341	PB1 PB1-F2 N40	RNA-dependent RNA polymerase (transcription and replication) Apoptosis, inflammation, enhances polymerase activity Unknown
3	2233	PA PA-X	Endonuclease Shutoff of host protein expression at late stage of infection
4	1778	HA	Hemagglutinin, receptor binding, fusion
5	1565	NP	Nucleoprotein, cytoplasm-nucleus translocation of v-RNP
6	1413	NA	Neuraminidase: virion release (main)
7	1027	M1 e M2	M1= matrix M2= ion channel
8	890	NS1 e NS2	NS1 = host function interference, absent inside the virion NS2 or NEP = v-RNP nuclear export

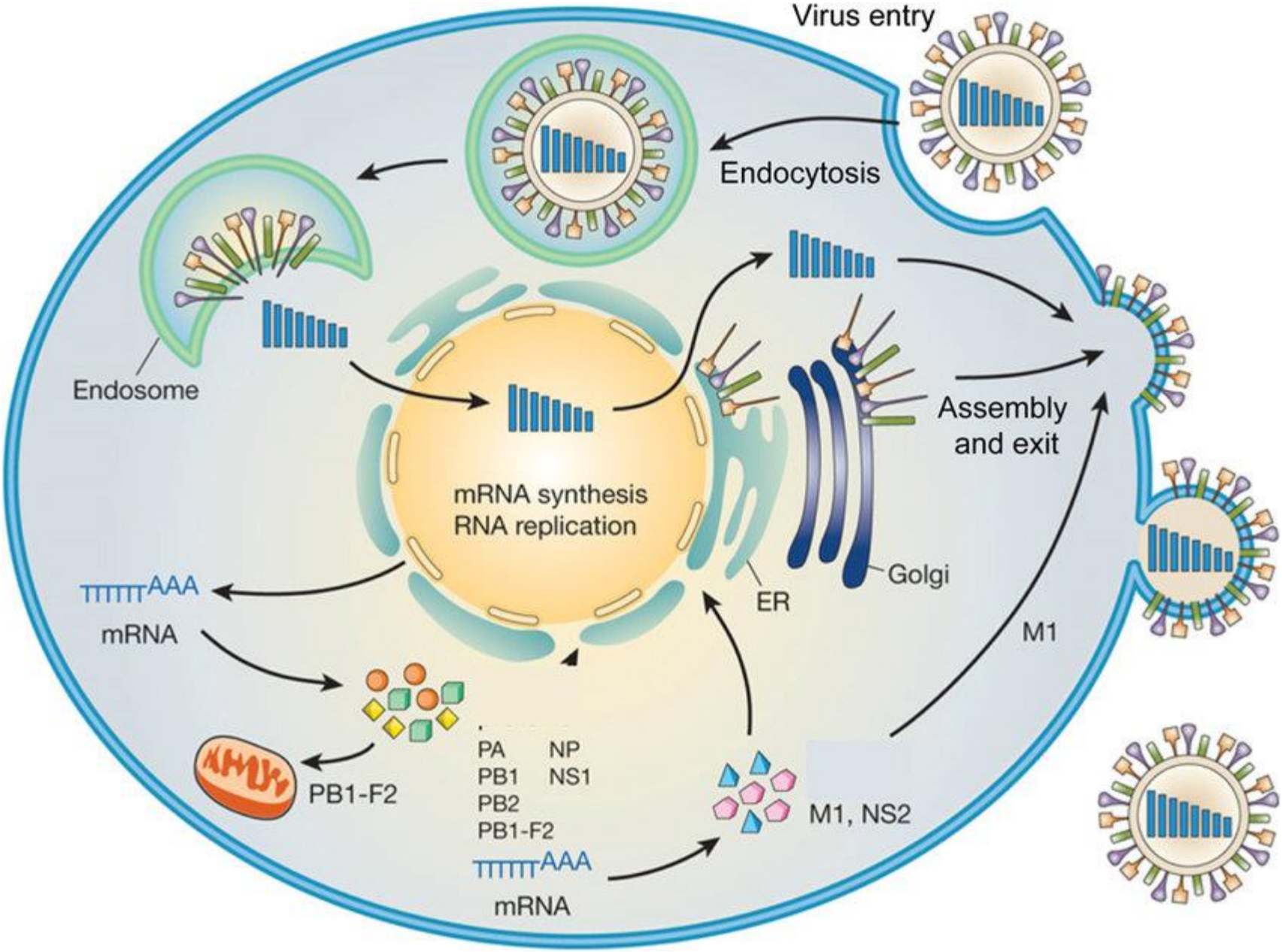
LIFE CYCLE

Among the RNA(-) viruses, the influenza A virus represents an exception, as the synthesis of the RNAs (during both transcription and replication) and the assembly of the vRNPs takes place inside the nucleus of the infected cell.

Replication of Influenza viruses occurs in the nucleus

After uncoating, the virus is transported to the nucleus, where the synthesis of mRNAs takes place. The mRNAs are then transported to the cytoplasm to be translated; in later stages, some of the viral proteins are transported back into the nucleus.

These viruses require cellular enzymatic functions that are located in the nucleus. Two important processes involved in their gene expression make them dependent on nuclear localization: the initiation of transcription and splicing.



Nuclear phase for transcription and replication

Influenza A temporal regulation

All viral genes are transcribed throughout every phase of the IAV life cycle, but a certain temporal regulation can be observed.

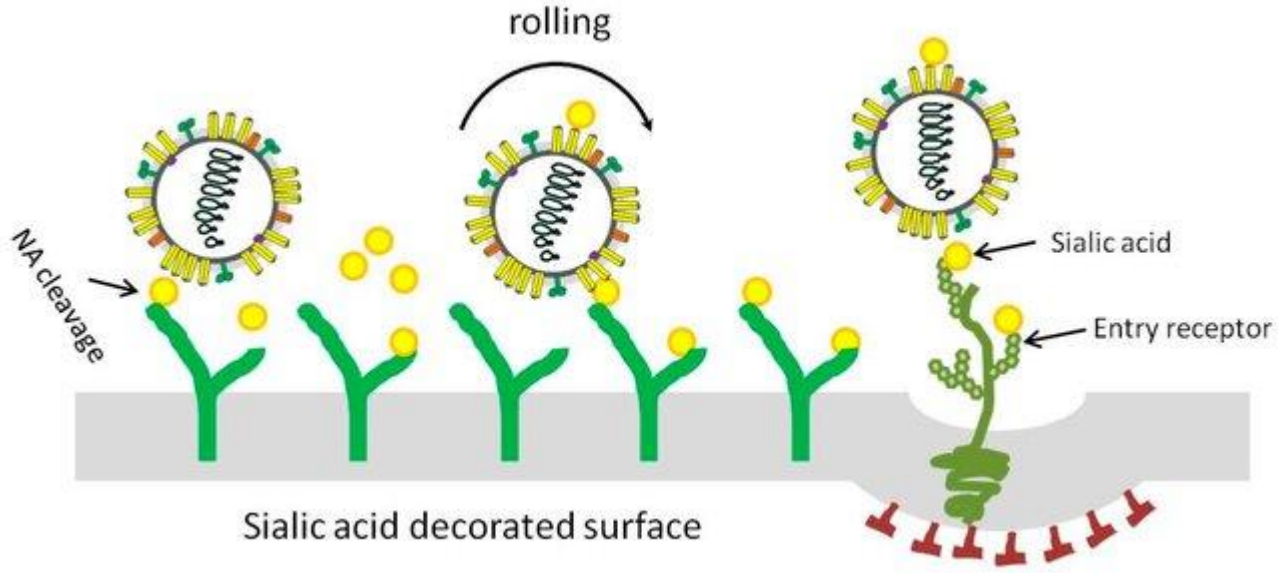
During the early stages of infection, similar amounts of each viral mRNA are produced.

About one hour after infection, higher levels of the mRNA encoding the nucleoprotein and of the mRNA encoding **NS1** begin to appear.

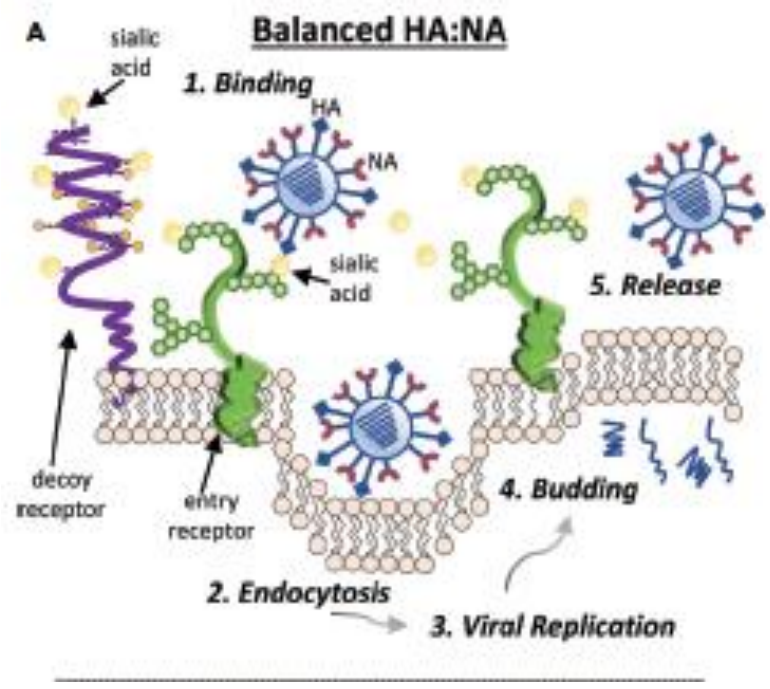
Following genome replication, we instead observe an increased production of the mRNAs encoding the structural proteins **HA, NA, and M1.**

Life Cycle: Attachment

The attachment of the virion to the host cell is mediated by the interaction of HA with glycoproteins containing sialic acid residues.



The relative activity of the HA and NA needs to be balanced to maintain the ability of the virion to efficiently infect and be released from cells (HA:NA=6:1)



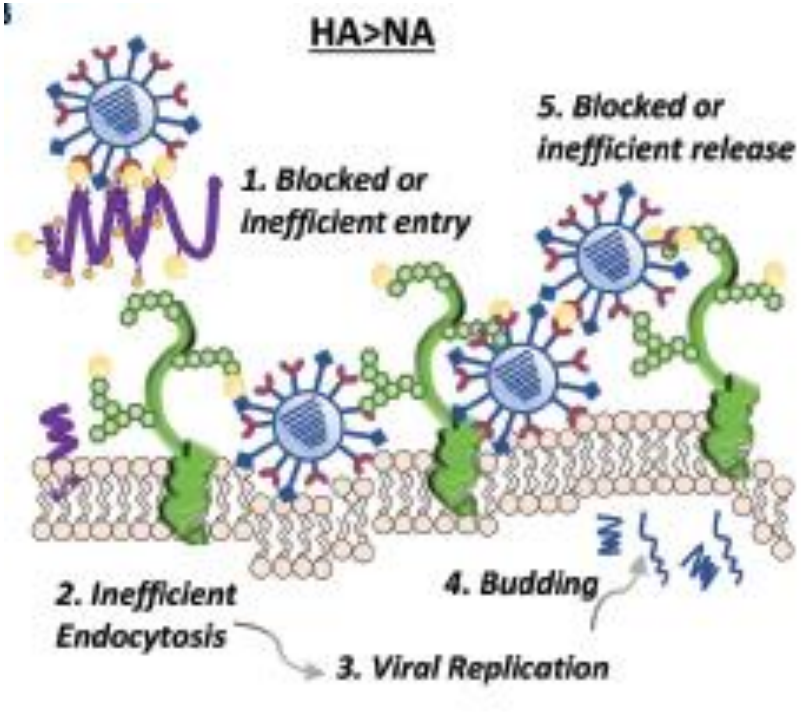
Efficient cleavage of sialic acids from decoy receptors (such as cell-surface mucins) by NA enables HA access to sialic acids expressed by entry receptors and efficient endocytosis.

After entry, transcription and replication new virions assemble at the cell surface and are released from the cell by budding.

As the viral components bud from the cell, NA cleaves sialic acids from receptors near the budding site to prevent virions binding back to the dying cell. NA cleavage of sialic acids from the carbohydrate side chains of nascent HA and NA also prevents newly budded virus from clumping together. Both these functions enable efficient release of the nascent virions from the cell.

HA and NA activities need to be balanced

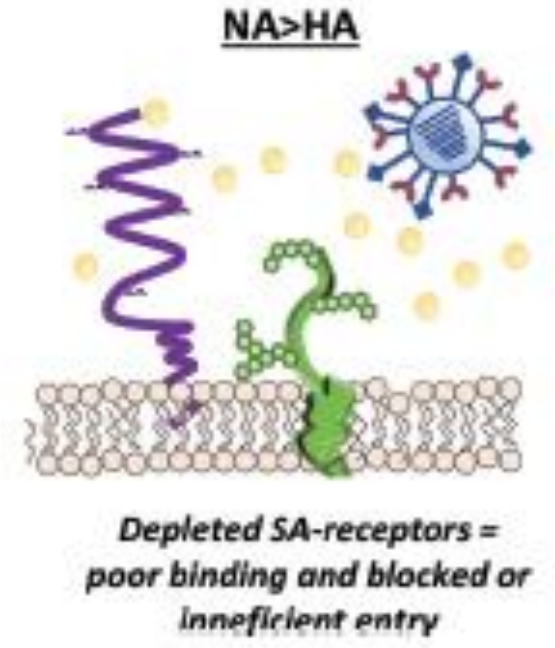
If the HA and NA are mismatched and have mutations in important binding or catalytic sites that alter function, the relative activity of the two proteins may be imbalanced.



If the sialidase function of NA is suboptimal, virus may remain bound by decoy receptors, which may shed and block virus entry into the cell.

As the virus buds from the cell, an imbalance of HA and NA function may result in the lack of release of the virions due to the binding of HA to the sialic acids expressed at the cell surface that have not been removed by the NA.

HA and NA activities need to be balanced



Alternatively, if the sialidase activity of NA is too strong when compared to the HA-binding
activity, sialic acids may be removed from receptors at the expense of the HA being able to bind and trigger endocytosis.

Life Cycle

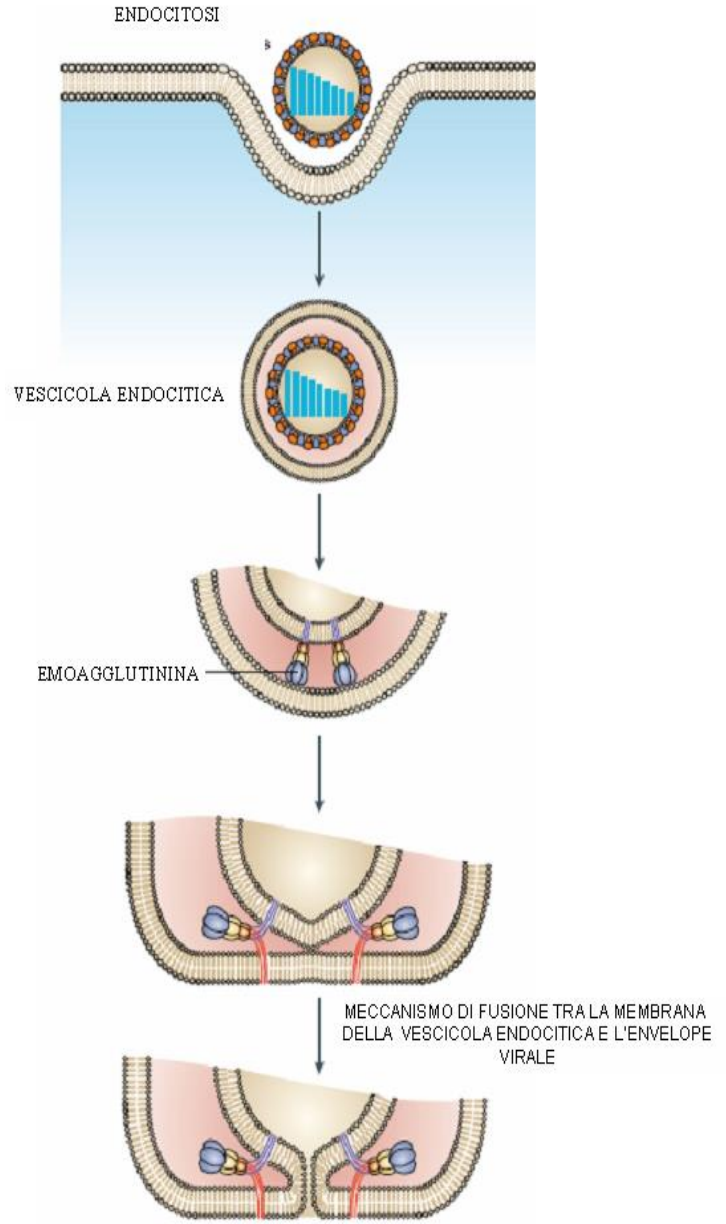
•After binding, the virus is incorporated into an endocytic vesicle. The endosome is acidified by the cell;

•The acidic pH determines a conformational modification of the HA molecules with activation of the fusion domain (located in the HA2 fragment) and subsequent fusion of the viral envelope with the endosome membrane, which determines the passage of the nucleocapsid into the cytoplasm.

• The ion channels formed by the M2 protein contribute to the virus uncoating.

Life Cycle: endocytosis

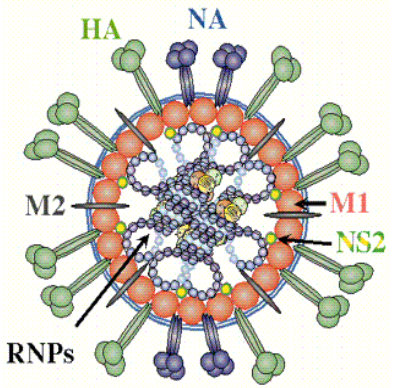
Hemagglutinin mediates the binding of the virus to sialic acid present on the glycoprotein on the cell surface. Through endocytosis, the virus enters the host cell. The acidification of the endocytic vesicle induces a conformational change of the HA exposing the fusion peptide in the HA2 fragment, which thus mediates the fusion between the viral envelope and the membrane of the endocytic vesicle.

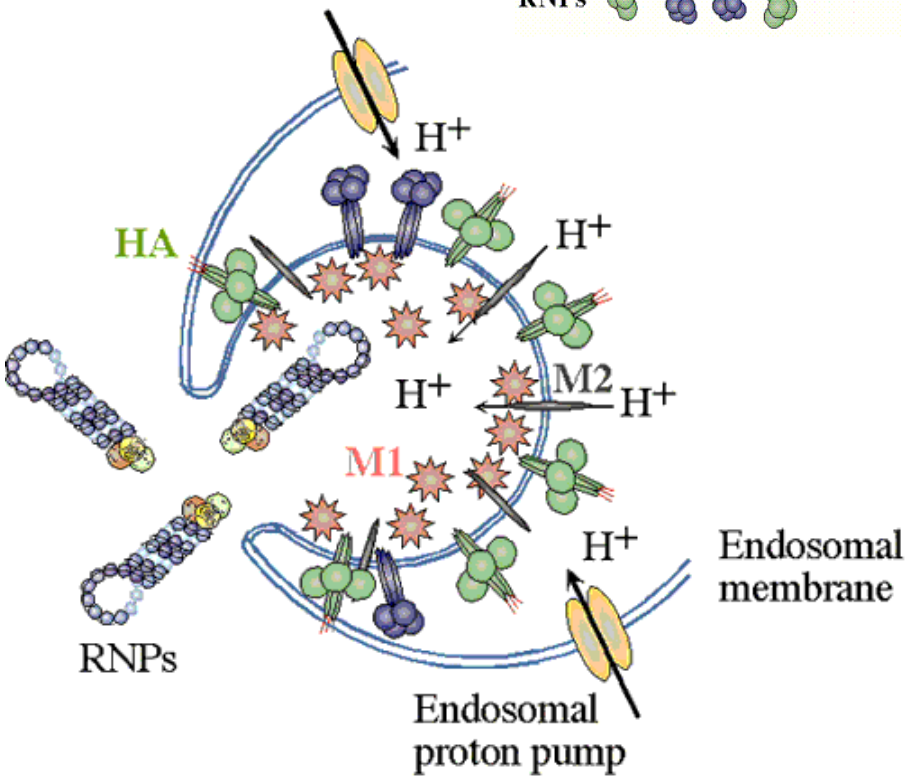


Life cycle: uncoating

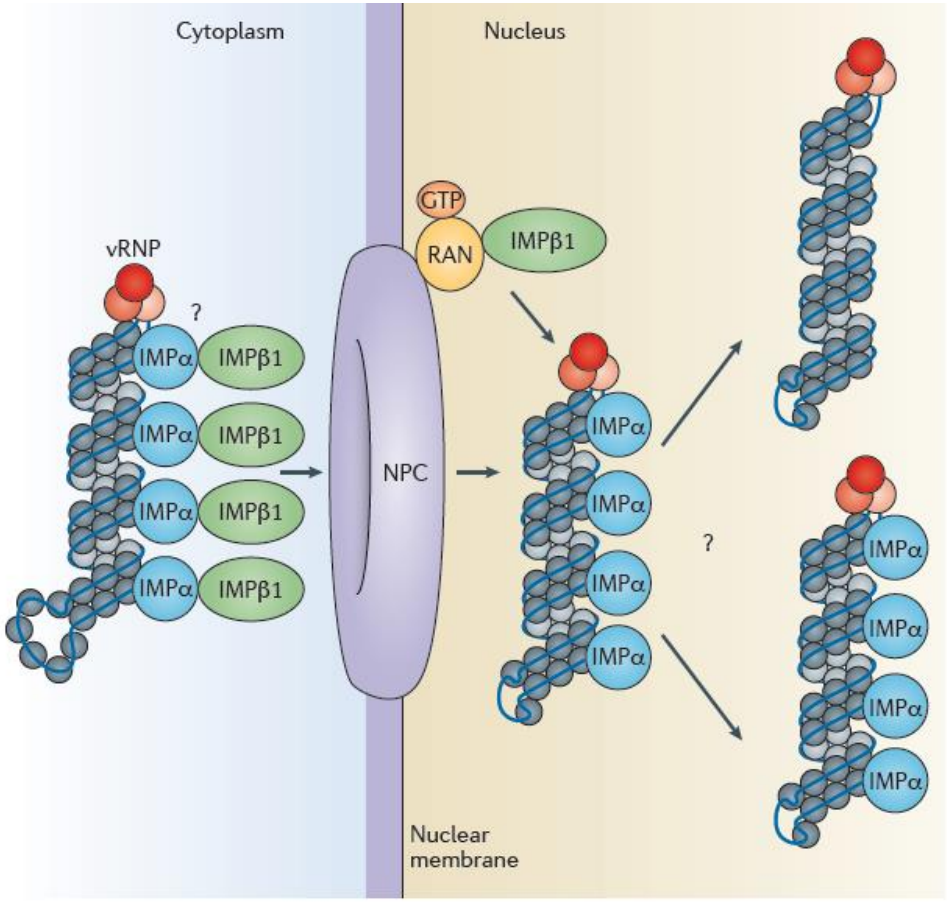
Uncoating and release of vRNP into the cytoplasm

Endocytic vesicle acidification allows proton entry inside the virion through the M2 ion channels. Virion acidification induces detachment of vRNP from the M1 matrix protein and their release into the cytoplasm.





Model for vRNP nuclear import

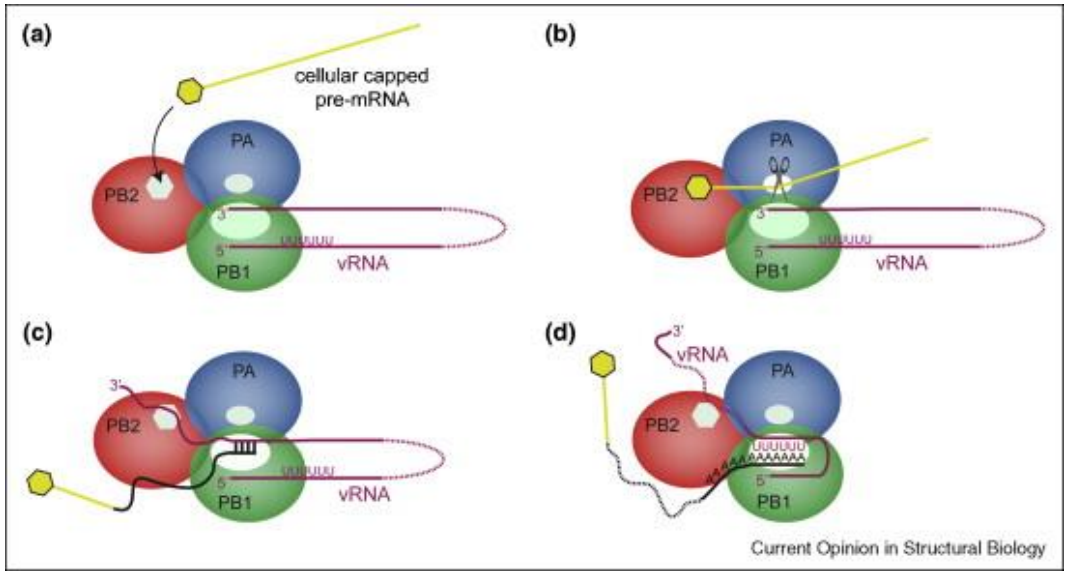


Uncoated, cytoplasmic viral ribonucleoproteins (vRNPs) with exposed nucleoprotein (NP) nuclear localization sequence (NLS) motifs associate with importin-α (IMPα), which in turn associates with IMPβ1. The entire complex docks at the nuclear pore complex (NPC) and is transported into the nucleus, where RAN–GTP binds to IMPβ1 and facilitates vRNP release into the nucleoplasm to initiate transcription and replication. Whether multiple IMPα and IMPβ1 molecules associate with each vRNP is unknown, as is the fate of the vRNP-associated IMPα once the vRNP cargo is released into the nucleoplasm.

Nature Reviews Microbiology **volume 13**, pages 28–41 (2015)

LIFE CYCLE: Transcription

Genome segments are transcribed by the polimerase complex

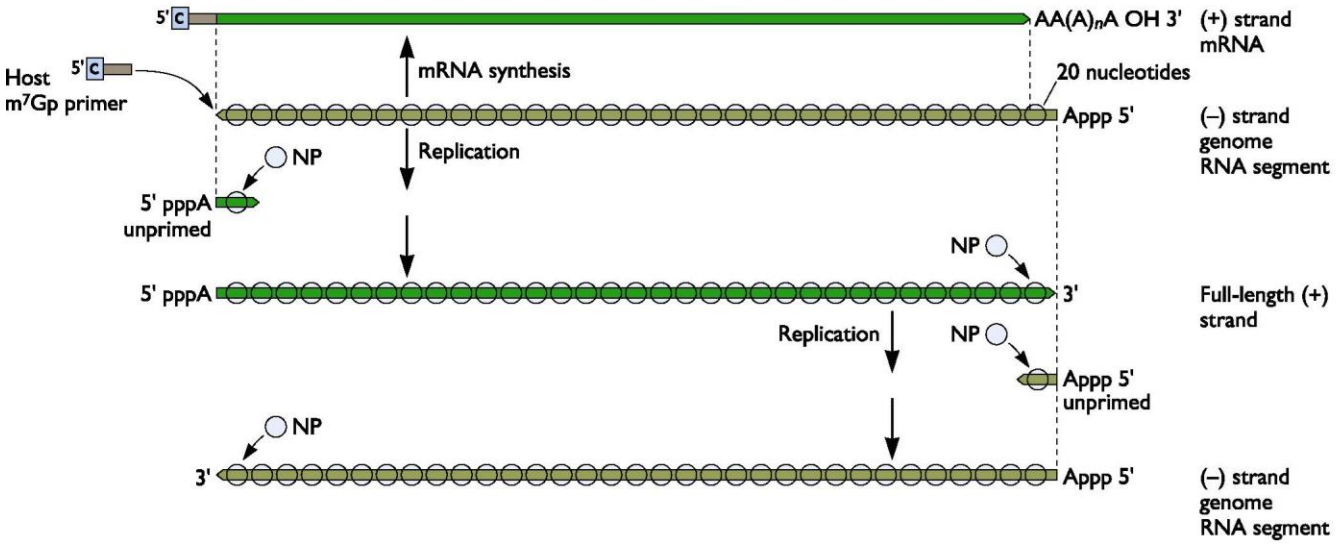


Schematic diagram showing steps in cap-dependent transcription by influenza virus polymerase. (a) Binding of host pre-mRNAs (yellow) by the cap binding domain located in the PB2 subunit. (b) Cleavage of the host mRNA after 10–13 nucleotides by the endonuclease located in the PA subunit. (c) Elongation of the chimeric viral mRNA by the nucleotidyl-transferase site in the PB1 subunit using the vRNA as template. (d) Poly-adenylation of the viral mRNA by polymerase stuttering at the oligo-U sequence near the 5' end of the vRNA.

Current Opinion in Structural Biology 2010, 20:104–113

LIFE CYCLE: genome replication

The transition from transcription to replication of genomic RNA depends, in part, on the abundance of the NP protein and on the polymerase acquisition of the ability to initiate a primer-independent RNA synthesis.

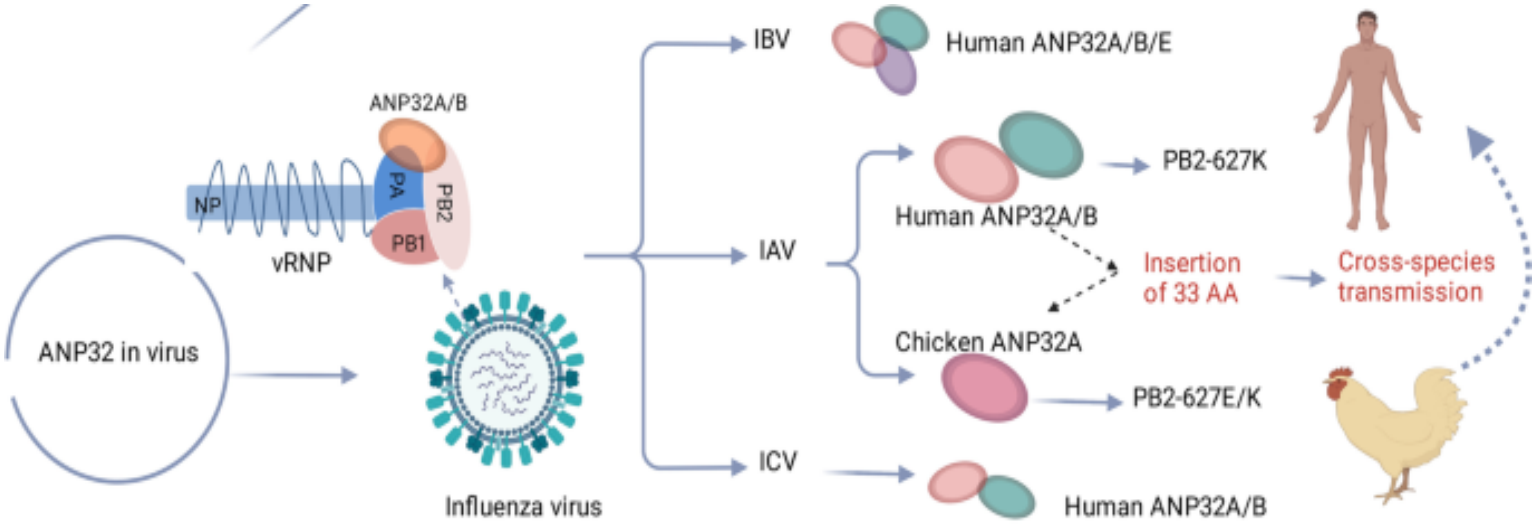


From Flint et al. *Principles of Virology* (2000), ASM Press

REVIEW Open Access

Roles of ANP32 proteins in cell biology and viral replication

Mengmeng Yu[1], Yuxing Qu[1], Haili Zhang[2*] and Xiaojun Wang[1*]



Article

Host ANP32A mediates the assembly of the influenza virus replicase

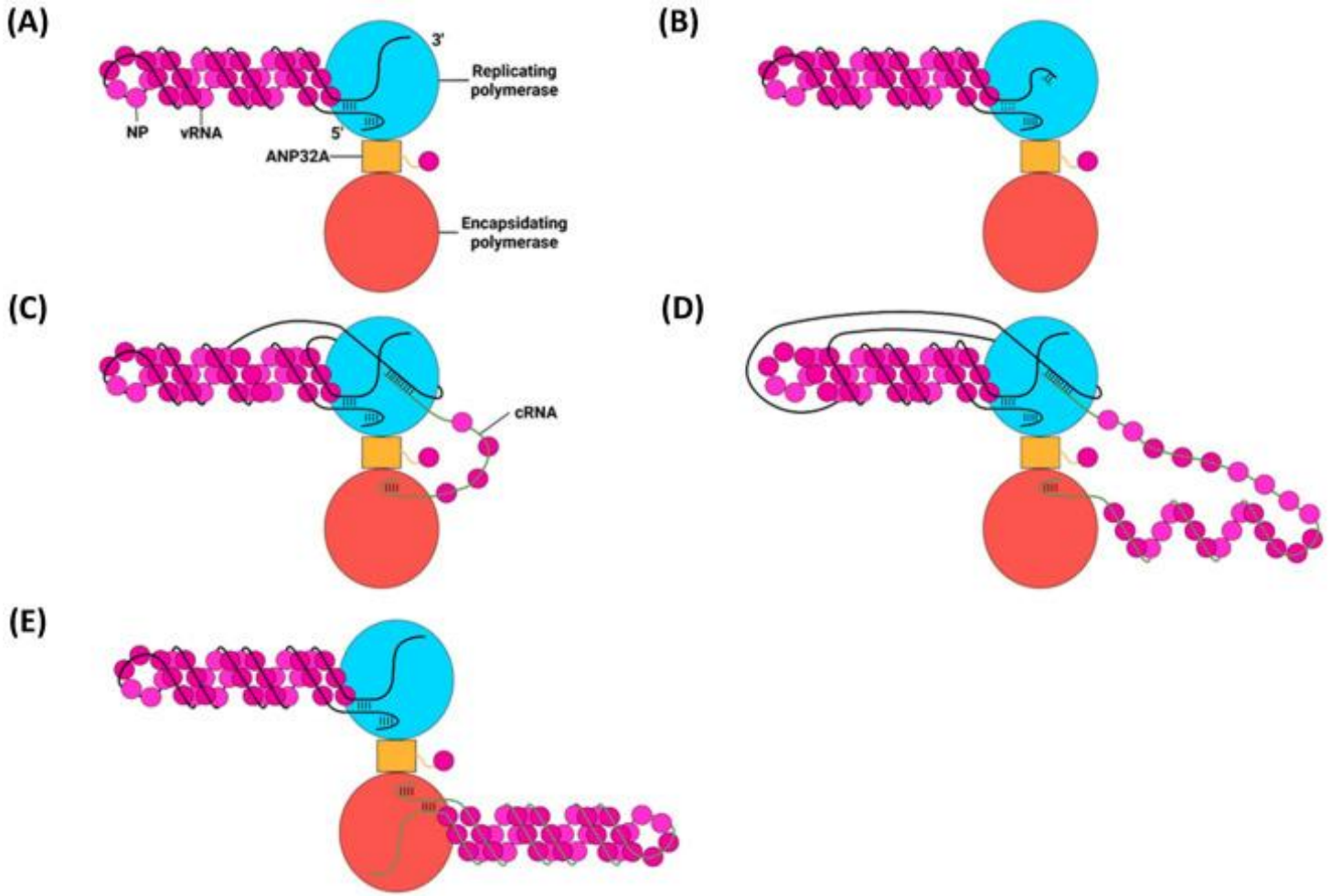
https://doi.org/10.1038/s41586-020-2927-z

Received: 7 February 2020

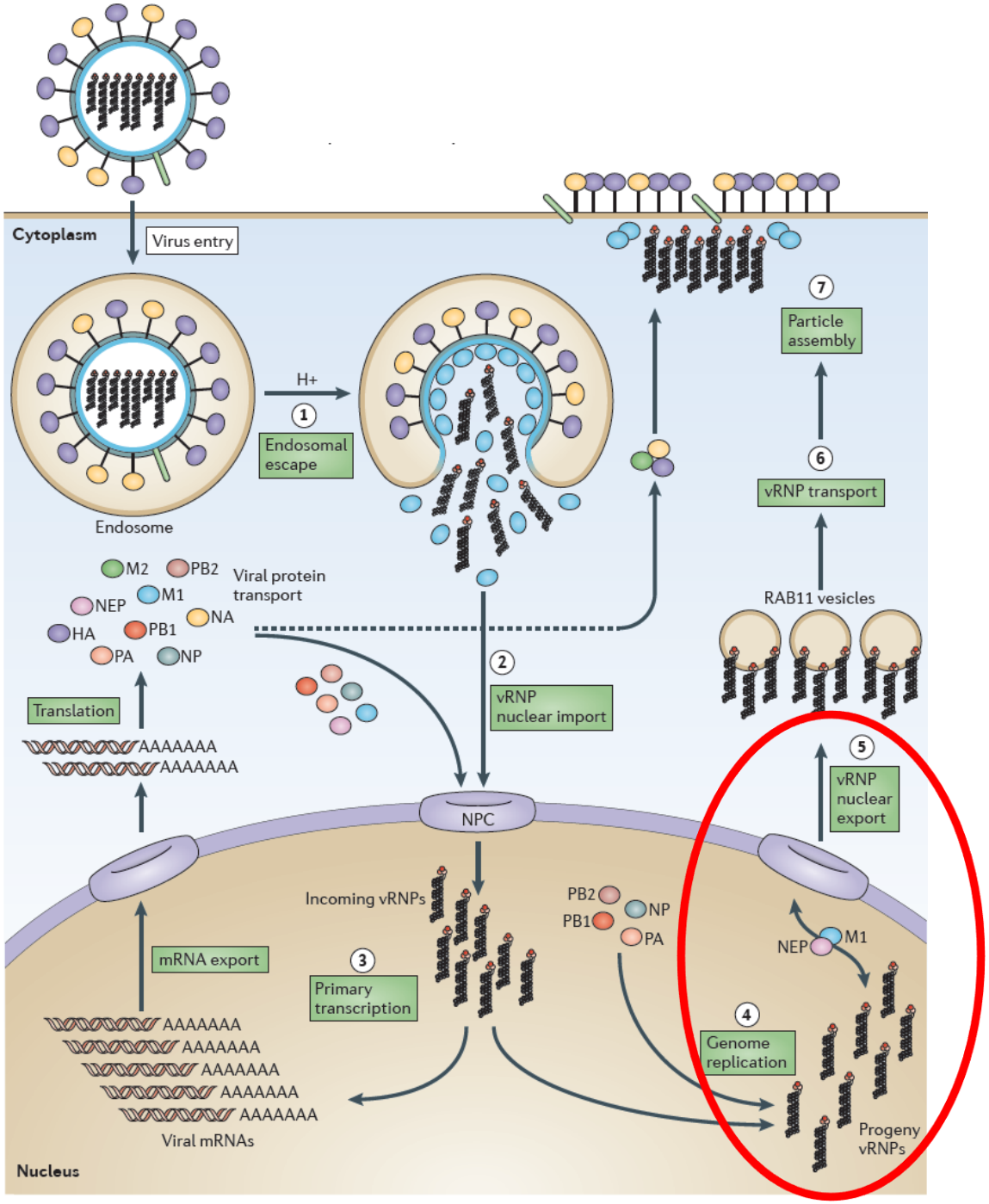
Loïc Carrique[1,6], Haitian Fan[2,6], Alexander P. Walker[2,6], Jeremy R. Keown[1,6], Jane Sharps[2], Ecco Staller[3,5], Wendy S. Barclay[3], Ervin Fodor[2,7]✉ & Jonathan M. Grimes[1,4,7]✉

ANP32 Proteins Are Essential for Influenza Virus Replication in Human Cells

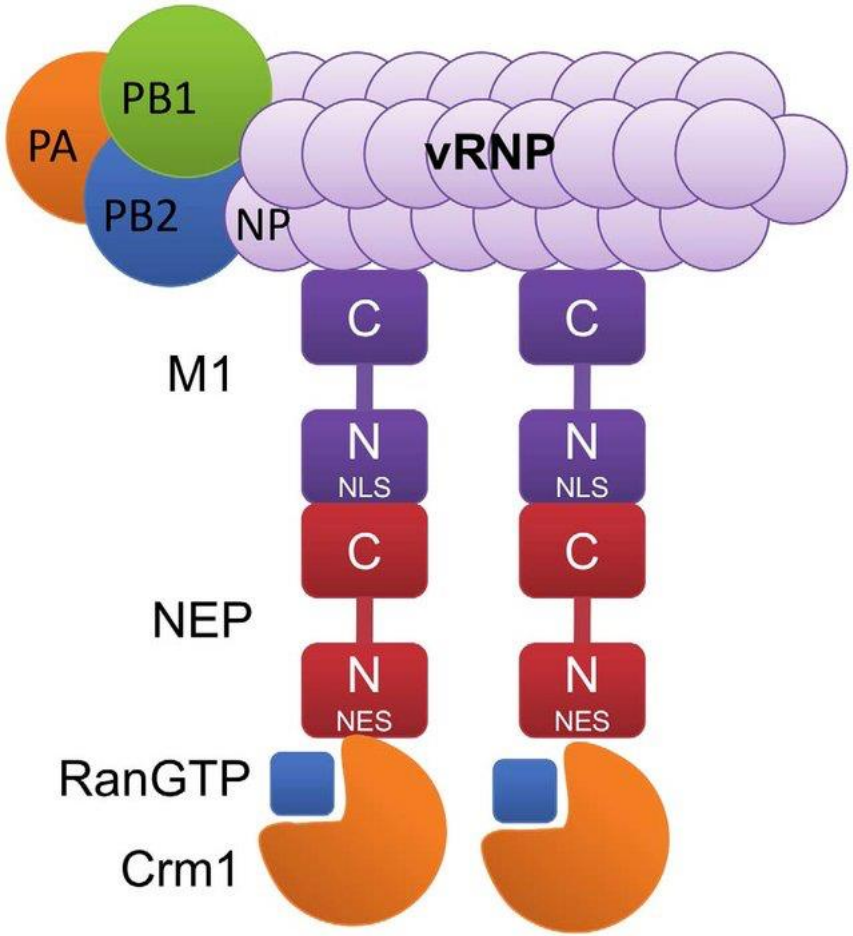
Ecco Staller,[a] Carol M. Sheppard,[a] Peter J. Neasham,[a] Bhakti Mistry,[a] Thomas P. Peacock,[a] Daniel H. Goldhill,[a] Jason S. Long,[a] Wendy S. Barclay[a]



Life Cycle


Cytoplasm
Virus entry
Endosome
H+
1
Endosomal escape
7
Particle assembly
6
vRNP transport
RAB11 vesicles
M2
PB2
M1
NEP
NA
HA
PB1
PA
NP
Viral protein transport
Translation
AAAAAAA
AAAAAAA
2
vRNP nuclear import
NPC
5
vRNP nuclear export
Incoming vRNPs
PB2
PB1
NP
PA
NEP
M1
mRNA export
3
Primary transcription
4
Genome replication
AAAAAAA
AAAAAAA
AAAAAAA
AAAAAAA
AAAAAAA
Viral mRNAs
Progeny vRNPs
Nucleus

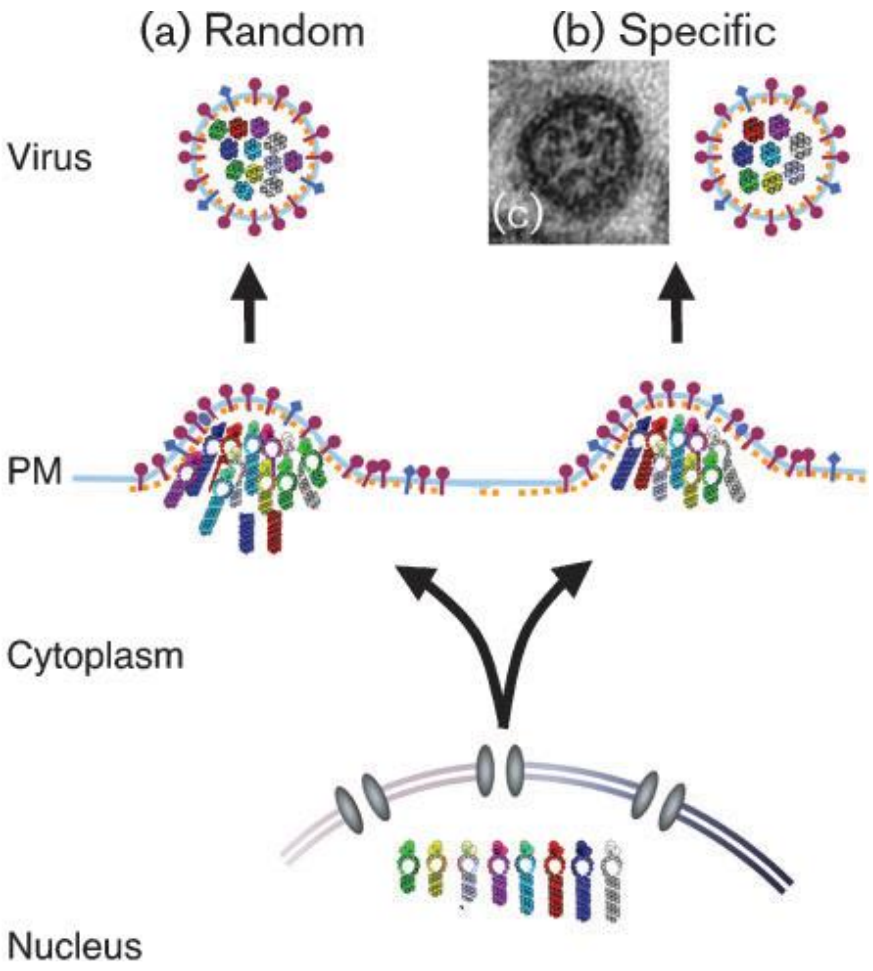
vRNP nuclear export



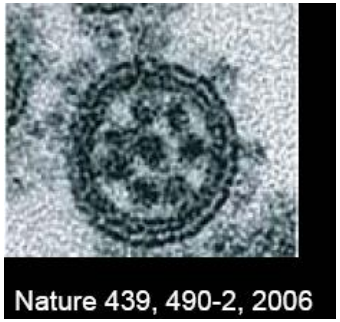
NEP-mediated nuclear export of influenza virus vRNPs. The b-importin Crm1 mediates export of the vRNP complex by binding to the N-terminal domain of NEP, as well as to its cofactor, the small GTPase Ran. The C-terminus of NEP binds to the nuclear localisation signal (NLS) on the N-terminal domain of the viral matrix protein M1. The C-terminus of M1 in turn binds strongly to the vRNP through interaction with NP

PLOS Pathogens | December 2012 | Volume 8 | Issue 12 | e1003019

Genome packaging

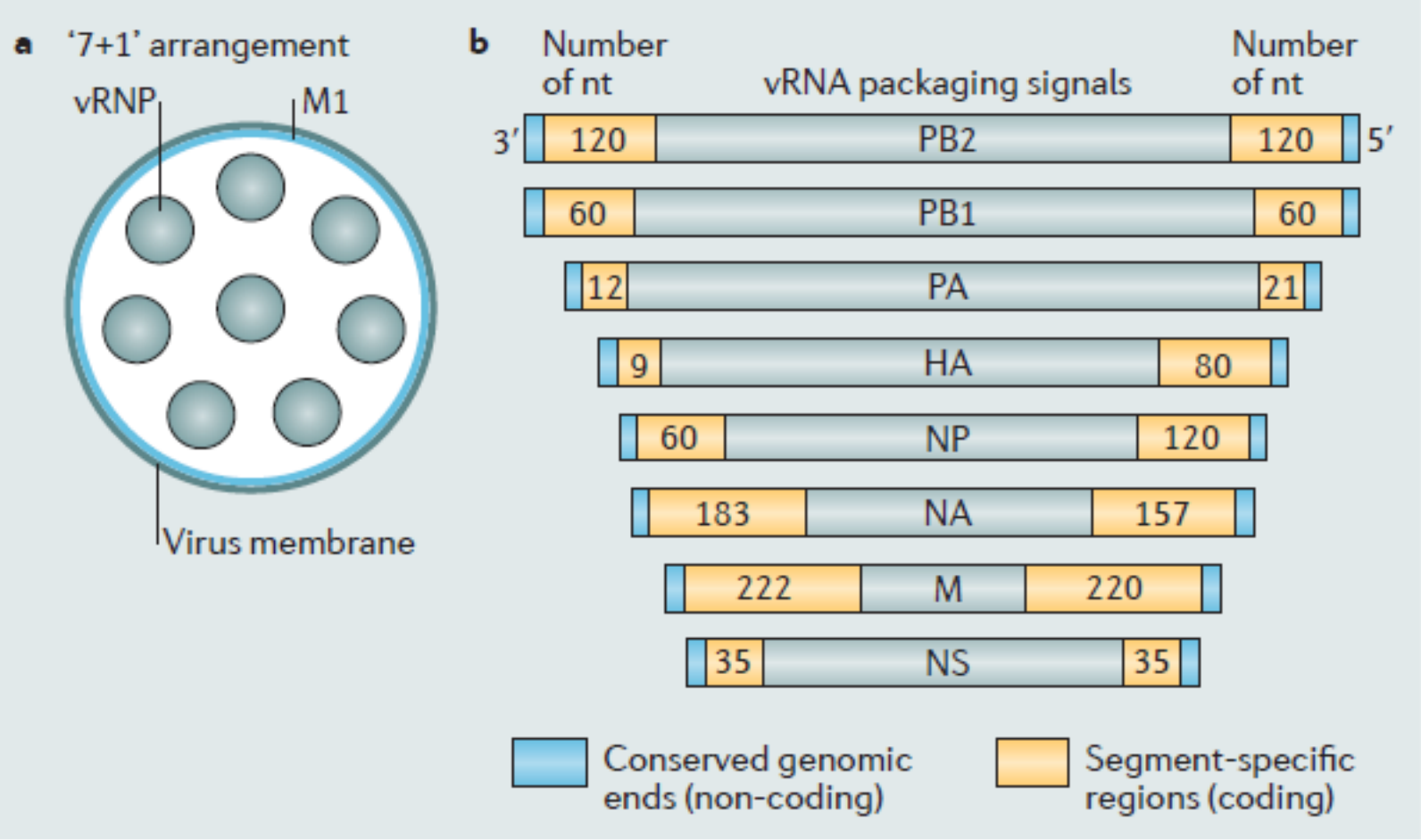


The eight individual segments (differentiated by colour) are replicated independently in the nucleus before being exported to the cytoplasm and migrating to the apical plasma membrane (PM). There, they interact with other viral structural proteins and new virus particles form by budding. (a) The random model for genome packaging proposes that more than eight RNPs are incorporated in a segment non-specific manner such that a reasonable proportion of virions contain at least one copy of each segment. (b) The specific model proposes that unique segment specific packaging signals operate to form a defined array of eight RNPs containing one copy of each segment. (c) A negatively stained EM section through an influenza virion showing the distinctive 7+1 array of RNPs.



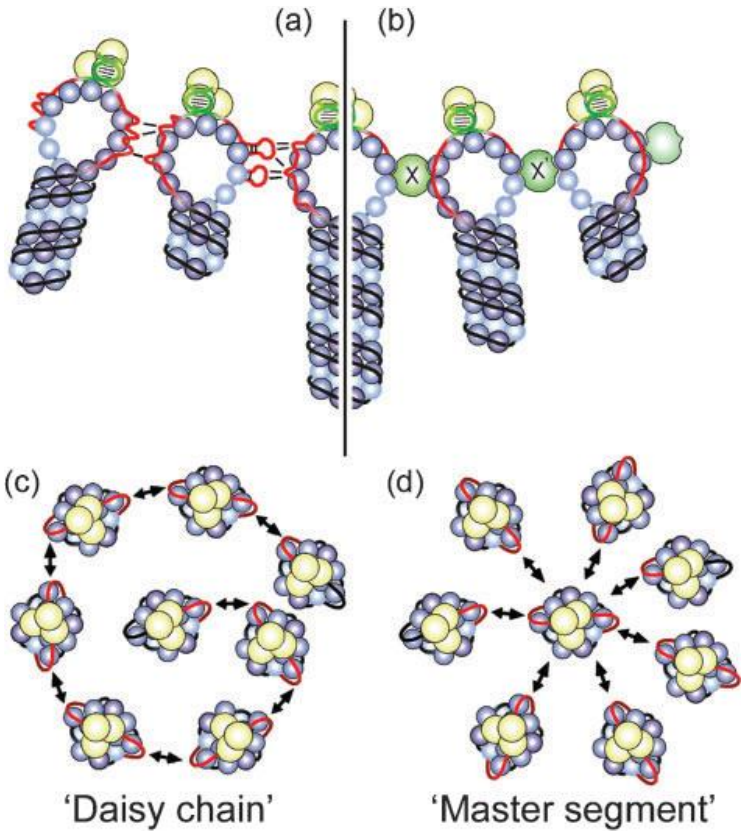
Journal of General Virology (2010), 91, 313–328

The packaging signals



Models of possible selective packaging methods for influenza A virus

It is proposed that the selective packaging of the segments is brought about by the assembly of a non-covalently linked higher-order genome complex, containing each of the eight vRNAs. The formation of this complex is mediated by specific interactions between the packaging signals of the segments, either (a) by direct RNA–RNA interactions between the packaging signals, possibly with the involvement of short secondary structures or (b) by as-yet-unidentified protein factors (X, X' etc).
(c, d) Possible organizations of the 7+1 genome complex utilizing the minimum possible number of between-segment interactions (arrows).



Journal of General Virology (2010), 91, 313–328

ARTICLE

OPEN

Selective flexible packaging pathways of the segmented genome of influenza A virus

Ivan Haralampiev[1,8,11], Simon Prisner[1,11], Mor Nitzan [2], Matthias Schade[1], Fabian Jolmes [1,9], Max Schreiber [3], Maria Loidolt-Krüger[4,9], Kalle Jongen[1], Jasmine Chamiolo[5], Niklaas Nilson[1], Franziska Winter [4,10], Nir Friedman [2,6], Oliver Seitz [5], Thorsten Wolff [7] ✉ & Andreas Herrmann [1] ✉

NATURE COMMUNICATIONS | (2020)11:4355 | https://doi.org/10.1038/s41467-020-18108-1 | www.nature.com/naturecommunications

The genome of influenza A viruses (IAV) is encoded in eight distinct viral ribonucleoproteins (vRNPs) that consist of negative sense viral RNA (vRNA) covered by the IAV nucleoprotein. Previous studies strongly support a selective packaging model by which vRNP segments are bundling to an octameric complex, which is integrated into budding virions. However, the pathway(s) generating a complete genome bundle is not known. We here use a multiplexed FISH assay to monitor all eight vRNAs in parallel in human lung epithelial cells. Analysis of 3.9 × 105 spots of colocalizing vRNAs provides quantitative insights into segment composition of vRNP complexes and, thus, implications for bundling routes. The complexes rarely contain multiple copies of a specific segment. The data suggest a selective packaging mechanism with limited flexibility by which vRNPs assemble into a complete IAV genome. We surmise that this flexibility forms an essential basis for the development of reassortant viruses with pandemic potential.

Li *et al. Virol J (2021) 18:36* https://doi.org/10.1186/s12985-021-01504-4

Packaging signal of influenza A virus

Xiuli Li, Min Gu, Qinmei Zheng, Ruyi Gao and Xiufan Liu*

Influenza A virus (IAV) contains a genome with eight single-stranded, negative-sense RNA segments that encode 17 proteins. During its assembly, all eight separate viral RNA (vRNA) segments are incorporated into virions in a selective manner. Evidence suggested that the highly selective genome packaging mechanism relies on RNA-RNA or protein- RNA interactions. The specific structures of each vRNA that contribute to mediating the packaging of the vRNA into virions have been described and identified as packaging signals. Abundant research indicated that sequences required for genome incorporation are not series and are varied among virus genotypes. The packaging signals play important roles in determining the virus replication, genome incorporation and genetic reassortment of influenza A virus. In this review, we discuss recent studies on influenza A virus packaging signals to provide an overview of their characteristics and functions.

Life cycle: budding

The new viral particles are gradually released by budding from the cell for a period of few hours. The cells are not lysed, but eventually die (due to interference with the normal synthesis of cellular macromolecules)

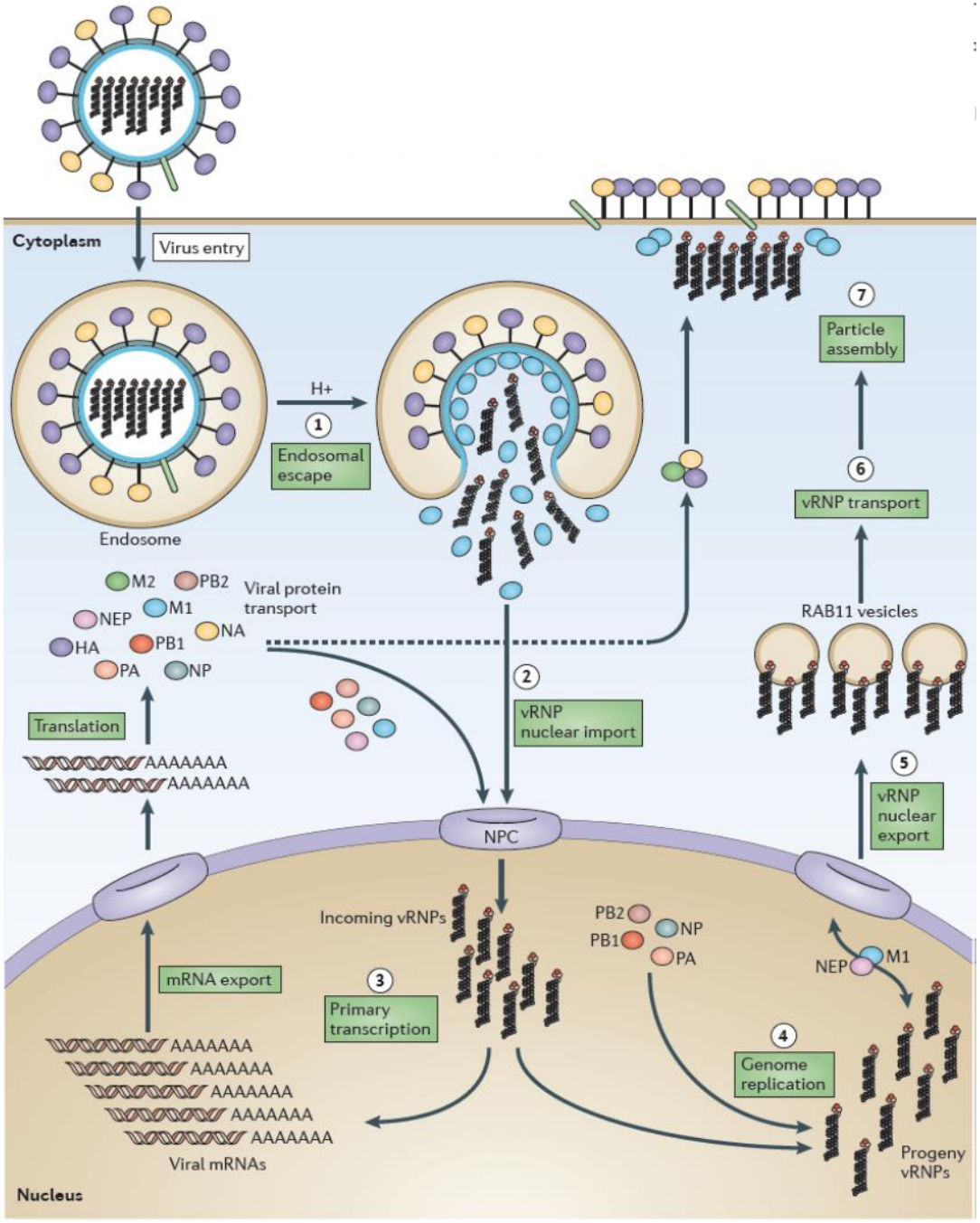
Nature **439**, 490-492(26 January 2006)

Life Cycle

Influenza A viruses (IAVs) predominantly enter cells by endocytosis after the viral haemagglutinin (HA) protein. Following internalization, endosomal acidification activates conformational changes in HA, which leads to fusion between the virion and endosomal membranes, providing the virus genome to the cytoplasm (step 1). The viral M2 ion channel concurrently promotes acidification of the virion interior, which dissociates the M1 matrix protein from the viral genome.

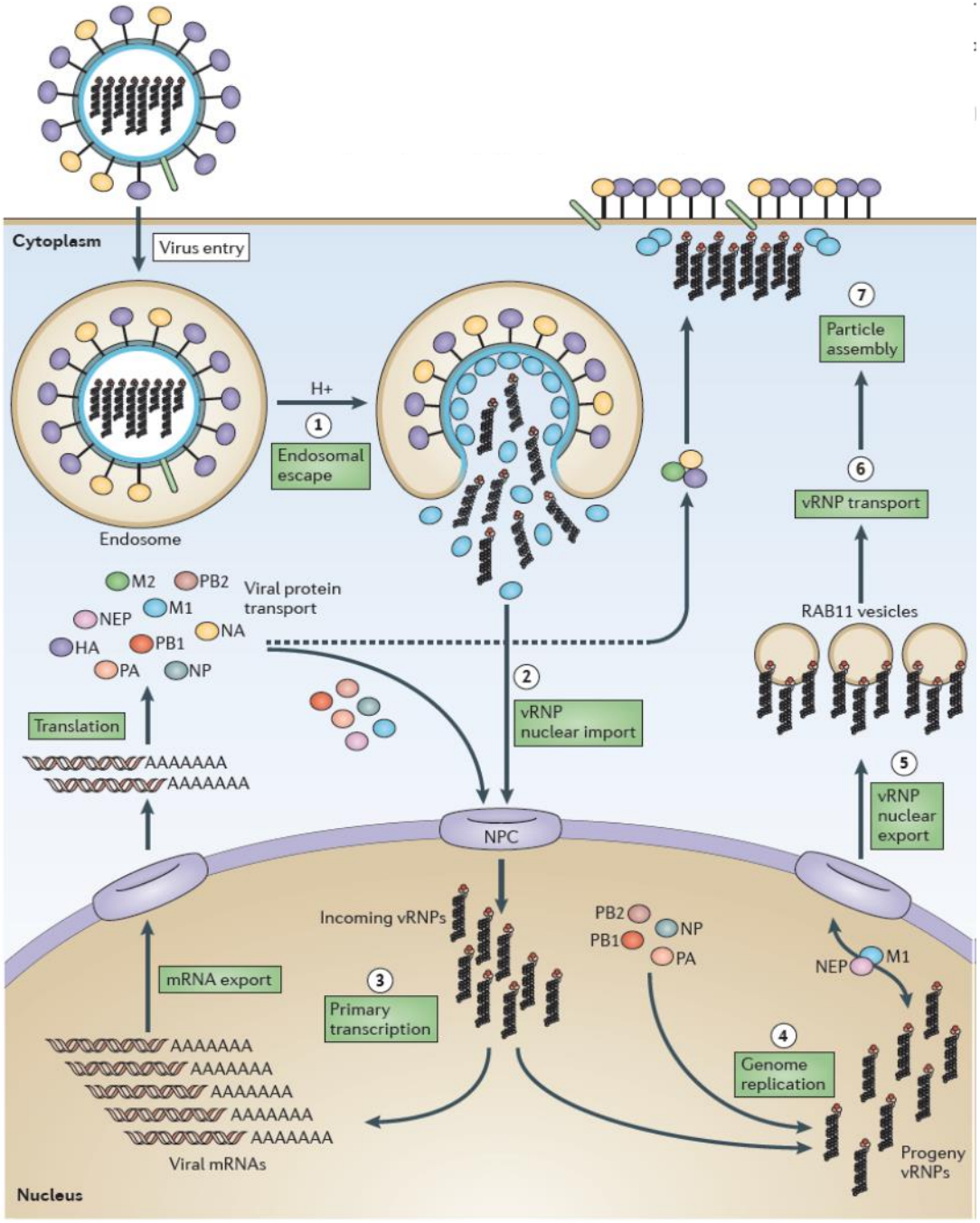
vRNPs that are released from endosomes are transported into the nucleus through the nuclear pore complex (NPC) (step 2),

and primary transcription results in the production of viral mRNAs, which are exported to the cytoplasm and translated into proteins by cellular ribosomes (step 3).

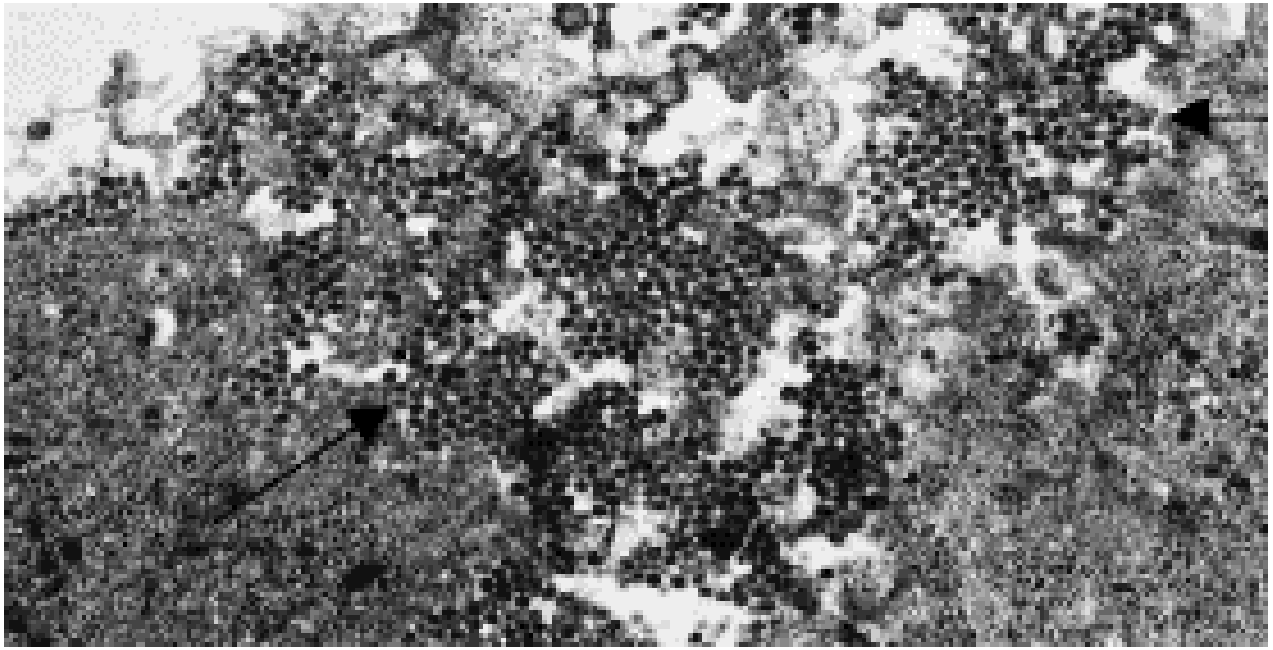


Life Cycle

Newly translated viral proteins are transported to the nucleus
After translation and nuclear entry of PB1, PB2, PA and NP, genome replication (that is, the production of progeny vRNPs) ensues (step 4). Progeny vRNPs are then exported to the cytoplasm with the assistance of the M1 and NEP proteins (step 5).
Newly exported vRNPs are subsequently trafficked to the plasma membrane on RAB11 vesicles (step 6), and vRNPs are incorporated into progeny virus particles containing HA, NA, M2 and M1 (step 7).
Virus release from the plasma membrane is mediated by the activities of at least two virion surface proteins, M2 and NA: M2 promotes scission of budding viruses from the plasma membrane, whereas NA prevents virus aggregation at the cell surface.



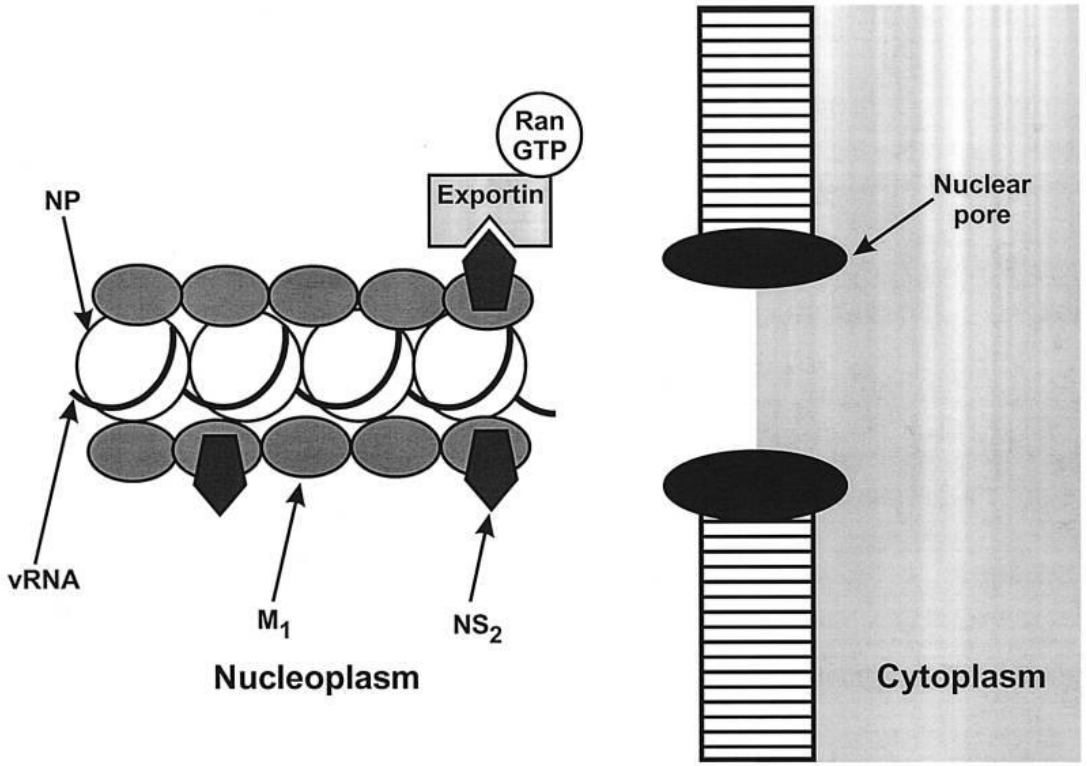
- L'attività enzimatica della Neuroaminidasi (NA) rimuove gli acidi neuroaminici (sialici) dalle catene di oligosaccaridi dei recettori prevenendo l'auto-aggregazione delle particelle della nuova progenie virale.

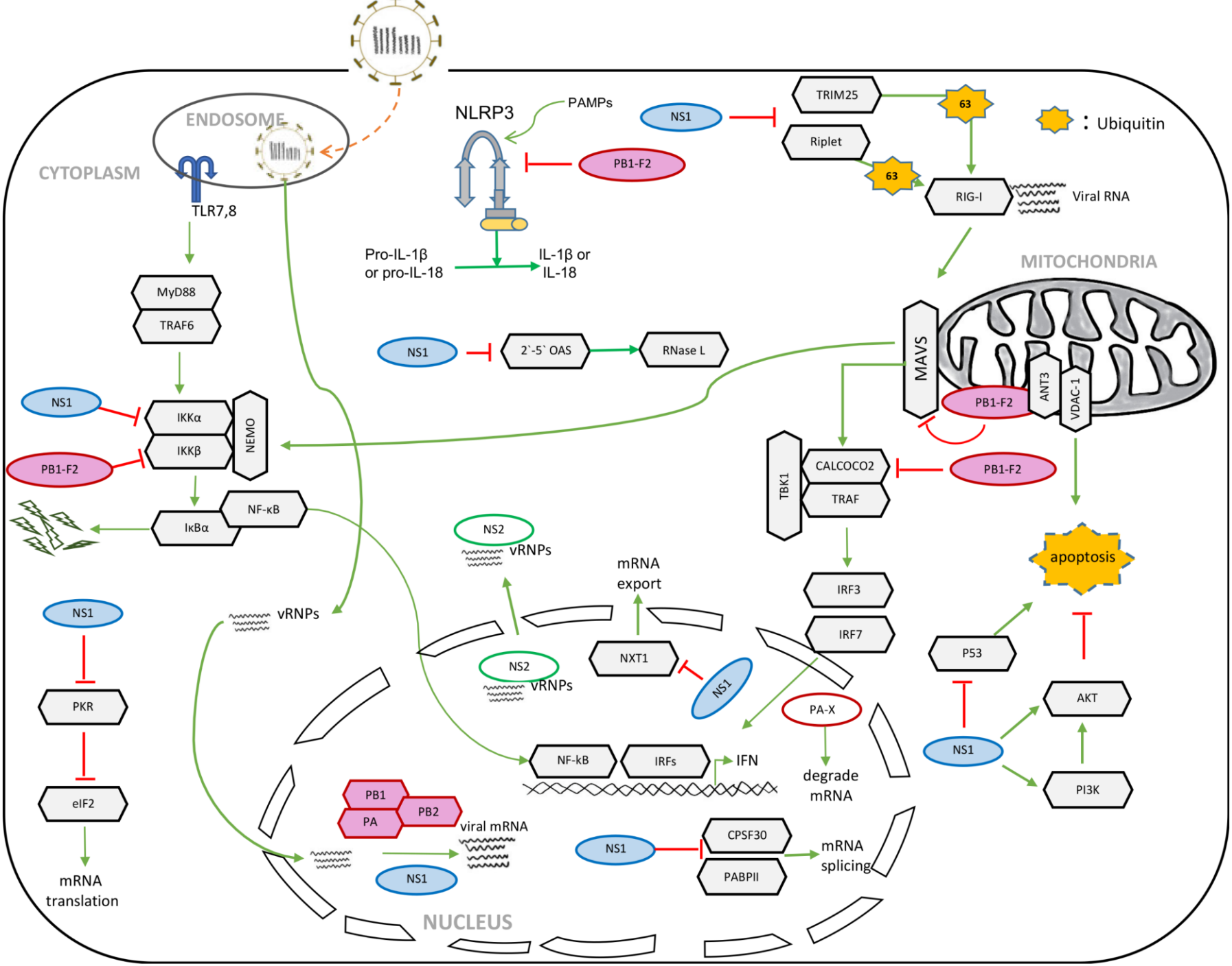


- Quando il virus viene propagato in presenza di inibitori della NA (Zanamivir and Oseltamivir (Tamiflu):, alla superficie della cellula infettata si formano aggregati della progenie virale.

vRNP nuclear export

- Un segnale di esporto nucleare in NS2 interagisce con un membro della famiglia delle esportine cellulari. Verosimilmente l'esportina è associata con la proteina RAN-GTP per mediare l'esporto del complesso RNP-M1-NS2 dal nucleo.




ENDOSOME
CYTOPLASM
TLR7,8
MyD88
TRAF6
NS1
IKKα
IKKβ
NEMO
PB1-F2
IκBα
NF-κB
NLRP3
PAMPs
PB1-F2
Pro-IL-1β
or pro-IL-18
IL-1β or
IL-18
NS1
2`-5` OAS
RNase L
TRIM25
63
Riplet
63
RIG-I
Viral RNA
: Ubiquitin
MITOCHONDRIA
MAVS
PB1-F2
ANT3
VDAC-1
TBK1
CALCOCO2
TRAF
PB1-F2
IRF3
IRF7
apoptosis
P53
AKT
NS1
PI3K
NS2
vRNPs
mRNA
export
NXT1
NS1
NS2
vRNPs
vRNPs
NS1
PKR
eIF2
mRNA
translation
PA-X
degrade
mRNA
NF-kB
IRFs
IFN
PB1
PB2
PA
viral mRNA
NS1
NS1
CPSF30
PABPII
mRNA
splicing
NUCLEUS