Gene therapy project

29/11/23 Project empowerment

Title Background Aim Materials and methods Results Discussion References Pitfall and solutions Budget

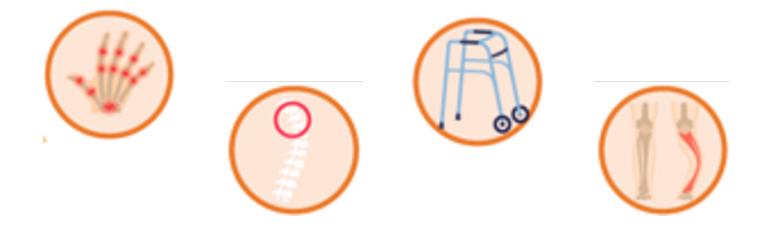
Gene therapy project

Theme I: Aging Group A: Bernardi, Ilie, Colonnelli, Bastianelli *Charcot marie tooth – pmp22* Group B: Hazrati, Bartolini, Glaudo, Montrone, Pourali *Werner syndrome*

Theme II: Cancer Group C: Belvedere, Jeong, Majaliwa, Virgilio *dCAS9 as a treatment for thyroid cancer*

Group D: Santacroce, Pace, Serra, Fanelli, Duarte *Hepatic cancer – RACGAP1*

Charcot Marie Tooth 1A



Silencing of PMP22 promoter 2 using a CRISPR/dCas9 combined with DNMT3A

Bastianelli, Bernardi, Colonnelli, Ilie

Background

Peripheral Nervous System Disease	Weakness in lower leg muscles, foot deformities ecc
Incidence of CMT	1/2500 \rightarrow 80% of which are type 1A
Development	Ageing related
<u>No resolutive</u> <u>treatment</u> available	Only palliative

Duplication of *PMP22*

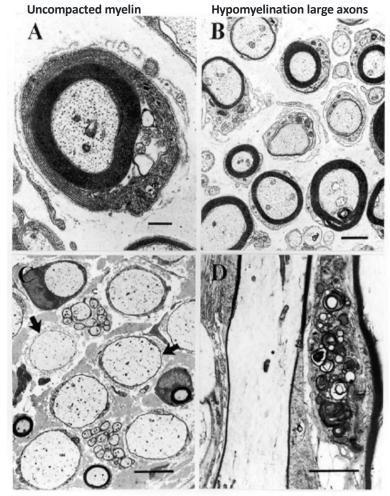
Overload of the Endoplasmic Reticulum (ER)

PMP22 aggregates

Onion Bulb formation

Dys and Demyelination

Secondary axonal degeneration



Thin myelin or lack (arrows)

Macrophage-demyelination

Huxley et al, Human Molecular Genetics, 1998.

Molecular basis of the disease

Aim of the project

Use of CRISPR-dCas9 associated with DNMT3A to perform an epigenetic silencing of the Promoter 2 of PMP22



C61 heterozygotic mice PMP22* Schwann cells

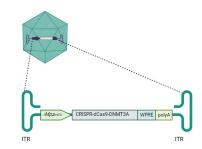
 \rightarrow Copy number of *PMP22:* 4

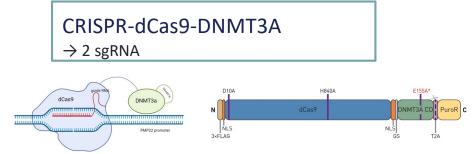
 \rightarrow MCV: 25 m/s

 \rightarrow Histology: mid demyelination

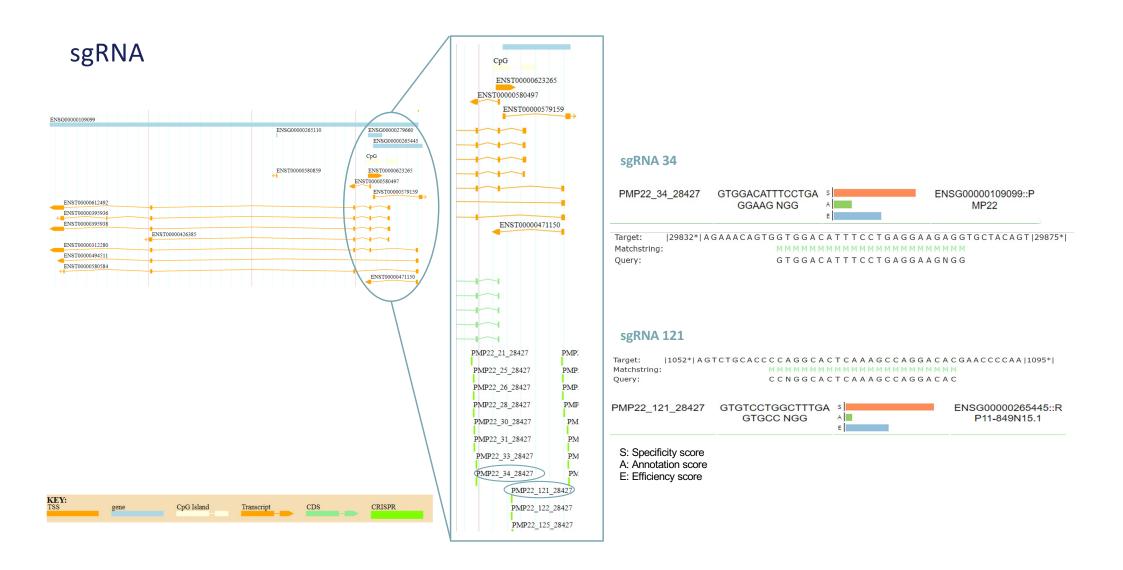
AAV 2/9

 \rightarrow MPZ promoter-SC specific \rightarrow Intrathecal lumbar injection

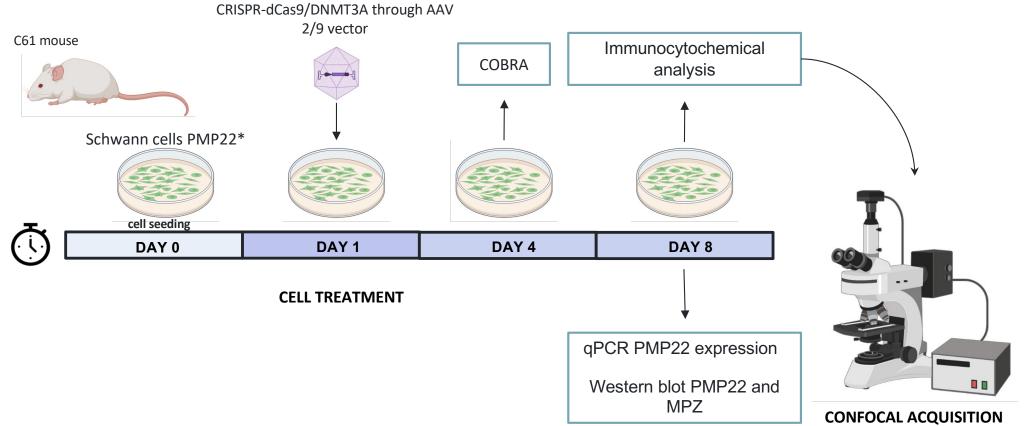




Adapted from Vojta et al. Nucleic acids research, 2016.



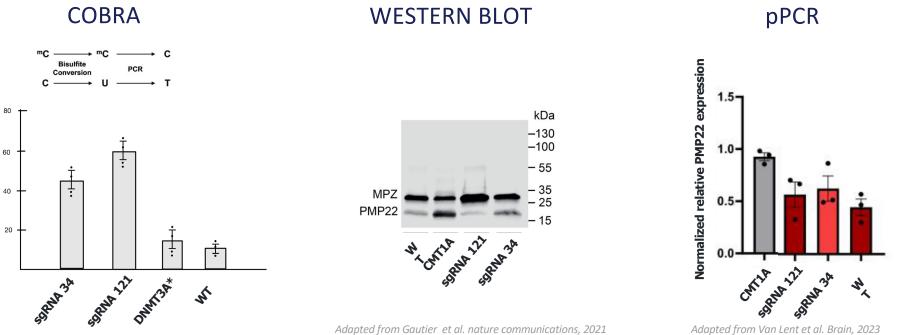
In vitro



RESULTS: in vitro

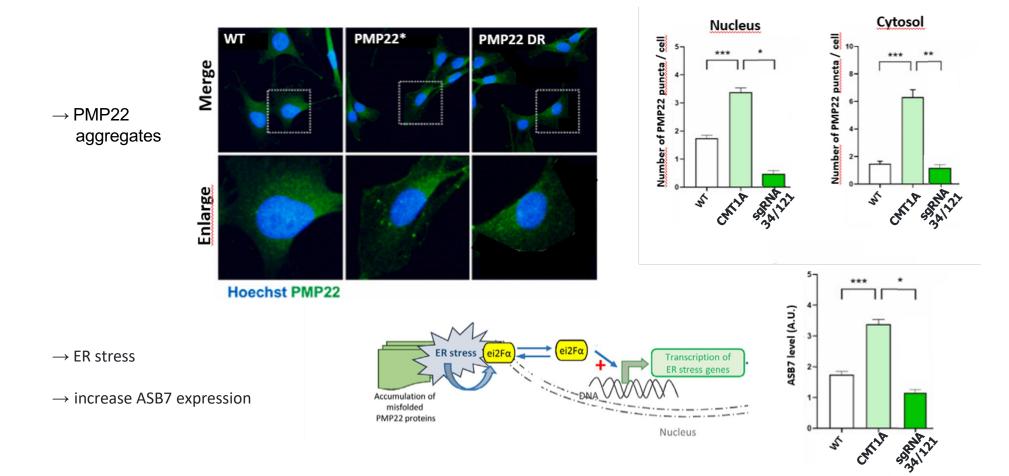
% methylation

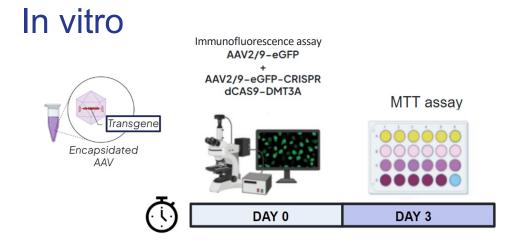
Does Methylation downregulate expression?



Adapted from Gautier et al. nature communications, 2021

Accumulation of *PMP22*

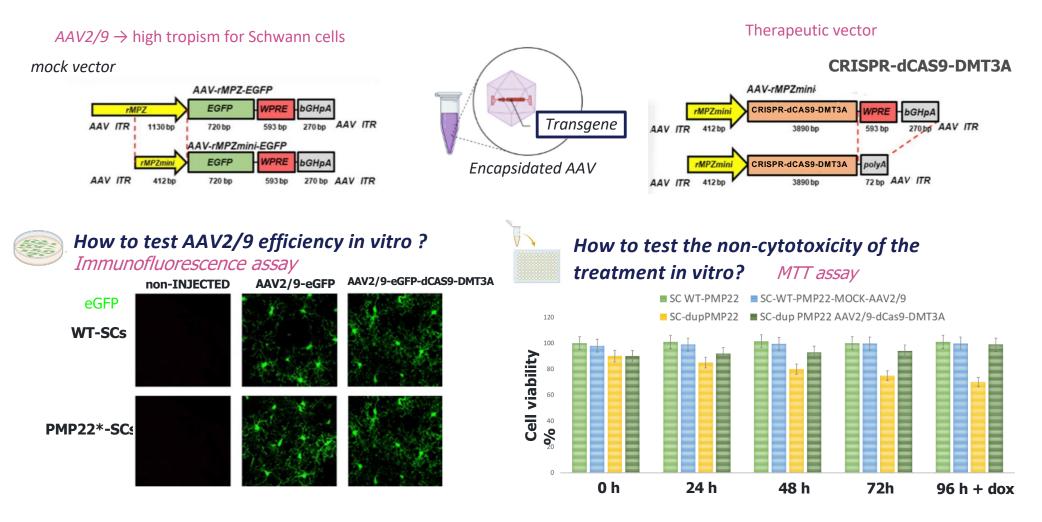




In vivo

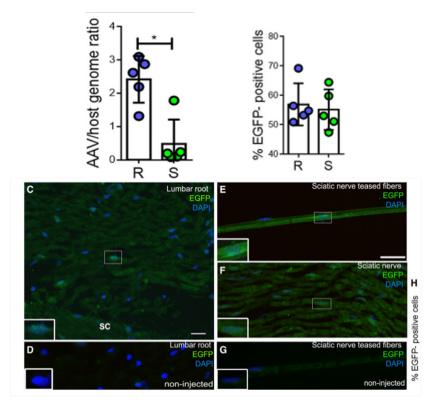
immunofluorescence	Histological onion bulb on axon	Western blot	EMG
assay AAV2/9-eGFP	CMT1A CMT1A	PMP22	
+ AAV2/9-eGFP-CRISPR dCAS9-DMT3A	Treated Treated	MPZ	
	ß	WT	comparison
	E (00)		
			00 -
5 WEEKS	DAY 1	DAY 3	up to 12 weeks
	AAV2/9-eGFP + AAV2/9-eGFP-CRISPR dCAS9-DMT3A	immunofluorescence assay AAV2/9-eGFP + AAV2/9-eGFP-CRISPR dCAS9-DMT3A	immunofluorescence assay AAV2/9-eGFP AAV2/9-eGFP-CRISPR dCAS9-DMT3A

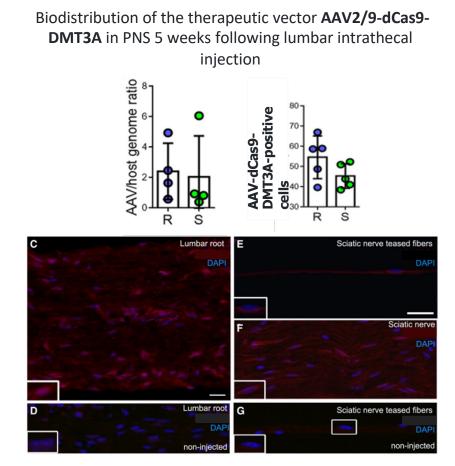
What is the system of delivery?



In vivo

Biodistribution of the mock **AAV2/9-EGFP** in PNS 5 weeks following lumbar intrathecal injection





Adapted from Georgiou E. et al., 2023 Molecular Therapy

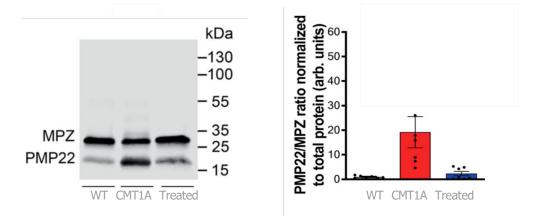
How to check the treatments efficiency?

WESTERN BLOT

Myelin protein zero (MPZ):

- \rightarrow expressed by Schwann cells
- \rightarrow main structural component of the myelin sheath.

Pmp22 was upregulated relative to the myelin marker Mpz in CMT1A, resulting in higher expression of Pmp22

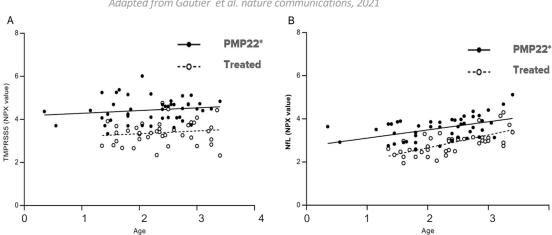


Adapted from Gautier et al. nature communications, 2021



High NPX value equals a high protein concentration. Circulating Biomarker:

- Nf-L (marker for axonal degeneration);
- TMPRSS5 (biomarker for myelinating).

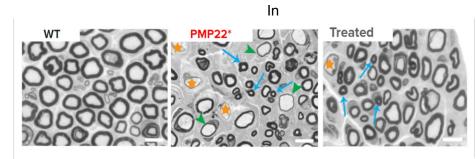


Δ

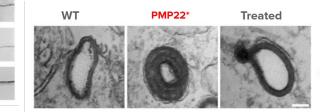
Adapted from Hongge Wang, et al. 2020

Histological

In CMT1A the PMP22 overexpression causes decreased myelination, recovery of axon myelination after treatment



karge demyelinated axons
 large hypomyelinated axons
 small hypermyelinated axons

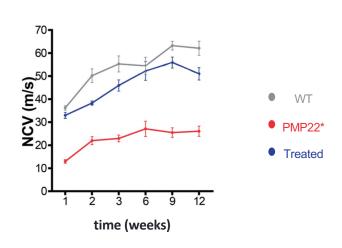


Adapted from Gautier et al. nature communications, 2021

EMG

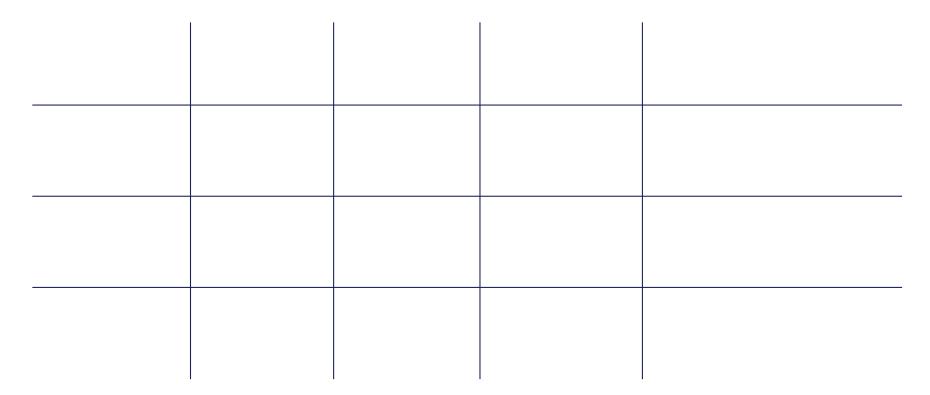
The loss of myelin in CMT1A causes a delay in impulse transmission

After the treatment we can see an axonal recovery of the impulse, due to the correct reformation of the myelin



Adapted from Gautier et al. nature communications, 2021

Budget



Thank you

& images by **Freepik**

References

- Kagiava A, Richter J, Tryfonos C, Leal-Julià M, Sargiannidou I, Christodoulou C, Bosch A, Kleopa KA. Efficacy of AAV serotypes to target Schwann cells after intrathecal and intravenous delivery. Sci Rep. 2021 Dec 2;11(1):23358. doi: 10.1038/s41598-021-02694-1.

-Marina Stavrou , Kleopas A. Kleopa: CMT1A current gene therapy approaches and promising biomarkers. N.R.R. 2022 Nov 25; doi: 10.4103/1673-5374.361538.

-Hongge Wang, Matthew Davison et al. **Transmembrane protease serine 5: a novel Schwann cell plasma marker for CMT1A.** ANA.2020; 7(1): 69–82. doi: 10.1002/acn3.50965 -Boe SG, Antonowicz NM, Leung VW et al. (2010). High inter-rater reliability in analyzing results of decomposition based quantitative electromyography in subjects with or without neuromuscular disorder. J Neurosci Methods 1992: 138–145

-Jennifer A. TracyPeter J. Dyck[...]P. James B. Dyc. Onion-bulb patterns predict acquired or inherited demyelinating polyneuropathy Muscle and Nerve (2019). doi:10.1002/mus.26452 -Bilichak, Andriy, and Igor Kovalchuk. "The Combined Bisulfite Restriction Analysis (COBRA) Assay for the Analysis of Locus-Specific Changes in Methylation Patterns." In Plant Epigenetics, edited by Igor Kovalchuk, 1456:63–71. Boston, MA: Springer US, 2017. https://doi.org/10.1007/978-1-4899-7708-3_5.

-Li, Jun, Brett Parker, Colin Martyn, Chandramohan Natarajan, and Jiasong Guo. "The PMP22 Gene and Its Related Diseases." Molecular Neurobiology 47, no. 2 (April 2013): 673–98. https://doi.org/10.1007/s12035-012-8370-x.

-Moore, Lisa D, Thuc Le, and Guoping Fan. "DNA Methylation and Its Basic Function." Neuropsychopharmacology 38, no. 1 (January 2013): 23–38. https://doi.org/10.1038/npp.2012.112. -Park, Hanseul, Jaein Shin, Yunkyung Kim, Takashi Saito, Takaomi C. Saido, and Jongpil Kim. "CRISPR/DCas9-Dnmt3a-Mediated Targeted DNA Methylation of APP Rescues Brain Pathology in a Mouse Model of Alzheimer's Disease." Translational Neurodegeneration 11, no. 1 (September 15, 2022): 41. https://doi.org/10.1186/s40035-022-00314-0.

-Stavrou, Marina, and KleopasA Kleopa. "CMT1A Current Gene Therapy Approaches and Promising Biomarkers." Neural Regeneration Research 18, no. 7 (2023): 1434. https://doi.org/10.4103/1673-5374.361538.

-Van Lent, Jonas, Leen Vendredy, Elias Adriaenssens, Tatiana Da Silva Authier, Bob Asselbergh, Marcus Kaji, Sarah Weckhuysen, Ludo Van Den Bosch, Jonathan Baets, and Vincent -Timmerman. "Downregulation of PMP22 Ameliorates Myelin Defects in IPSC-Derived Human Organoid Cultures of CMT1A." Brain 146, no. 7 (July 3, 2023): 2885–96. https://doi.org/10.1093/brain/awac475. -Vojta, Aleksandar, Paula Dobrinić, Vanja Tadić, Luka Bočkor, Petra Korać, Boris Julg, Marija Klasić, and Vlatka Zoldoš. "Repurposing the CRISPR-Cas9 System for Targeted DNA Methylation." Nucleic Acids Research 44, no. 12 (July 8, 2016): 5615–28. <u>https://doi.org/10.1093/nar/gkw159</u>.

-Georgiou E, Kagiava A, Sargiannidou I, Schiza N, Stavrou M, Richter J, Tryfonos C, Heslegrave A, Zetterberg H, Christodoulou C, Kleopa KA. AAV9-mediated SH3TC2 gene replacement therapy targeted to Schwann cells for the treatment of CMT4C. Mol Ther. 2023 Nov 1;31(11):3290-3307. doi: 10.1016/j.ymthe.2023.08.020. Epub 2023 Aug 28. PMID: 37641403; PMCID: PMC10638072. -Gao, F.; Zhang, Y.; Wu, D.; Luo, J.; Gushchina, S.; Bo, X. Combination of Engineered Expression of Polysialic Acid on Transplanted Schwann Cells and in Injured Rat Spinal Cord Promotes Significant Axonal Growth and Functional Recovery. *Neuroglia* **2023**, *4*, 222-238. <u>https://doi.org/10.3390/neuroglia4040016</u>

-Guo C, Ma X, Gao F, Guo Y. Off-target effects in CRISPR/Cas9 gene editing. Front Bioeng Biotechnol. 2023 Mar 9;11:1143157. doi: 10.3389/fbioe.2023.1143157. PMID: 36970624; PMCID: PMC10034092.

-Gautier, Benoit, Helene Hajjar, Sylvia Soares, Jade Berthelot, Marie Deck, Scarlette Abbou, Graham Campbell, et al. "AAV2/9-Mediated Silencing of PMP22 Prevents the Development of Pathological Features in a Rat Model of Charcot-Marie-Tooth Disease 1 A." Nature Communications 12, no. 1 (April 21, 2021): 2356. <u>https://doi.org/10.1038/s41467-021-22593-3</u>.

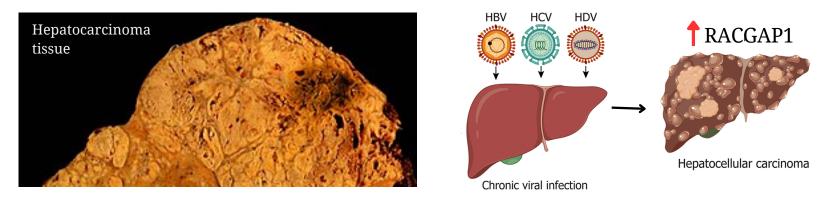
RACGAP1 competitive inhibition in hepatocellular carcinoma via vector based mRNA transfection

Lavinia Pace Miriana Santacroce Luigi Fanelli

Antonio Duarte

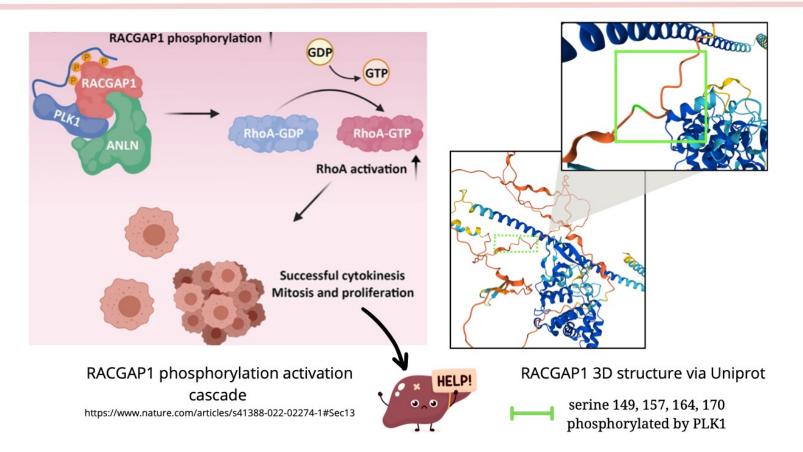
Ernest Serra

Background: panoramic on Hepatocellular Carcinoma (HCC)



- Liver cancer is the **third most lethal cancer** globally. **Infection by hepatitis B****C** viruses is the main risk factor for HCC development
- The median age: > **60** years
- HCC **recurrence** is significantly associated with **RACGAP1 upregulation**: activation of RACGAP1/Rho/ERK signaling axis

Background: RACGAP1 pathway



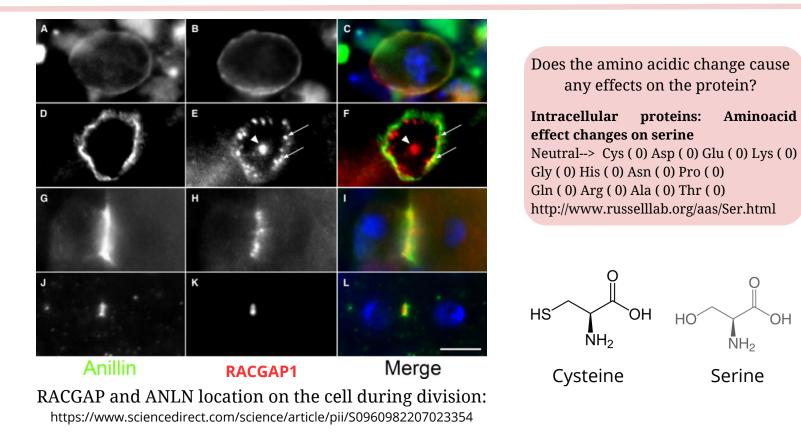
Aim of the project

- Induce a competitive inhibition of RACGAP1 by mutating its phosphorylation sites
- Leading to a reduced activation of RhoA
- Inhibition of self proliferation

Experimental plan In silico • Sequence analysis In vitro • Mutation of 4 serines into cysteins • Cloning of mRNA seq • Sequence alignment In vivo • Transfection into HCCLM3 cells • Structure analysis • mRNA expression • Co-ChIP for complex formation • Tumor mass analysis • Levels of phosphorylation • Tumorigenesis assay RhoA activation

• Apoptosis and cell cycle analysis

Where are RACGAP1 and ANLN located?

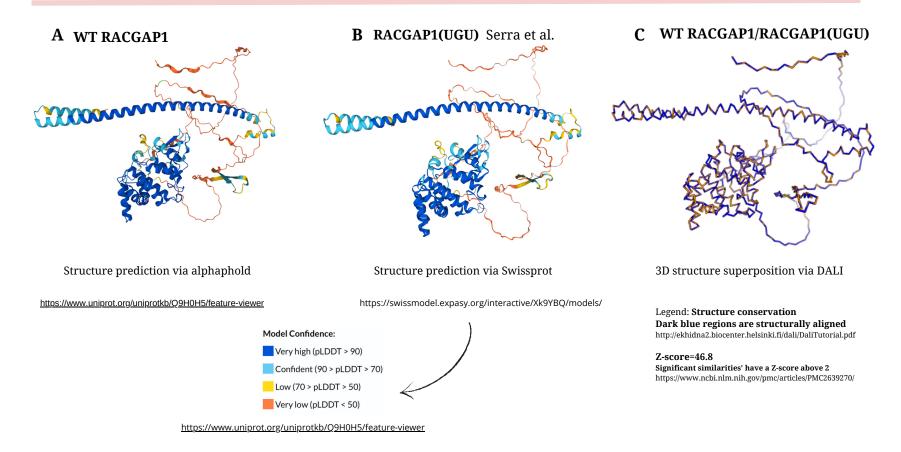


OH

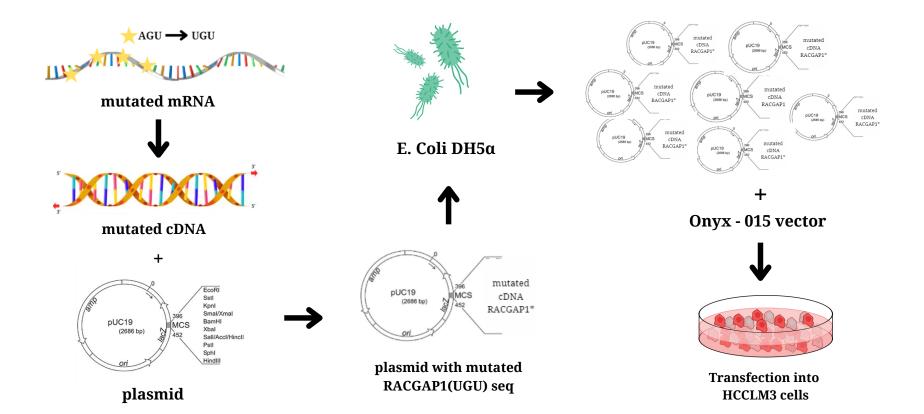
AMINOACID MODIFICATIONS - sequence

WT modified	AETERSALDVKLKHARNQVDVEI AETERSALDVKLKHARNQVDVEI *******	KRRQRAEADCEK	LERQIQLIRE	EMLMCDTSGSIQI	LSEE 119		
WT modified	QKSALAFLNRGQPSSSNAGNKRL QKSALAFLNRGQPSSSNAGNKRL ******	STIDE <mark>C</mark> GSILSD	ICFDKTDECI	LDWDS <mark>C</mark> LVKTFKI	LKKR 179		
WT modified	EKRRSTSRQFVDGPPGPVKKTRSIGSAVDQGNESIVAKTTVTVPNDGGPIEAVSTIETVP24EKRRSTSRQFVDGPPGPVKKTRSIGSAVDQGNESIVAKTTVTVPNDGGPIEAVSTIETVP23***********************************						
	MSA using ClustalW from ebi https://www.ebi.ac.uk						
			150	160	170		
RACGAP1_HUN	AN_MODIFIED_SEQUENCE	LSTIDEC	ĠSILSĎI	CFDKTDEC			
S	p Q9H0H5 RGAP1_HUMAN	LSTIDES	GSILSDI	SFDKTDES	LDWDSSLV		
Number of conserved phys properties /10 https://www.jalview.org/help/html/calcular	K	* * * * * * 7	* * * * * * *	• 7 * * * * * * 7	* * * * * 7 * *		
ml		MSA using T-	coffe via Jalv	iew			

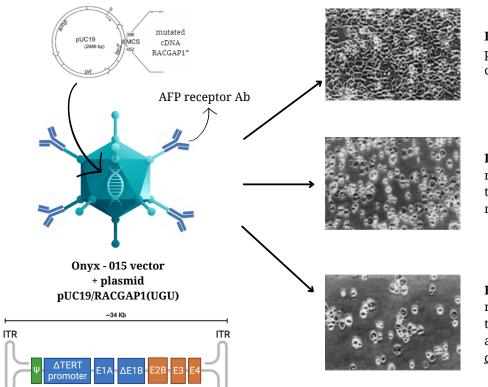
Aminoacid modifications - structural predictions



Cloning and transfection of RACGAP(UGU) mRNA



What happens to the cells using ONYX-015?

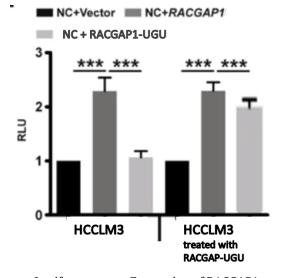


Healthy liver cells p53 wt and no alpha-feto protein receptor. The vector doesn't enter the cells and <u>no healthy liver cells</u> are <u>killed</u>

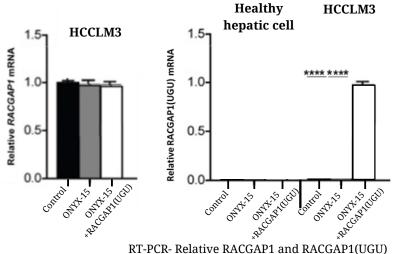
HCC cells p53 wt and with alpha-feto protein receptor on the cell surface. The vector enters the cells and delivers the RACGAP1(UGU) mRNA. The <u>cells die by cytokinetic failure</u>

HCC cells Δ p53 and with alpha-feto protein receptor on the cell surface. The vector enters the cells, delivers the RACGAP1(UGU) mRNA and activates the Δ p53 onyx pathway. The <u>cells</u> <u>die</u> by both <u>cytokinetic failure</u> and <u>cell lysis</u>

In vitro: Is RACGAP1(UGU) expressed?

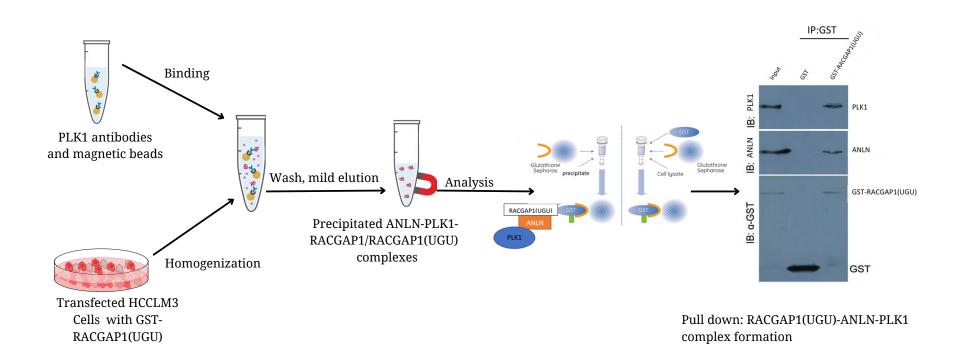


Luciferase essay- Expression of RACGAP1 and RACGAP1(UGU) in wt HCCLM3 cells and HCCLM3 cells after transfection with RACGAP1(UGU) mRNA Adapted from https://www.nature.com/articles/s41419-019-1666-2



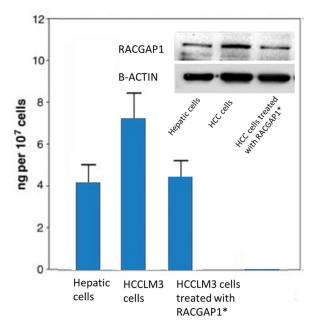
RI-PCR- Relative RACGAP1 and RACGAP1(UGU mRNA level in HCCLM3. The expression of RACGAP1 and RACGAP1(UGU) is not influeced by the presence of the other

In vitro: Does the RACGAP1(UGU)/ANLN/PLK1 complex form?

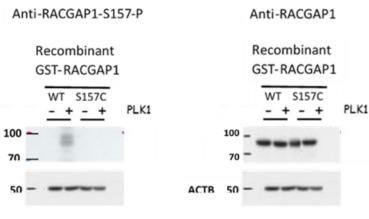


In vitro: Is RACGAP1(UGU) phosphorylated?

ACTB

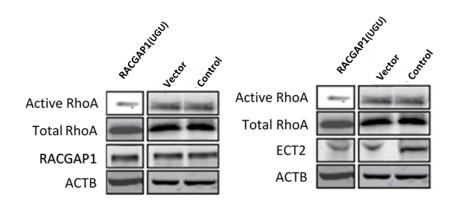


Phosphorylation assay - ELISA and Western Blot -Normal levels of RACGAP1P in healthy cells, elevetated levels of RACGAP1P in HCCLM3, reduced levels of RACGAP1P(UGU) in HCCLM3

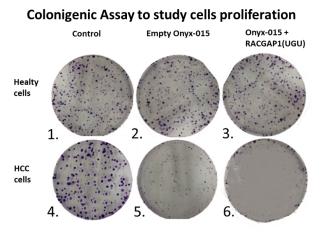


Anti-Phospho Ser157 of GST-RACGAP1 in presence of RACGAP1 (WT) or RACGAP1 (S157C) Anti-RACGAP1 of GST-RACGAP1 in presence of RACGAP1 (WT) or RACGAP1 (S157C)

In vitro: Is RohA activity decreased?



Western blot- Detection of RhoA activity and also ECT2 and RACGAP1 expression in HCCLM3 after trasfection of RACGAP1(UGU) Adapted from (Yang et al., 2018)



Clonogenic assay:

1. Healthy Hepatic cells,

2. Healthy Hepatic cells with transfection of empty Onyx-015,

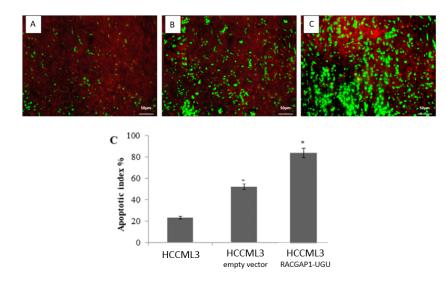
3. Healthy Hepatic cells with transfection of RACGAP1(UGU) mutated protein,

4. Hepatocarcinoma HCCLM3 cells,

5. Hepatocarcinoma HCCLM3 with transfection of empty Onyx-015,

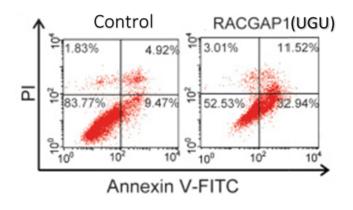
6. Hepatocarcinoma HCCLM3 with transfection of RACGAP1(UGU) mutated protein,

In vitro: Does RACGAP1(UGU) cause apoptosis?



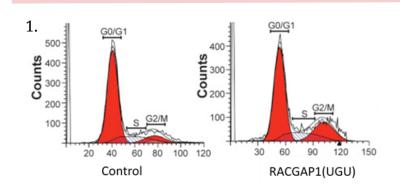
TUNEL assay - A. HCCLM3 non treated and no apoptotic cells are detected. B. HCCLM3 treated with emptyvector, no apoptotic cells are detected C. HCCLM3 treated with the mutated RACGAP1(UGU), incresead levels of apoptotic cells

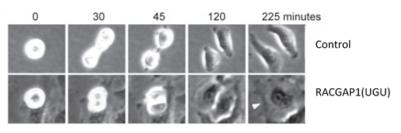
Adapted from https://www.researchgate.net/publication/335679404_In_Vivo_Anti-Tumor_Effects_of_Citral_on_4T1_Breast_Cancer_Cells_via_Induction_of_Apoptosis_and_Downregulation_of_Aldehyde_Dehyd romansae_Artivity



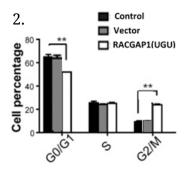
Flow cytometry analysis- Staining cells with the apoptosis marker Annexin V (FITC) and propidium iodide allows the discrimination of intact cells (FITC-PI-), early apoptotic (FITC+PI-) and late apoptotic or necrotic cells (FITC+PI+). Adapted from (Yang et al., 2018)

In vitro: Does RACGAP1(UGU) cause cytokinesis failure?

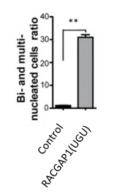




Cytokinesis analysis - Selected frames from time-lapse imaging of RACGAP1(UGU) and control HCCLM3 cells.



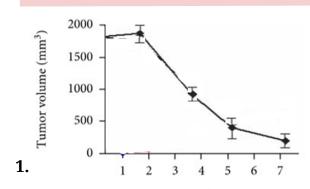
Cell cycle analysis - 1. Cell count in different cell cycle phases, RACGAP1(UGU) vs control HCCLM3 cells. 2. Cell percentage in different cell cycle pahses, RACGAP1(UGU) vs control HCCLM3 cells.



Statistics of bi- and multi-nucleated HCCLM3 cells after trasfection

Adaptedfrom (Yang et al., 2018)

In vivo: Is there a tumor mass decrease?

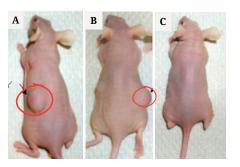






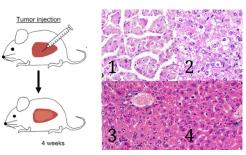
 Decrease of tumor mass during weeks with the treatment of RACGAP1(UGU) on BALB/c nude mice
 Representative images of tumors

removed from mice https://www.hindawi.com/journals/omcl/2022/3034150/



A. Nude mouse injected with HCCLM3 cells. Evident tumor mass. B. Nude mouse injected with HCCLM3 cells transfected with empty vector, smaller tumor mass C. Nude mouse injected with HCCLM3 cells transfected with RACGAP1(UGU) vector. No evidence of tumor growth.

Adapted from https://bmccancer.biomedcentral.com/articles/10. 1186/1471-2407-11-425/figures/7



HCC histological samples from BALB/c nude mouse

1. HCC tissue

2. Tissue sample injected with empty vecotr

3. Tissue sample injected with

RACGAP1(UGU) vector

4. Healthy liver tissue https://translationalmedicine.biomedcentral.com/articles/10.1186/s12967-017-1247z/figures/4

2.

REFERENCES

Chen, J., Li, Z., Jia, X., Song, W., Wu, H., Zhu, H., Xuan, Z., Du, Y., Zhu, X., Song, G., Dong, H., Bian, S., Wang, S., Zhao, Y., Xie, H., Zheng, S., Song, P., 2022. Targeting anillin inhibits tumorigenesis and tumor growth in hepatocellular carcinoma via impairing cytokinesis fidelity. Oncogene 41, 3118–3130. https://doi.org/10.1038/s41388-022-02274-1

RACGAP1 - Rac GTPase-activating protein 1 - Homo sapiens (Human) | Feature viewer | UniProtKB | UniProt [WWW Document], n.d. URL https://www.uniprot.org/uniprotkb/Q9H0H5/feature-viewer (accessed 11.29.23).

Serine [WWW Document], n.d. URL http://www.russelllab.org/aas/Ser.html (accessed 11.29.23).

Institute, E.B., n.d. EMBL-EBI homepage [WWW Document]. URL https://www.ebi.ac.uk/ (accessed 11.29.23).

Holm, L., Kääriäinen, S., Rosenström, P., Schenkel, A., 2008. Searching protein structure databases with DaliLite v.3. Bioinformatics 24, 2780–2781. https://doi.org/10.1093/bioinformatics/btn507

Gregory, S.L., Ebrahimi, S., Milverton, J., Jones, W.M., Bejsovec, A., Saint, R., 2008. Cell Division Requires a Direct Link between Microtubule-Bound RacGAP and Anillin in the Contractile Ring. Current Biology 18, 25–29. https://doi.org/10.1016/j.cub.2007.11.050

https://www.google.com/url?

sa=t&rct=j&q=&esrc=s&source=web&cd=&cad=rja&uact=8&ved=2ahUKE wiyv7vCtNeCAxUiVPEDHRnQDoUQFnoECBkQAQ&url=https%3A%2F%2F www.spandidos-

publications.com%2Fmmr%2F3%2F4%2F589%2Fdownload&usg=AOvVa w0ZL3ZXEycqcznFa7YRgGL7&opi=89978449

Kageyama, K., Ohara, M., Saito, K., Ozaki, S., Terai, M., Mastrangelo, M.J., Fortina, P., Aplin, A.E., Sato, T., 2017. Establishment of an orthotopic patient-derived xenograft mouse model using uveal melanoma hepatic metastasis. J Transl Med 15, 1–19. https://doi.org/10.1186/s12967-017-1247-z

464. Humanized AAV Vectors: Attaching a Human Protein to the Surface of AAV Vectors To Reduce Binding of Neutralizing and Opsonizing Antibodies, 2009. . Molecular Therapy 17, S180–S181. https://doi.org/10.1016/S1525-0016(16)38822-0 Jiang, F., Liu, T., He, Y., Yan, Q., Chen, X., Wang, H., Wan, X., 2011. MiR-125b promotes proliferation and migration of type II endometrial carcinoma cells through targeting TP53INP1 tumor suppressor in vitro and in vivo. BMC Cancer 11, 425. https://doi.org/10.1186/1471-2407-11-425

Wang, M.-Y., Chen, D.-P., Qi, B., Li, M.-Y., Zhu, Y.-Y., Yin, W.-J., He, L., Yu, Y., Li, Z.-Y., Lin, L., Yang, F., Lin, Z.-R., Liu, J.-Q., 2019. Pseudogene RACGAP1P activates RACGAP1/Rho/ERK signalling axis as a competing endogenous RNA to promote hepatocellular carcinoma early recurrence. Cell Death Dis 10, 1–12. https://doi.org/10.1038/s41419-019-1666-2

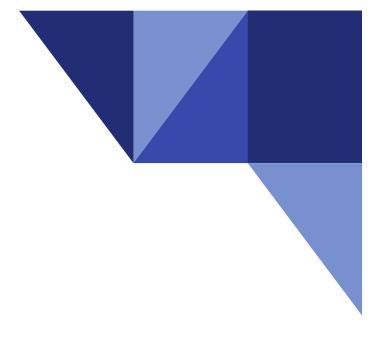
Anti-Phospho RacGAP1 (Ser157), Polyclonal [pS157 HsCyk-4] [WWW Document], n.d. . Ximbio. URL https://ximbio.com/reagent/151587/anti-phospho-racgap1-ser157-polyclonal-ps157-hscyk-4 (accessed 11.29.23).

Rahman, H., 2019. In Vivo Anti-Tumor Effects of Citral on 4T1 Breast Cancer Cells via Induction of Apoptosis and Downregulation of Aldehyde Dehydrogenase Activity. Molecules. Shi, D.-M., Dong, S.-S., Zhou, H.-X., Song, D.-Q., Wan, J.-L., Wu, W.-Z., 2023. Genomic and transcriptomic profiling reveals key molecules in metastatic potentials and organ-tropisms of hepatocellular carcinoma. Cellular Signalling 104, 110565. https://doi.org/10.1016/j.cellsig.2022.110565

Yang, X.-M., Cao, X.-Y., He, P., Li, J., Feng, M.-X., Zhang, Y.-L., Zhang, X.-L., Wang, Y.-H., Yang, Q., Zhu, L., Nie, H.-Z., Jiang, S.-H., Tian, G.-A., Zhang, X.-X., Liu, Q., Ji, J., Zhu, X., Xia, Q., Zhang, Z.-G., 2018. Overexpression of Rac GTPase Activating Protein 1 Contributes to Proliferation of Cancer Cells by Reducing Hippo Signaling to Promote Cytokinesis. Gastroenterology 155, 1233-1249.e22. https://doi.org/10.1053/j.gastro.2018.07.010

http://ekhidna2.biocenter.helsinki.fi/dali/DaliTutorial.pdf

Gu, Y., Chen, B., Guo, D., Pan, L., Luo, X., Tang, J., Yang, W., Zhang, Y., Zhang, L., Huang, J., Duan, R., Wang, Z., 2022. Up-Regulation of RACGAP1 Promotes Progressions of Hepatocellular Carcinoma Regulated by GABPA via PI3K/AKT Pathway. Oxidative Medicine and Cellular Longevity 2022, e3034150. https://doi.org/10.1155/2022/3034150



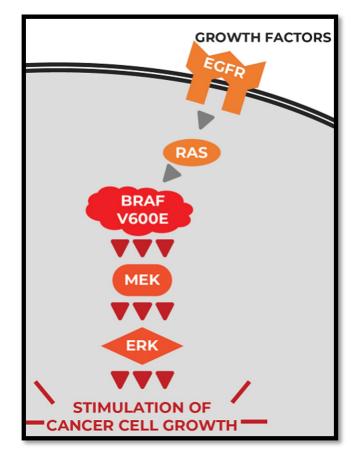
CRISPRi: A THERAPEUTIC APPROACH IN MANAGING ANAPLASTIC THYROID CANCER

Gabriele Virgilio, Alessandro Belvedere, Emanuela Unhe Jeong, Nashon Majaliwa

BACKGROUND

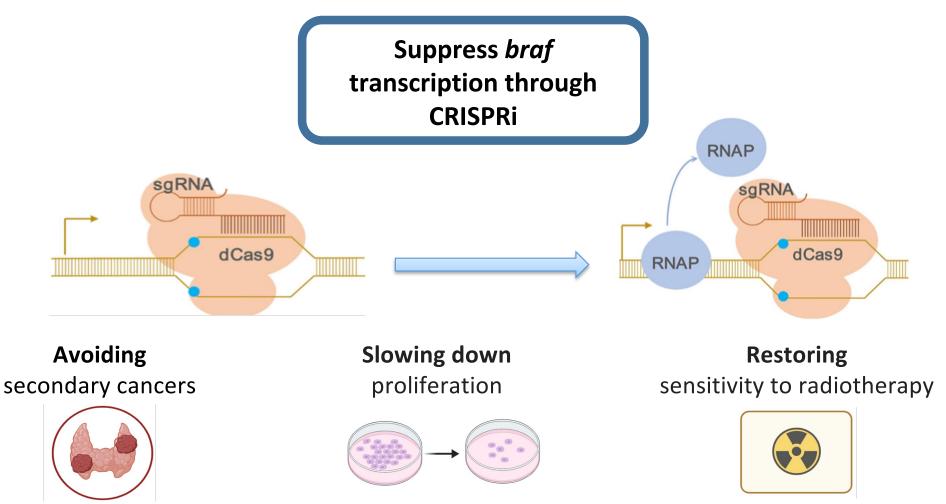
Anaplastic thyroid cancers (ATCs) are highly aggressive tumors and account for **30%** of US thyroid cancer deaths.

- **BRAF** is a proto-oncogene involved in the activation of the MAPK pathways;
- The **BRAF**^{V600E} **point mutation** is a common early molecular event and is detected in **45%** of ATC cases;
- Cancer cells expressing BRAF V600E are less responsive to radioactive iodine therapy due to downregulation of the sodium iodide symporter (NIS);
- BRAF chemical inhibitors can cause **paradoxical BRAF activation**, leading to the formation of secondary cancers.

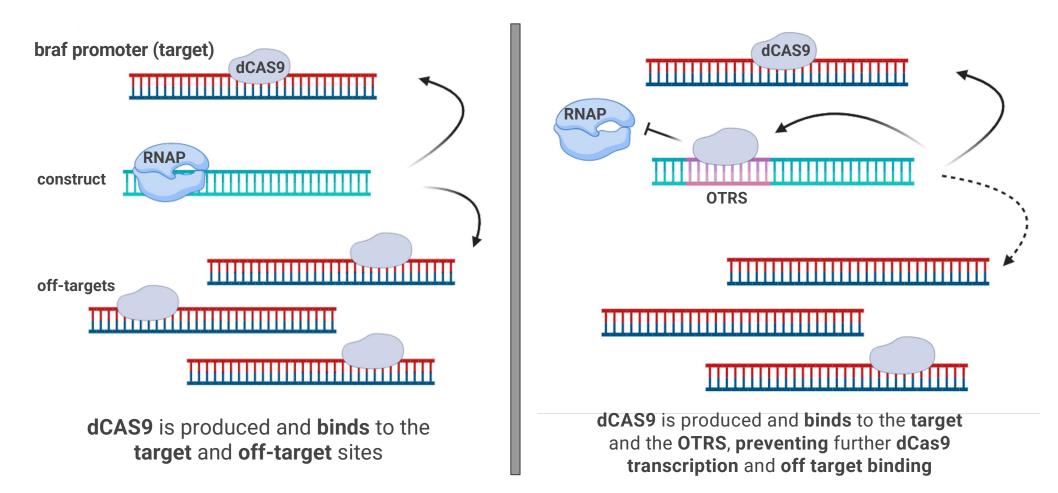


tttps://www.researchgate.net/publication/320341777_PIK3CAH1047Rnduced_paradoxical_ERK_activation_results_in_resistance_to_BRAFV600E_specif publication_provided and the provided and the pro

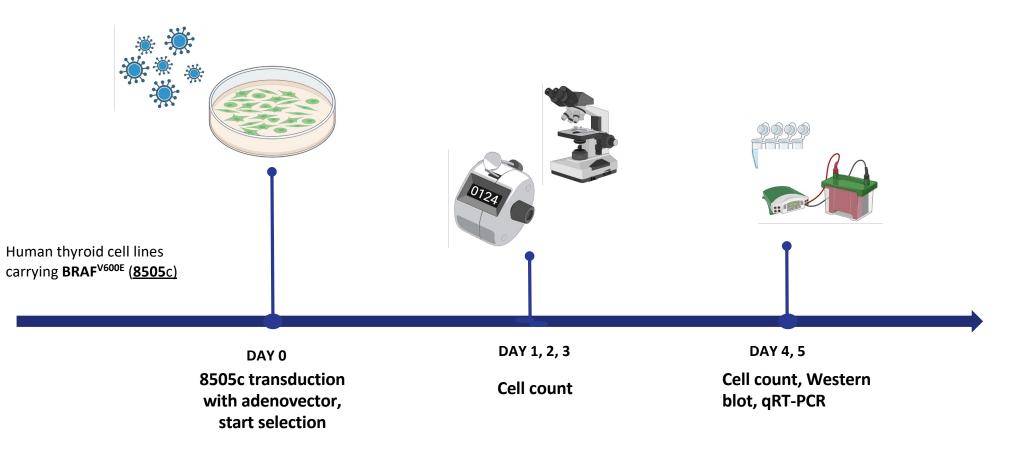
AIM OF THE PROJECT AND STRATEGY



REDUCING SIDE EFFECTS Off-Target Regulatory Sequence (OTRS)

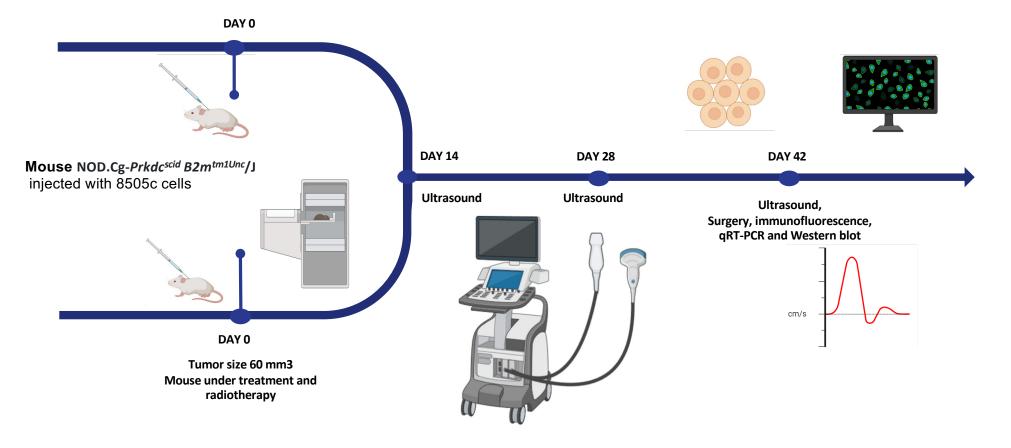


IN VITRO EXPERIMENTAL PLAN

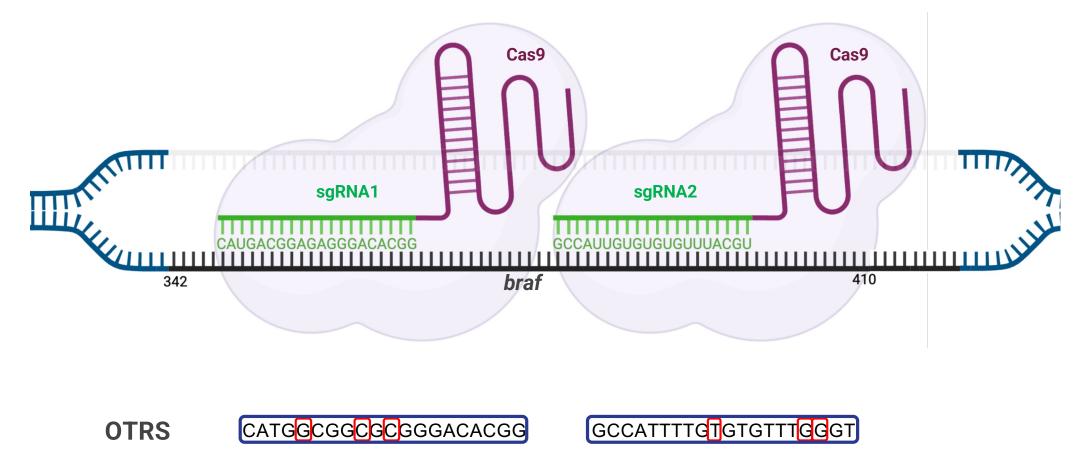


IN VIVO EXPERIMENTAL PLAN

Tumor size 60 mm3 Mouse under treatment

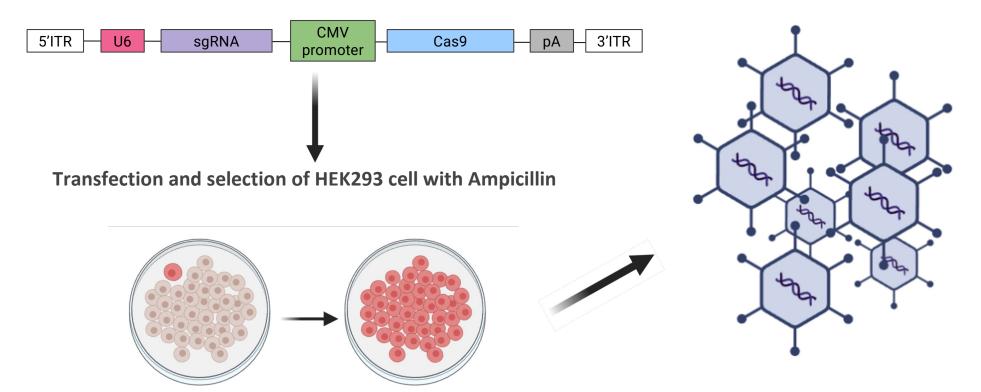


sgRNA DESIGN



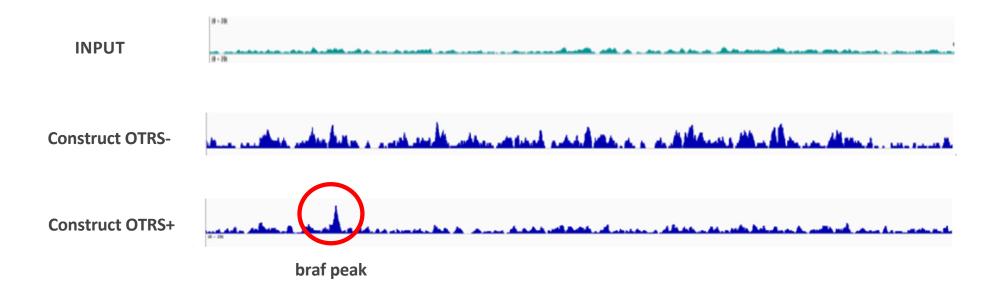
ADENOVECTOR DESIGN & PRODUCTION

Construct: dCas9 + sgRNA

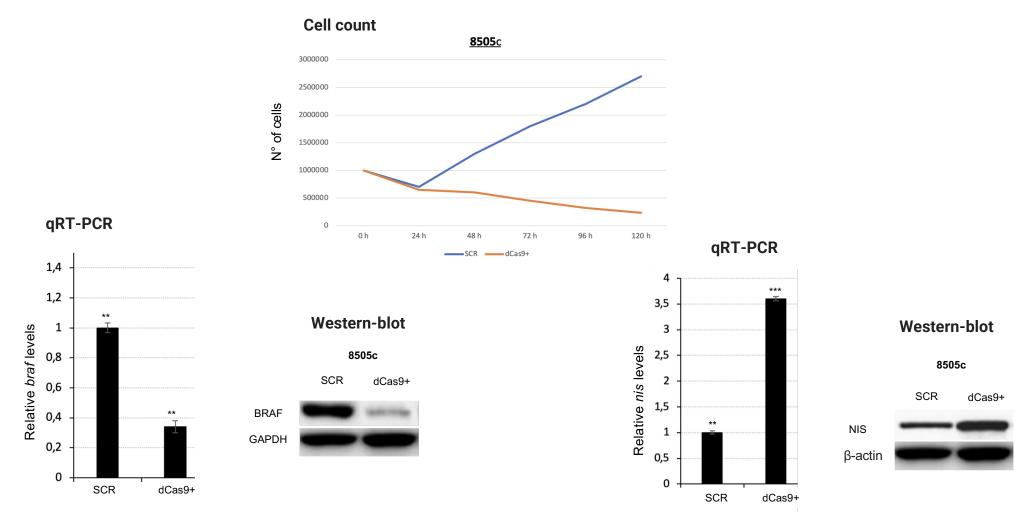


PROOF THAT THE SYSTEM WORKS

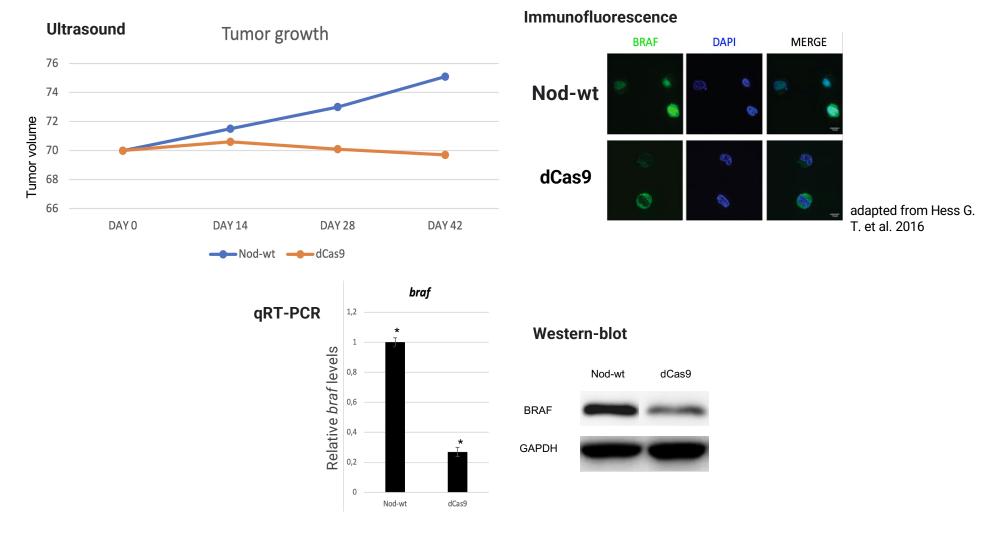
ChIP-seq: targeting dCAS9



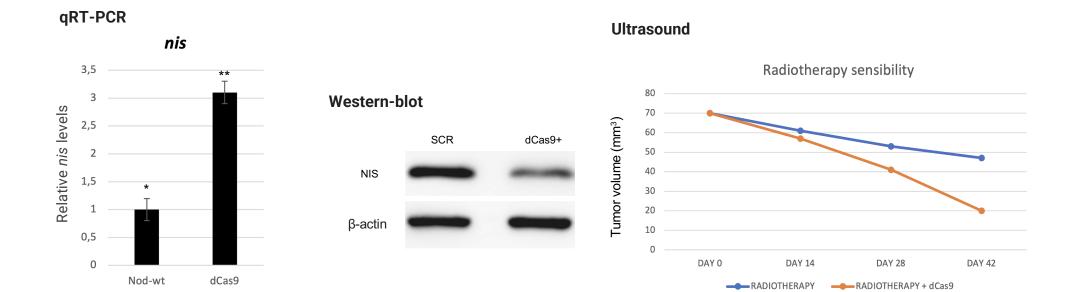
EXPERIMENT IN VITRO



EXPERIMENT IN VIVO



EXPERIMENT IN VIVO



BIBLIOGRAPHY

- Larson MH, Gilbert LA, Wang X, Lim WA, Weissman JS, Qi LS. CRISPR interference (CRISPRi) for sequence-specific control of gene expression. Nat Protoc. 2013 Nov;8(11):2180-96. doi: 10.1038/nprot.2013.132. Epub 2013 Oct 17. PMID: 24136345; PMCID: PMC3922765. Using iCRISPR
- Giuliano CJ, Lin A, Girish V, Sheltzer JM. Generating Single Cell-Derived Knockout Clones in Mammalian Cells with CRISPR/Cas9. Curr Protoc Mol Biol. 2019 Sep;128(1):e100. doi: 10.1002/cpmb.100. PMID: 31503414; PMCID: PMC6741428. USO DI DUE GUIDE
- <u>https://www.cellosaurus.org/CVCL_1054</u> Cell line used
- Shimamura M, Shibusawa N, Kurashige T, Mussazhanova Z, Matsuzaki H, Nakashima M, Yamada M, Nagayama Y. Mouse models of sporadic thyroid cancer derived from BRAFV600E alone or in combination with PTEN haploinsufficiency under physiologic TSH levels. PLoS One. 2018 Aug 7;13(8):e0201365. doi: 10.1371/journal.pone.0201365. PMID: 30086162; PMCID: PMC6080762. Mouse model
- Park KS, Saindane M, Yang EY, Jin T, Rallabandi HR, Heil A, Nam SE, Yoo YB, Yang JH, Kim JB, Park SY, Park WS, Youn YK. Selective inhibition of V600E-mutant BRAF gene induces apoptosis in thyroid carcinoma cell lines. Ann Surg Treat Res. 2021 Mar;100(3):127-136. doi: 10.4174/astr.2021.100.3.127. Epub 2021 Feb 26. PMID: 33748026; PMCID: PMC7943282. Effect of braf inhibitors on 8505c cell proliferation
- Oh HS, Kwon H, Song E, Jeon MJ, Kim TY, Lee JH, Kim WB, Shong YK, Chung KW, Baek JH, Kim WG. *Tumor Volume Doubling Time in Active Surveillance of Papillary Thyroid Carcinoma*. Thyroid. 2019 May;29(5):642-649. doi: 10.1089/thy.2018.0609. Epub 2019 Apr 8. PMID: 30864894.
 Tumor growth speed
- Zhang C, Chai J, Jia Q, Tan J, Meng Z, Li N, Yuan M. Evaluating the therapeutic efficacy of radiolabeled BSA@CuS nanoparticle-induced radiophotothermal therapy against anaplastic thyroid cancer. IUBMB Life. 2022 May;74(5):433-445. doi: 10.1002/iub.2601. Epub 2022 Feb 15. PMID: 35112451. Injection of tumor cells into mice for in vivo experiments
- Wächter S, Roth S, Gercke N, Schötz U, Dikomey E, Engenhart-Cabillic R, Maurer E, Bartsch DK, Di Fazio P. Anti-Proliferative Effect of Radiotherapy and Implication of Immunotherapy in Anaplastic Thyroid Cancer Cells. Life (Basel). 2023 Jun 15;13(6):1397. doi: 10.3390/life13061397. PMID: 37374179; PMCID: PMC10301015. Radiotherapy efficacy
- Hess GT, Frésard L, Han K, Lee CH, Li A, Cimprich KA, Montgomery SB, Bassik MC. *Directed evolution using dCas9-targeted somatic hypermutation in mammalian cells*. Nat Methods. 2016 Dec;13(12):1036-1042. doi: 10.1038/nmeth.4038. Epub 2016 Oct 31. PMID: 27798611; PMCID: PMC5557288. Immunofluorescence image

Materials

- HEK293T cell
- 8505c cell
- 4 mice
- Empty vector
- Vector with dCAS9 with the off-target
- Vector with dCAS9 without the off-target
- sgRNA1
- sgRNA2