CRISPR/Cas9 APPLICATIONS

CRISPR/Cas engineering is enabling a broad range of applications



(Hsu et al., Cell, 2014)



Crygc mutation (dominant inheritance)



(Wu et al., Cell 2013)

1 bp deletion in exon 3 of Crygc gene leads to cataract

In vitro



(Wu et al., Cell 2013)

In vitro

sgRNA leads to HDR mediated repair

sgRNA	E14 ESC clones		m <i>Crygc</i> (<i>Crygc</i> ^{+/-}) ESC clones								
	Cleavage at 1 Allele/Total	Cleavage at 2 Alleles/Total	Cleavage at WT Allele/Total	Cleavage at Mutant Allele/Total	HDR-mediated Repair/Total						
						sgRNA-1	4/36	0/36	0/36	10/36	7/36
						sgRNA-2	23/36	7/36	17/36	25/36	2/36
sgRNA-3	3/36	0/36	0/36	7/36	5/36						
sgRNA-4	0/36	0/36	0/36	11/36	16/36						
sgRNA-5	4/36	26/36	27/36	26/36	0/36						

(Wu et al., Cell 2013)

sgRNA4 show high specificity for mCrygc allele and mediates HDR

In vivo



(Wu et al., Cell 2013)

CRISPR/Cas9 system leads to gene correction via HDR using wt allele on the homologous chromosome

	WT allele AGTACCGGCGCTTCCAGGACTGGGGCTCTG
	Mutant allele AGTACCGGC – CTTCCAGGACTGGGGCTCTG
DR iated	WT allele AGTACCGGCGCTTCCAGGACTGGGGCTCTG
Нре	Mutant allele AGTACCGGCGCTTCCAGGACTGGGGCTCTG HDR (×4)
	WT allele AGTACCGGCGCTTCCAGGACTGGGGCTCTG
air	Mutant allele AGTACCGGCCTTCC a AGGACTGGGGCTCTG +1
NHE redia	WT allele AGTACCGGCGCTTCCAGGACTGGGGCTCTG
E	Mutant allele AGTACCcagAGGACTGGGGCTCTG -8 +3
-	WT allele AGTACCGGCGCTTCCAGGACTGGGGCTCTG
spai	Mutant allele AGTACCGGC
Dn-re	WT allele AGTACCGGCGCTTCCAGGACTGGGGCTCTG
рл	Mutant allele AGTACCGGCCTGGGGGCTCTG -9 (×2)
Ξ	WT allele AGTACCGGCGCTTCCAGGACTGGGGCTCTG
z	Mutant allele AGTACCGGCCTTGGACTGGGGCTCTG -4

(Wu et al., Cell 2013)

NHEJ events can lead to correct reading frame

Is it possible to improve CRISPR/Cas9 sgRNA4 gene correction?



(Wu et al., Cell 2013)

Insertion of Oligo-1 that mimic wt allele and Oligo-2 that contains specific in frame mutation

Is it possible to improve CRISPR/Cas9 sgRNA4 gene correction?



(Wu et al., Cell 2013)

Insertion of Oligo-1 that mimic wt allele and Oligo-2 that contains specific synonymous mutations

Article

In vivo base editing rescues Hutchinson– Gilford progeria syndrome in mice



Adapted from Baek, Erikson, 2013

Does ABE approach correct HGPS mutation in patient cells?





Lentiviral mediated delivery ABE7-10fused with optimized SpCas9-VRQR+ SgRNA targeting HGP mutation

84-85% of correction 10 days 87-91% of correction 20 days

Does ABE approach correct HGPS mutation in patient cells?



Does ABE approach correct HGPS mutation in patient cells?



1.8 fold reduction of nuclear abnormalities

Base editing to correct LMNA c.1824 C \rightarrow T mutation in cells from HGPS patients rescues the molecular and phenotypic consequences of the mutation

Does LMNA c.1824 C \rightarrow ABE correction approach induce off targets?



Does LMNA c.1824 C \rightarrow ABE correction approach induce off targets?



ABE treated transcriptomes cluster with unaffected Fb

Treating cells with LMNA-targeting sgRNA and ABEmax-VRQR didn't results in off-target DNA or RNA editing despite high level of ontarget editing

In vivo ABE delivery in mice with progeria

Progeria mouse model:

C57BL/6-tg(LMNA*G608G)HCIns/J: includes complete human LMN c.1824C>T



Systemic in vivo delivery of ABE and sgRNA characterized in cells

Varga et al, PNAS 2006

For the first 4 months after birth, homozygotes exhibit a slower rate of weight gain when compared to hemizygotes. In addition to developing kyphosis, hair loss, tight skin, loss of subcutaneous fat and joint contracture, homozygotes develop a significantly more severe vascular damage with VSMC loss in aortic vessel walls, calcification, and periadventitial thickening. Homozygotes die at an average of 7-8 months of age as the result of aortic stiffening and impaired cardiovascular functioning. The Donating Investigator reports that homozygous females are infertile and that a lower than expected number of homozygotes are born.

From Jackson Lab

In vivo ABE delivery in mice with progeria



A single in vivo AB-encoding AAV results in 10-60% of point mutation correction in the various organs

6 months after injection



Increase in gene editing at 6 months, mainly for p3 mice but remains higher for p14 mice injected with higher doses off ABE encoding AAVs

Base editing persists in vivo

In vivo ABE delivery in mice with progeria. Long term ABE treatment



In vivo ABE treatment led to decrease in progerin transcript abundance and robust reduction of progerin protein

ABE correction of LMNA c.1824C>T alleli in mice can reduce progerin mRNA and protein levels in several clinically relevant tissues



Bystander editing and indels observed at very low frequency compared to on target editing

ABE treatment improves vascular pathology



ABE treatment improves vascular pathology

Increase in VSMCs count



Base editing of around 25% in aorta in enough to rescue two key vascular defects of progeria, preserving the expression of Lamin A/C and reducing progerin abundance

ABE treatment extends progeria mouse lifespan



Conclusions

- Patients-derived cells ABE correct pathogenic allele (97-91% of on target editing) with minimal degree of off-target. Reduces mis-splicing and progerin protein resuces nuclear morphology defects
- A single dose of dual AAV9 encoding ABE and sgRNA in a progeria mouse model resulted in durable correction of pathogenic allele, reduction of progerin protein with different efficient in diverse tissue and organs. Treatment of p14 mice greatly improved cardiac phenotype and aortic health.
- A single dose of dual AAV9 encoding ABE and sgRNA in a progeria mouse model increases mice lifespan

Concerns:

- Immune response to treatment
- Timing of treatment to take account of the time of diagnosis
- Base editing amelioration to obtain results in other tissues
- No direct correlation between base editing, RNA levels and proteins levels