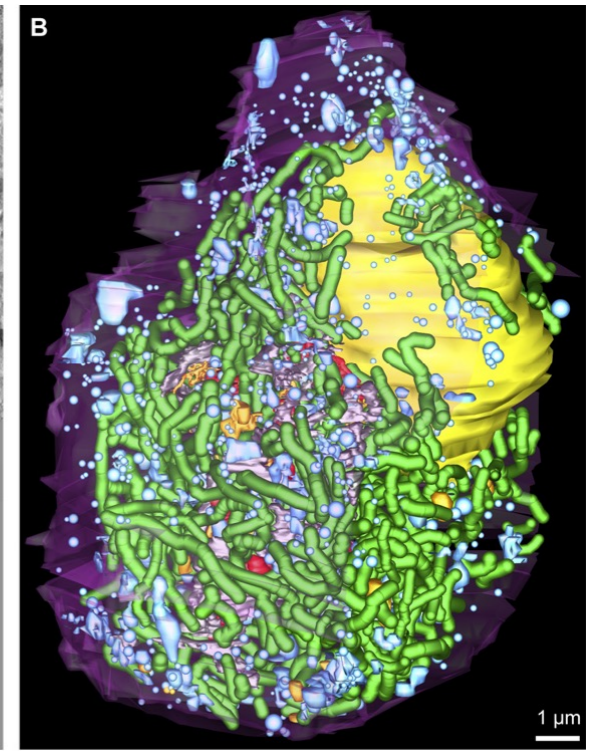
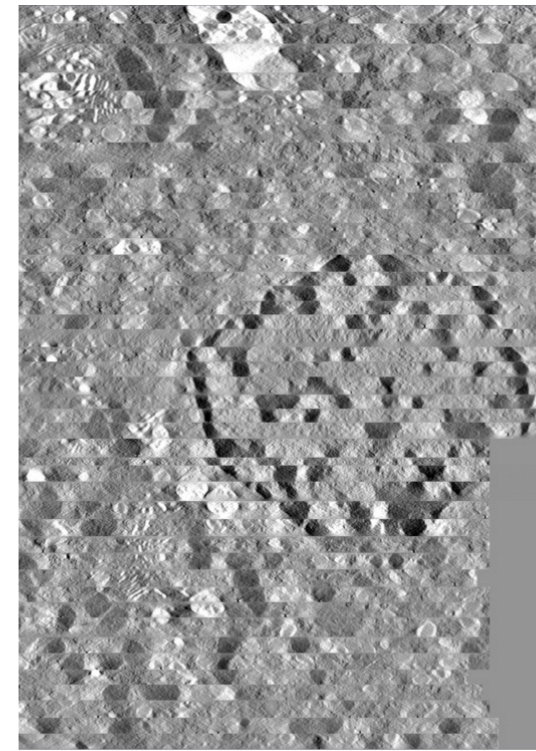
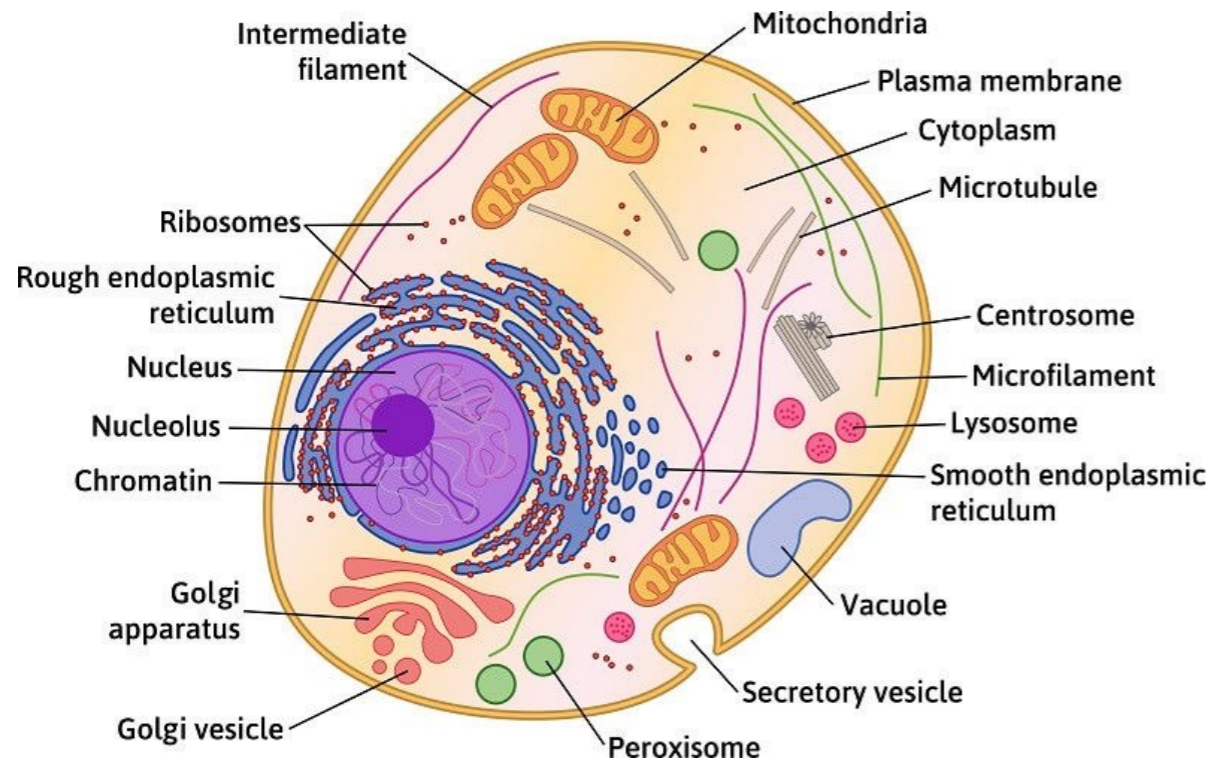


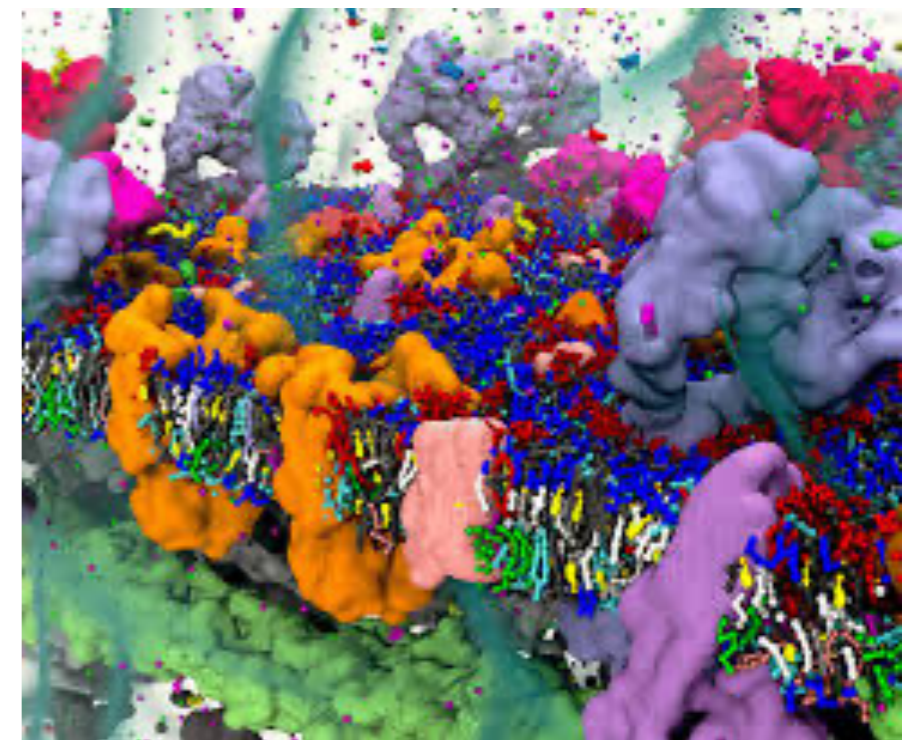
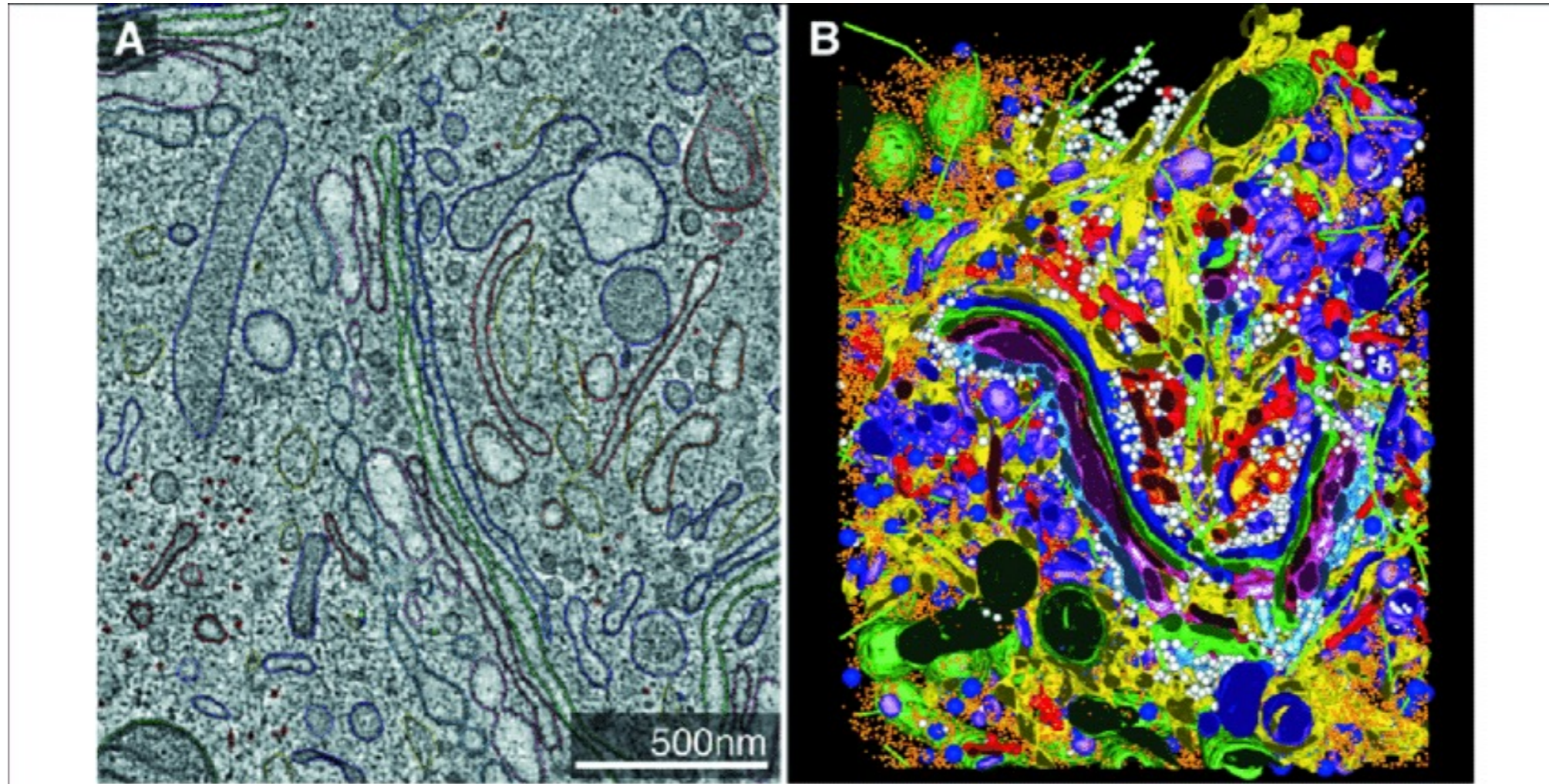
# **THE INTERACTOME STUDY**

October  
2022

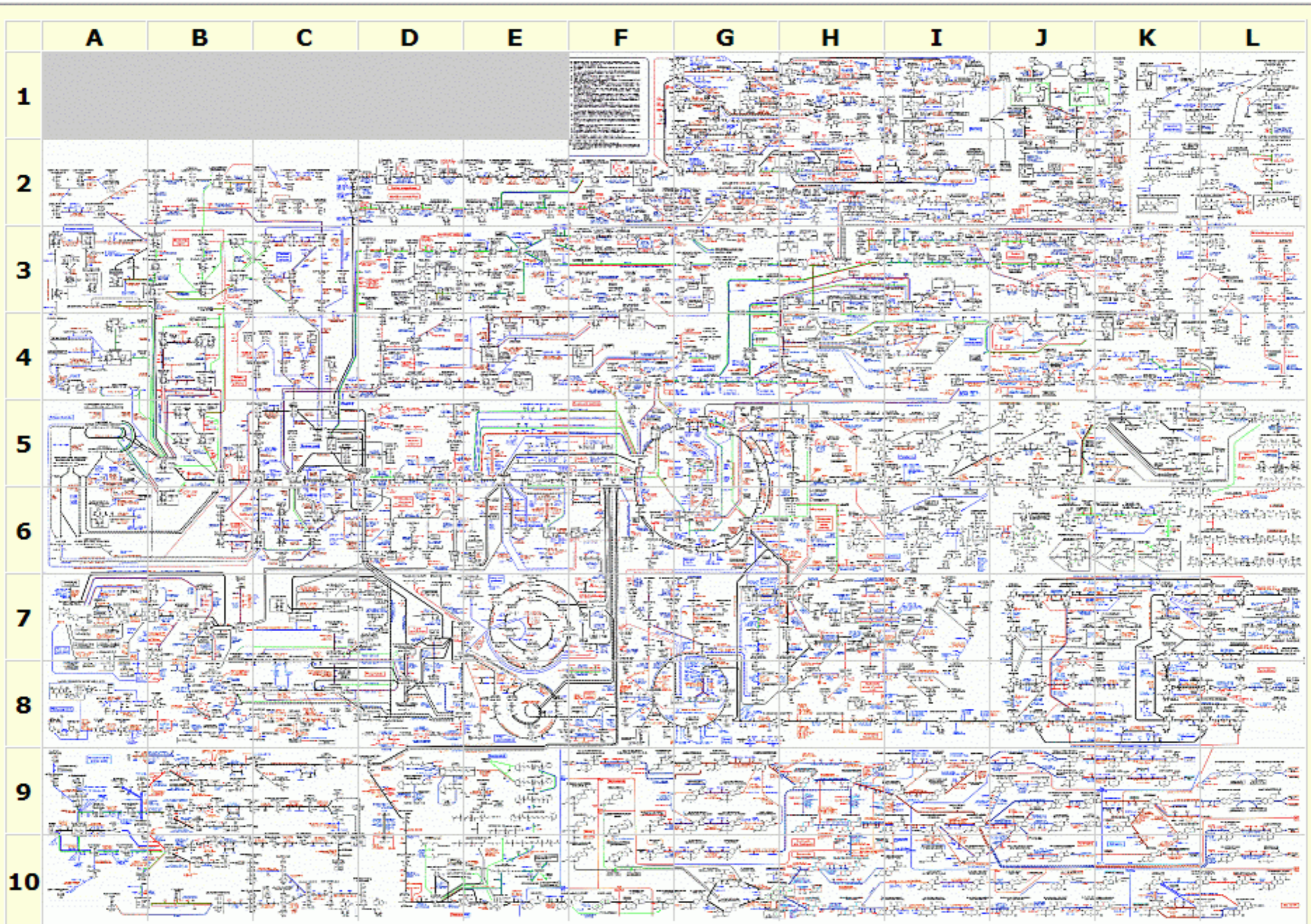
# Cells are complex



# Cells are complex



# The molecular level is not better





# Enrichment methods: the basics

## Co-IP

Prepare

Protein complex

Primary antibody

Enrich

Protein A/G  
magnetic  
beads

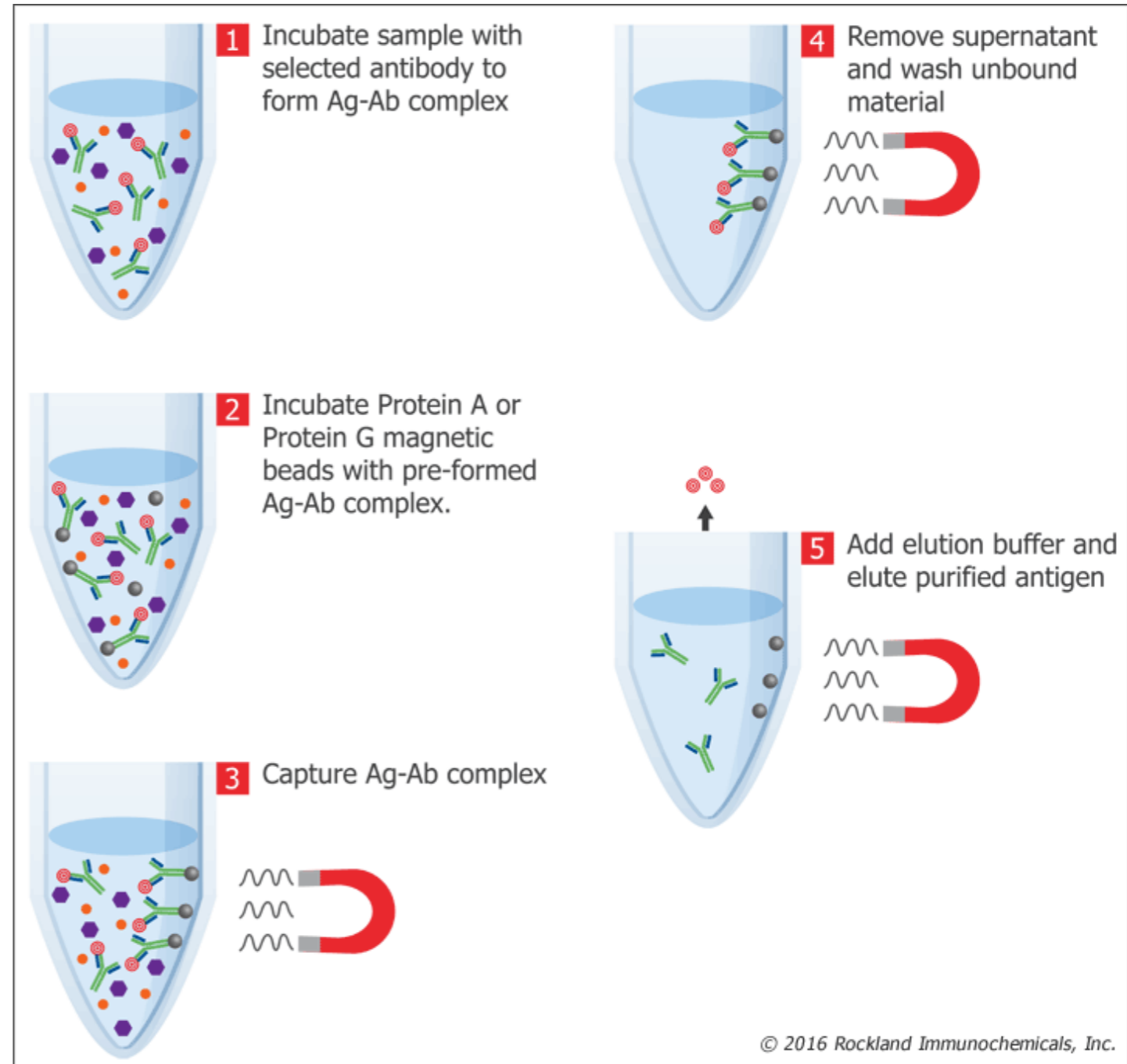
Wash

Washing

Analyse

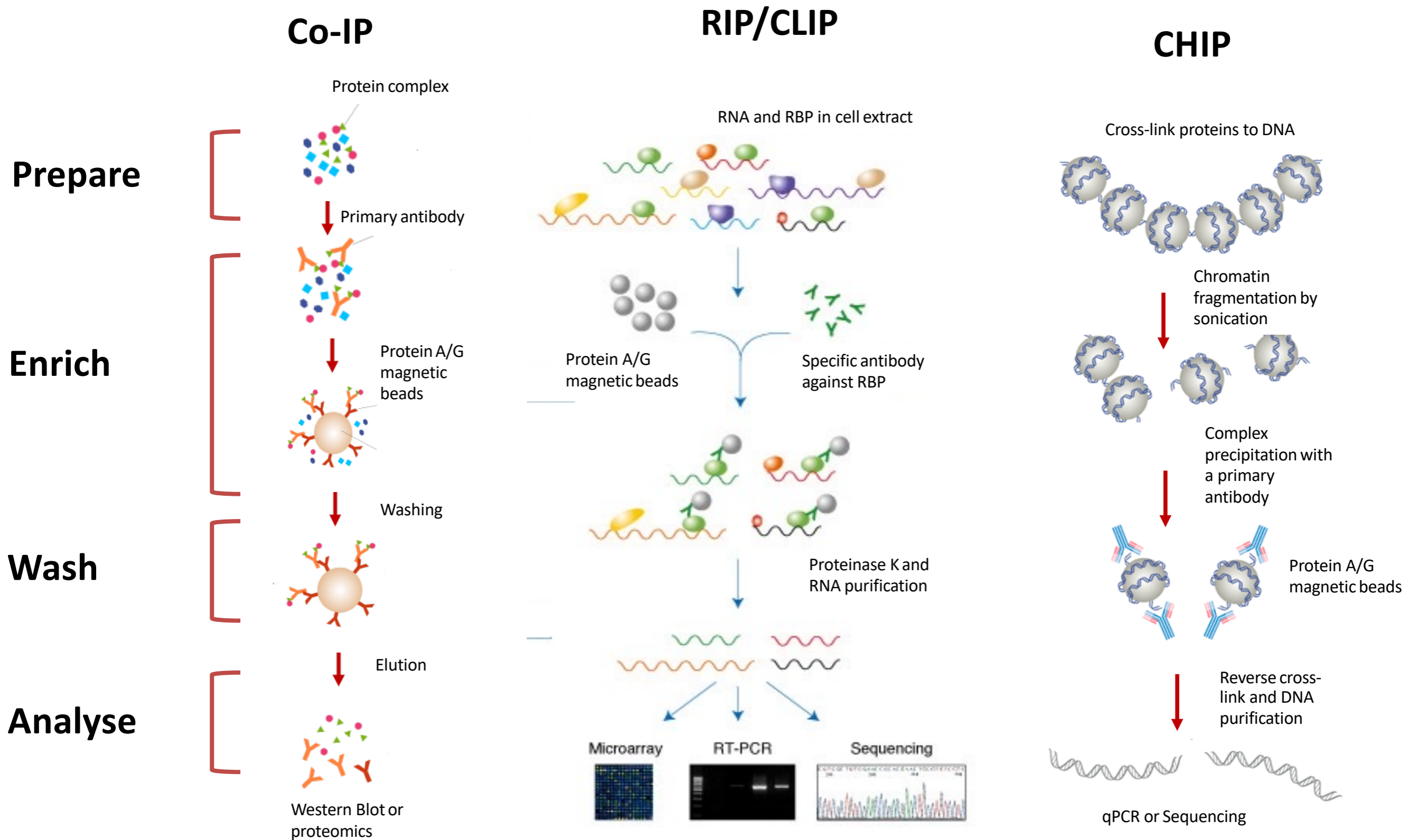
Elution

Western Blot or  
proteomics



# Enrichment methods

## Finding molecular partners:



# Types of interaction

		OUTPUT (what we analyse)		
		Protein	RNA	DNA
BAIT (What we enrich)	Protein	<b>CO-IP</b> (co-immunoprecipitation)	<b>RIP/CLIP</b> (RNA-Immunoprecipitation)	<b>ChIP</b> (Chromatin Immunoprecipitation)
	RNA	<b>Exogenous RNA pull Down</b> <b>RAP-Protein</b> (RNA antisense purification)	<b>RAP-RNA</b> (RNA antisense purification)	<b>ChIRP</b> (Chromatin isolation by RNA purification)
	DNA	<b>DNA pull down</b>		<b>Conformation capture 3C</b>



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	DNA	<b>DNA pull down</b>		<b>Conformation capture 3C</b>

# Considerations:

Crosslinked vs native.

Types of baits.

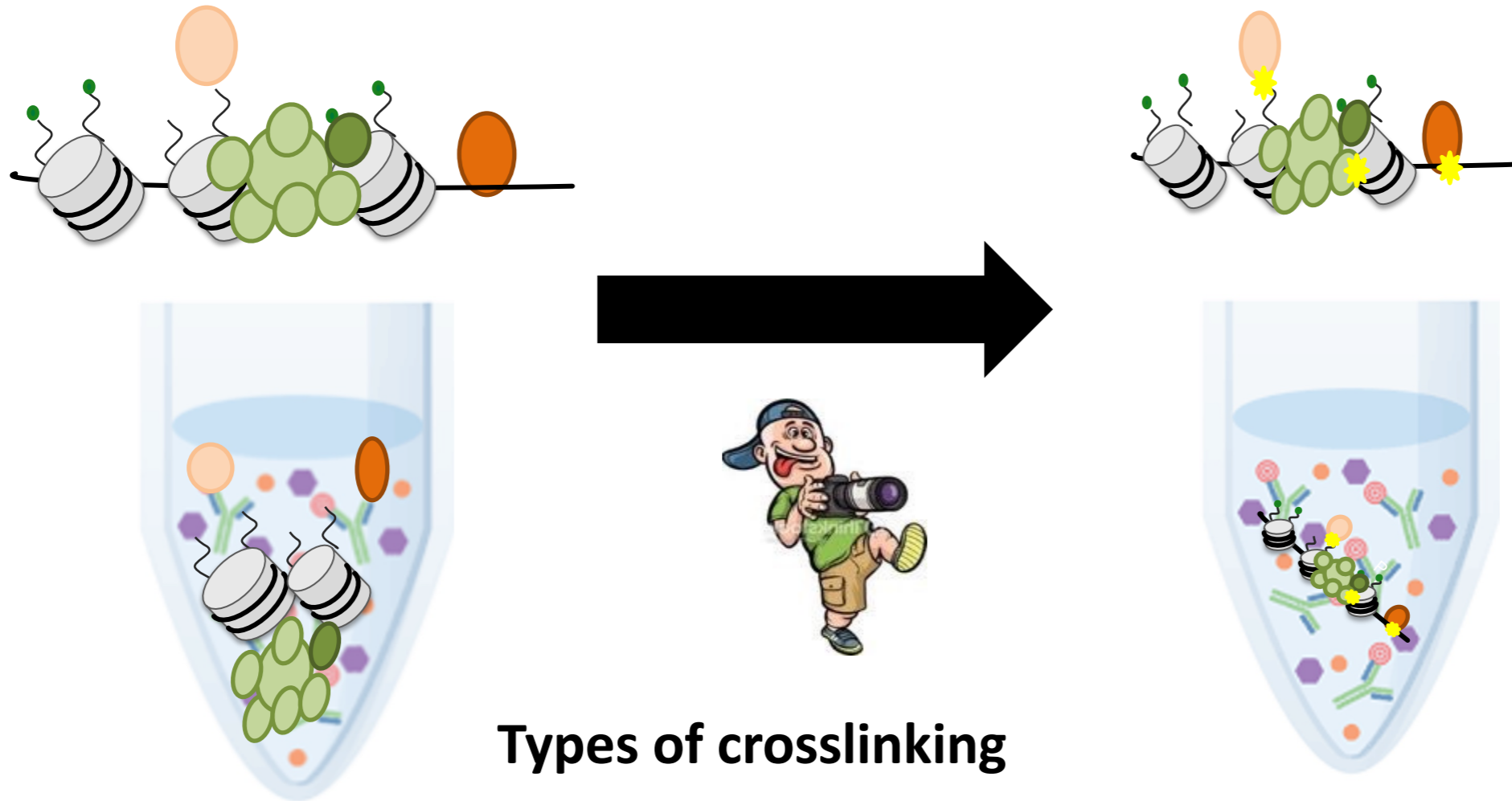
Types of output.

# Crosslinked vs native

**Crosslinking: establishing molecular bonds between molecules.**

**Avoid non specific interaction**

**Capture interaction in a timeframe**



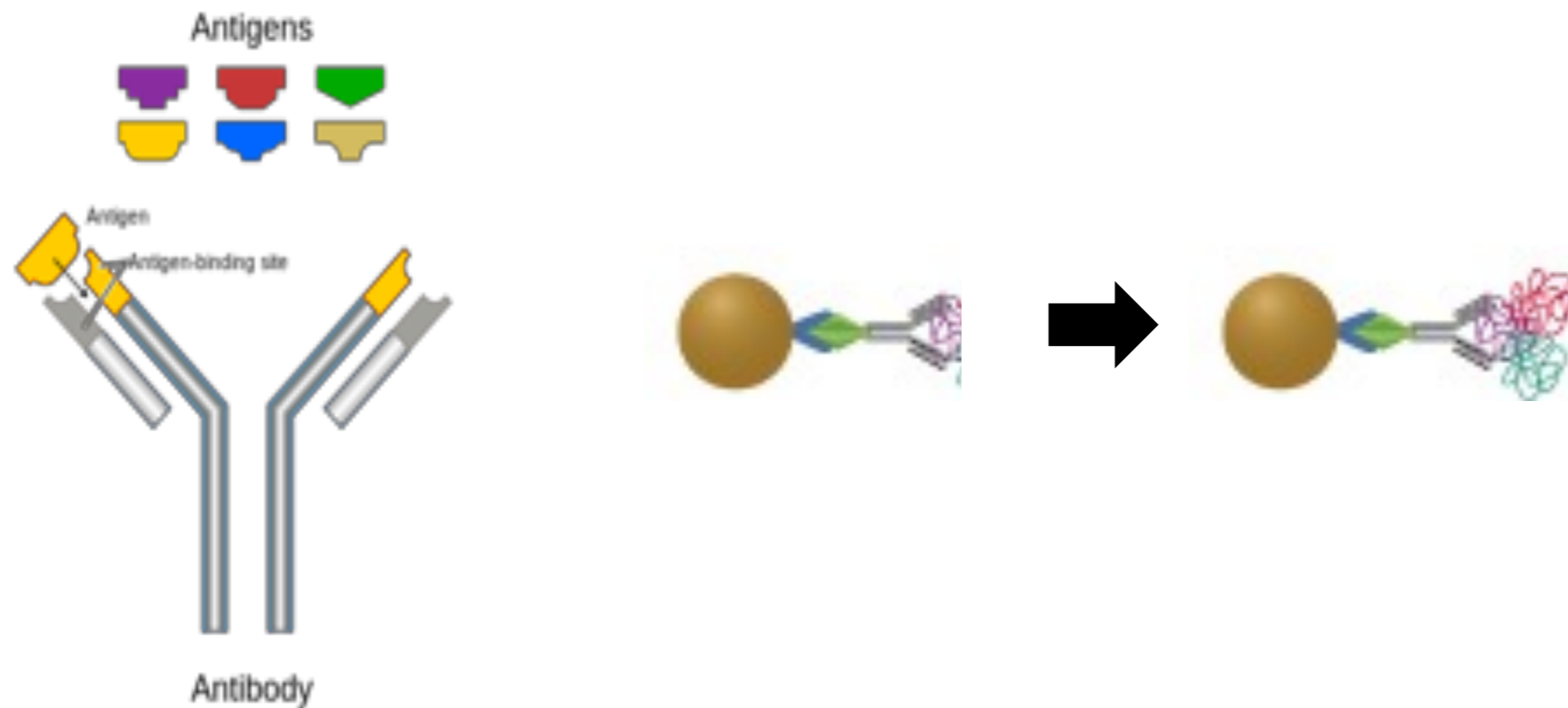
## Types of crosslinking

- UV light
- Formaldehyde
- Gluthaladehyde
- Psoralen

## Types of baits.

### Antibodies + prot G → to catch proteins

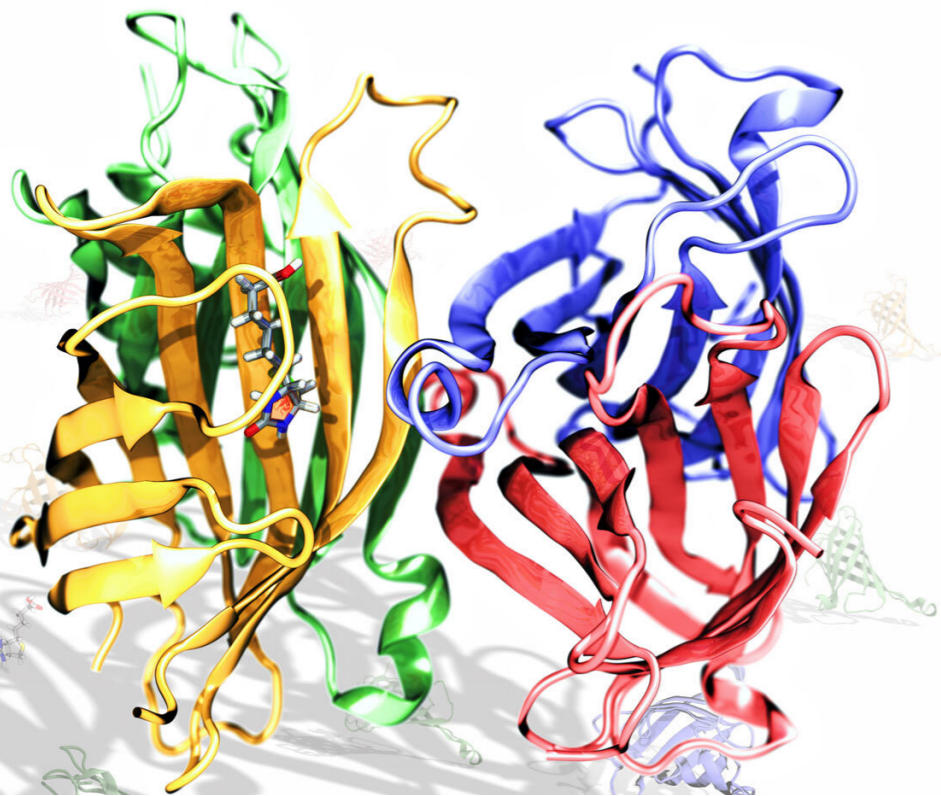
- Monoclonal: 1 clone, 1 epitope
- Polyclonal: several clones, several epitopes



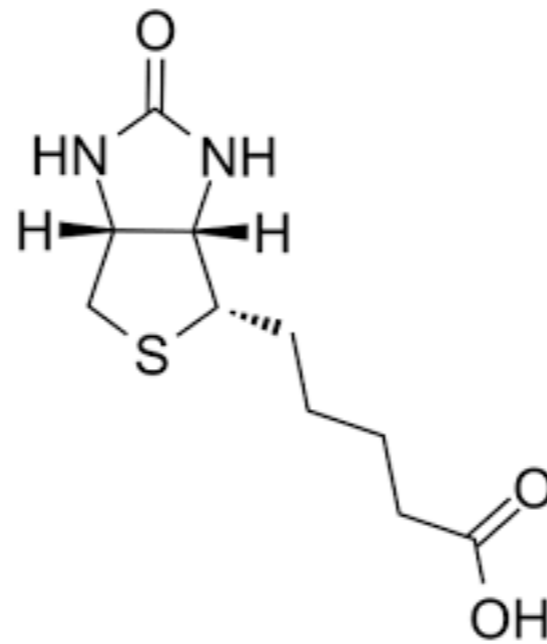
## Types of baits.

### Biotinylated Nucleic acid + streptavidin beads

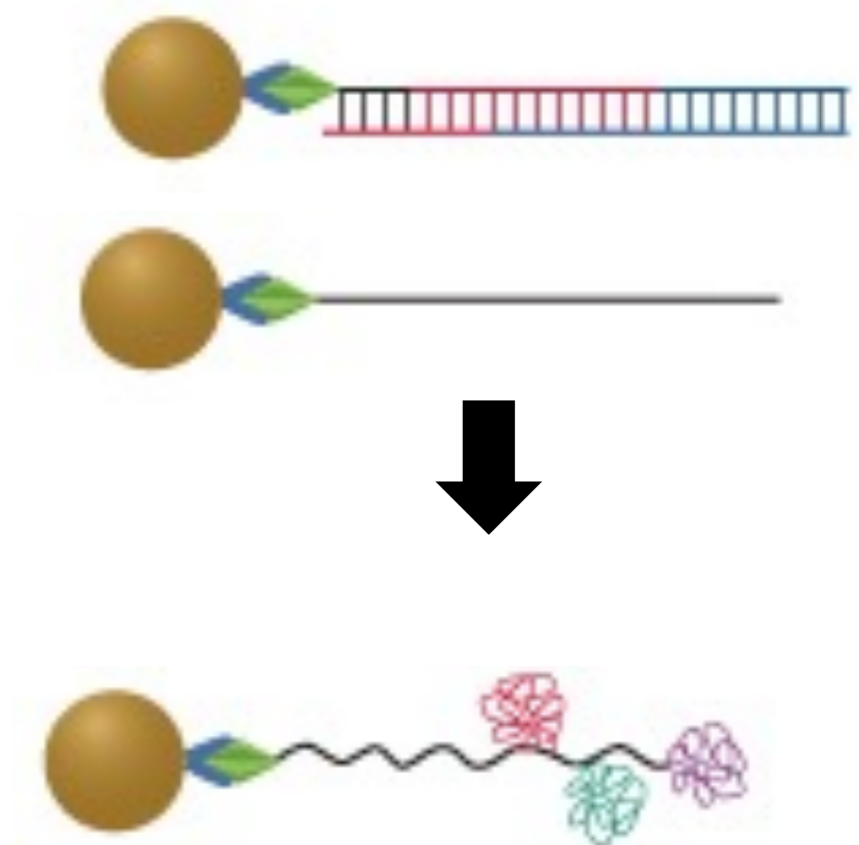
- Biotinylated DNA/RNA
- We can modify nucleotides :LNA, sulfur bonds..
- Specific base-base interaction.



Streptavidin



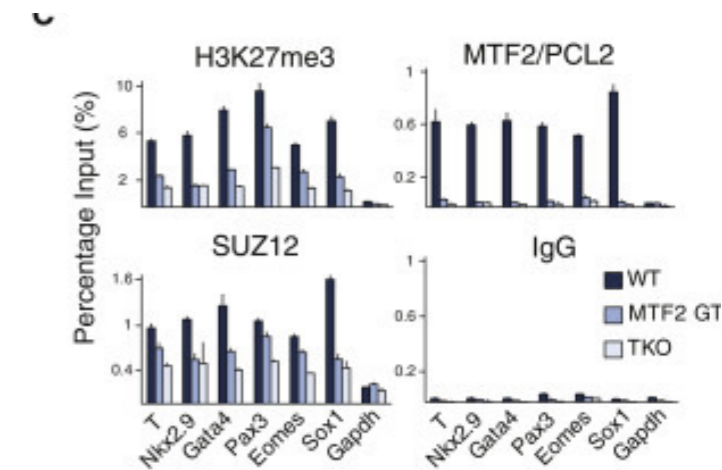
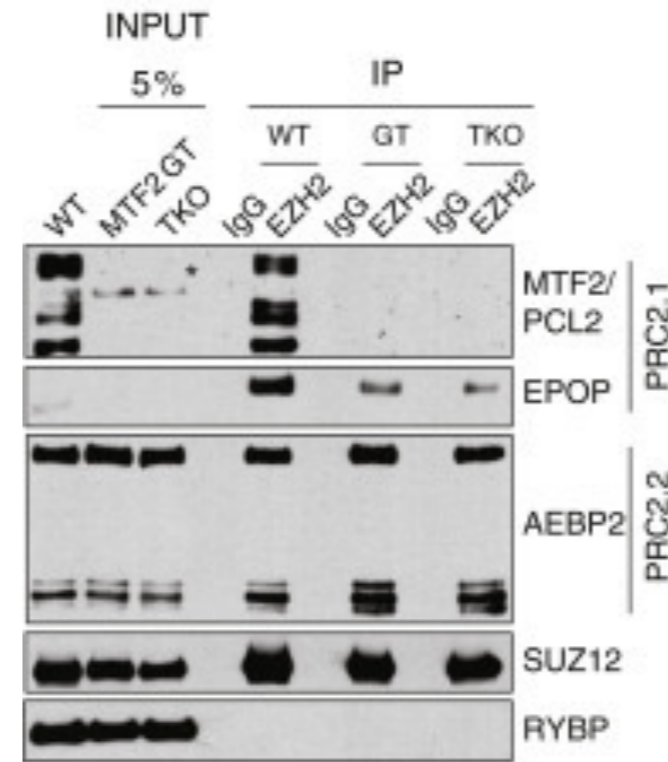
Biotin



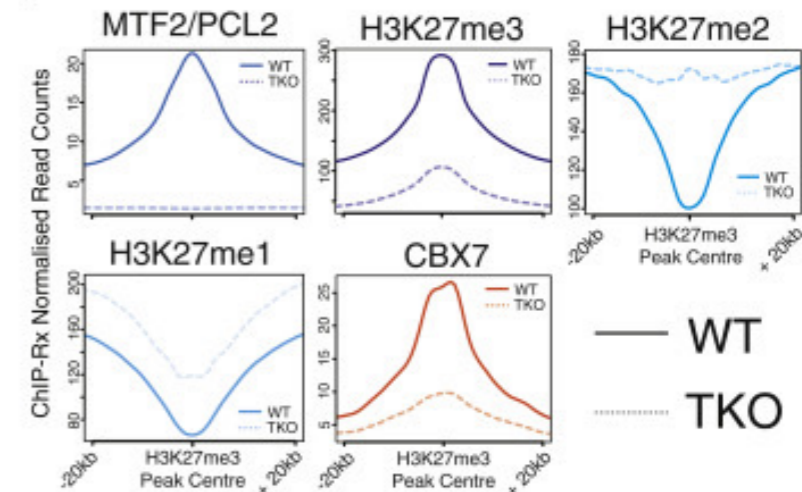
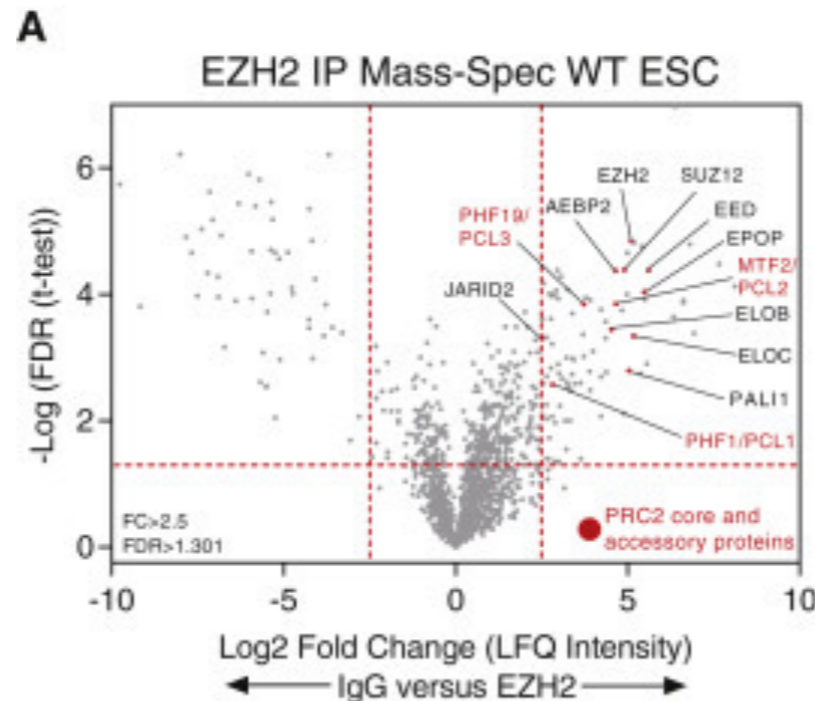
# Types of outputs.

Low throughput: we analyse the interaction of our bait with few genes

## Western Blot/PCR



High throughput: we analyse the interaction of our bait with all the genome/proteome in the cell.



## Mass spectrometry/ Sequencing

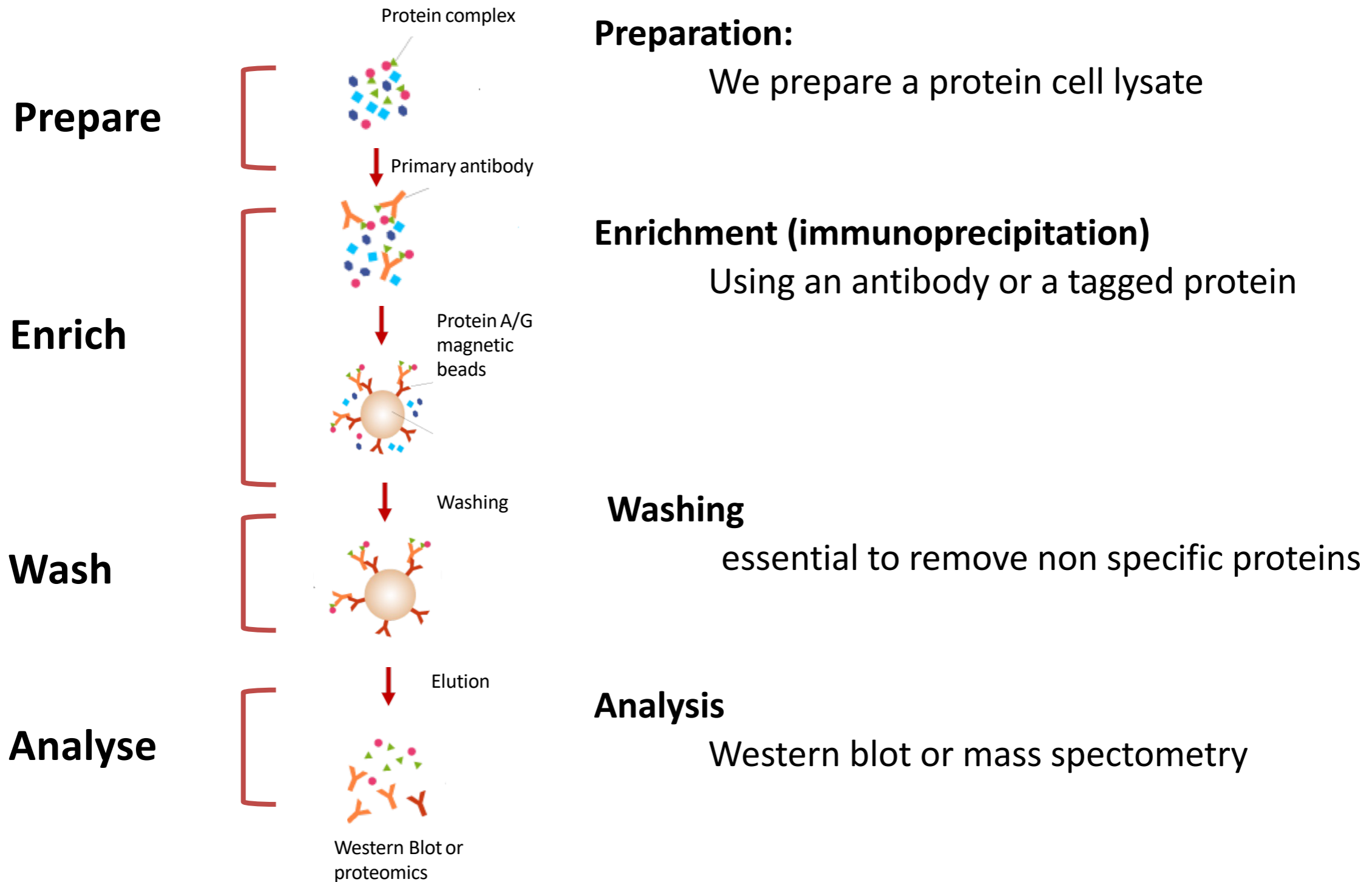
# Types of interaction

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		Protein	RNA	DNA
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	DNA	<b>DNA pull down</b>		<b>Conformation capture 3C</b>

# Co-immunoprecipitation

**AIM: Identification of protein interactions. Bait: protein/output: protein)**

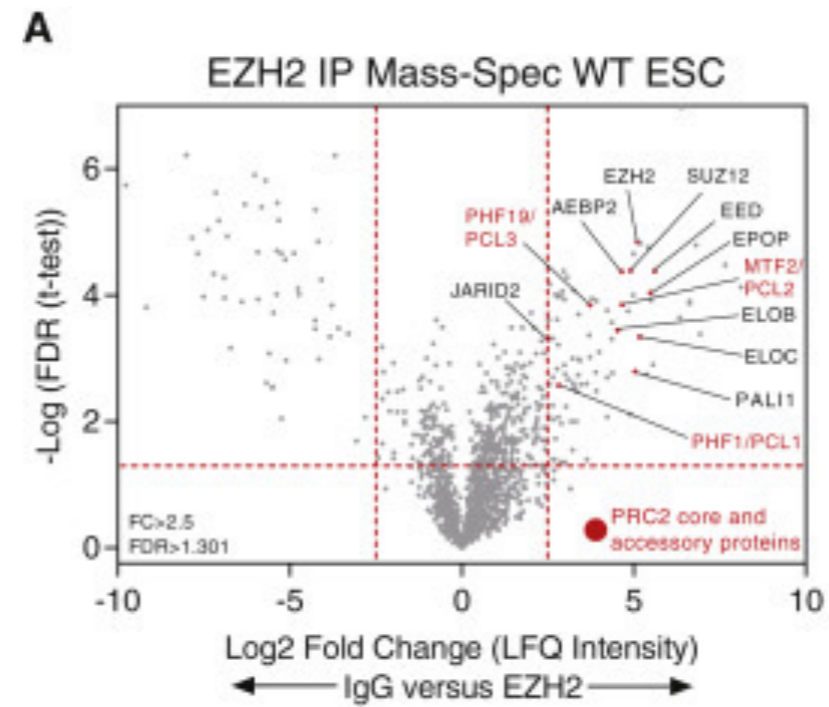
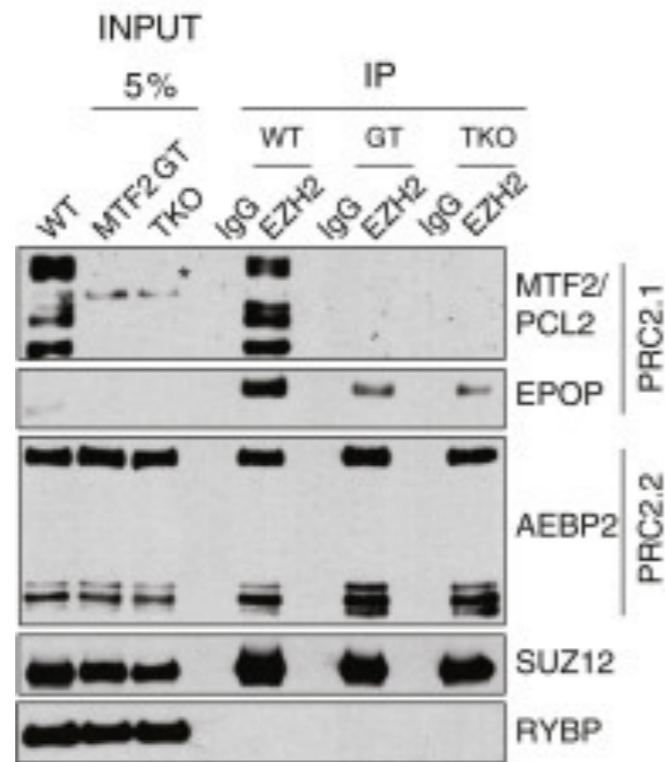
**Co-Immunoprecipitation (COIP)** is an immunoprecipitation technique used to investigate the interaction between proteins.





# Co-immunoprecipitation

## Example: PRC2 complex



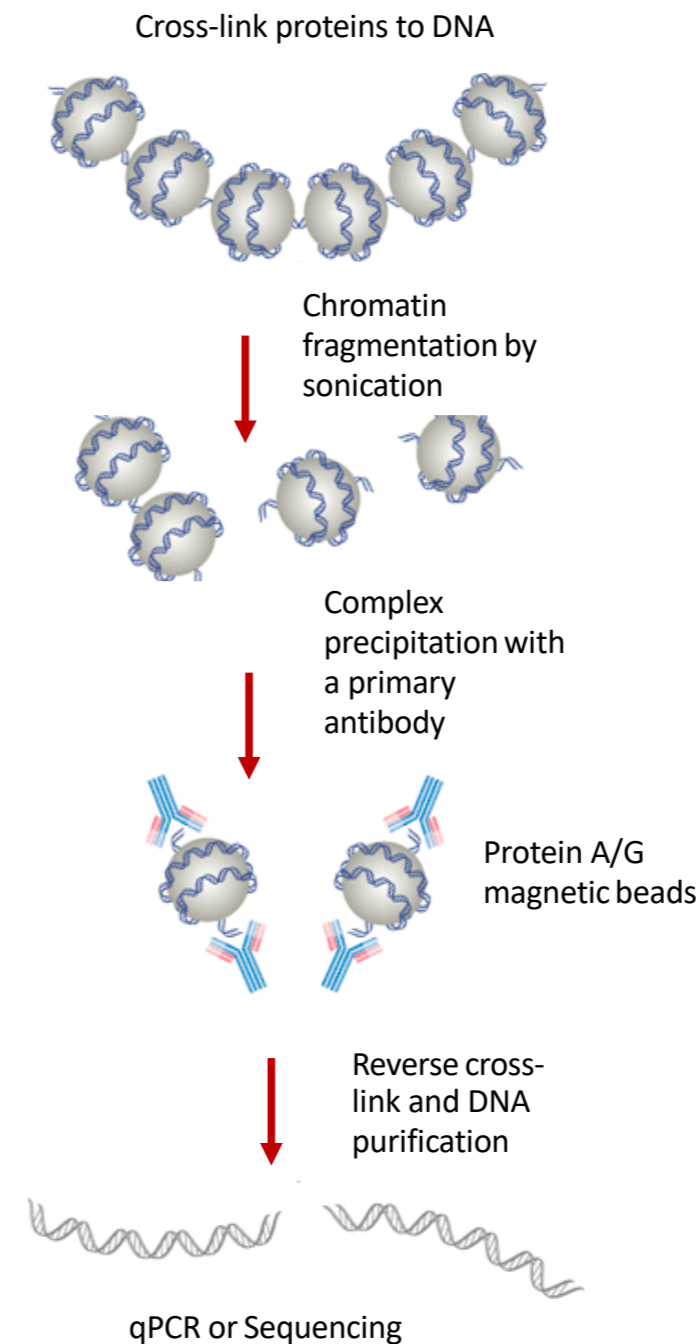
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# ChIP

**AIM: Identification of the genomic loci bound to a DNA binding protein. (Bait Protein/ output :DNA)**

**Chromatin Immunoprecipitation (ChIP)** is an immunoprecipitation technique used to investigate the interaction between proteins and DNA in the cell. It aims to determine whether specific proteins are associated with specific genomic regions, such as transcription factors on promoters.

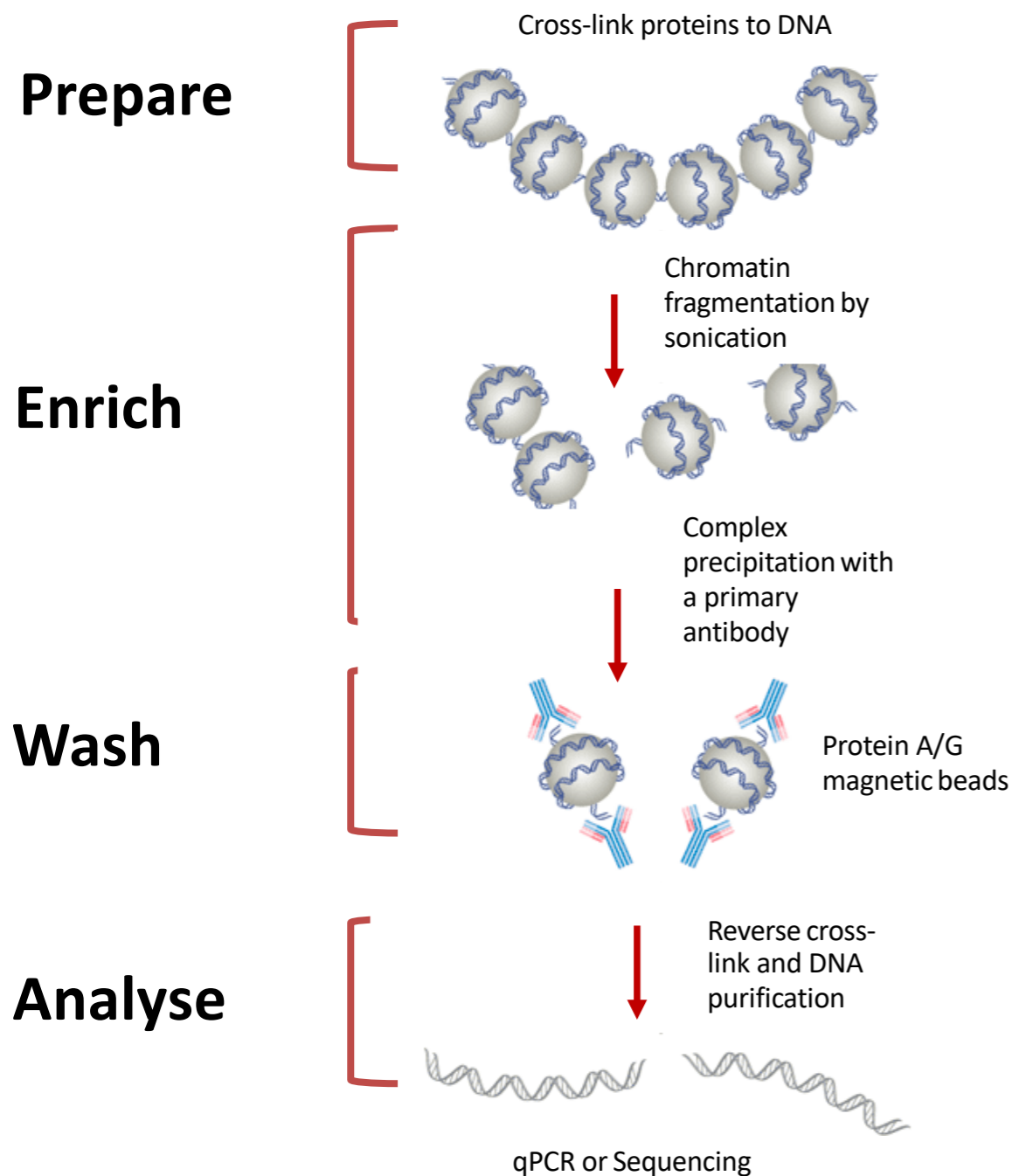


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## Preparation: Possible crosslinking



**Native-ChIP:** Native chromatin is used as substrate

- only proteins tightly associated with DNA can be immunoprecipitated
- antigens cannot be occurred or modified by chemical cross-linking
- the specificity of the antibody binding to un9ixed chromatin is more predicta

**X-ChIP:** cross-linked chromatin is used as substrate

- also proteins weakly or not directly associated with DNA
- antigens can be obscured or modified by the formaldehyde cross-linking
- more widely used than Native ChIP

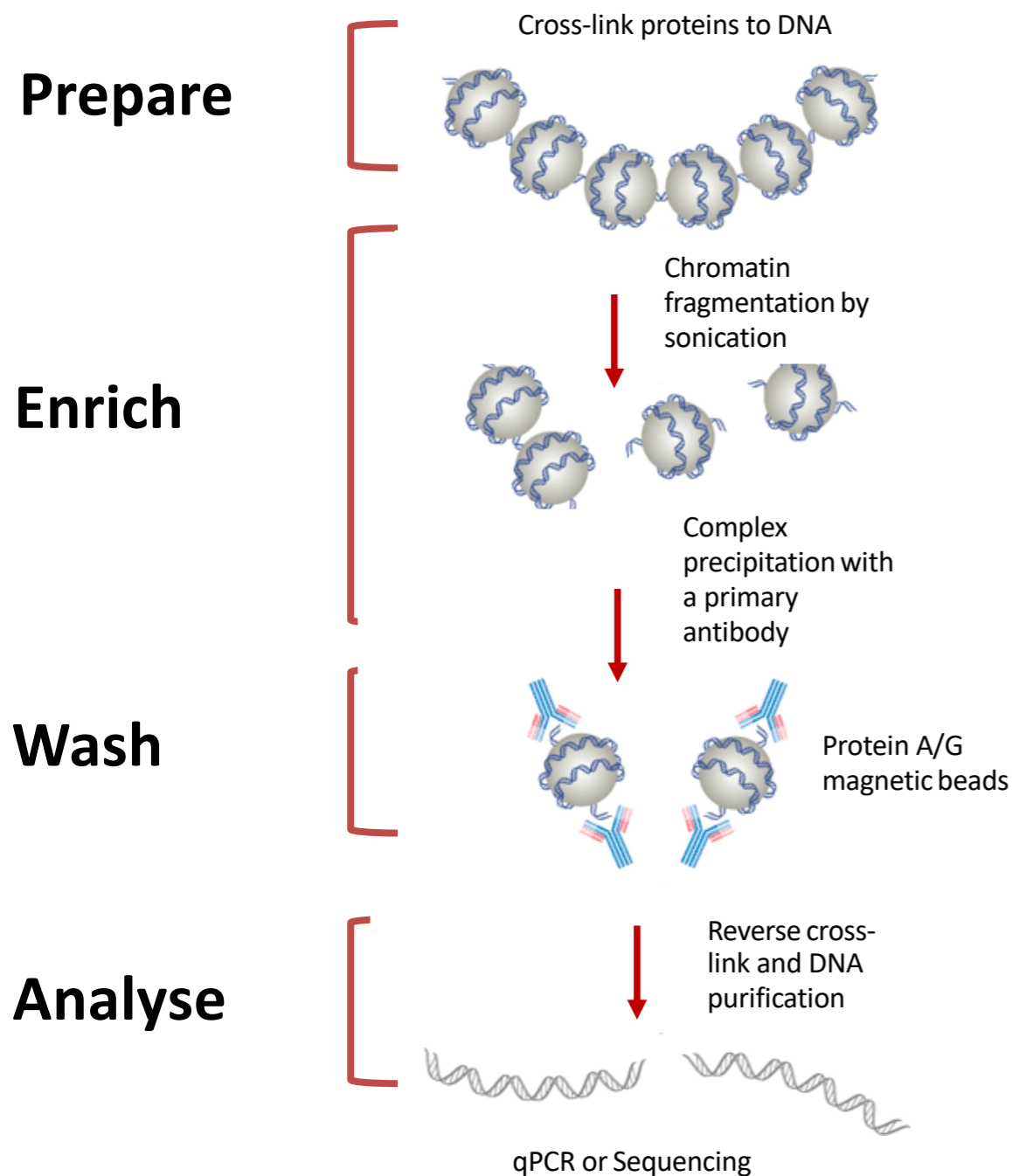
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## 1.Preparation: crosslinking

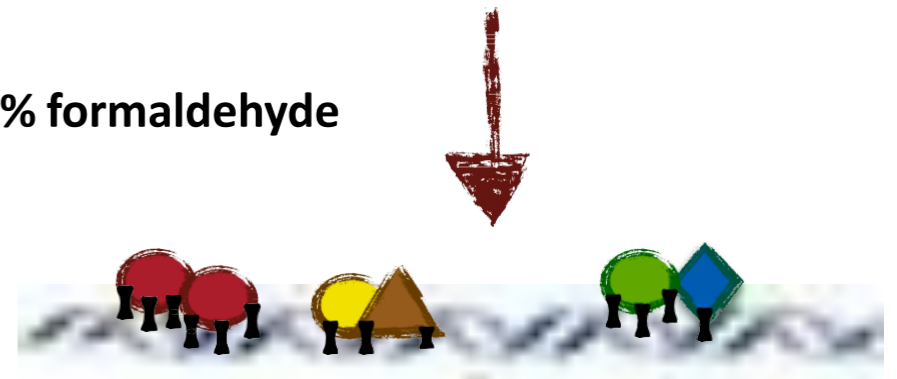
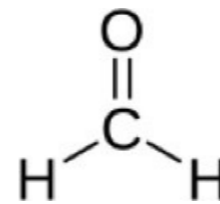
The cross-linking is an experimental procedure that convert in **covalent** all the the weak and non-covalent interactions between DNA - PROTEINS and PROTEIN-PROTEIN



UV-Crosslinking  
Formaldehyde  
Glutaraldehyde



1% formaldehyde

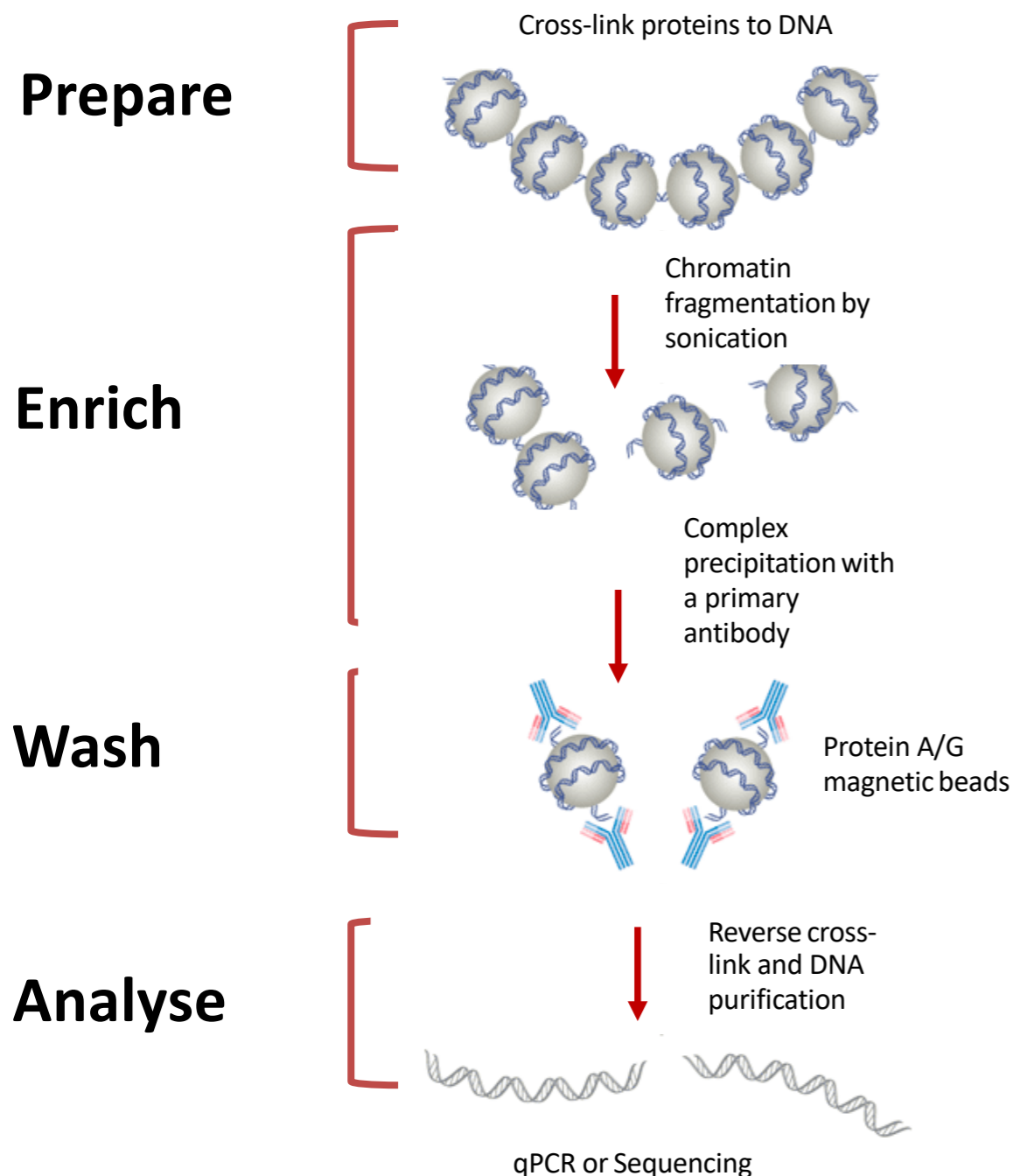


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## 1. Preparation: crosslinking



## Formaldehyde Cross-linking

- Formaldehyde is an organic compound. It is water soluble and penetrates biological membranes. It targets **primary aminogroups** (i.e. lysines in proteins, side chains of A,C,G in DNA)
- It crosslinks both **protein-nucleic acids**, **nucleic acids-nucleic acids** and **protein-protein**
- The crosslinking is reversible (65.C reverse protein-DNA; 100.C reverse protein-protein)
- Reaction is stopped by providing an excess of primary amino groups (0.125M glycine)

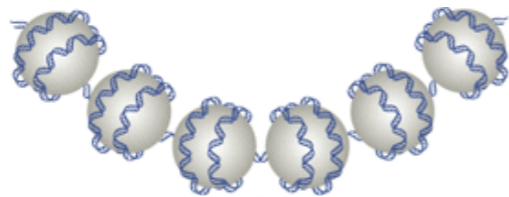
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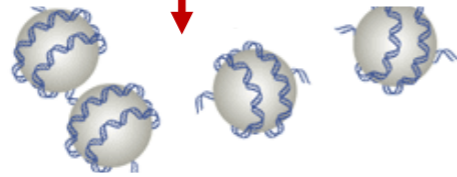
**Prepare**

Cross-link proteins to DNA



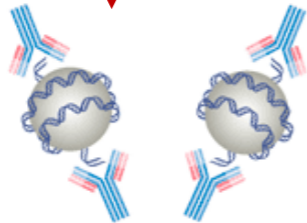
**Enrich**

Chromatin fragmentation by sonication



**Wash**

Complex precipitation with a primary antibody



Protein A/G magnetic beads

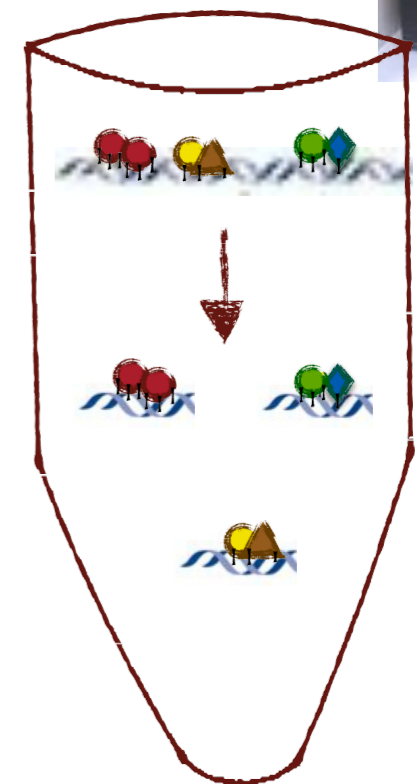
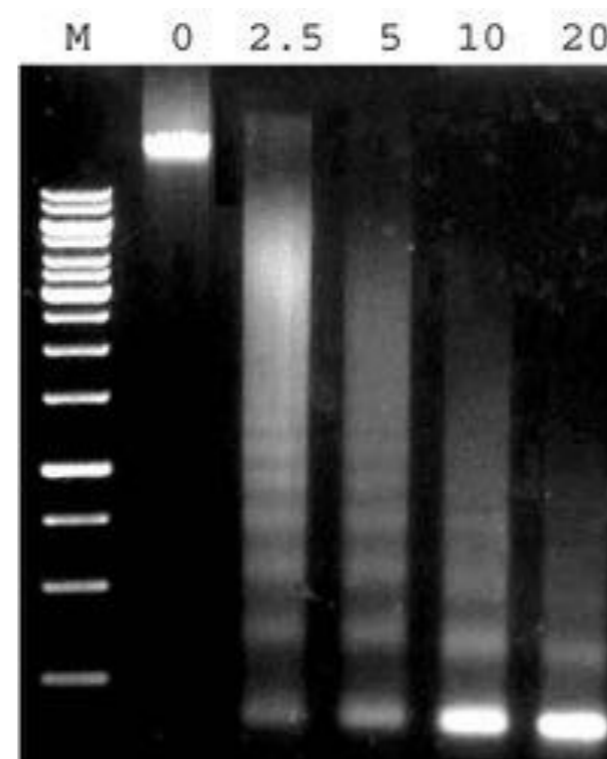
Reverse cross-link and DNA purification



qPCR or Sequencing

## 2. Preparation: sonication

The DNA-protein complexes (chromatin-protein) are then sheared into ~500 bp DNA fragments by **sonication** or (nuclease digestion).



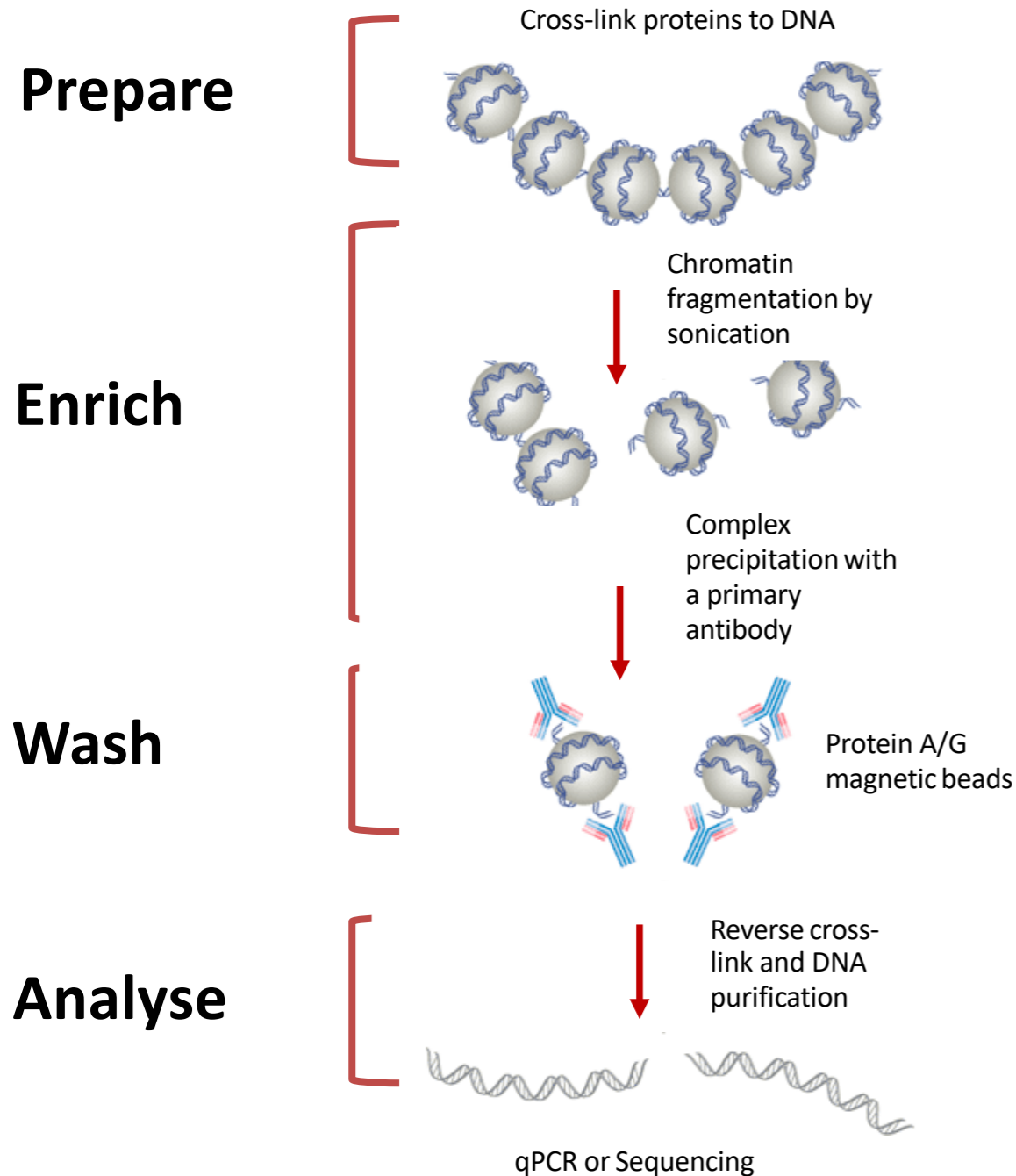
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AIM: Identification of the genomic loci bound to a DNA binding protein. (Bait Protein/ output :DNA)

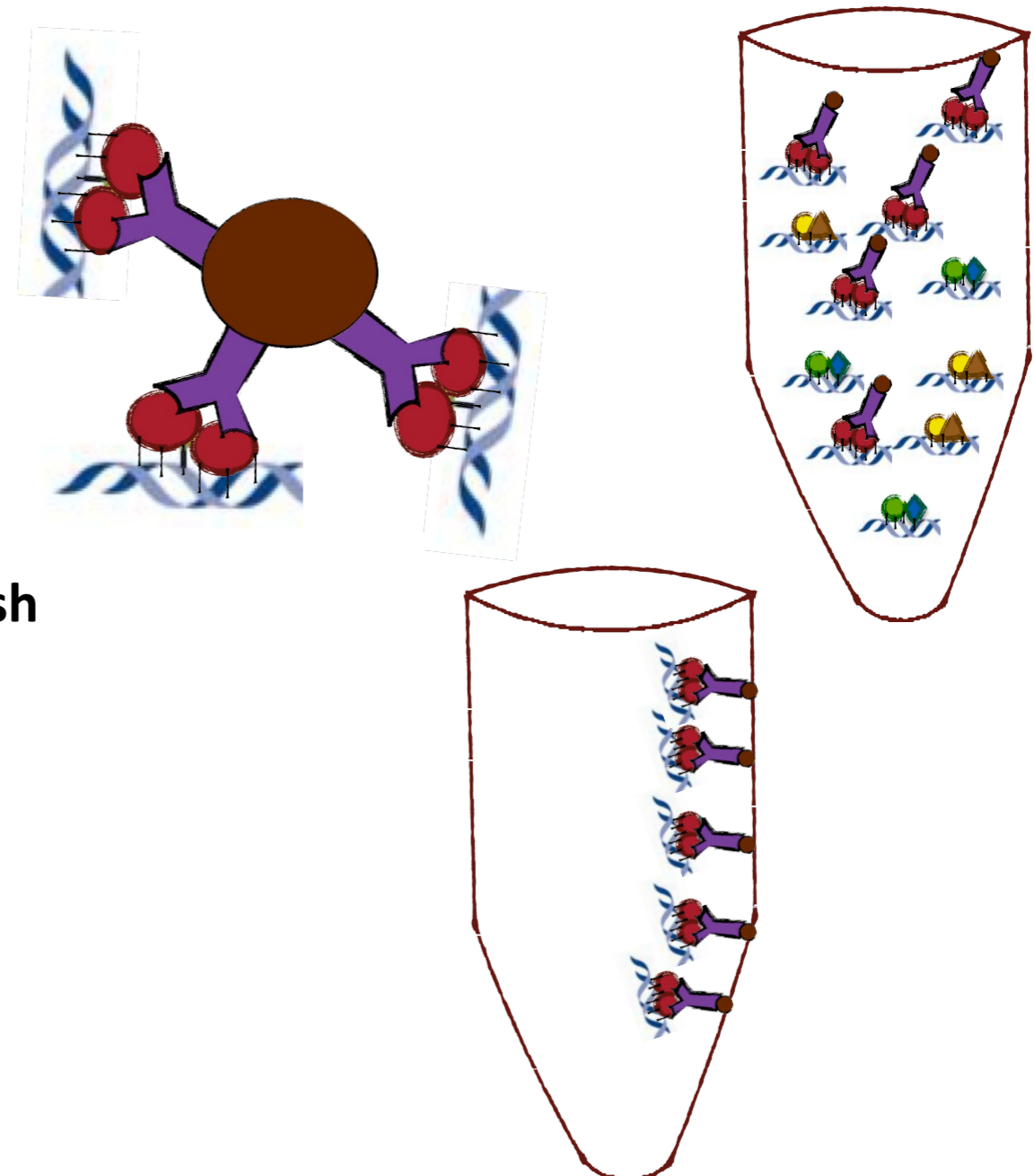
**Chromatin Immunoprecipitation (ChIP)** is an immunoprecipitation technique used to investigate the interaction between proteins and DNA in the cell. It aims to determine whether specific proteins are associated with specific genomic regions, such as transcription factors on promoters.

## 2. Antibody binding

Antibody binding several hours (or over night) at 4 degrees.



## 3. Wash





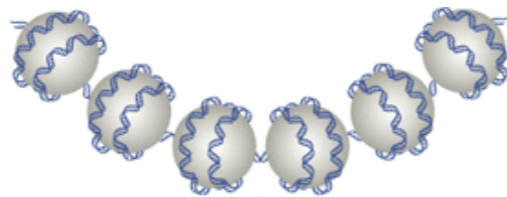
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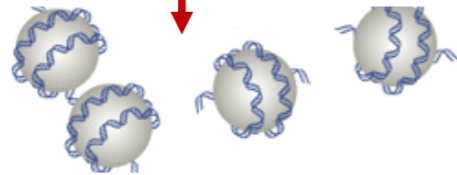
**Prepare**

Cross-link proteins to DNA

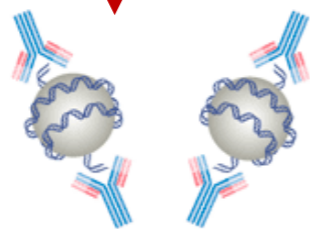


**Enrich**

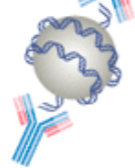
Chromatin fragmentation by sonication



Complex precipitation with a primary antibody



Protein A/G magnetic beads



**Wash**

**Analyse**

Reverse cross-link and DNA purification

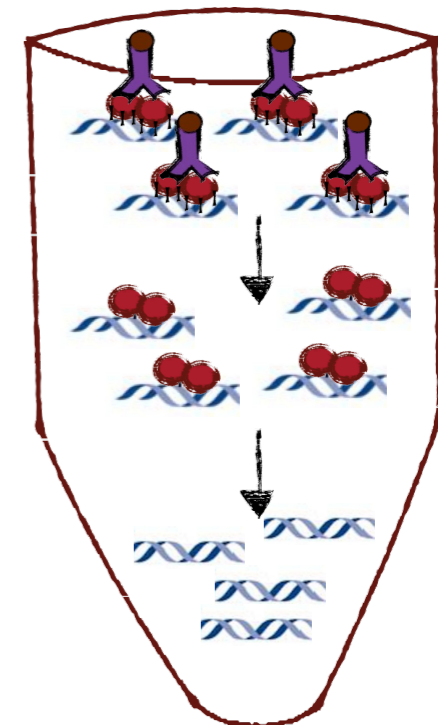


qPCR or Sequencing

## 4. Reverse crosslinking

The cross-linking with formaldehyde is able to be removed through the incubation of the extract at High temperature. (65.C reverse protein-DNA; 100.C reverse protein-protein this step allows the detach of the protein from the DNA that is subsequently purified and analyzed

## 5. Proteinase K treatment, Rnase A treatment and DNA purification

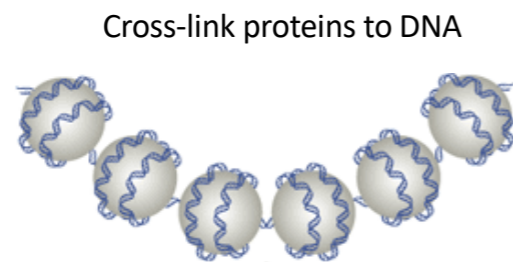


# ChIP

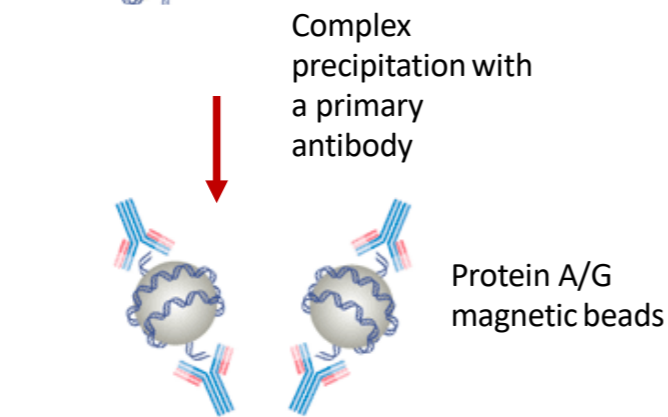
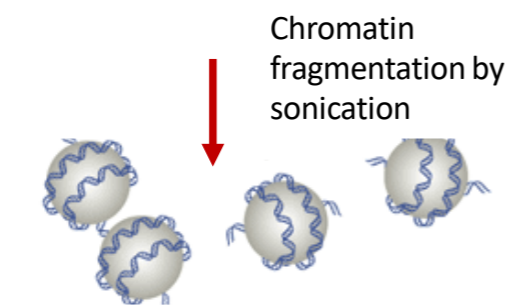
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## Prepare

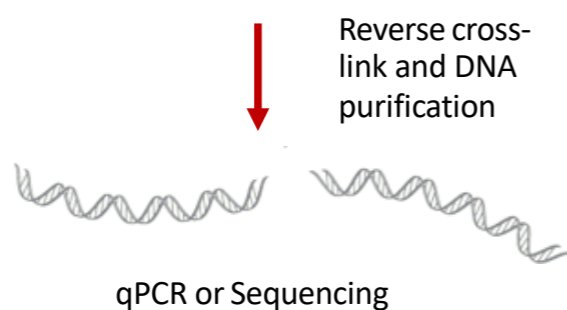


## Enrich



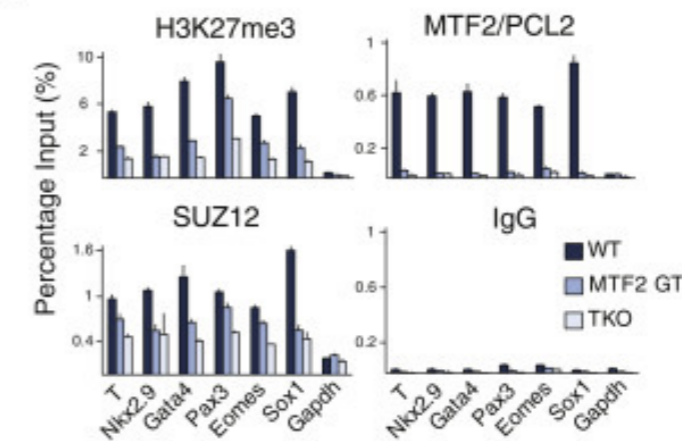
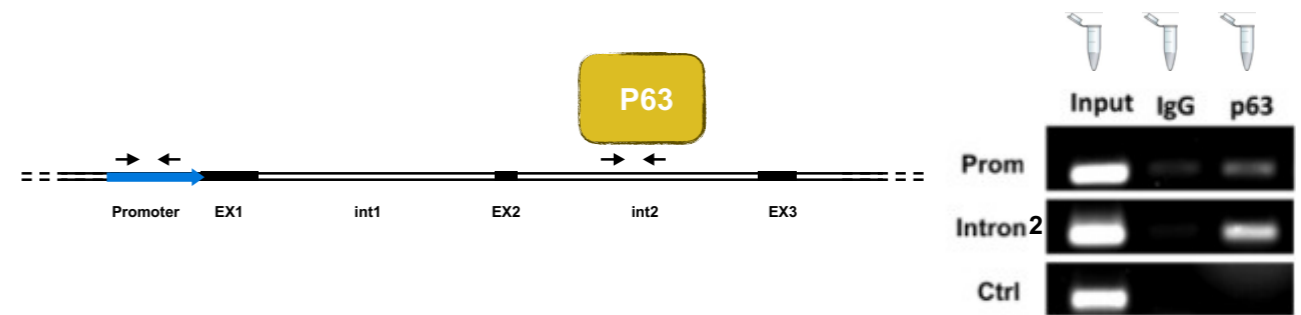
## Wash

## Analyse



## 6. DNA analysis PCR

The isolated DNA can be quantified by PCR using specific probes. This allows the analysis of a specific region in multiple samples



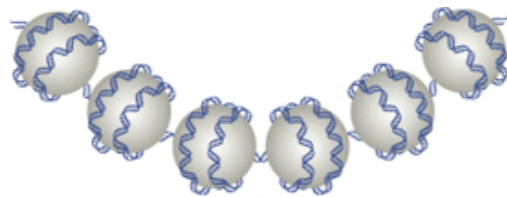
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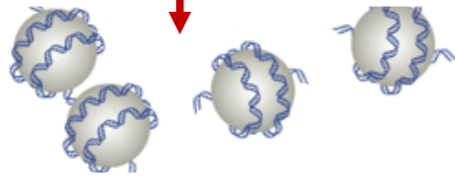
**Prepare**

Cross-link proteins to DNA

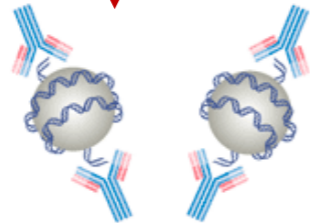


**Enrich**

Chromatin fragmentation by sonication

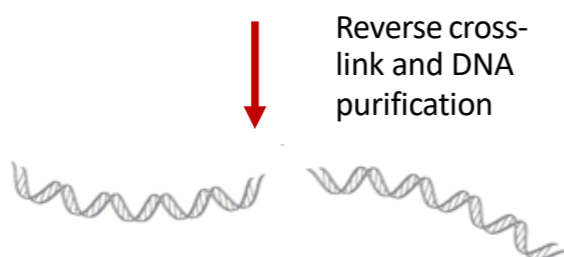


Complex precipitation with a primary antibody



**Wash**

Protein A/G magnetic beads

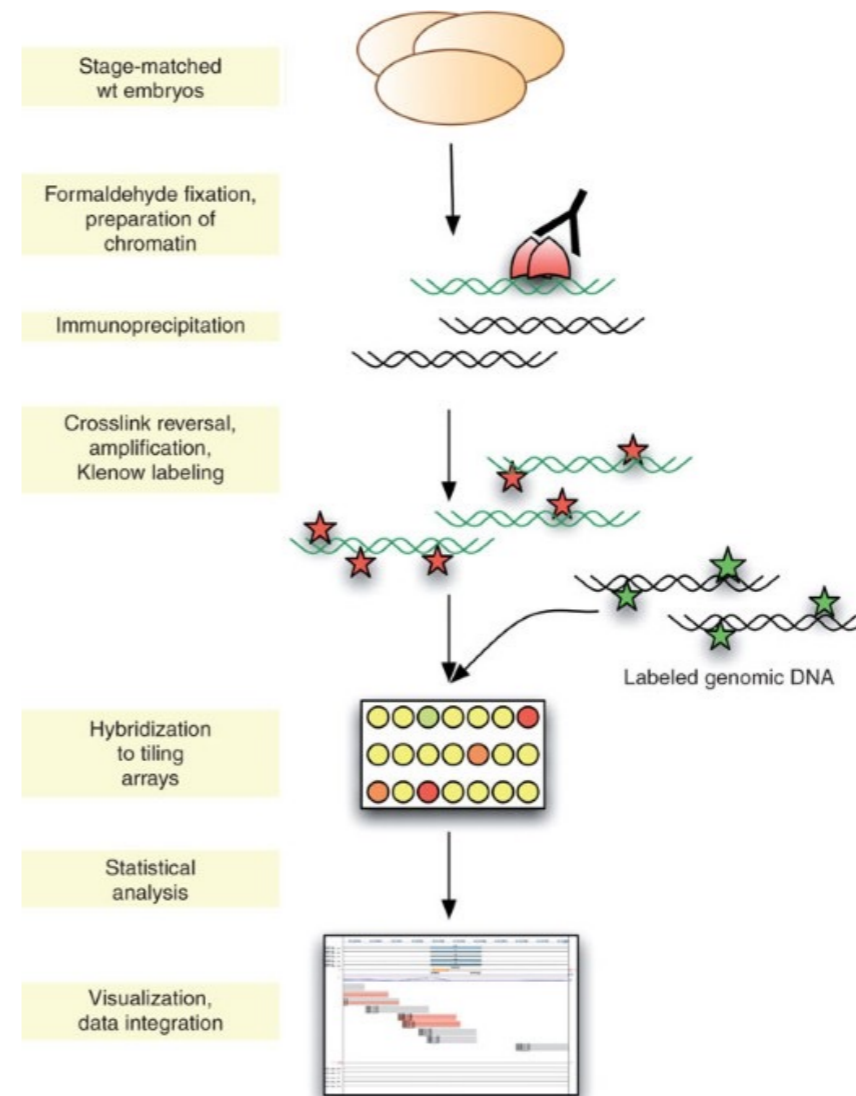


**Analyse**

Reverse cross-link and DNA purification

## 6. DNA analysis ChIP on chip

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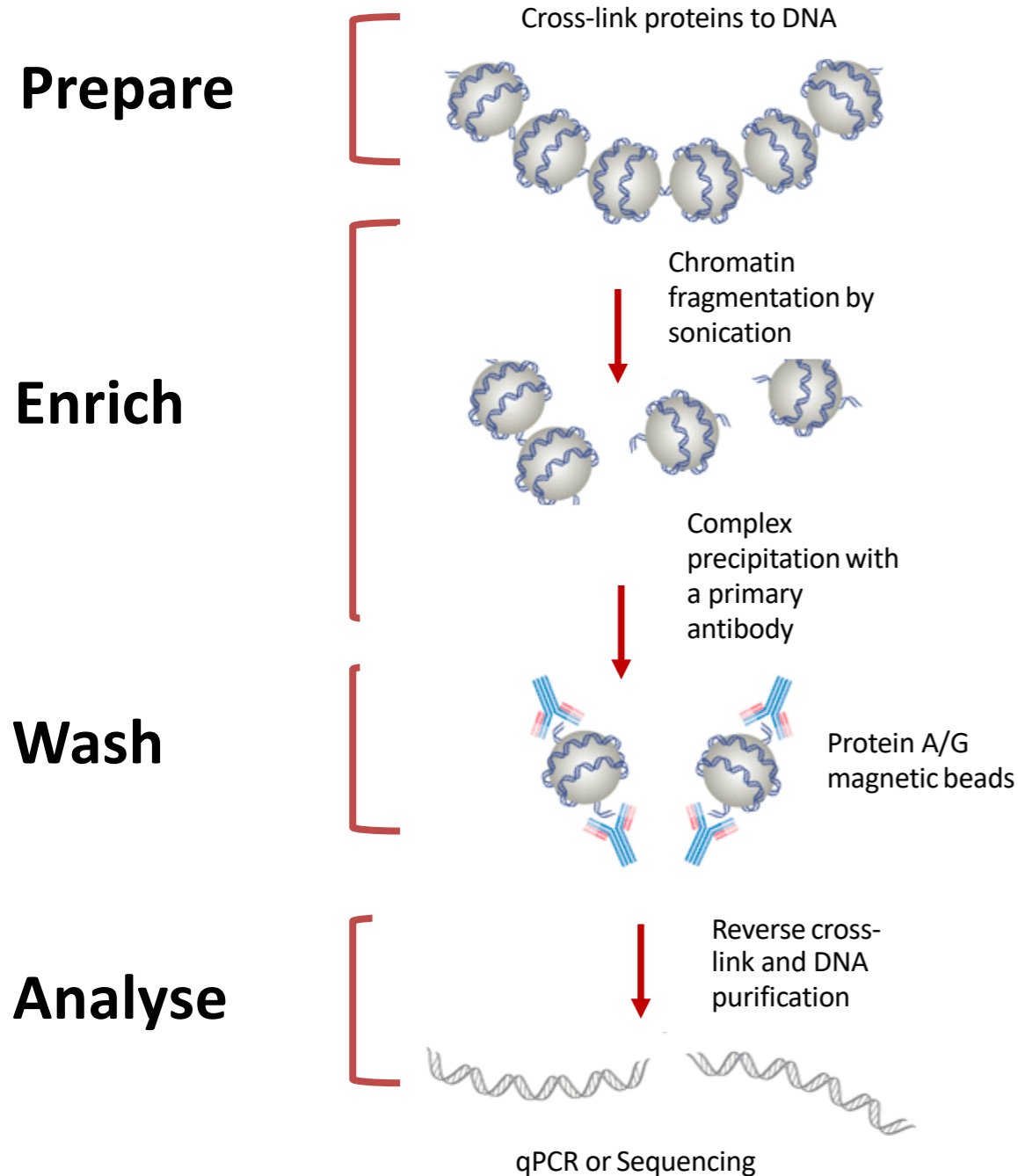
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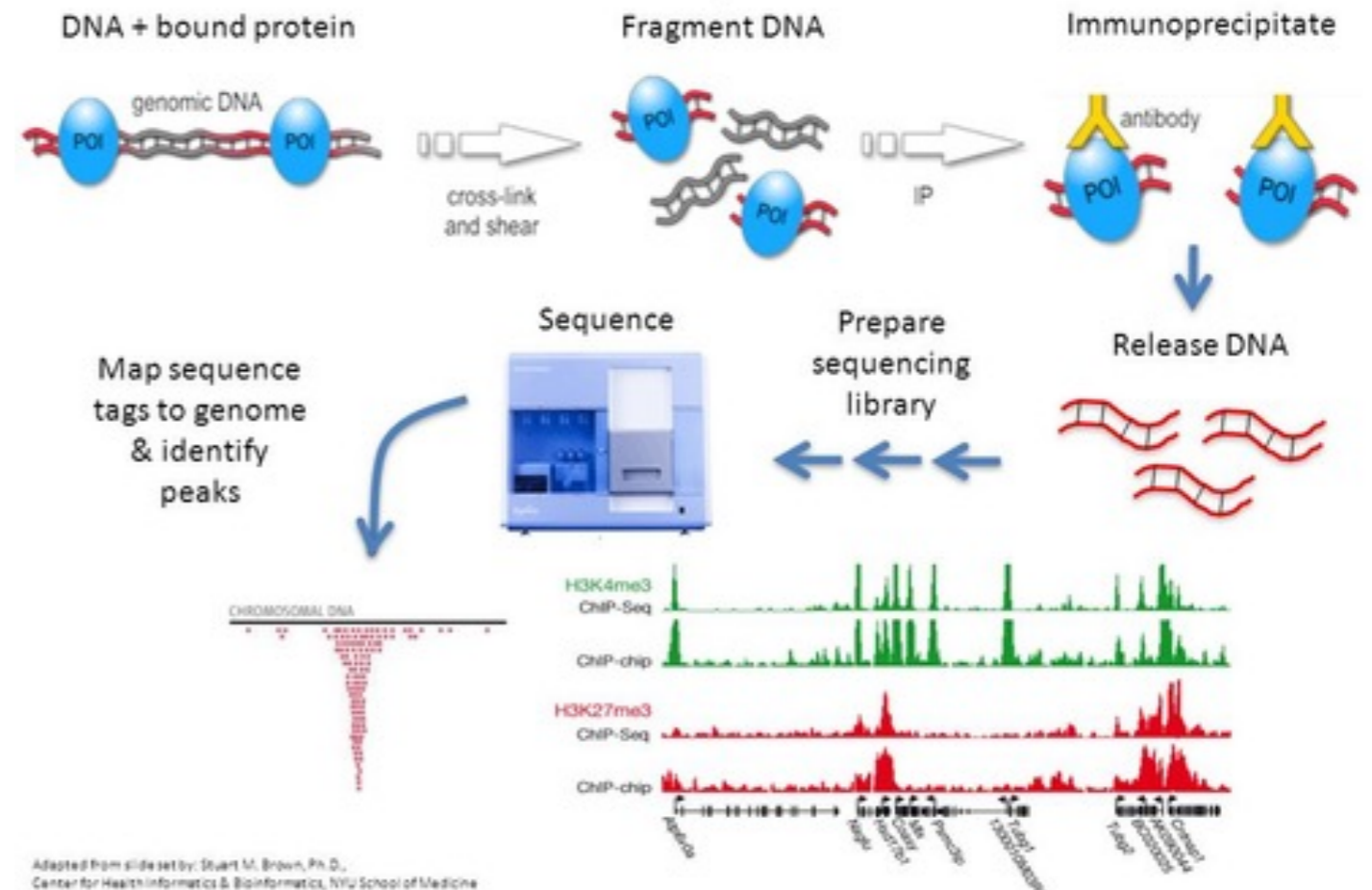
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## 6. DNA analysis ChIP seq

We can isolate the DNA and sequence every each single piece of DNA attached to this protein



## ChIP-seq overview



Adapted from slide set by Stuart M. Brown, Ph.D., Center for Health Informatics & Bioinformatics, NYU School of Medicine

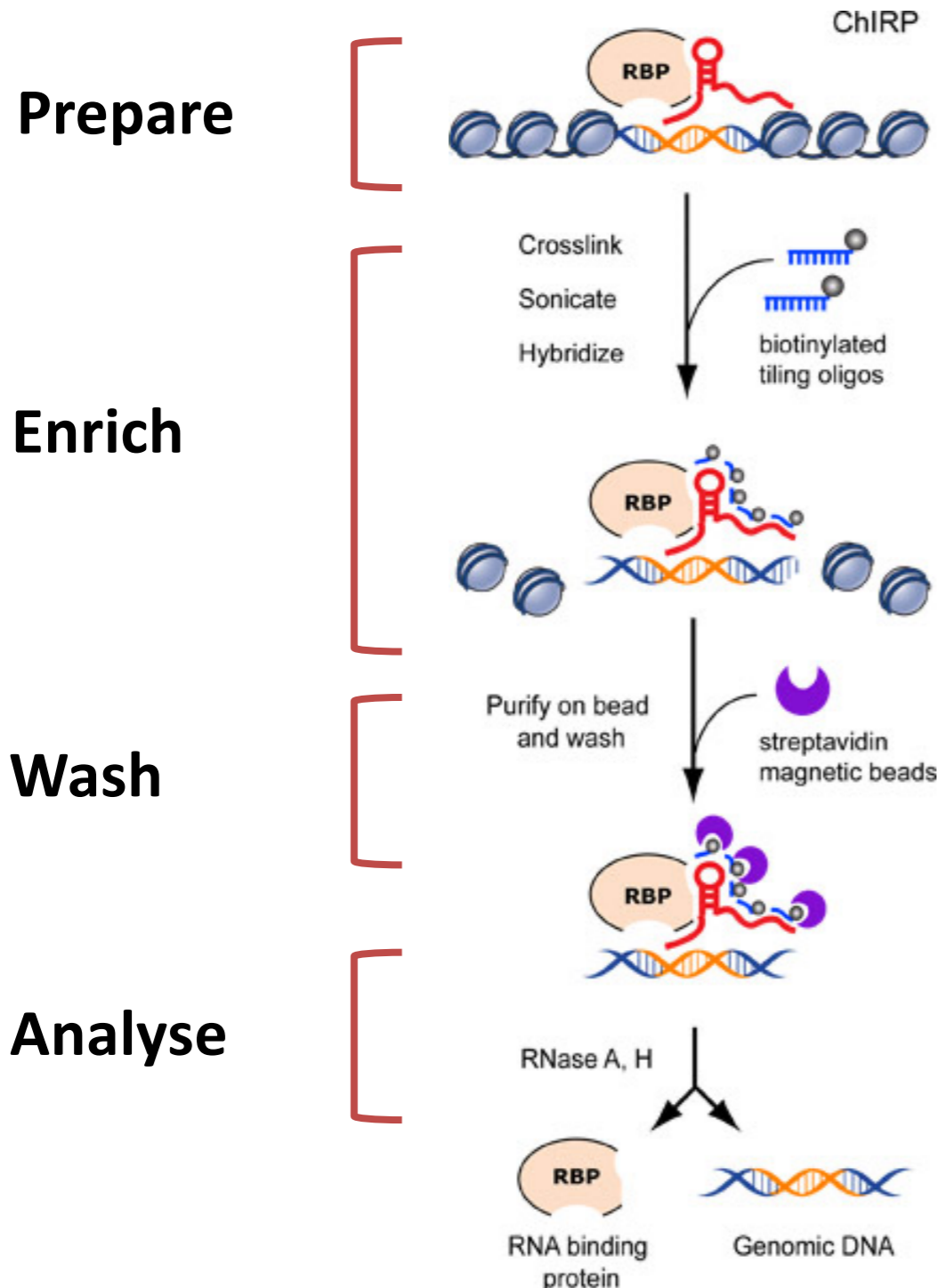
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# ChIRP

**AIM: Identification of RNA interactions with the chromatin. Bait: RNA output: DNA/protein)**

**ChIRP (Chromatin Isolation by RNA Purification)** a pull down technique used to investigate the interaction between RNA (usually ncRNAs and chromatin). Not only serve to determine interaction but also place of the interaction.



- Chromatin associated lncRNAs
- Discrimination between *cis* and *trans* action
- The amount of cellular extract depends on the abundance of the lncRNA

# ChIRP

**AIM: Identification of RNA interactions with the chromatin. Bait: RNA output: DNA/protein)**

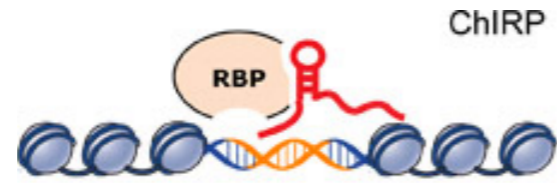
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## 1. Crosslinking

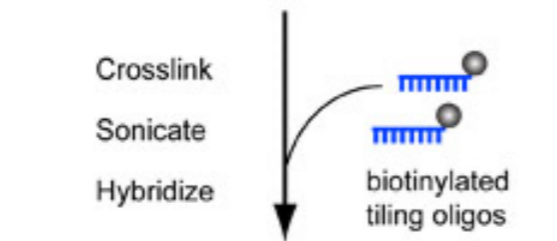
**1% of glutaraldehyde**

*Keep RNA*

**Prepare**



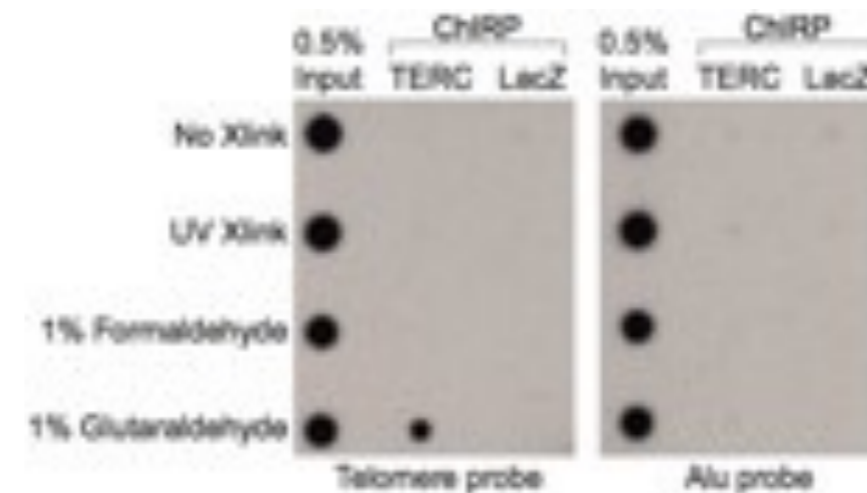
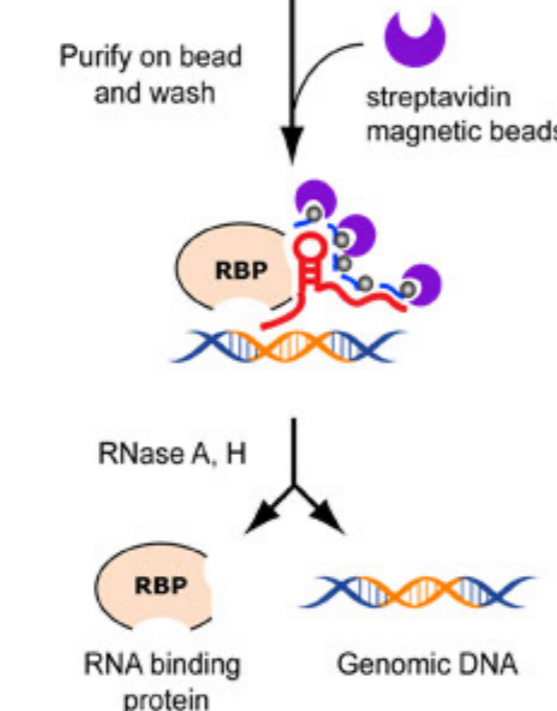
**Enrich**



**Wash**



**Analyse**



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**ChIRP (Chromatin Isolation by RNA Purification)** a pull down technique used to investigate the interaction between RNA (usually ncRNAs and chromatin). Not only serve to determine interaction but also place of the interaction.

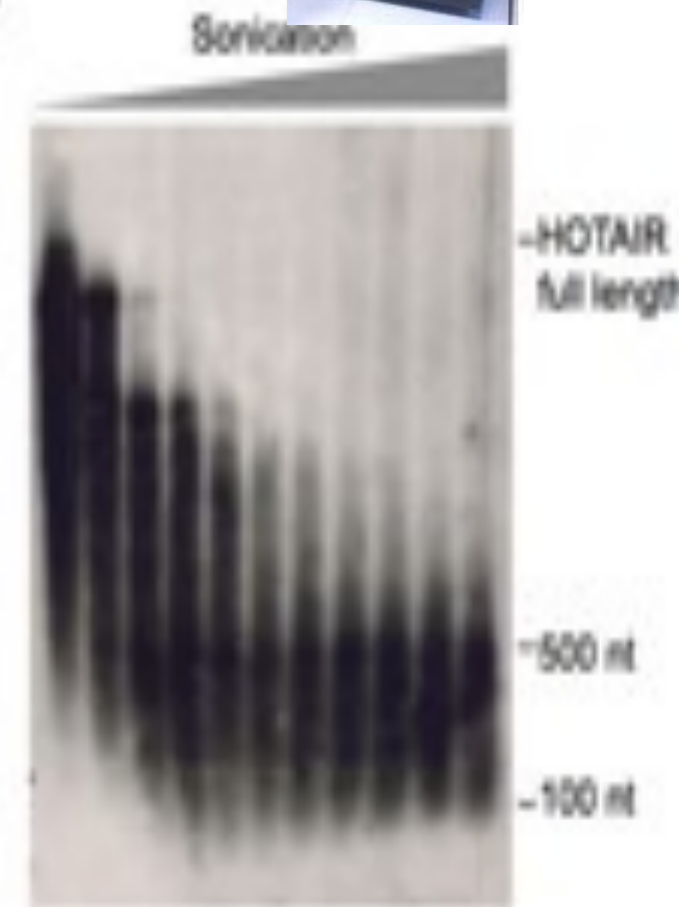
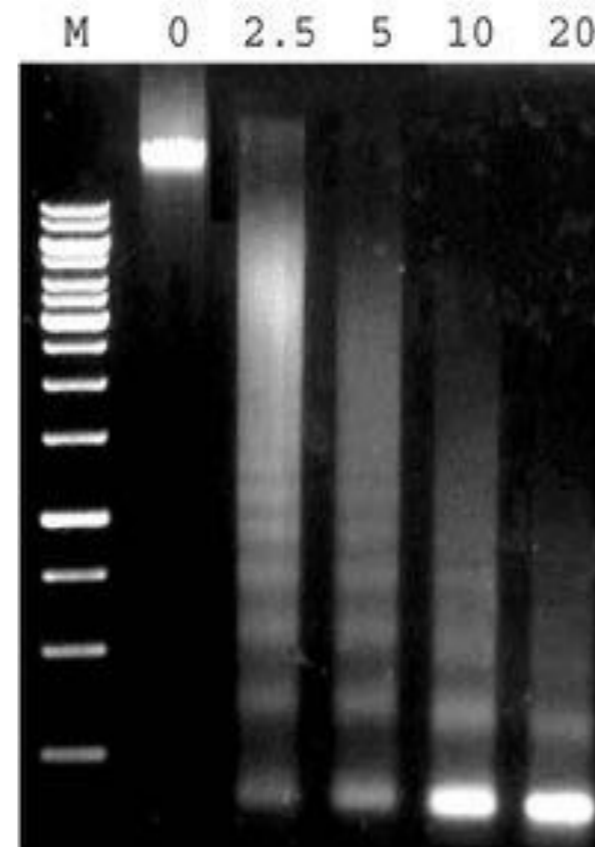
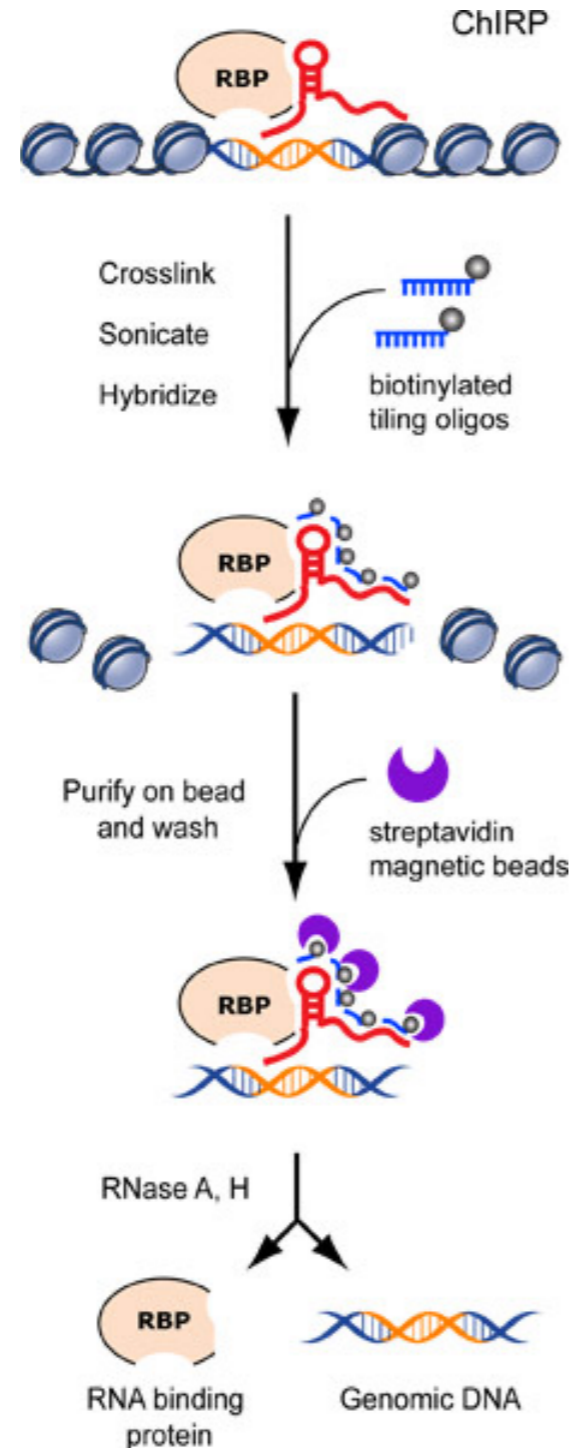
## 1. Sonication

Prepare

Enrich

Wash

Analyse



Northern Blot





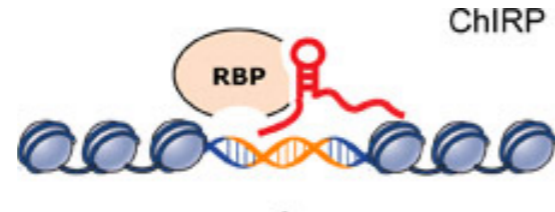
# ChIRP

AIM: Identification of RNA interactions with the chromatin. Bait: RNA output: DNA/protein)

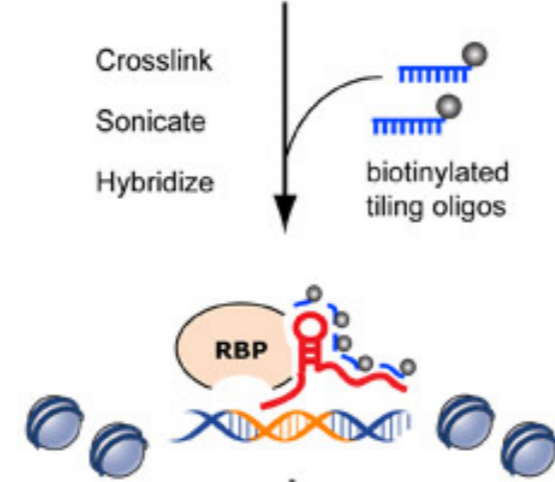
**ChIRP (Chromatin Isolation by RNA Purification)** a pull down technique used to investigate the interaction between RNA (usually ncRNAs and chromatin). Not only serve to determine interaction but also place of the interaction.

## 1. Probe design

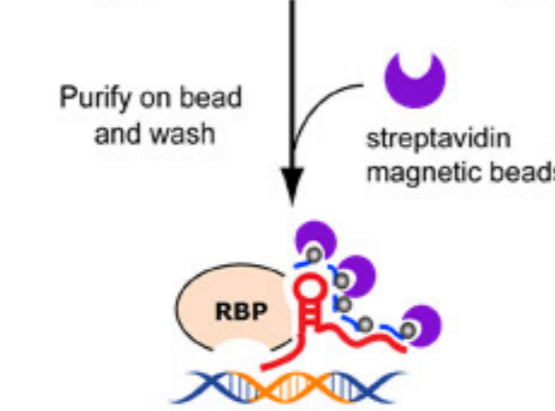
Prepare



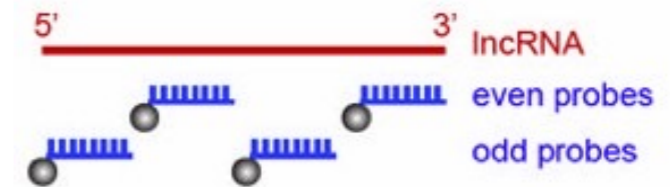
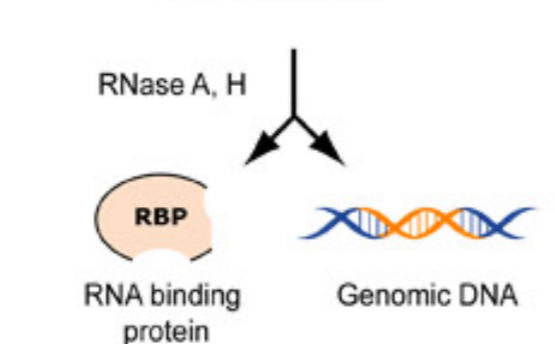
Enrich



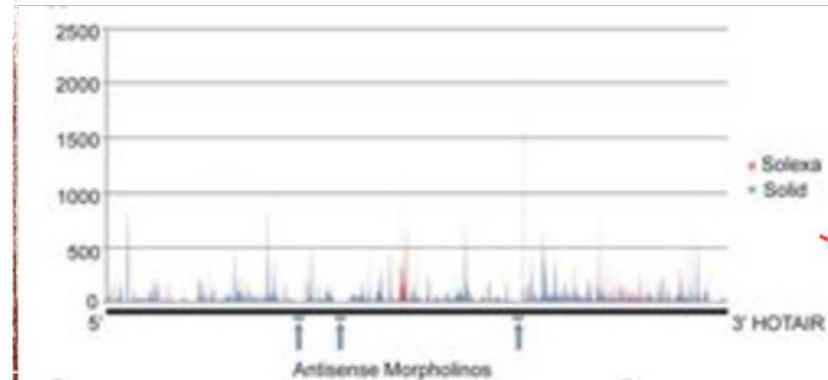
Wash



Analyse



HOTAIR ChIRP



Antisense morpholinos are designed against **structurally open regions** of HOTAIR RNA. Peaks denote secondary structures previously determined by Parallel Analysis of RNA Structure (PARS) (Kertesz et al., 2010).

random design

Ci Chu et al. 2011

Custom Probe Sets

General Info Technical Specs Related Links Literature References

Custom Probe Sets

Design your own Custom Stellaris® RNA FISH Probe Set to detect your RNA of interest. By using our Stellaris RNA FISH Probe Designer, you can design up to 48 individually fluorescently labeled oligonucleotides to bind along and visualize your target RNA. Before you begin designing your Stellaris RNA FISH Probe Set, check to see if we have already designed a set against your target of interest by entering your target into our [TargetCheck](#) page. We also offer Stellaris® RNA FISH Probe Sets to be used as controls for your experiment and [Stellaris Beads](#), which are the perfect accompaniment to your probe sets, allowing for easy, more robust RNA detection and analysis.

STELLARIS RNA FISH PROBE DESIGNER

Product Information: One set of Stellaris RNA FISH Probe Sets contains a blend of up to 48 oligos labeled with a fluorophore. Probe sets are purified and ready to use.

Minimum Amount: 1 unit of purified oligos (200-400 hybridization experiments depending on optimal working dilutions for each target)

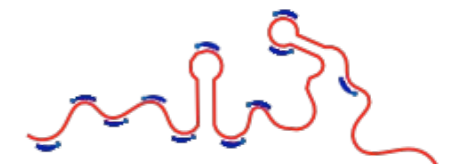
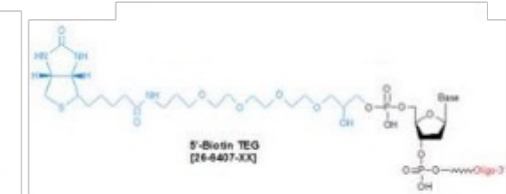
Price: \$175 for all custom Stellaris probe sets.

Turnaround time: 12 to 17 business days.

Learn more about the Stellaris RNA FISH technology by reading our [Stellaris Getting Started Guide](#) and [designing your probe sets](#).

STELLARIS PROBE SET EXCEL FORM

If you have the sequence ready to enter!



<https://www.biosearchtech.com/Account/Login?return=/stellarisdesigner/>

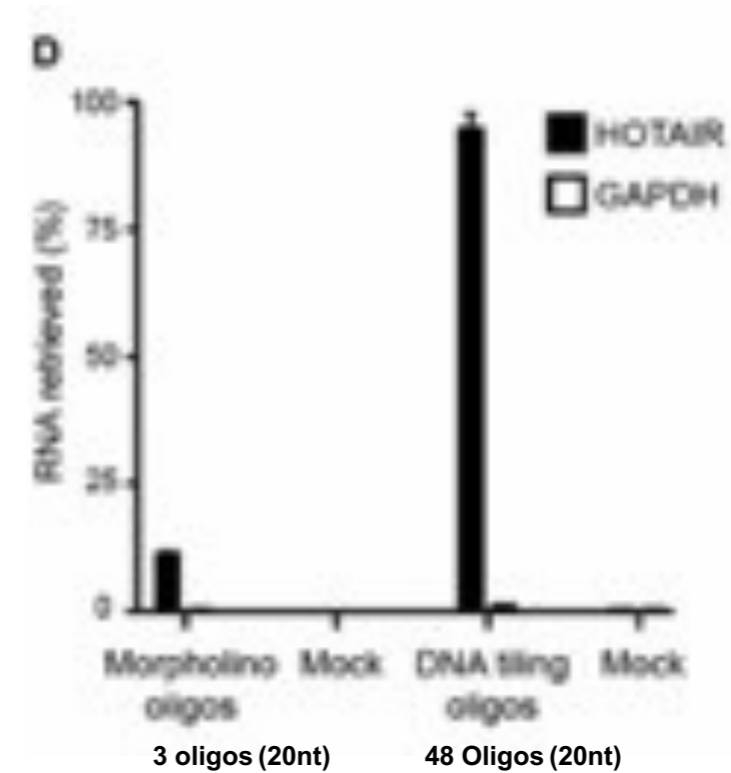
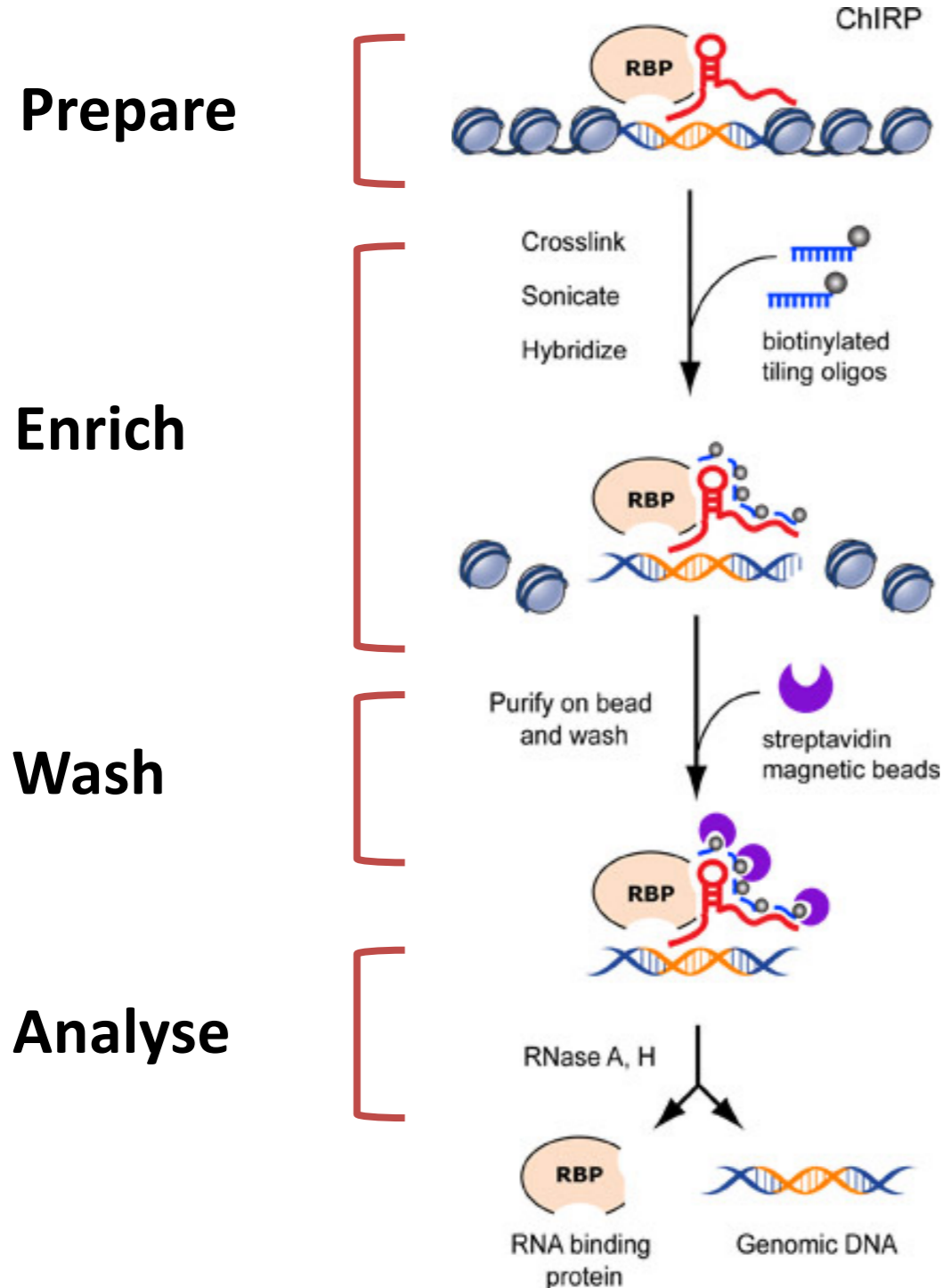
Ci Chu et al. 2011

# ChIRP

**AIM: Identification of RNA interactions with the chromatin. Bait: RNA output: DNA/protein)**

**ChIRP (Chromatin Isolation by RNA Purification)** a pull down technique used to investigate the interaction between RNA (usually ncRNAs and chromatin). Not only serve to determine interaction but also place of the interaction.

## 1. Probe design



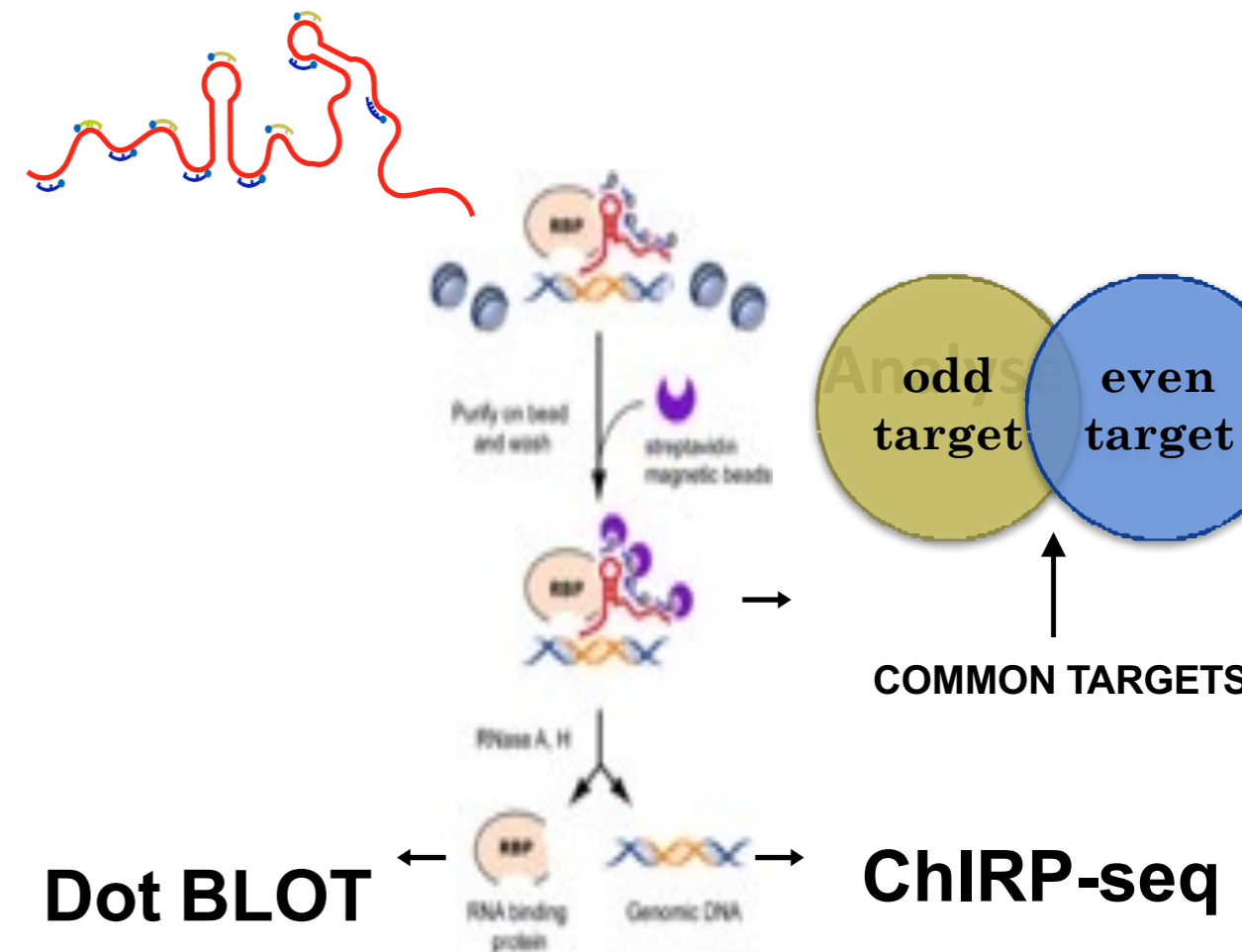
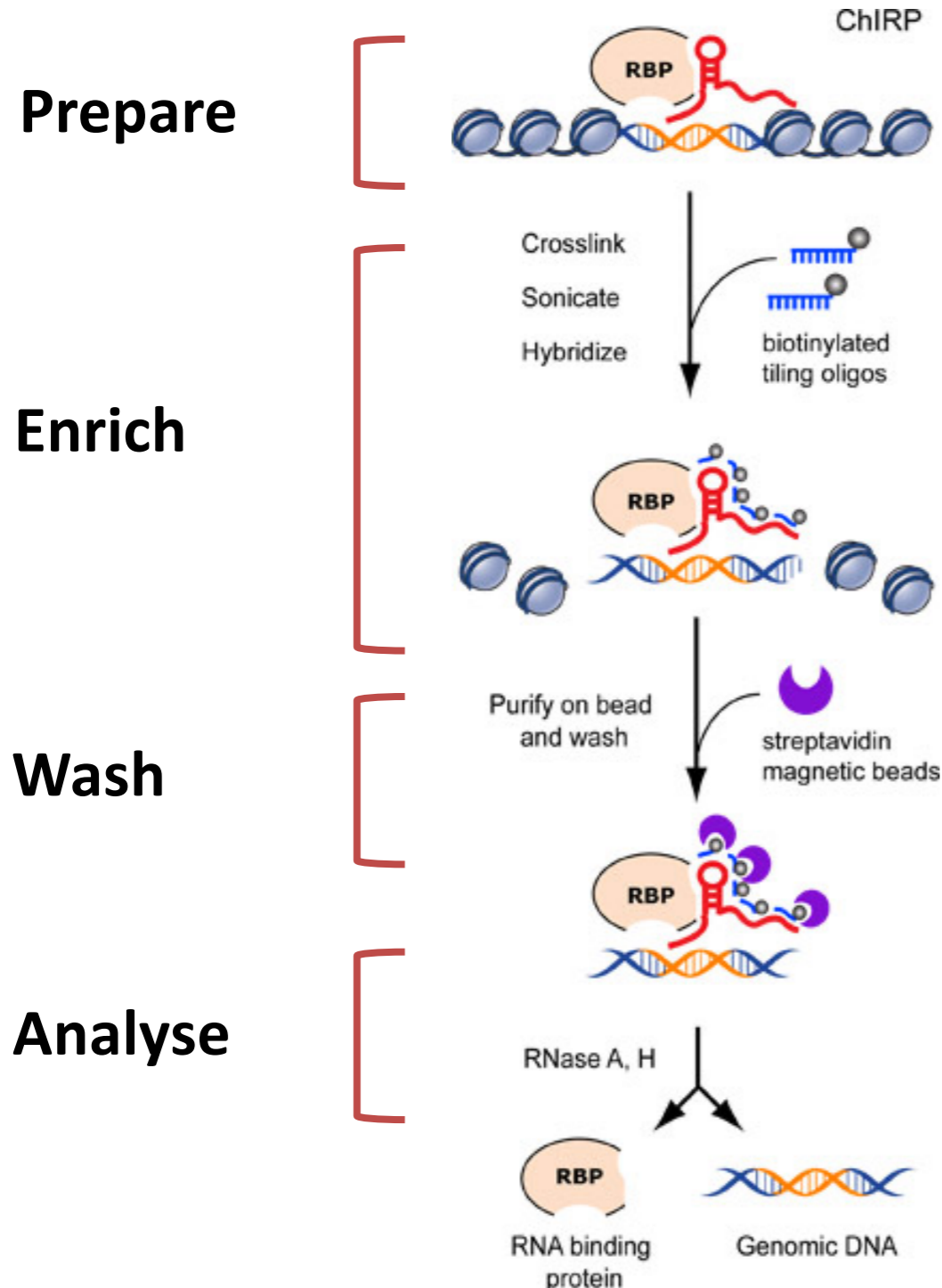
**The more probes you have, the better.**

# ChIRP

AIM: Identification of RNA interactions with the chromatin. Bait: RNA output: DNA/protein)

**ChIRP (Chromatin Isolation by RNA Purification)** a pull down technique used to investigate the interaction between RNA (usually ncRNAs and chromatin). Not only serve to determine interaction but also place of the interaction.

## 2. Precipitation and wash



# ChIRP

**AIM: Identification of RNA interactions with the chromatin. Bait: RNA output: DNA/protein)**

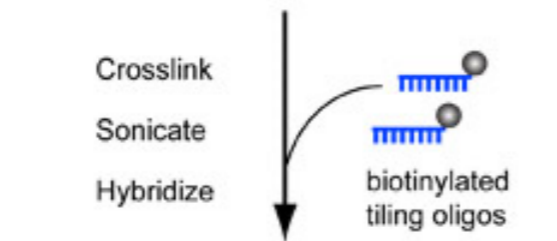
**ChIRP (Chromatin Isolation by Rna Purification)** a pull down technique used to investigate the interaction between RNA (usually ncRNAs and chromatin). Not only serve to determine interaction but also place of the interaction.

## 3. Analysis ChIRP-Seq

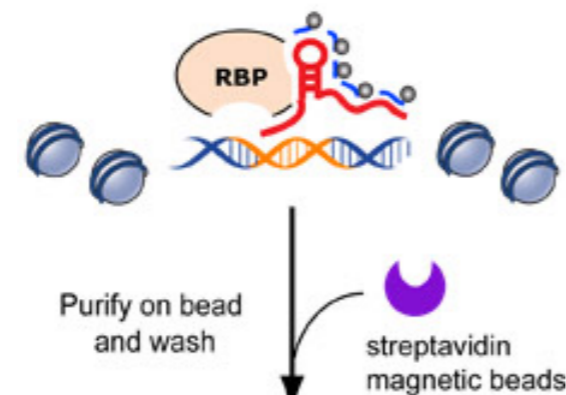
Prepare



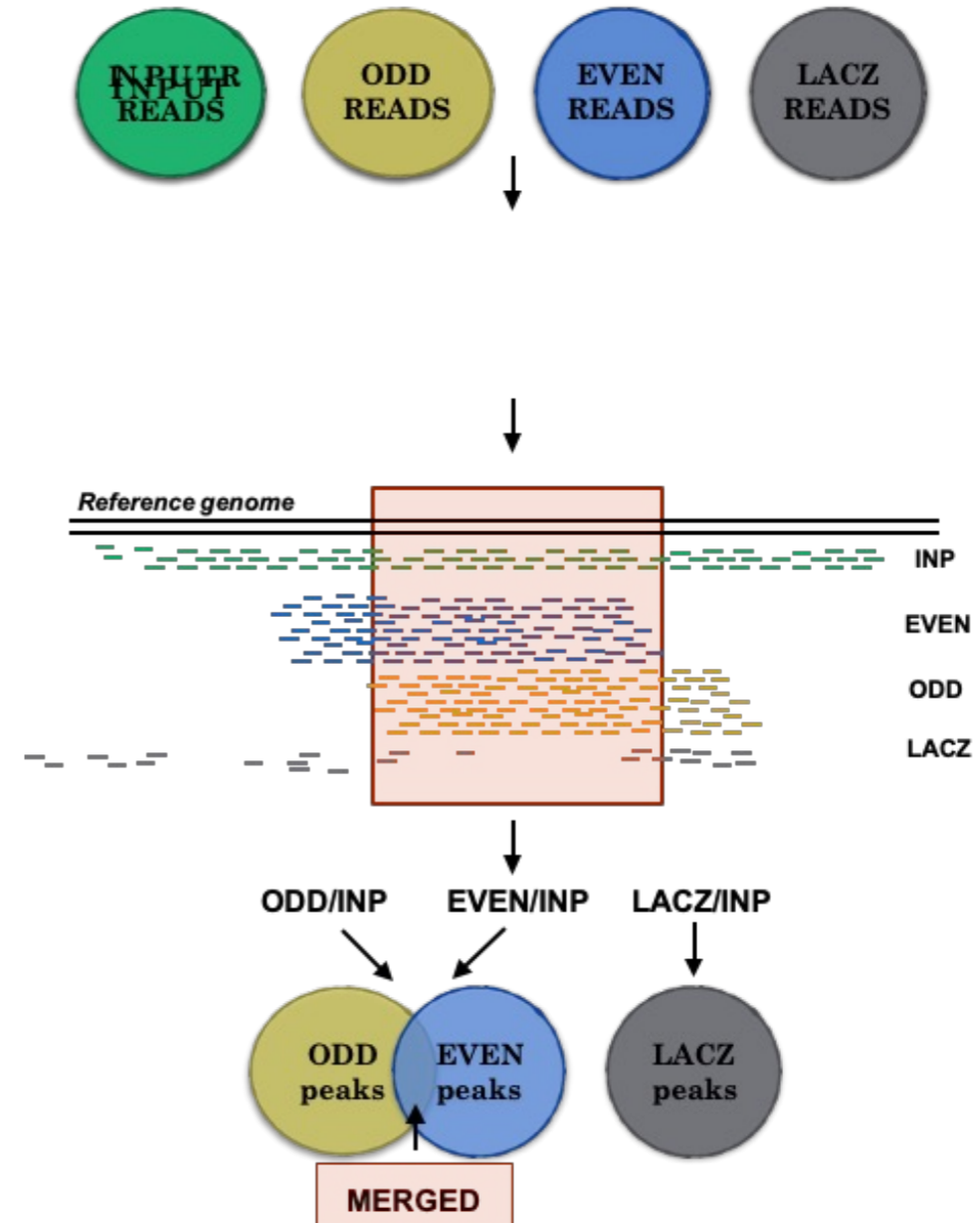
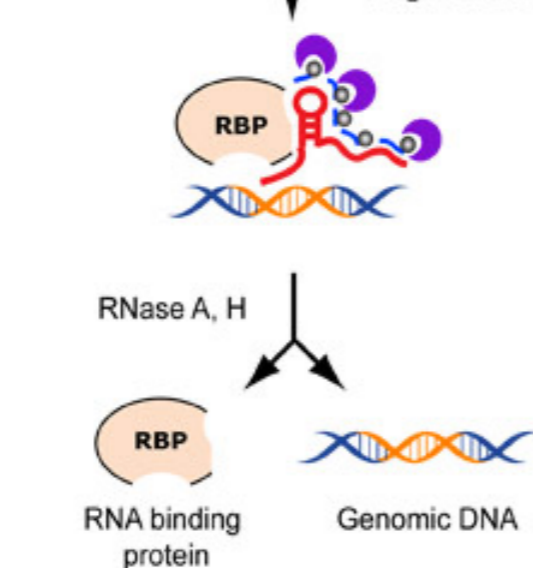
Enrich



Wash

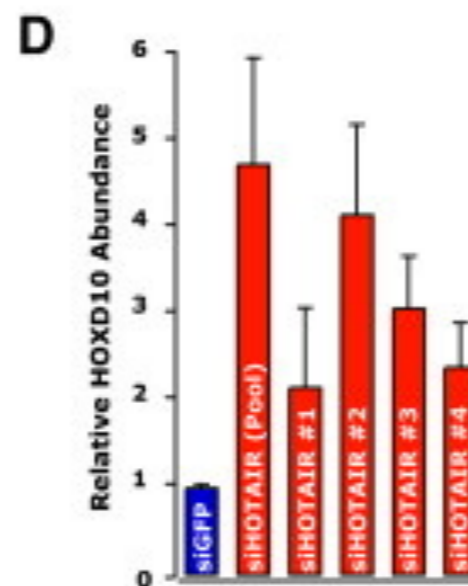
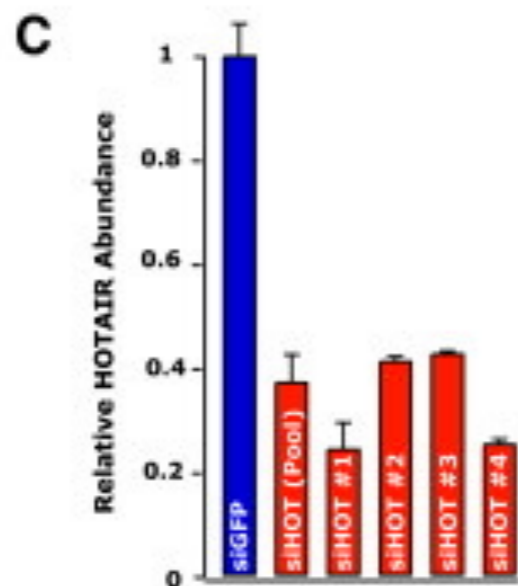
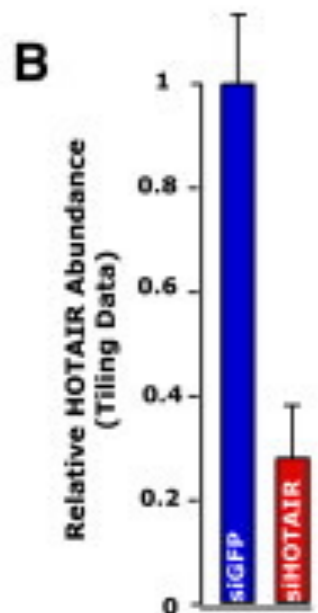
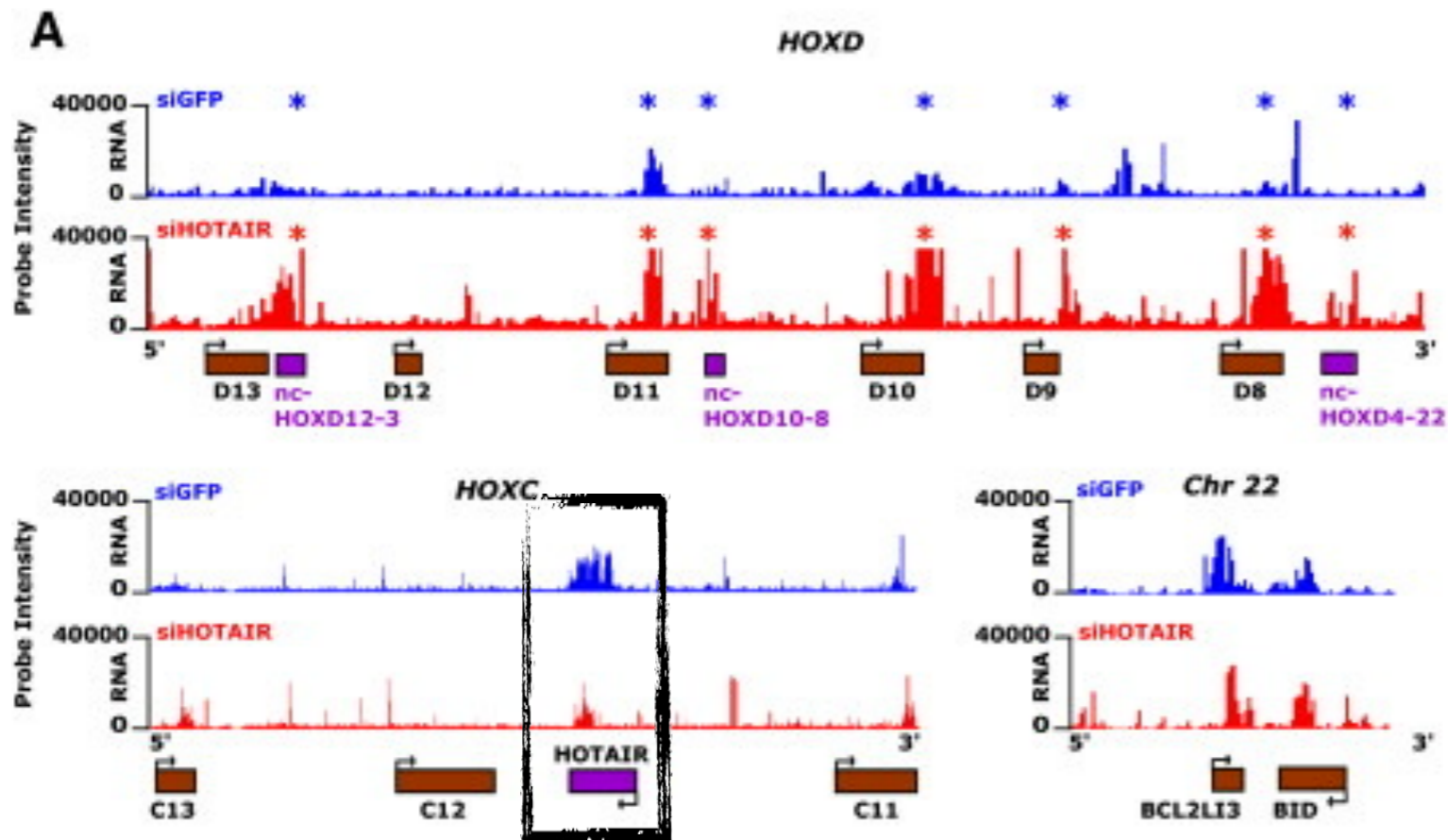


Analyse



# ChIRP: example HOTAIR

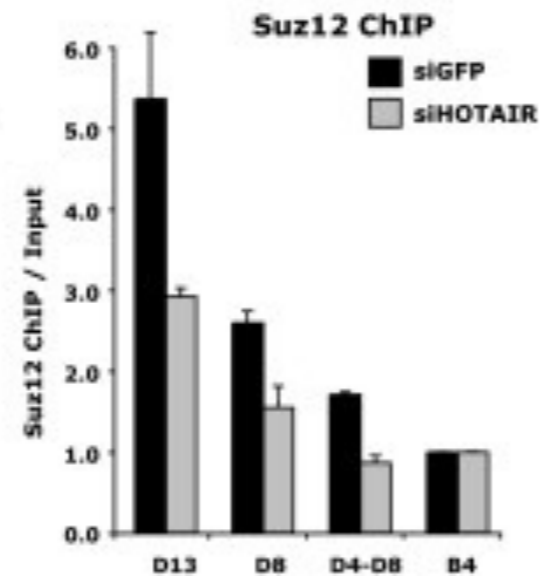
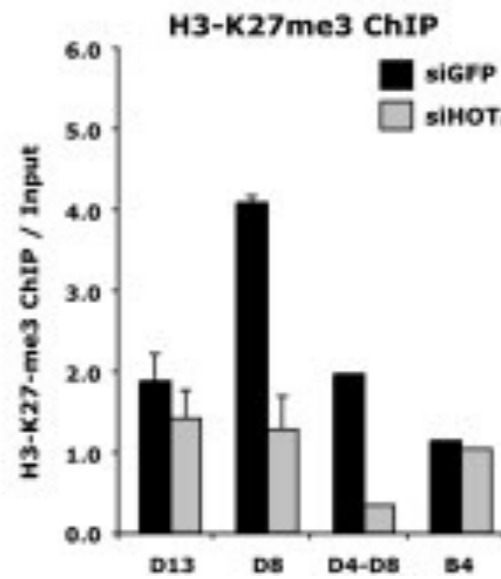
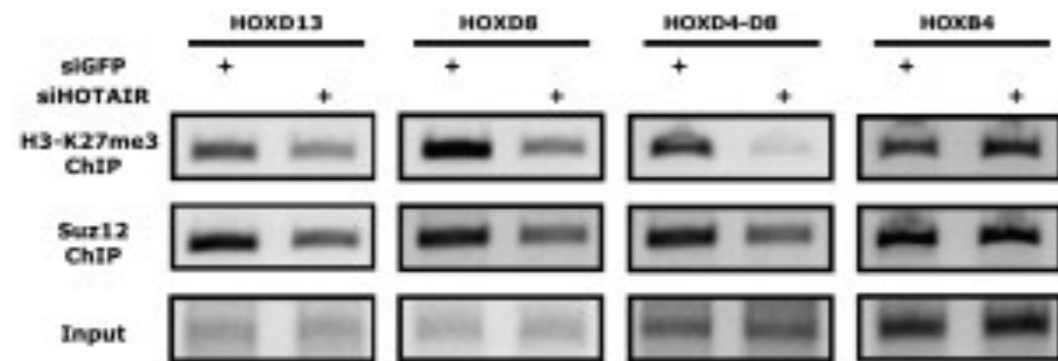
Functional data



# ChIP: example HOTAIR

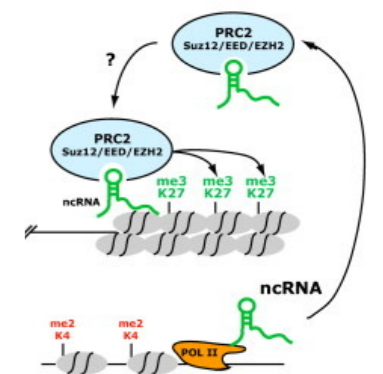
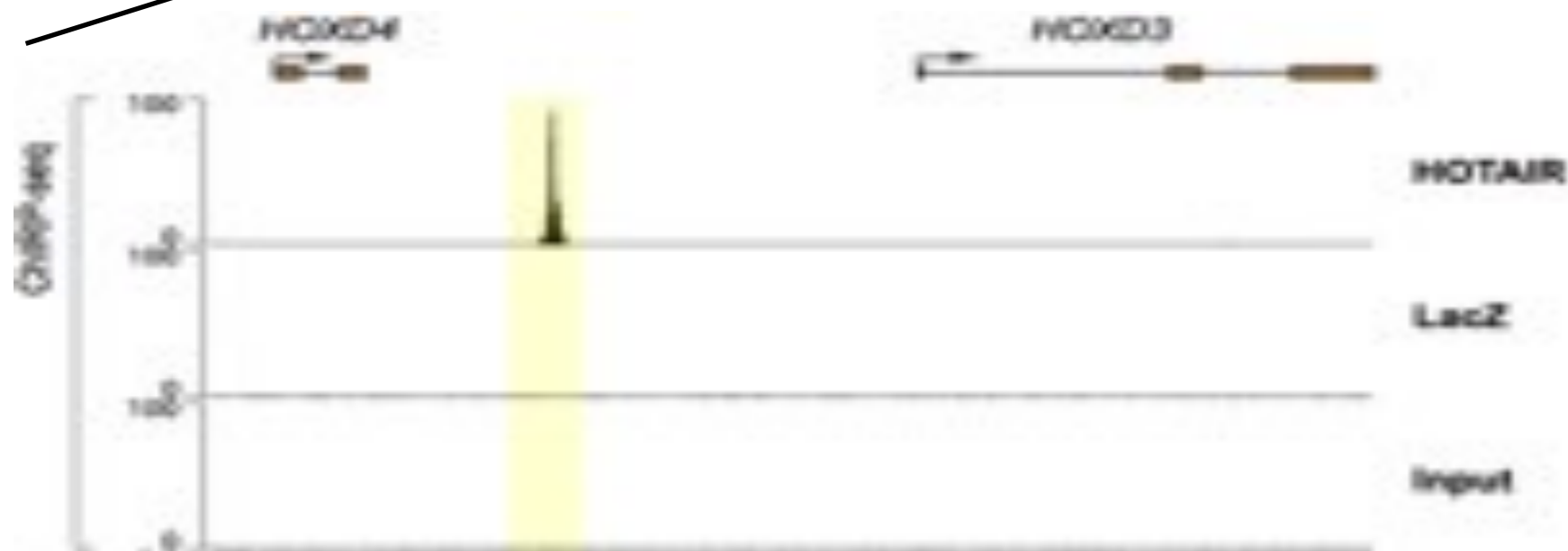


**B**



# ChIRP: example HOTAIR

*HOTAIR ChIRP*



*Ci Chu et al. 2011*

# Types of interaction

		OUTPUT (what we analyse)		
		Protein	RNA	DNA
BAIT (What we enrich)	Protein	<b>CO-IP</b> (co-immunoprecipitation)	<b>RIP/CLIP</b> (RNA-Immunoprecipitation)	<b>ChIP</b> (Chromatin Immunoprecipitation)
	RNA	<b>Exogenous RNA pull Down</b> <b>RAP-Protein</b> (RNA antisense purification)	<b>RAP-RNA</b> (RNA antisense purification)	<b>ChIRP</b> (Chromatin isolation by RNA purification)
	DNA	<b>DNA pull down</b>		<b>Conformation capture 3C</b>



# RAP: RNA pull down techniques

**AIM: Identification of the protein interactors of an RNA Bait: RNA output: Protein/RNA**

**RAP (RNA affinity pull down)** a pulldown technique used to investigate the interaction between RNA and proteins.

## **Precipitation of the RNA and PROTEINS checking**

Total Cytoplasmic or Nuclear extract

Huge amount of cellular extract is needed

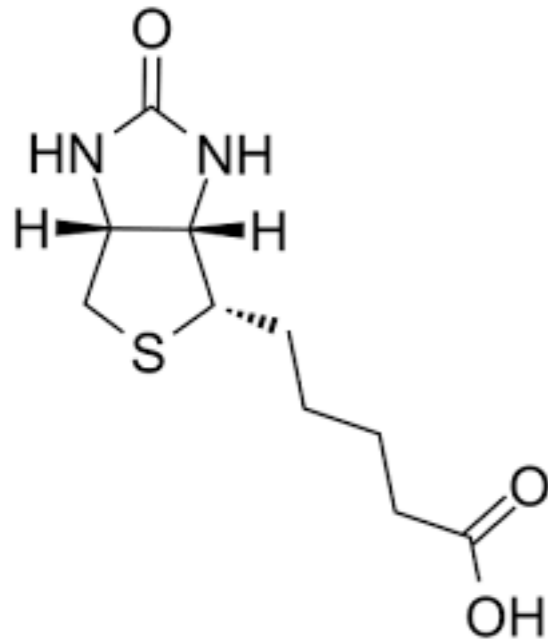
The efficiency depends on the abundance of the RNA

# Exogenous RNA capture

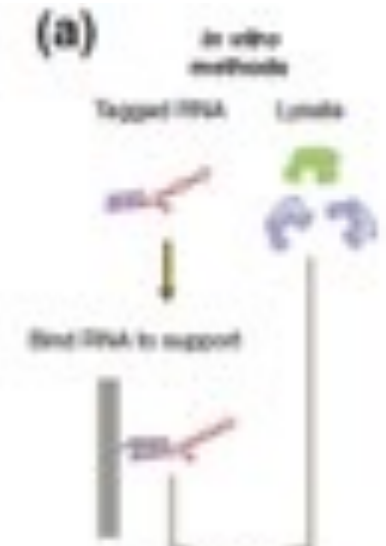
## RNA affinity capture methods

### Different Tags

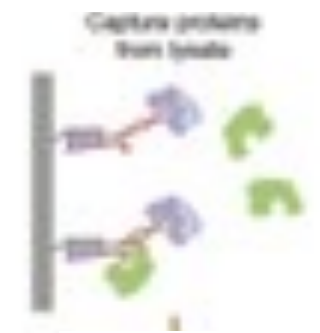
**MS2 viral protein** —> Loop stem loop  
**Cy4** —> RNA aptamer  
**STREPTAVIDIN** —> S1 aptamer



Biotin



Capture protein from lysate



Washing



Boil with SDS



Western Blot



Mass spectrometry



# Exogenous RNA capture

## Work flow

1. RNA tagging

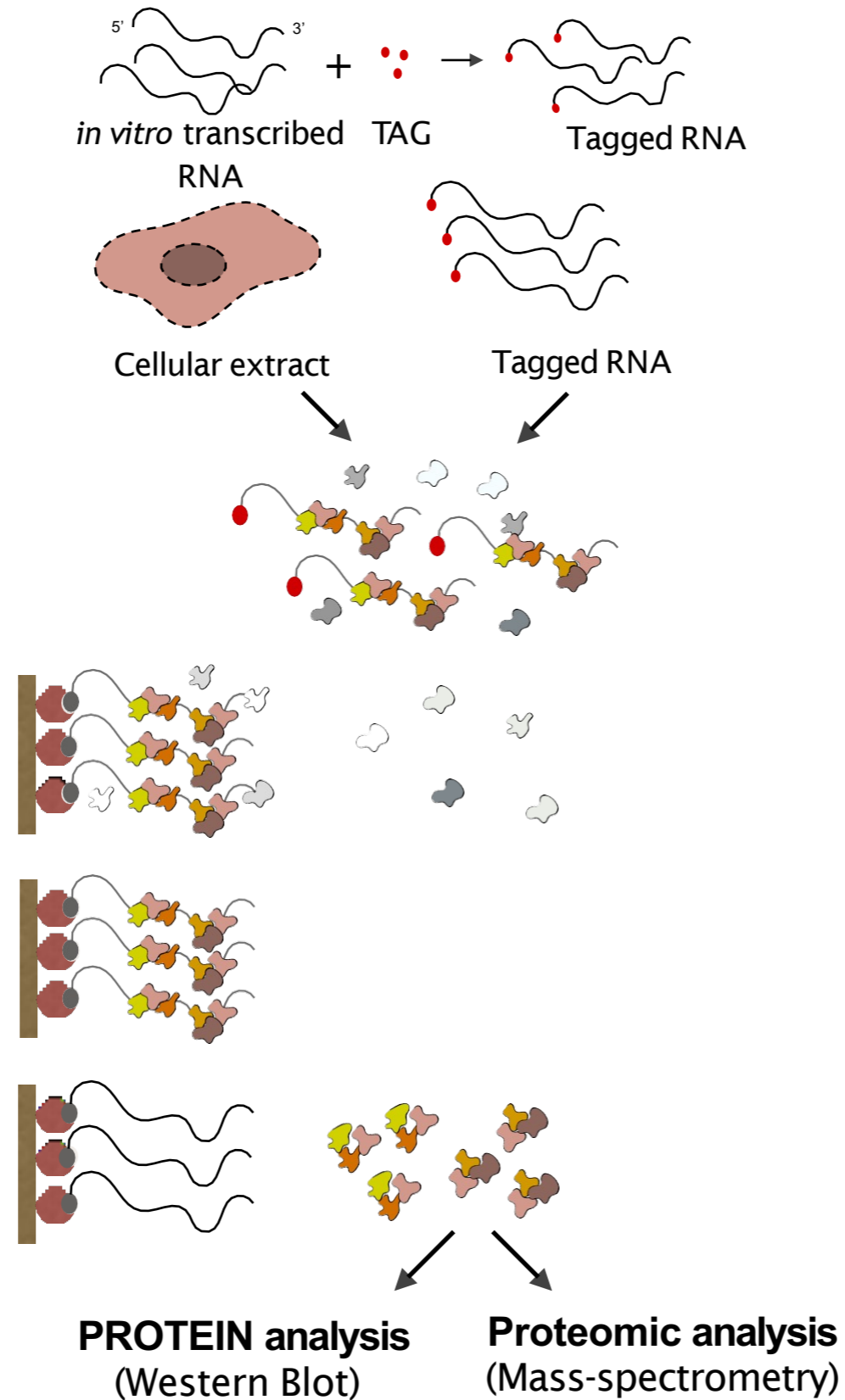
2. Cell lysis

3. Cell extract/tagged RNA incubation

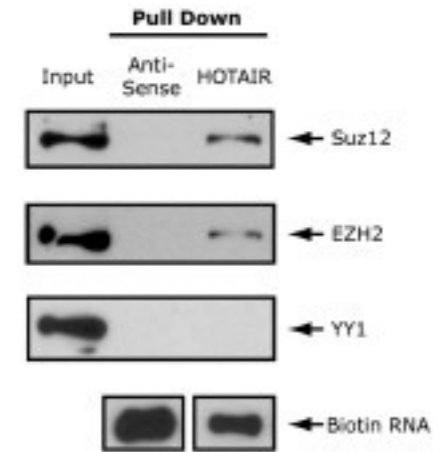
4. RNA/resin binding

5. Washing

6. PROTEIN elution



B



Rinn et al. *Cell* 129, 1311–1323, June 29, 2007

*HOTAIR exogenous pull-down*

*Cipriano and Ballarino, FMB 2018*

# Exogenous RNA capture

The SDS boiling phase will dissociate bound material from the resin, including complexes bound specifically through the tag and those bound non-specifically directly to the resin

## **NEGATIVE CONTROL**

With the exogenous RNA pulldown is difficult to discriminate between real and fake interactions since many interactions can occur just *in vitro*


# Exogenous RNA capture. It might not be specific

Article



THE  
EMBO  
JOURNAL

## PRC2 is dispensable for *HOTAIR*-mediated transcriptional repression

Manuela Portoso<sup>1,2</sup>, Roberta Ragazzini<sup>1,2</sup>, Živa Brenčič<sup>1,2</sup>, Arianna Moiani<sup>1,2</sup>, Audrey Michaud<sup>1,2</sup>, Ivaylo Vassilev<sup>1,2</sup>, Michel Wassef<sup>1,2</sup>, Nicolas Servant<sup>1,3</sup>, Bruno Sargueil<sup>4</sup> & Raphaël Margueron<sup>1,2,\*</sup> 

### Abstract

Long non-coding RNAs (lncRNAs) play diverse roles in physiological and pathological processes. Several lncRNAs have been suggested to modulate gene expression by guiding chromatin-modifying complexes to specific sites in the genome. However, besides the example of Xist, clear-cut evidence demonstrating this novel mode of regulation remains sparse. Here, we focus on *HOTAIR*, a lncRNA that is overexpressed in several tumor types and previously proposed to play a key role in gene silencing through direct recruitment of Polycomb Repressive Complex 2 (PRC2) to defined genomic loci. Using genetic tools and a novel RNA-tethering system, we investigated the interplay between *HOTAIR* and PRC2 in gene silencing. Surprisingly, we observed that forced overexpression of *HOTAIR* in breast cancer cells leads to subtle transcriptomic changes that appear to be independent of PRC2. Mechanistically, we found that artificial tethering of *HOTAIR* to chromatin causes transcriptional repression, but that this effect does not require PRC2. Instead, PRC2 recruitment appears to be a consequence of gene silencing. We propose that PRC2 binding to RNA might serve functions other than chromatin targeting.

regulation of chromatin structure, either through histone modifications or through chromatin compaction (Simon & Kingston, 2009). In *Drosophila*, four PcG complexes have been identified, while in mammals, only two complexes are well characterized so far: Polycomb Repressive Complex 2 (PRC2) and Polycomb Repressive Complex 1 (PRC1). The PRC2 is responsible for histone H3 lysine 27 (H3K27) di- and tri-methylation (Margueron & Reinberg, 2011).

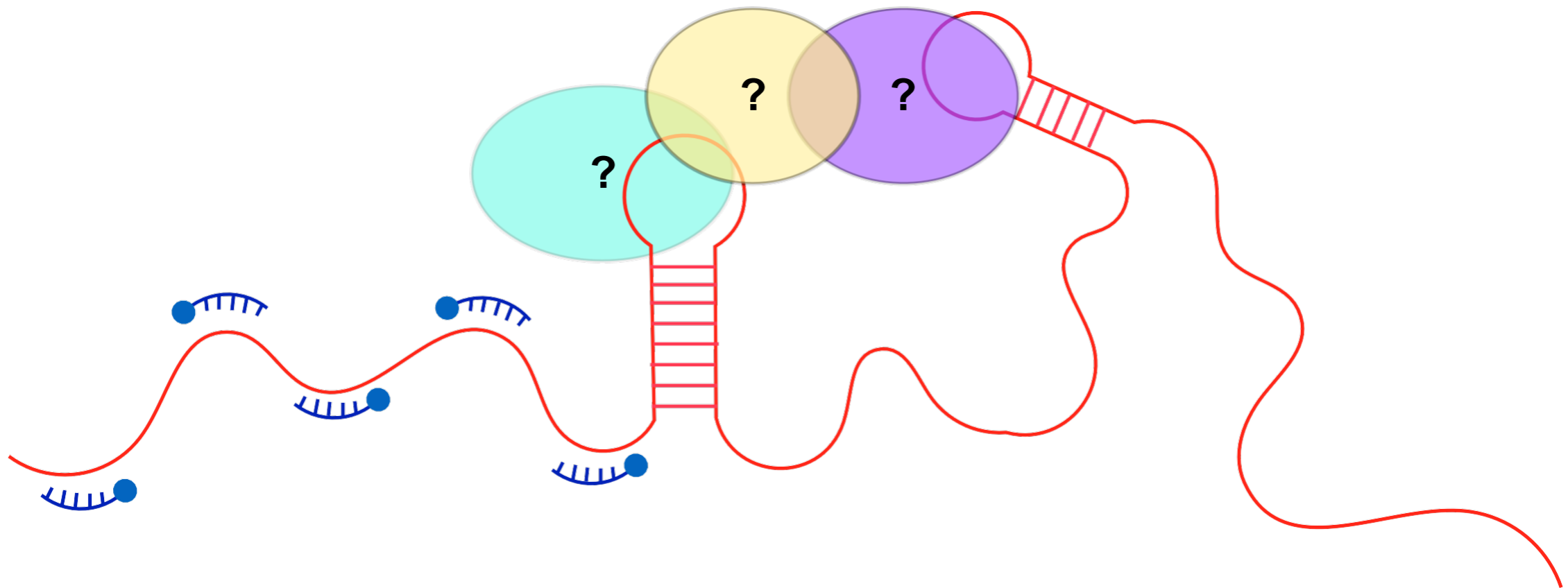
Although our understanding of how PRC2 contacts chromatin has improved, how it is specifically recruited to defined genomic loci is still only partially understood. The core PRC2 has no known sequence-specific DNA-binding domain. In *Drosophila*, DNA sequences known as Polycomb responsive elements (PREs) mediate PcG recruitment through a combination of specific transcription factors. Although similar mechanisms have been proposed in mammals (Arnold *et al.*, 2013; Sing *et al.*, 2009; Woo *et al.*, 2010), they do not appear to be the general rule. Indeed, the specific transcription factors found to bind these putative mammalian PREs do not act consistently as PRC2 genomewide recruiters. Importantly, GC-rich regions are frequently bound by PRC2 components (Ku *et al.*, 2008) and they are, in some instances, sufficient to mediate PRC2 recruitment (Mendenhall *et al.*, 2010; Jermann *et al.*, 2014), although once again this cannot account for the specificity and dynamics of

EPIC FAIL

# Endogenous RNA capture. RAP

**AIM: Identification of the protein interactors of an RNA Bait: RNA output: Protein/RNA**

**RAP (RNA affinity pull down)** a pulldown technique used to investigate the interaction between RNA and proteins.



# Endogenous RNA capture. RAP

AIM: Identification of the protein interactors of an RNA Bait: RNA output: Protein/RNA

RAP (RNA affinity pull down) a pulldown technique used to investigate the interaction between RNA and proteins.

2 Cell lysis

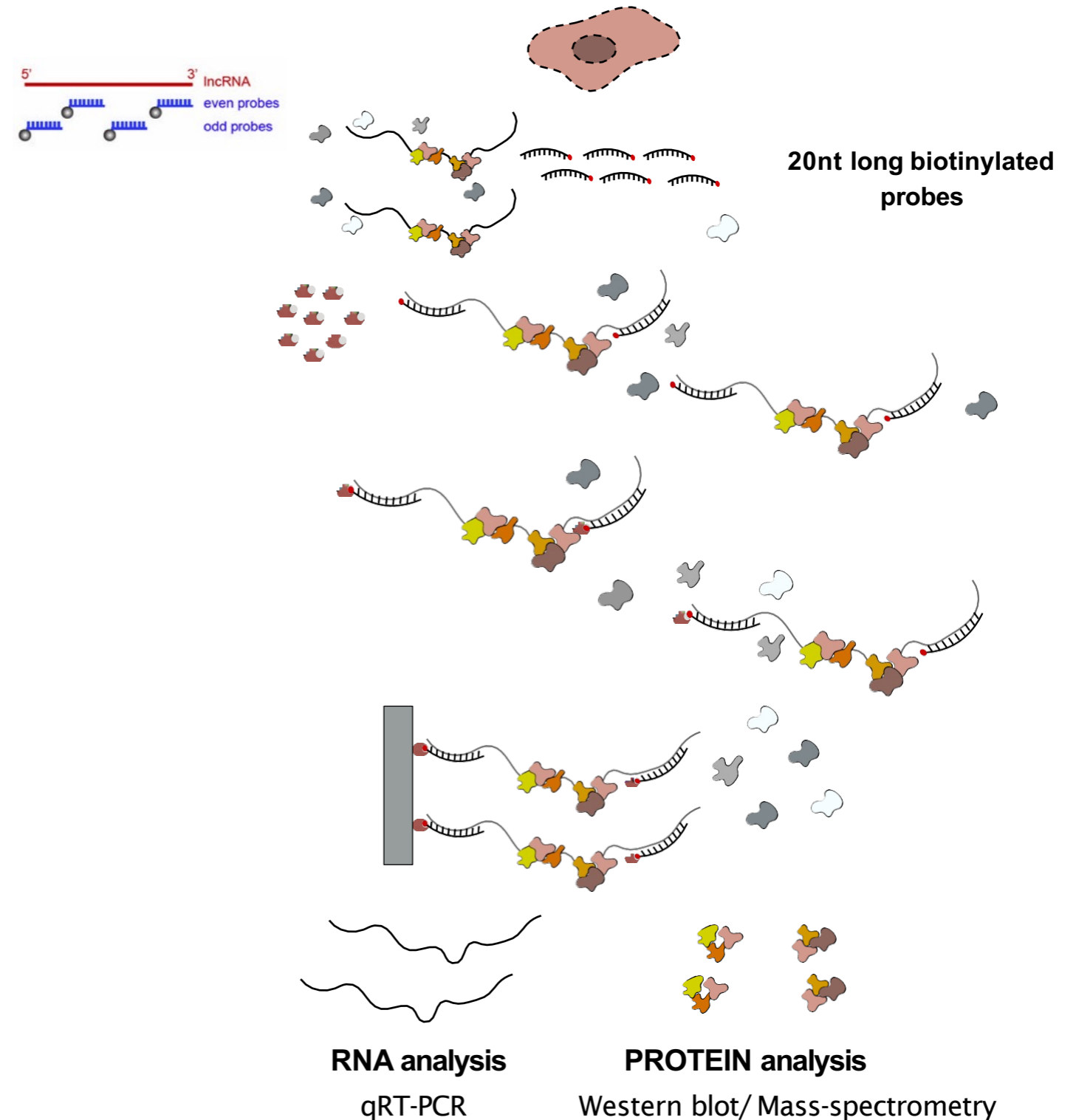
3. Cell extract/probes incubation

3. Binding step

4. Introduction of Streptavidin-magnetic beads and Capture RNA/probes complexes from lysate

5. Purification of RNA/probes complexes and washes with low salt buffers (150mM NaCl)

6. Protein and RNA elution and analysis

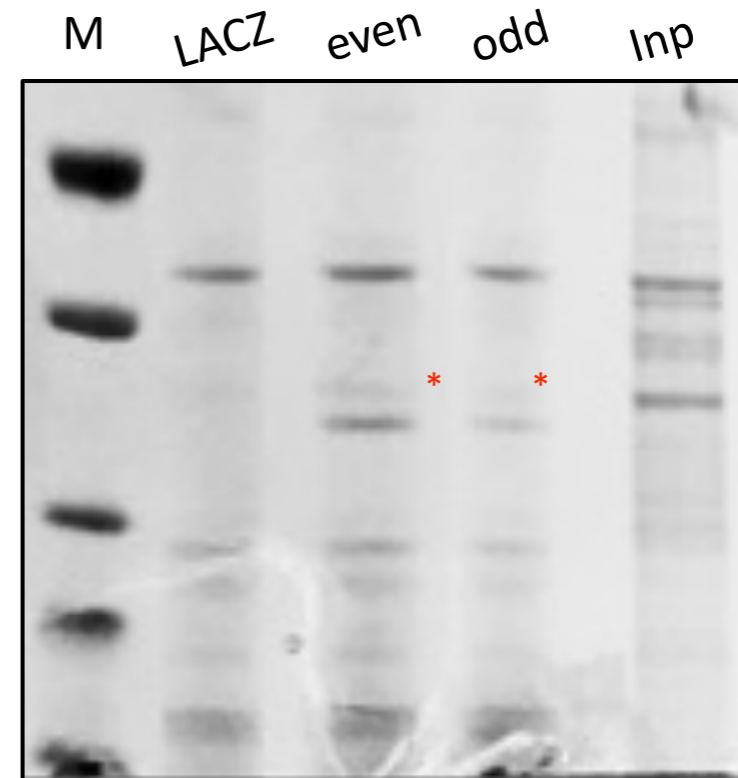
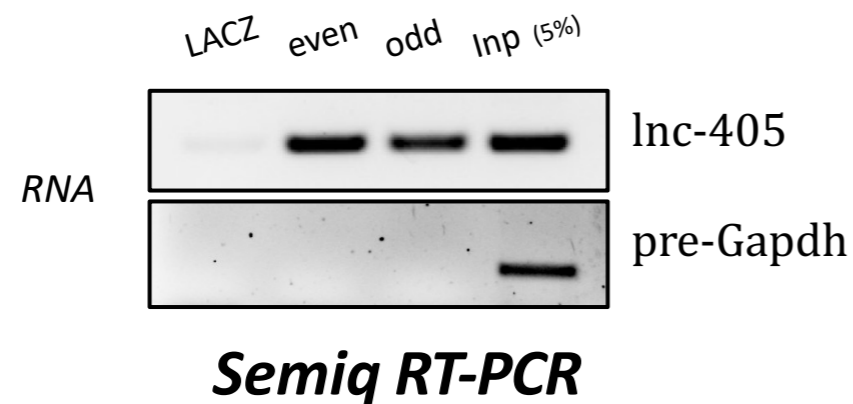


# Endogenous RNA capture. RAP

**AIM: Identification of the protein interactors of an RNA** Bait: RNA output: Protein/RNA

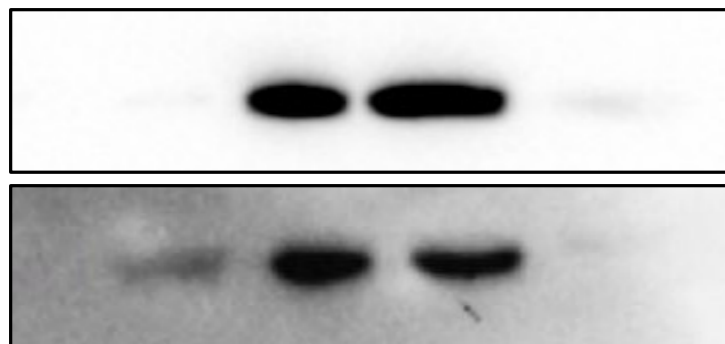
**RAP (RNA affinity pull down)** a pulldown technique used to investigate the interaction between RNA and proteins.

## *Lnc-405 endogenous pulldown*



**WB**

Inp (1%) ODD EVEN LACZ



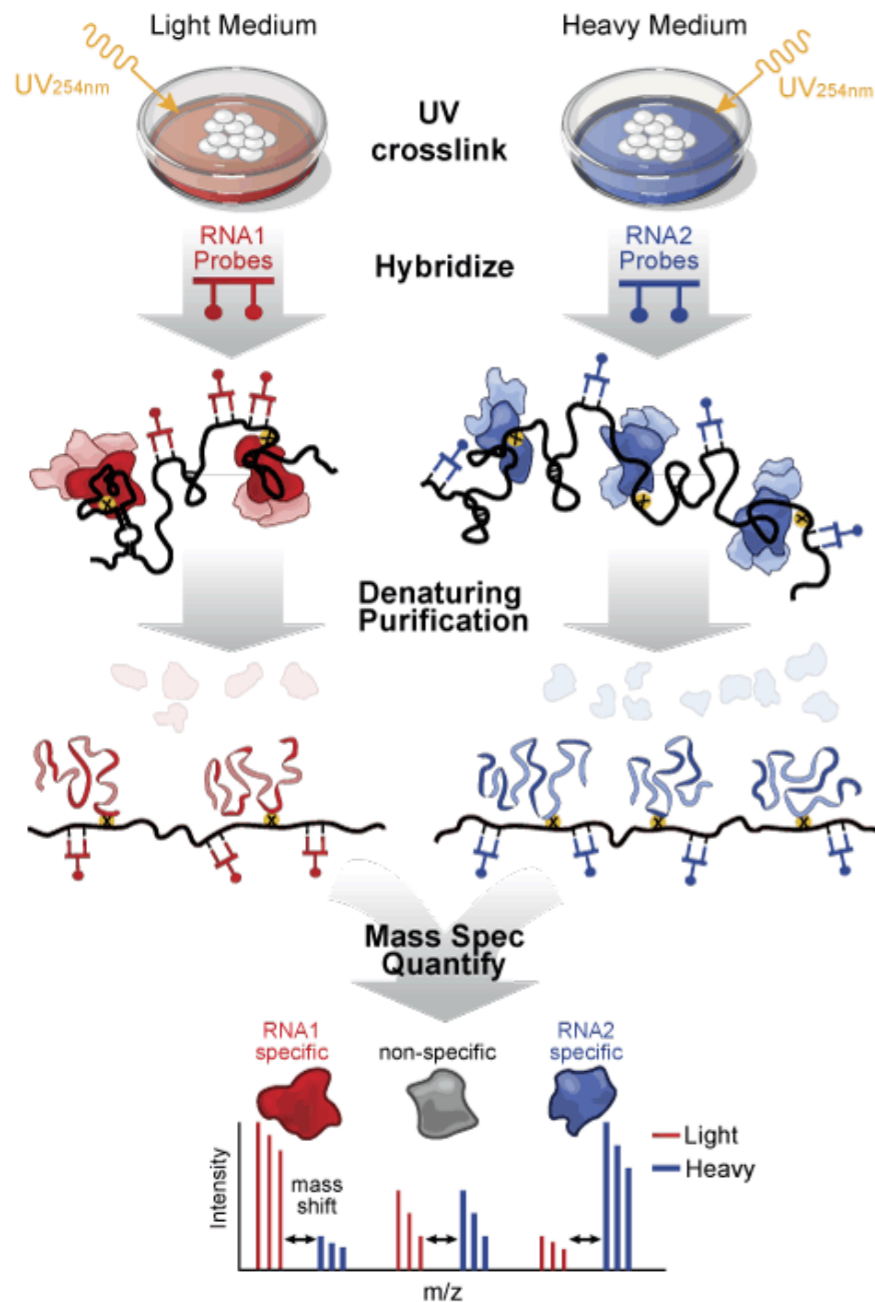
Accession	Description
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Q8VDD5	Myosin-9 OS=Mus musculus GN=Myh9 PE=1 SV=4 - [MYH9_MOUSE]
P62843	40S ribosomal protein S15 OS=Mus musculus GN=Rps15 PE=1 SV=2 - [RS15_MOUSE]
P62669	Transcriptional activator protein Pur alpha OS=Mus musculus GN=Pura PE=1 SV=1 - [PURA_MOUSE]
P62301	40S ribosomal protein S13 OS=Mus musculus GN=Rps13 PE=1 SV=2 - [RS13_MOUSE]
E9Q557	Desmoplakin OS=Mus musculus GN=Dsp PE=1 SV=1 - [DESP_MOUSE]
Q6P5H2	Nestin OS=Mus musculus GN=Nes PE=1 SV=1 - [NEST_MOUSE]
P62702	40S ribosomal protein S4, X isoform OS=Mus musculus GN=Rps4x PE=1 SV=2 - [RS4X_MOUSE]
P20152	Vimentin OS=Mus musculus GN=Vim PE=1 SV=3 - [VIME_MOUSE]
P31001	Desmin OS=Mus musculus GN=Des PE=1 SV=3 - [DESM_MOUSE]
Q93299	Transcriptional activator protein Pur beta OS=Mus musculus GN=Purb PE=1 SV=1 - [PURB_MOUSE]



# Endogenous RNA capture. RAP

AIM: Identification of the protein interactors of an RNA Bait: RNA output: Protein/RNA

RAP (RNA affinity pull down) a pulldown technique used to investigate the interaction between RNA and proteins.



## *In vivo* UV crosslinking

Longer probes (90nt)

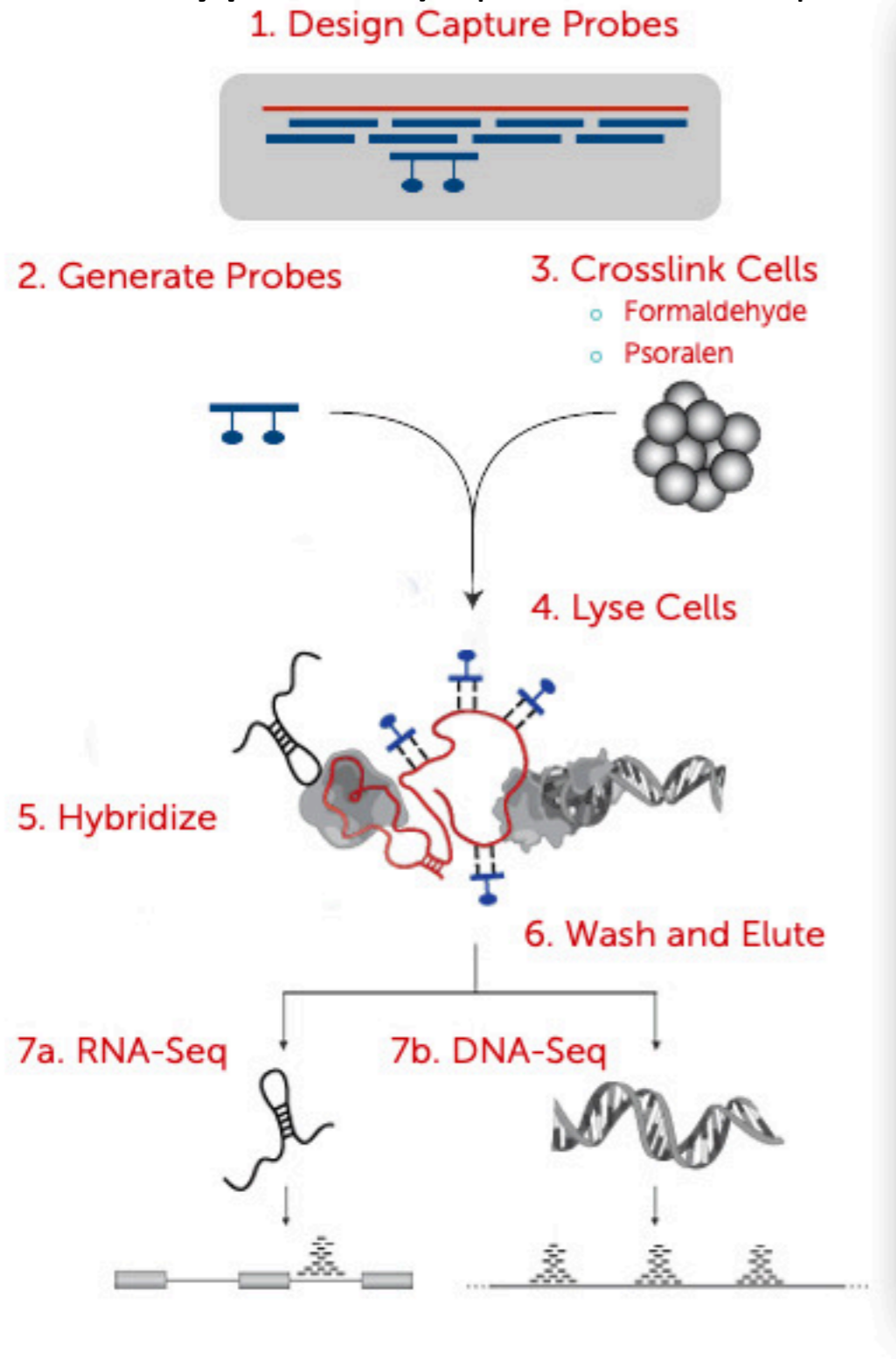
High stringency binding conditions

High stringency wash conditions

# Endogenous RNA capture. RAP

AIM: Identification of the protein interactors of an RNA Bait: RNA output: Protein/RNA

RAP (RNA affinity pull down) a pulldown technique used to investigate the interaction between RNA and proteins.



## *In vivo* UV crosslinking

Longer probes (90nt)

High stringency binding conditions

High stringency wash conditions

# Endogenous RNA capture. RAP

## Work flow

UV cross-linking

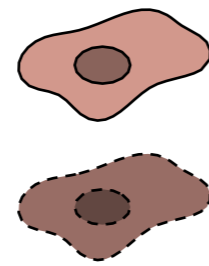
Cell lysis

RNA/probes binding to streptavidin magnetic beads

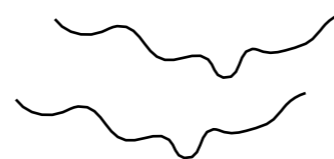
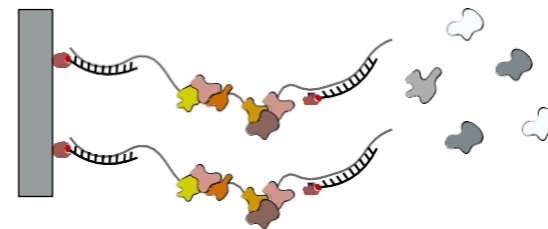
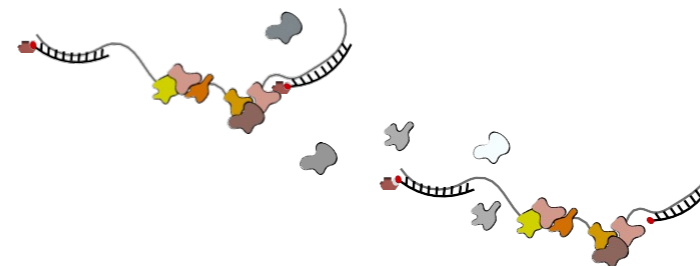
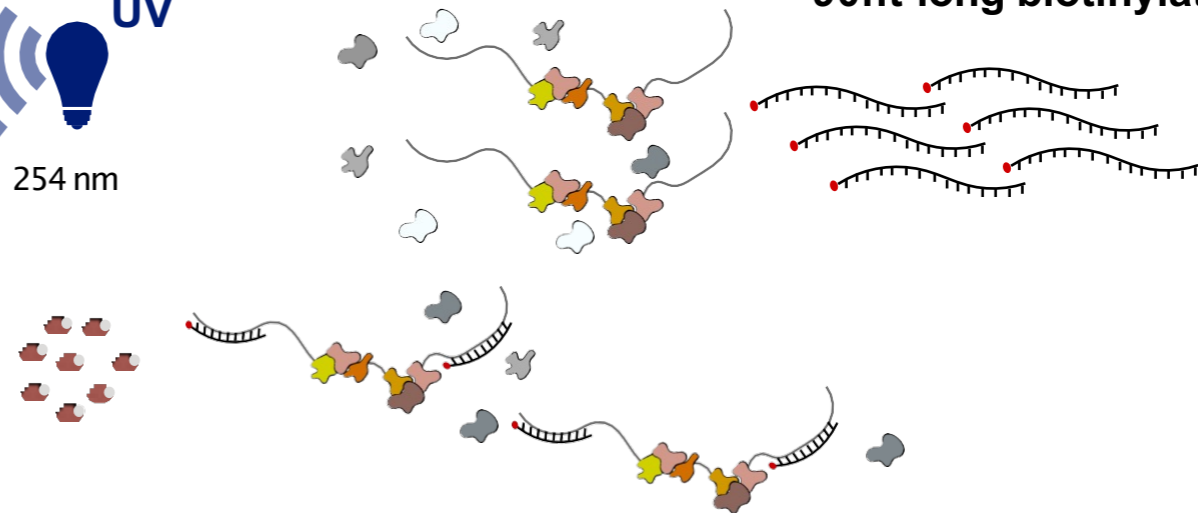
Capture RNA/probes complexes from lysate

Purification of RNA/probes complexes and washes in high salt buffers (1M LiCl)

RNA and PROTEIN elution



90nt long biotinylated probes



RNA analysis

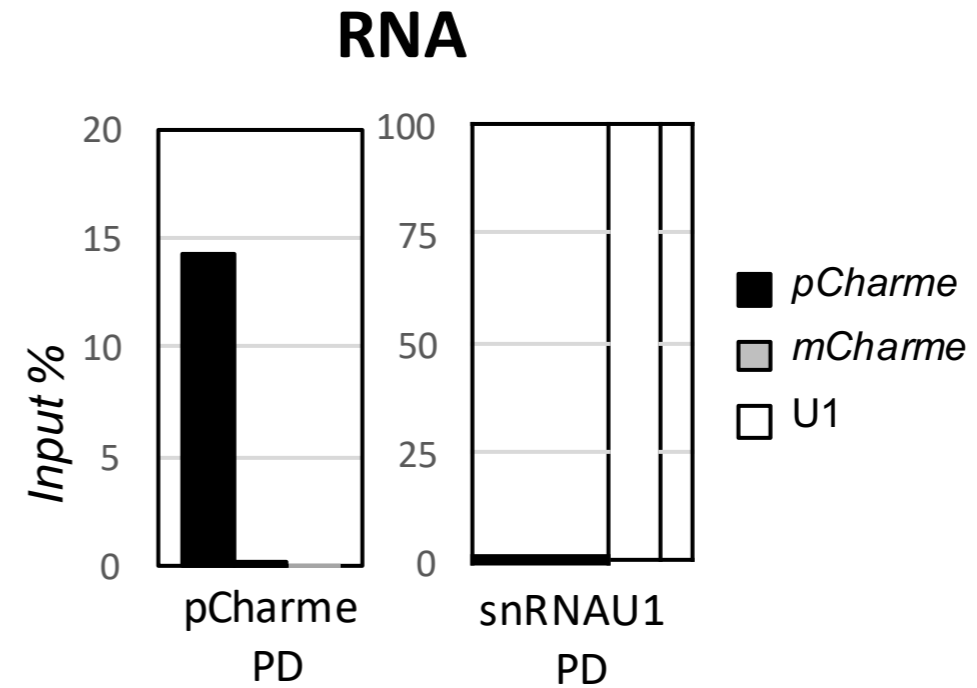
qRT-PCR



PROTEIN analysis

Western blot/ Mass-spectrometry

# Endogenous RNA capture. RAP



Accession	Description	pCharme	U1
Q8BGJ5	MCG13402, isoform CRA_a OS=Mus musculus GN=Ptbp1 PE=1 SV=1 - [Q8BGJ5_MOUSE]	273.10	50.91
Q8K310	Matrin-3 OS=Mus musculus GN=Matr3 PE=1 SV=1 - [MATR3_MOUSE]	165.04	35.42
Q61990	Poly(rC)-binding protein 2 OS=Mus musculus GN=Pcbp2 PE=1 SV=1 - [PCBP2_MOUSE]	57.12	19.65
A0A0G2JGW0	Polypyrimidine tract-binding protein 2 OS=Mus musculus GN=Ptbp2 PE=1 SV=1 - [A0A0G2JGW0_MOUSE]	22.18	0.00
B1B0C7	Basement membrane-specific heparan sulfate proteoglycan core protein OS=Mus musculus GN=Hspg2 PE=1 SV=1 -	16.66	0.00
A0A0R4J044	Poly(rC)-binding protein 4 OS=Mus musculus GN=Pcbp4 PE=1 SV=1 - [A0A0R4J044_MOUSE]	16.04	0.00
Q4FK66	Pre-mRNA-splicing factor 38A OS=Mus musculus GN=Prpf38a PE=1 SV=1 - [PR38A_MOUSE]	14.10	0.00
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Q62376	U1 small nuclear ribonucleoprotein 70 kDa OS=Mus musculus GN=Snrnp70 PE=1 SV=2 - [RU17_MOUSE]	0.00	432.94
Q62189	U1 small nuclear ribonucleoprotein A OS=Mus musculus GN=Snrpa PE=1 SV=3 - [SNRPA_MOUSE]	0.00	304.30
Q8K4Z5	Splicing factor 3A subunit 1 OS=Mus musculus GN=Sf3a1 PE=1 SV=1 - [SF3A1_MOUSE]	0.00	162.87
P62309	Small nuclear ribonucleoprotein G OS=Mus musculus GN=Snrpg PE=1 SV=1 - [RUXG_MOUSE]	0.00	121.09
P62317	Small nuclear ribonucleoprotein Sm D2 OS=Mus musculus GN=Snrpd2 PE=1 SV=1 - [SMD2_MOUSE]	0.00	68.35
Q6P4T2	U5 small nuclear ribonucleoprotein 200 kDa helicase OS=Mus musculus GN=Snrnp200 PE=1 SV=1 - [U520_MOUSE]	10.24	50.45
P62320	Small nuclear ribonucleoprotein Sm D3 OS=Mus musculus GN=Snrpd3 PE=1 SV=1 - [SMD3_MOUSE]	0.00	50.24
Q62241	U1 small nuclear ribonucleoprotein C OS=Mus musculus GN=Snrpc PE=1 SV=1 - [RU1C_MOUSE]	0.00	34.61

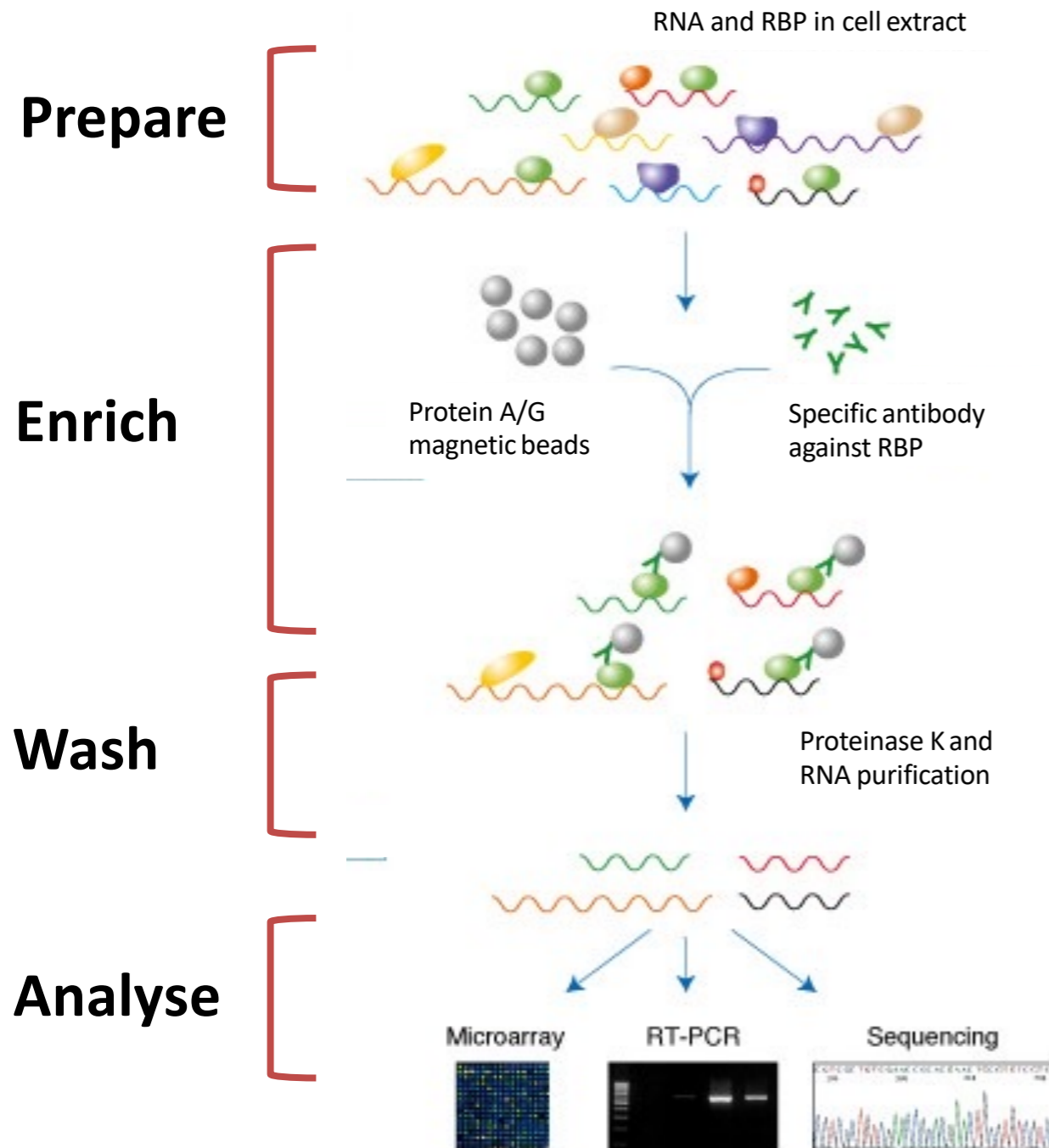
# Types of interaction

		OUTPUT (what we analyse)		
		Protein	RNA	DNA
BAIT (What we enrich)	Protein	<b>CO-IP</b> (co-immunoprecipitation)	<b>RIP/CLIP</b> (RNA-Immunoprecipitation)	<b>ChIP</b> (Chromatin Immunoprecipitation)
	RNA	<b>Exogenous RNA pull Down</b> <b>RAP-Protein</b> (RNA antisense purification)	<b>RAP-RNA</b> (RNA antisense purification)	<b>ChIRP</b> (Chromatin isolation by RNA purification)
	DNA	<b>DNA pull down</b>		<b>Conformation capture 3C</b>

# RIP/CLIP

**AIM: Identification of RNA interaction with known proteins. Bait: Proteins output: RNA**

**RIP (RNA immunoprecipitation) or CLIP (Crosslinked RNA immunoprecipitation)** an immunoprecipitation technique used to investigate the interaction between RNA and proteins. Not only serve to determine interaction but also place of the interaction.



Interaction between RNA and Proteins focusing on the proteins (protein focused)

A lot of protocols, same essence.

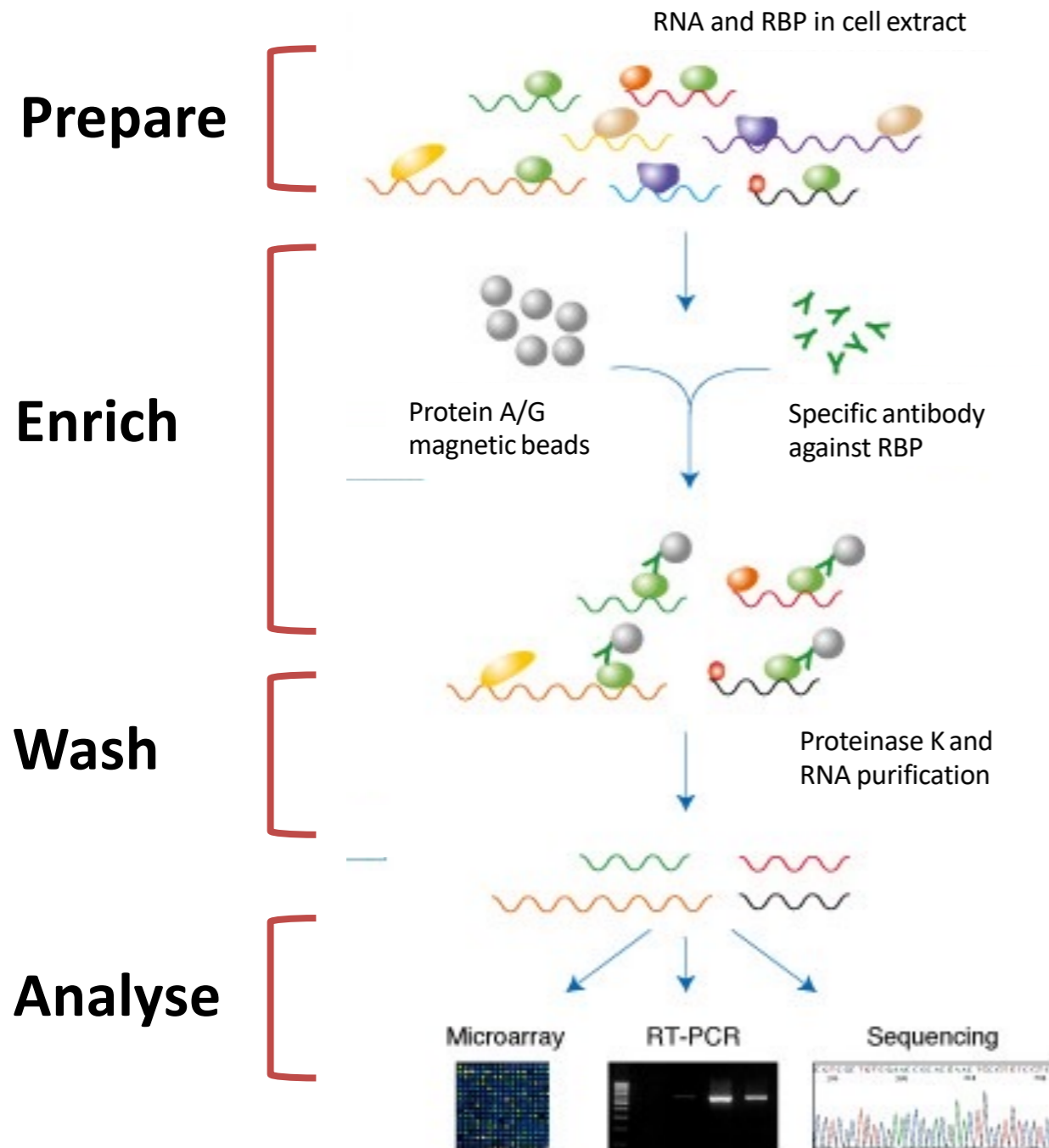
- Cytoplasmic or Nuclear extract
- Isolation of Ribonucleoprotein complexes
- The resulting data have a low resolution, also **not directly associated** RNAs could be immunoprecipitated, and the **binding site** in the co-purified RNA molecule remained unresolved.
- Variants:

**CLIP (UV-RIP)**  
**PAR-CLIP**  
**i-CLIP (CLIP-seq)**

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## Work flow

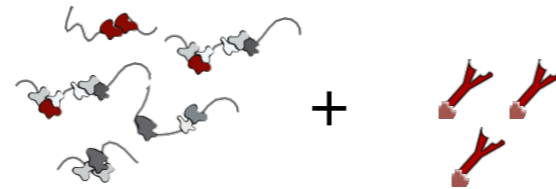
1. Lysis Cells and collect cell extract

2 Prebinding between AntiBody and Beads

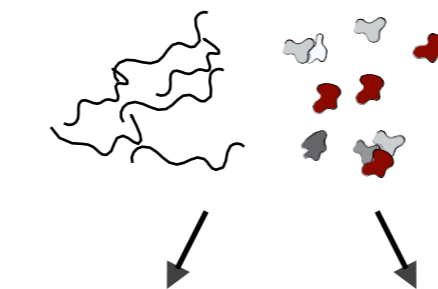
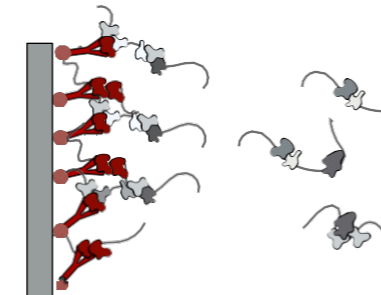
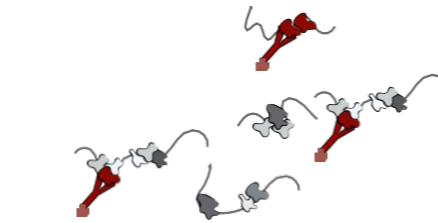
3. Introduction of cellular extract (Binding step)

4. Wash and Purification of RNA-protein complexes

5. Protein and RNA elution



No crosslink  
Capture specific and non specific interactions

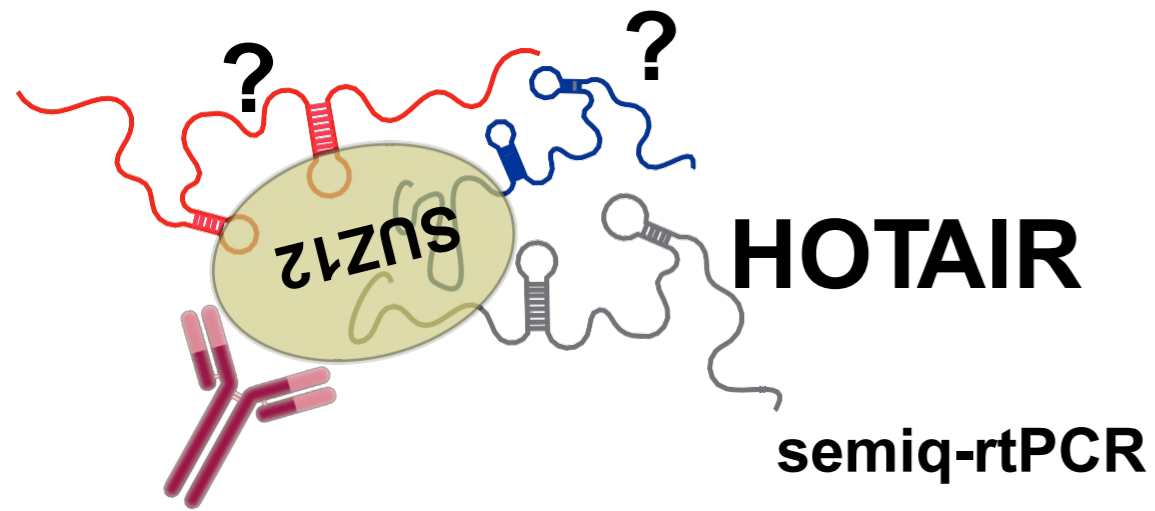


RNA analysis  
(qRT-PCR or RNA-seq)

PROTEIN analysis  
(Western Blot)



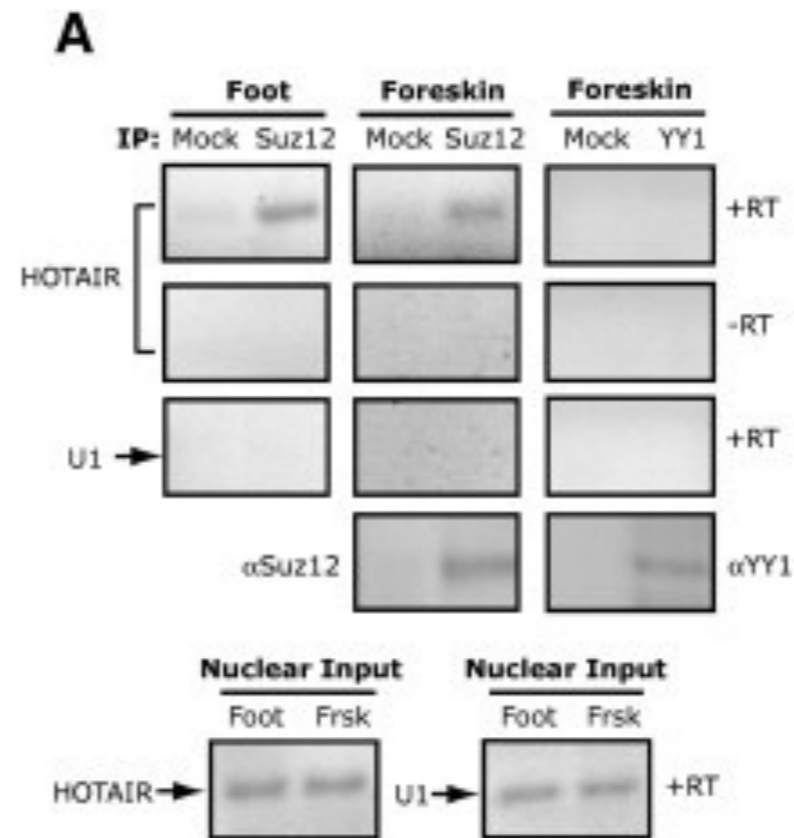
# RIP/CLIP



**Western Blot**

**semiq-rtPCR**

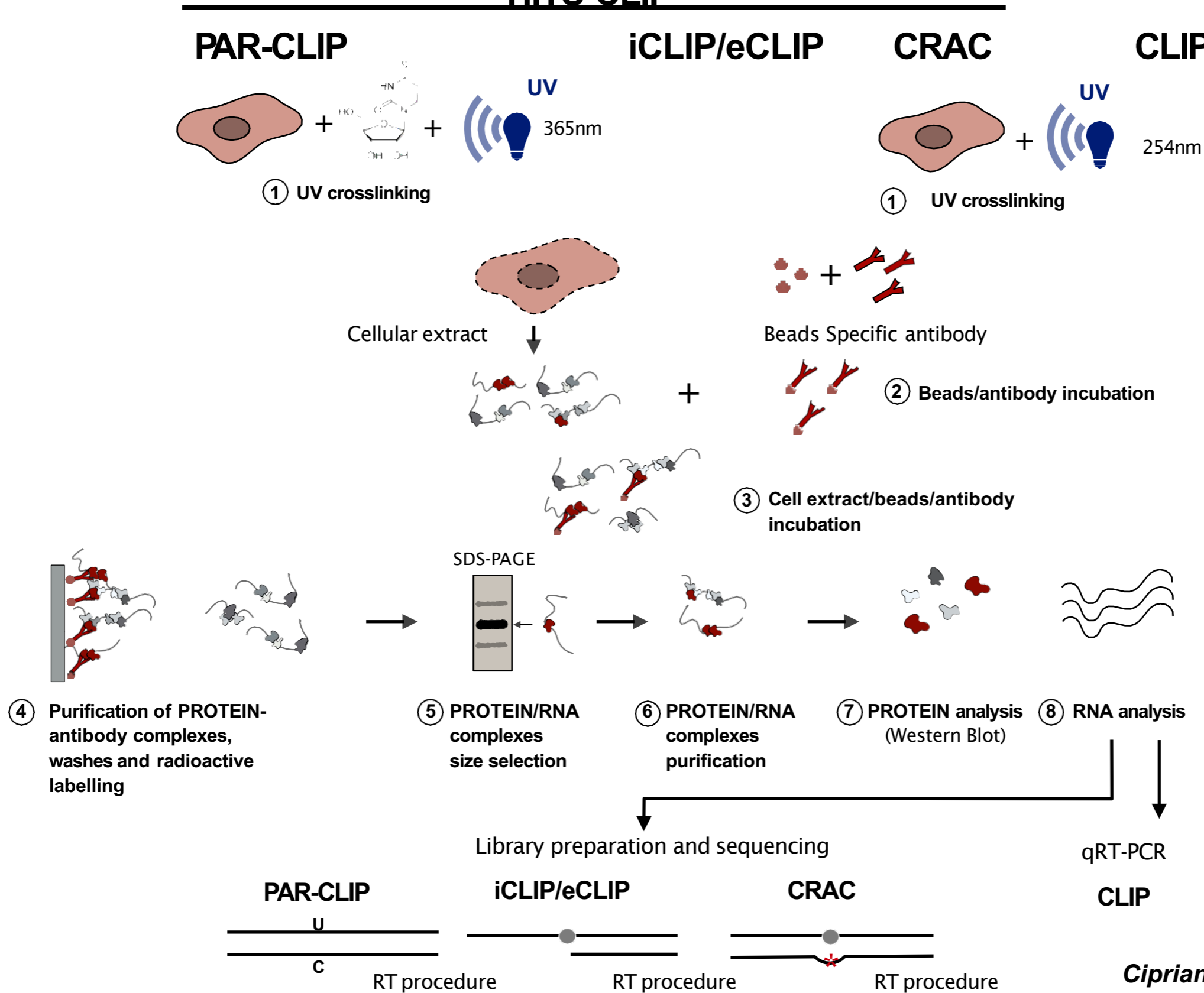
## SUZ12 RIP



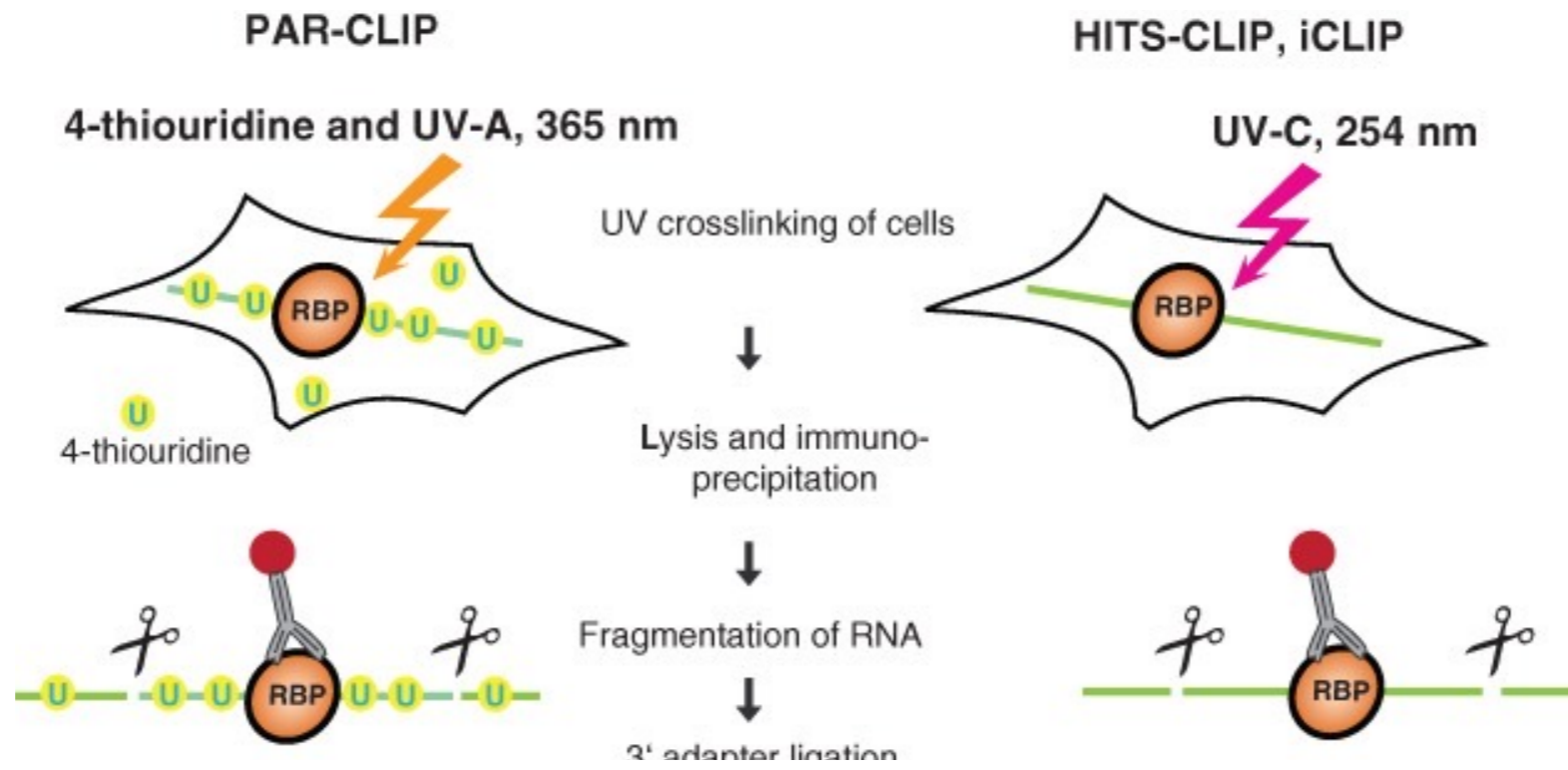
# CLIP

(in vivo and cross-linked)

## HITS-CLIP



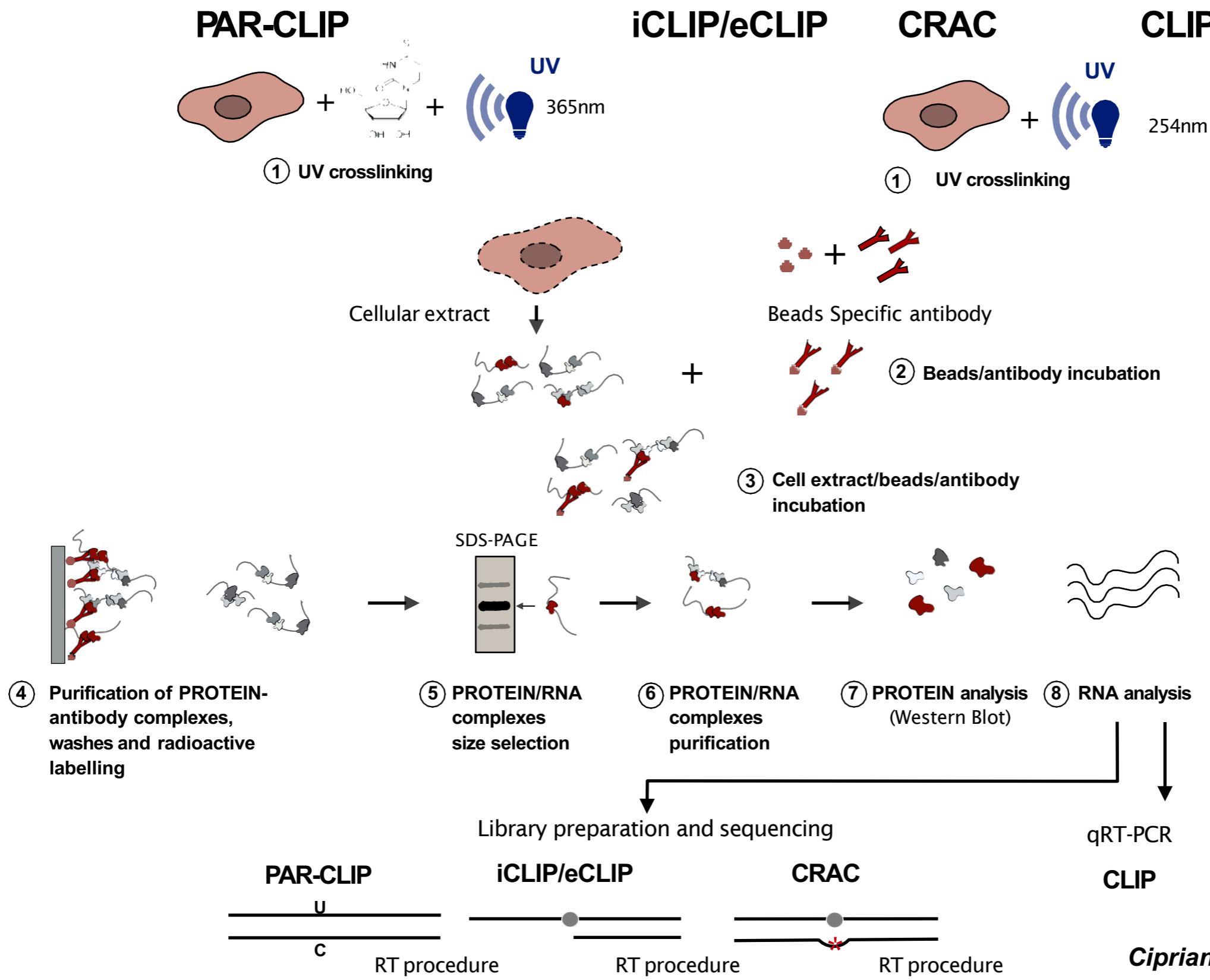
# CLIP



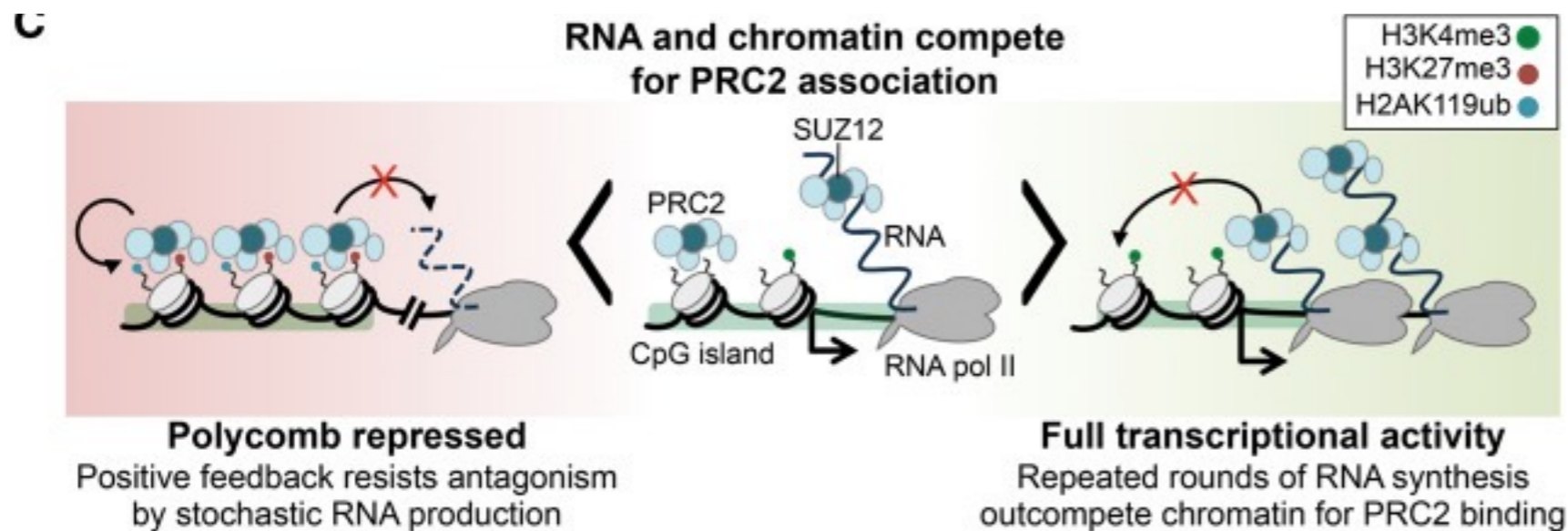
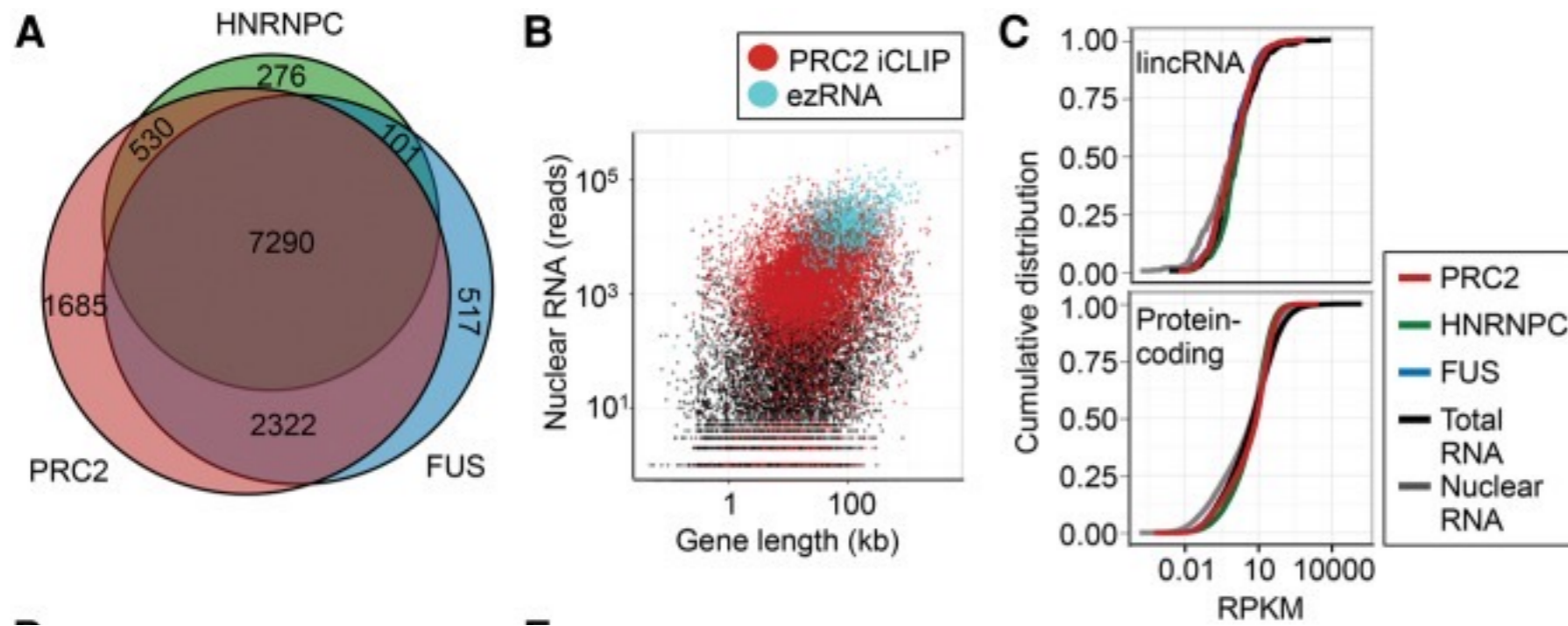
# CLIP

(in vivo and cross-linked)

## HITS-CLIP



# CLIP example

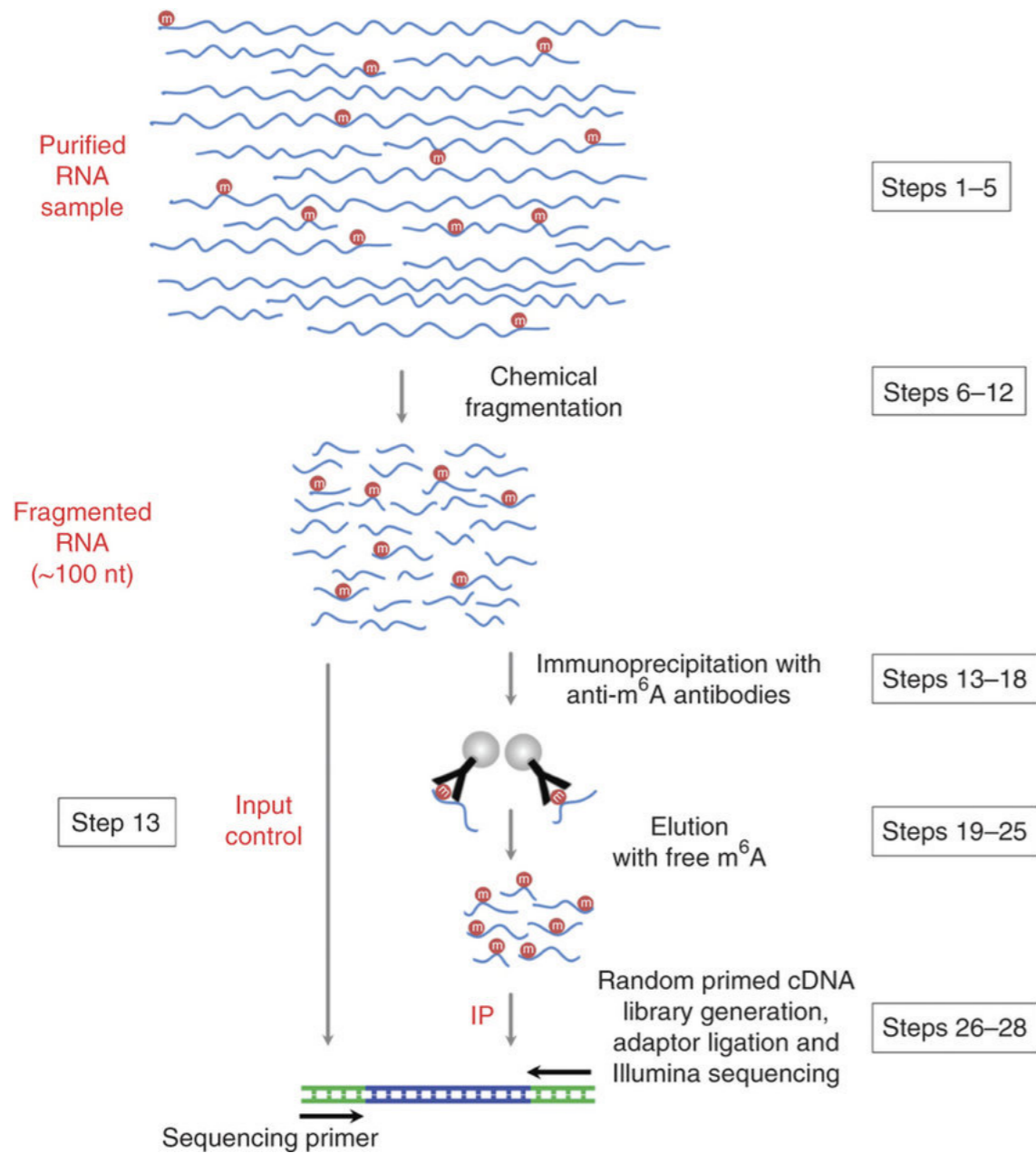


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**ALL of them are useful. But please make proper controls!**

# Bonus track: Antibodies against nucleic acids



(F) G4 ChIP-seq



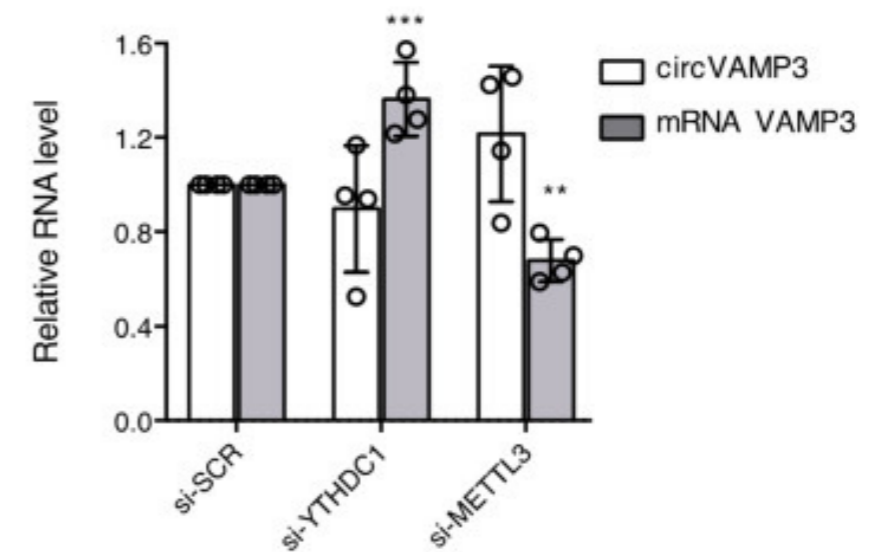
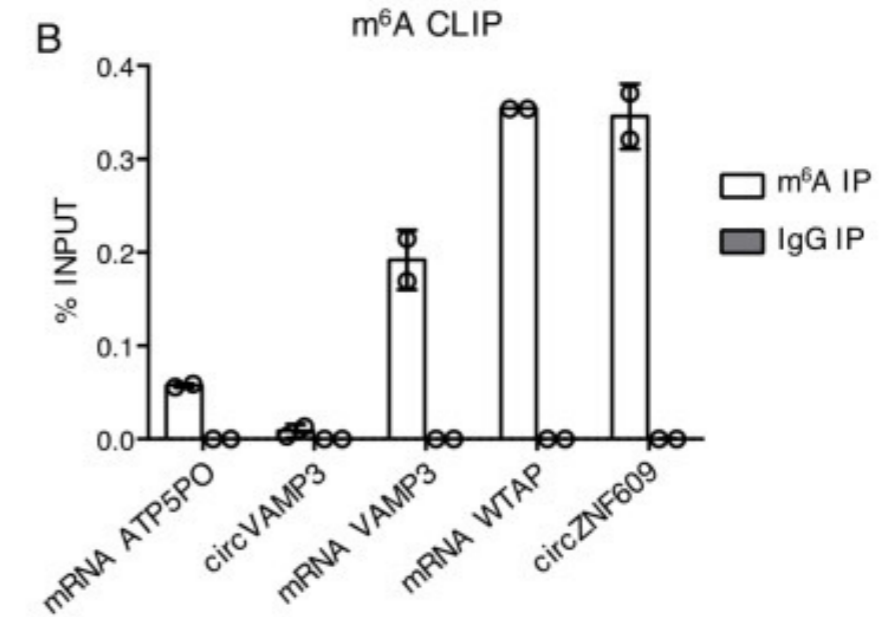
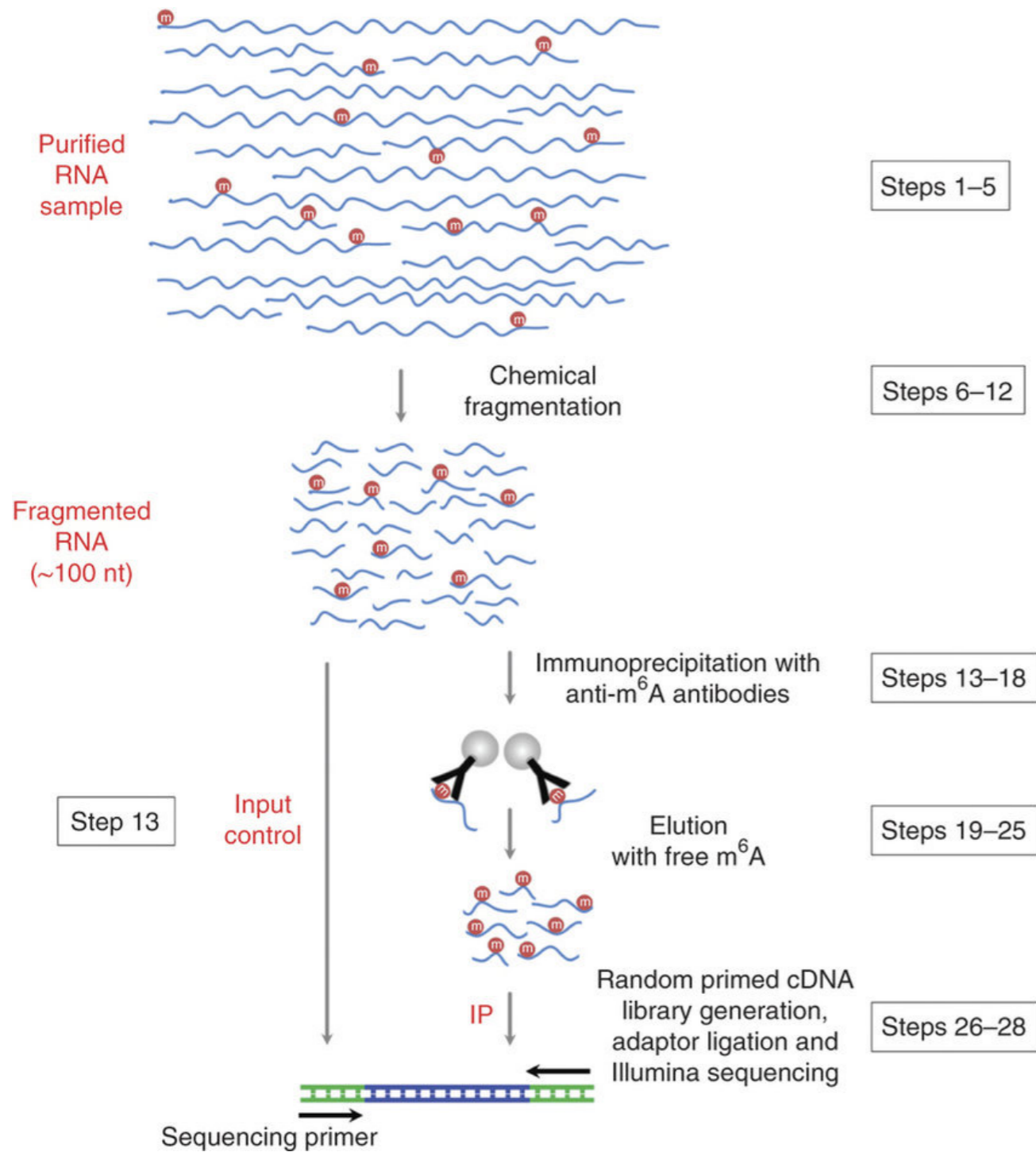
Trends in Chemistry

**We also can detect modification on the RNA/DNA structures using antibodies:**

**M6A RIP**

**G4- quadruplex CHIP**

# Bonus track: Antibodies against nucleic acids





# References

## SUMMARY

- Cipriano A and Ballarino M. “**The ever-evolving concept of the gene: the use of RNA/Protein experimental techniques to understand genome functions**”. *Front. Mol. Biosci* 2018 Mar 6;5:20 doi:10.3389/fmolb.2018.00020.

## COIP And ChiP

Healy et al., 2019, *Molecular Cell* 76, 437–452 November 7, 2019<sup>a</sup>2019 Elsevier Inc. <https://doi.org/10.1016/j.molcel.2019.08.012>

## CHiRP and RNA pulldown

- Ci Chu, Kun Qu, Franklin L. Zhong, Steven E. Artandi and Howard Y. Chang; **Genomic maps of lincRNA occupancy reveal principles of RNA-chromatin interactions**. *Mol Cell* 44, 667–678, November 18, 2011.
- John L. Rinn, Michael Kertesz, Jordon K. Wang, Sharon L. Squazzo, Xiao Xu, Samantha A. Brugmann, L. Henry Goodnough, Jill A. Helms, Peggy J. Farnham, Eran Segal, and Howard Y. Chang; **Functional Demarcation of Active and Silent Chromatin Domains in Human HOX Loci by Non-Coding RNAs**. *Cell* 129, 1311–1323, June 29, 2007.
- Colleen A. McHugh<sup>1</sup>, Chun-Kan Chen, Amy Chow, Christine F. Surka, Christina Tran, Patrick McDonel, Amy Pandya-Jones, Mario Blanco, Christina Burghard, Annie Moradian, Michael J. Sweredoski, Alexander A. Shishkin, Julia Su, Eric S. Lander, Sonja Hess, Kathrin Plath & Mitchell Guttman; **The Xist lincRNA interacts directly with SHARP to silence transcription through HDAC3**. *Nature* 521, 232–236 May 2015

## HITS-CLIP AND PAR-CLIP

- Zhen Wang, James Tollervey, Michael Briese, Daniel Turner, Jernej Ule; **CLIP: Construction of cDNA libraries for high-throughput sequencing from RNAs cross-linked to proteins *in vivo*** *Methods* 48 (2009) 287–293
- Julian König, Kathi Zarnack, Nicholas M. Luscombe and Jernej Ule; **Protein–RNA interactions: new genomic technologies and perspectives**. *Nature Reviews Genetics* 13, 77-83 February 2012.
- Colleen A McHugh, Pamela Russell and Mitchell Guttman; **Methods for comprehensive experimental identification of RNA-protein interactions**. *Genome Biology*, 15:203 2014

# Exercise: Interactome in the web

## Genome browser

<https://genome.ucsc.edu/>

<https://genome.ucsc.edu/s/mbeltran/G401>

## STRING

<https://string-db.org/>