

Ion Channels

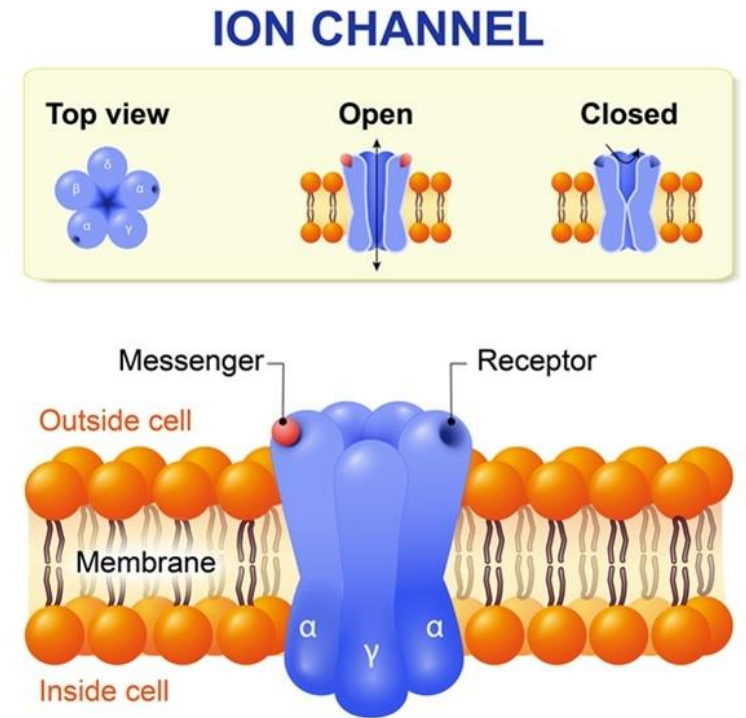
1- Voltage Gated Ion Channels

Ion Channels

Ion channels are pore-forming membrane proteins whose function is

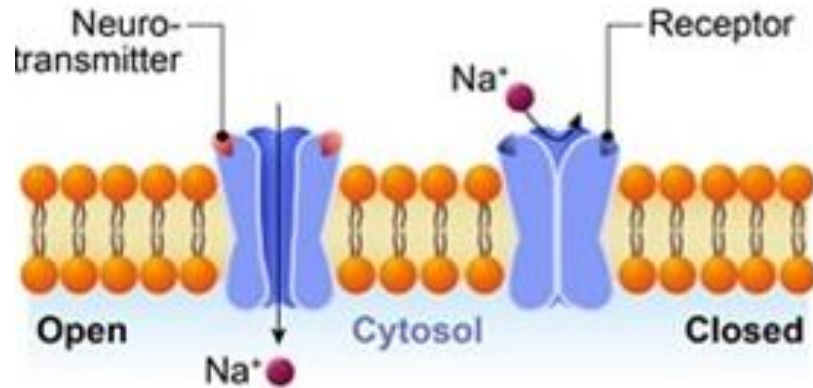
- establishing a resting membrane potential,
- shaping action potentials and other electrical signals by gating the flow of ions across the cell membrane,
- controlling the flow of ions across membranes,
- regulating cell volume.

Their activation translates into a rapid physiological effect

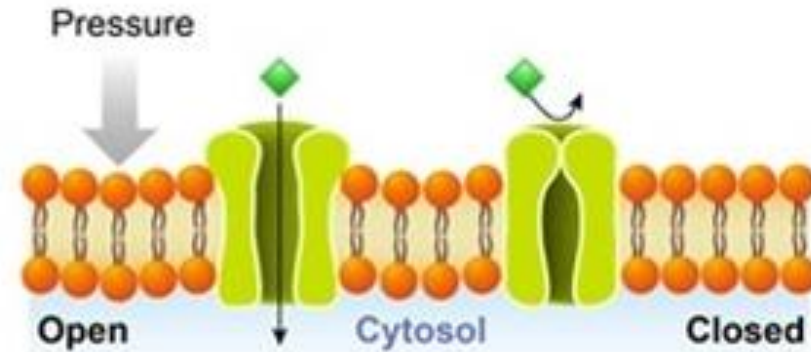


Different classes of ion channels

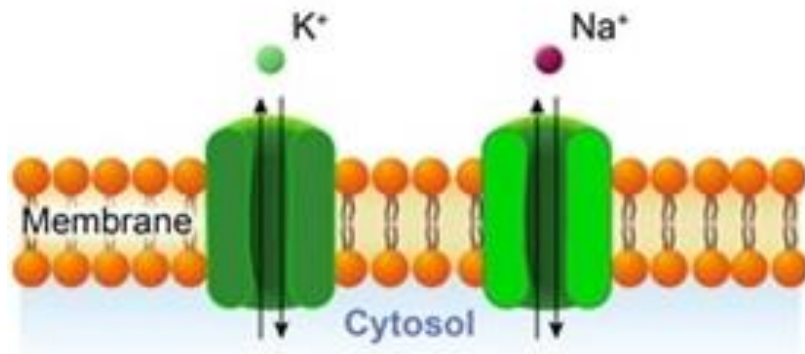
Ligand-gated



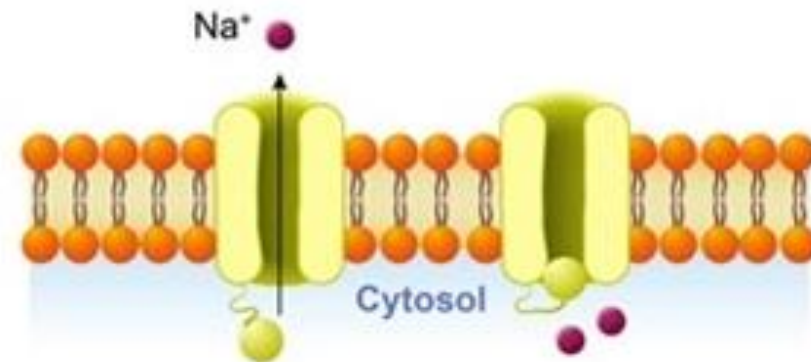
Mechanically-gated



Always open

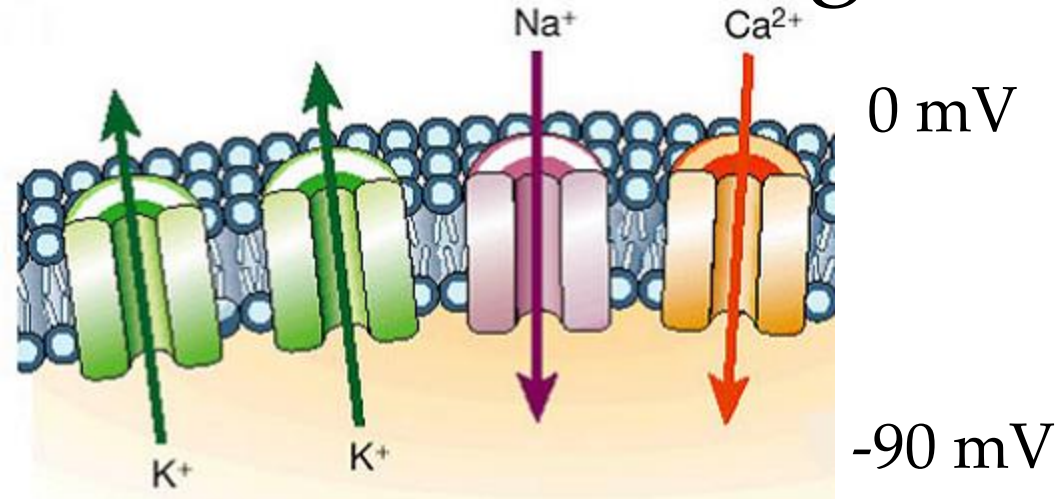


Voltage-gated



Which force moves the ions?

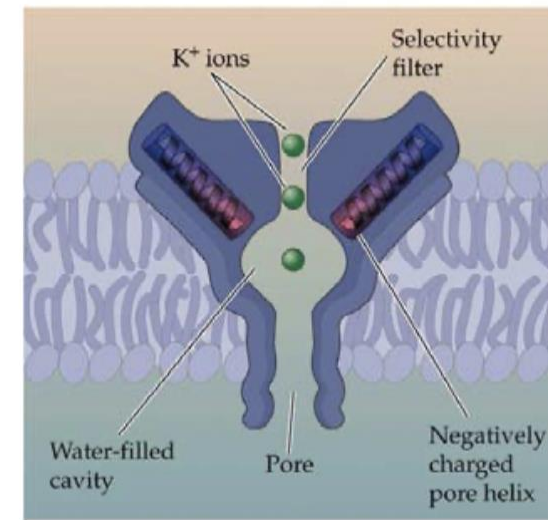
The electrochemical gradient



Ion	Intracellular concentration (mM)	Extracellular concentration (mM)
K^+	140	4
Na^+	15	145
Cl^-	4	110
Ca^{2+}	0.0001	5

What are Ion Channels ?

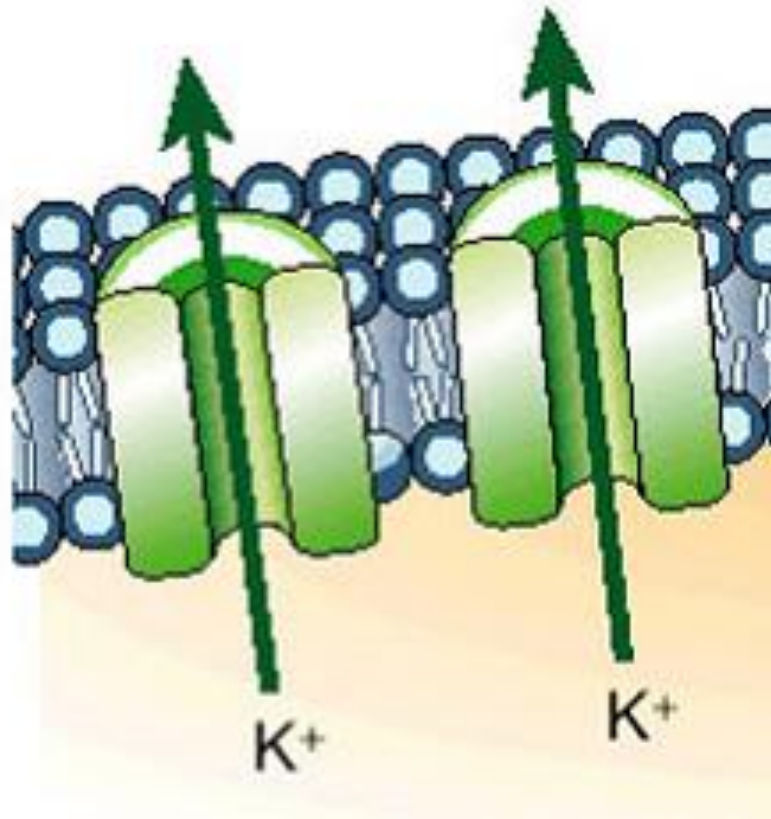
- **Ion channels - structure**
 - are proteins that span (or traverse) the membrane
 - have water-filled 'channel' that runs through the protein
 - ions move through channel, and so through membrane
- **Ion channel - properties**
 - Selectivity: Each specific ion crosses through specific channels
 - Gating: transition between states (closed $\leftrightarrow$ open $\leftrightarrow$ Inactivated)
 - Voltage-gated ; Ligand-gated
 - Channels mediate ion movement down electrochemical gradients.
 - Activation of channel permeable to ion **X** shifts membrane potential towards to its Equilibrium Potential, E_X



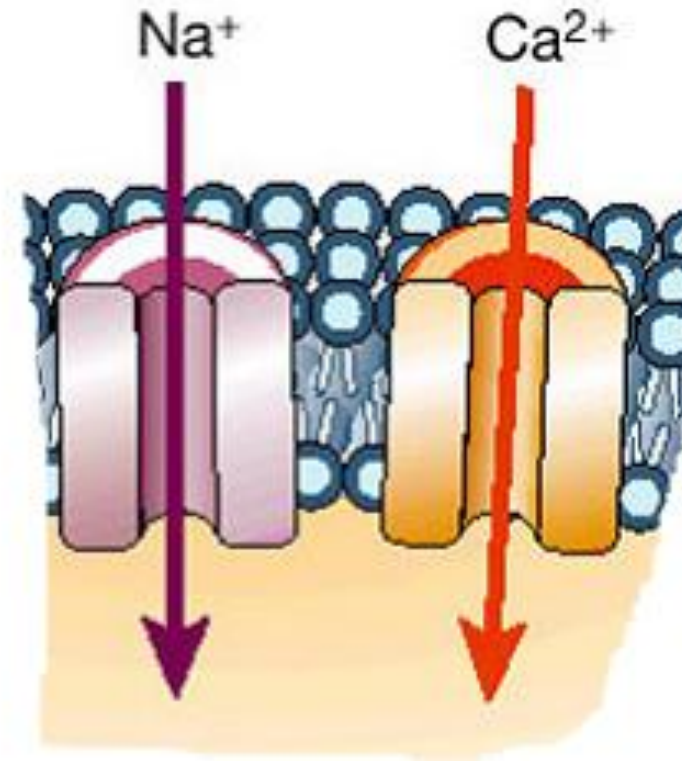
Physiological Function of Ion Channels

- **Maintain cell resting membrane potential:** inward rectifier K and Cl channels.
- **Action potential and Conduction of electrical signal:** Na, K, and Ca channels of nerve axons and muscles
- **Excitation-contraction (E-C) coupling:** Ca channels of skeletal and heart muscles
- **Synaptic transmission at nerve terminals:** glutamate, Ach receptor channels
- **Intracellular transfer of ion, metabolite, propagation:** gap junctions
- **Cell volume regulation:** Cl channel, aquaporins
- **Sensory perception:** cyclic nucleotide gated channels of rods, cones
- **Oscillators:** pacemaker channels of the heart and central neurons
- **Stimulation-secretion coupling:** release of insulin from pancreas (ATP sensitive K channel)

What happens when ions move?



$V_m \rightarrow -90 \text{ mV}$



$V_m \rightarrow +50 \text{ mV}$

Equilibrium Potential or Nernst Potential

The voltage at which there is zero net flux of a given ion
(Electrical gradient = a chemical concentration gradient)

For K^+ : $\sim -90 \text{ mV}$

$$E_K = \frac{RT}{ZF} \ln \frac{[K^+]_o}{[K^+]_i}$$

- R = gas constant
- F = Faraday constant
- T = temperature (K)
- Z = valence (charge) of ion ~



Hermann (Walther) Nernst
1864-1941
1920 Nobel Prize for chemistry



K current (I_{K1}) is the major contributor for RMP

Four Milestones in Ion Channel Research

1. Ionic conductance

Noble 1963 (Physiol/Medicine)



Alan L. Hodgkin



Andrew F. Huxley

2. Patch clamp methodology

Noble 1991 (Physiol/Medicine)



Erwin Neher



Bert Sakmann

3. Channel cloning sequencing (Ach receptor, Na, Ca channels)

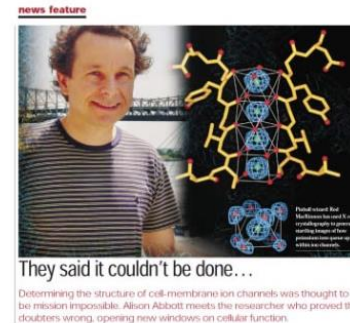


Japan Academy
Prize 1985

Shosaku Numa (沼 正作)

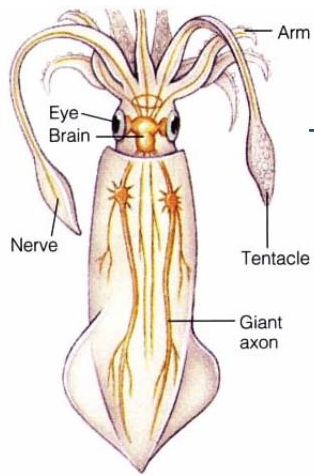
4. K channel structure

Noble 2003 (Chemistry)

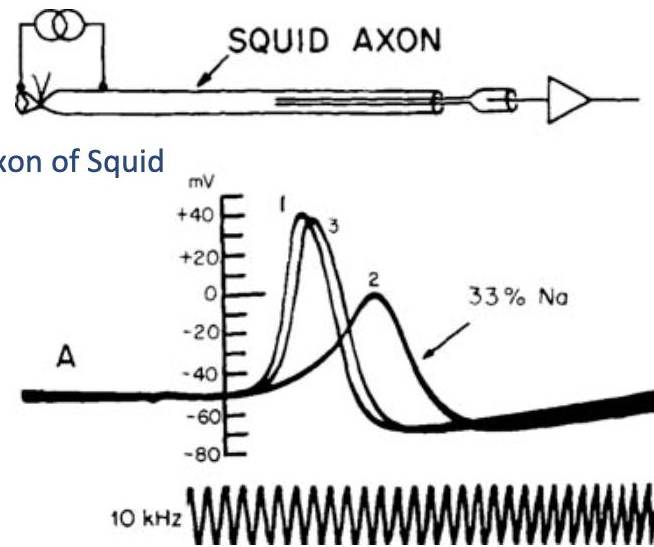


Rod MacKinnon

Hodgkin-Huxley Model Predicted the Existence of Ion Channels



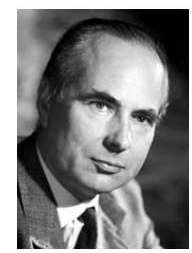
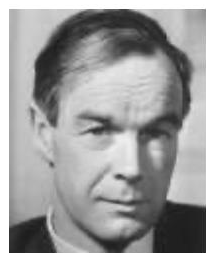
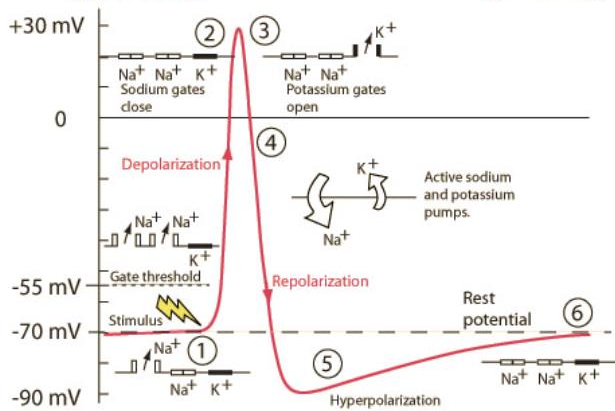
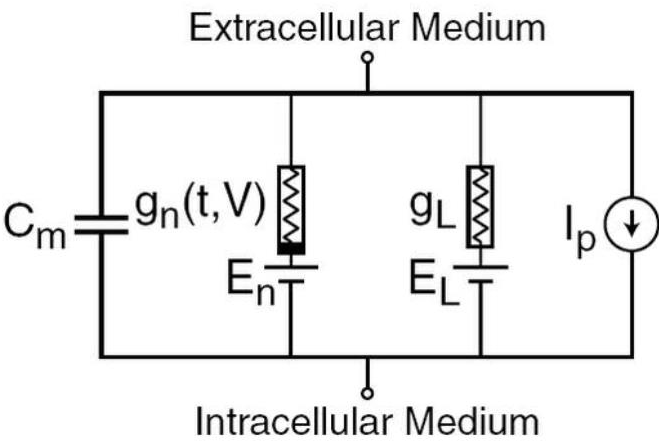
The Giant Axon of Squid



$$C \frac{dV}{dt} = \bar{g}_{Na} m^3 h (V_{Na} - V) + \bar{g}_K n^4 (V_K - V) + g_L (V_L - V) + I_{ext}$$

Na channel
gating

K channel
gating

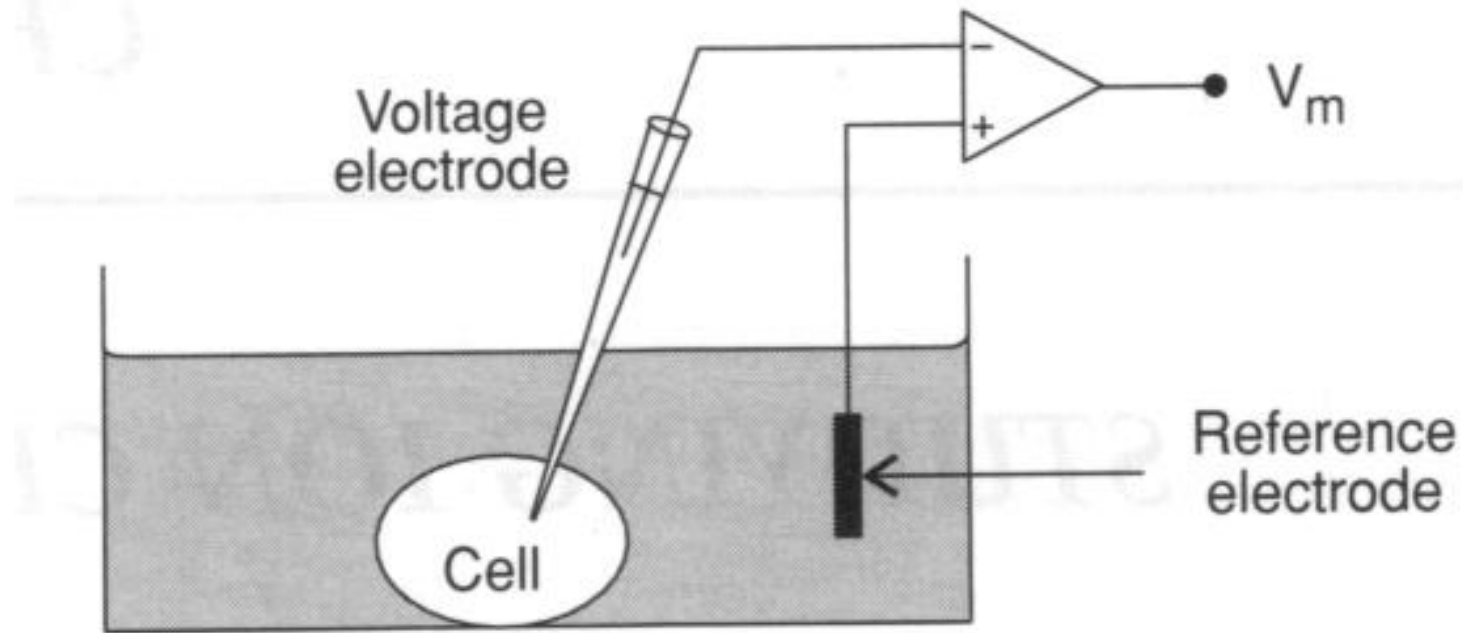


1963 noble Prize

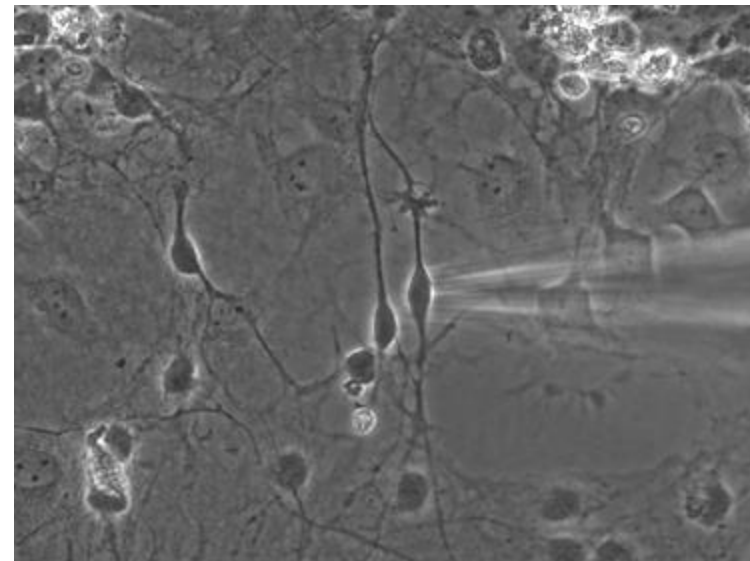
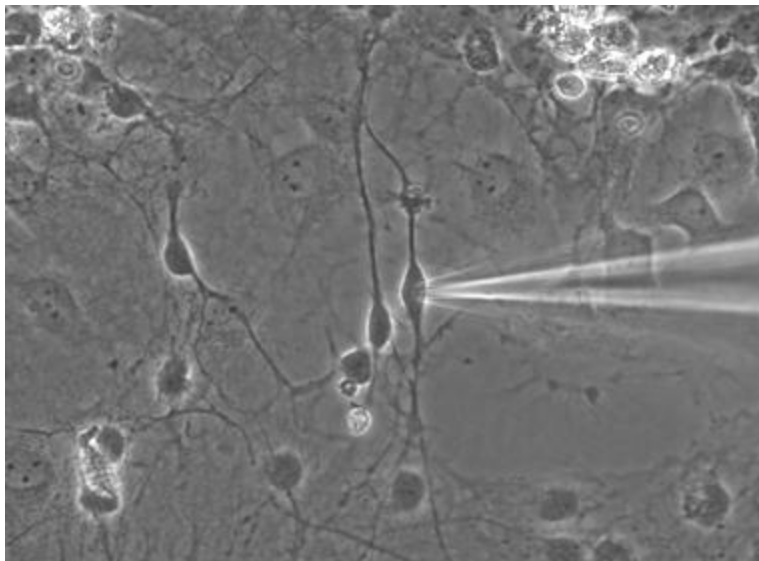
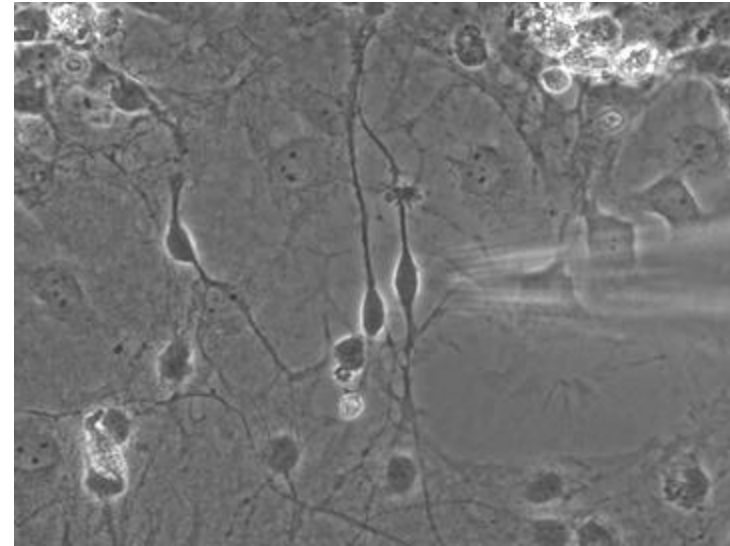
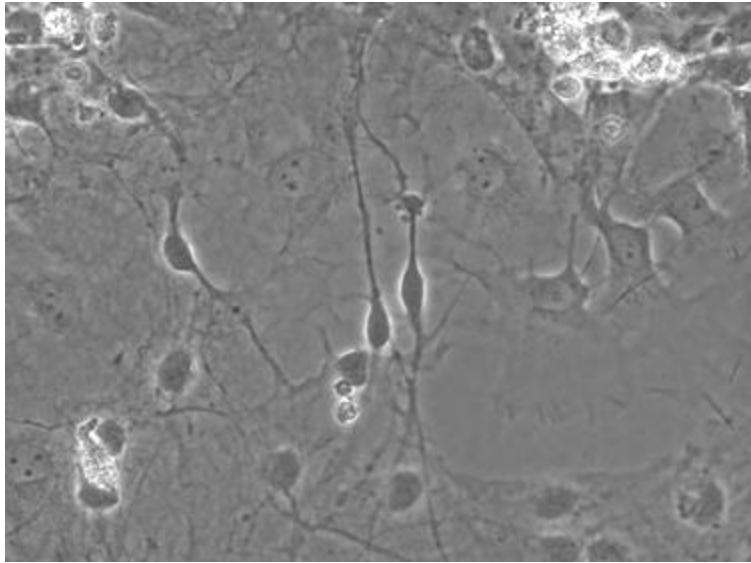
How to study ion channels?

How to study ion channel function?

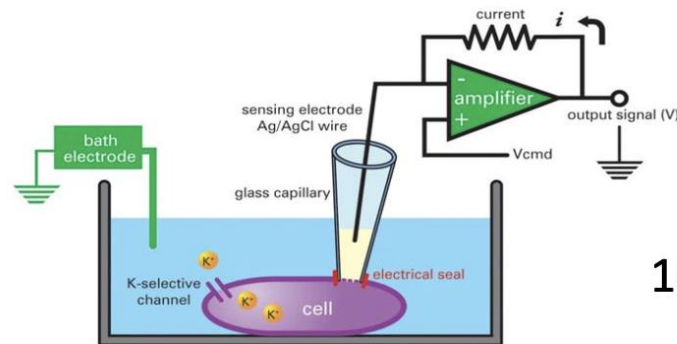
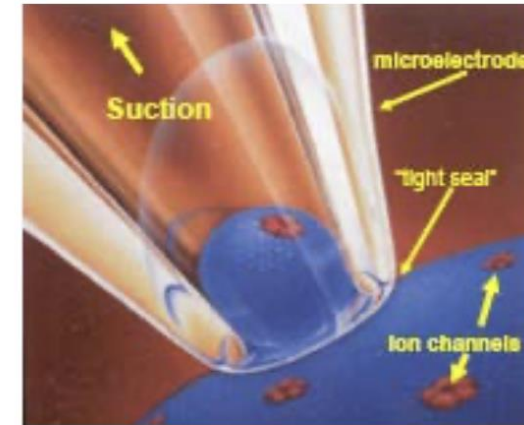
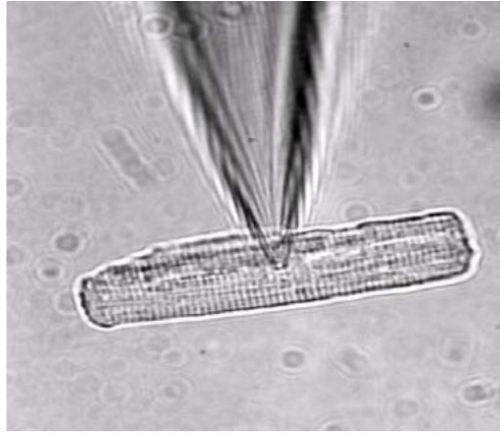
Electrophysiological approaches



Electrophysiological recordings

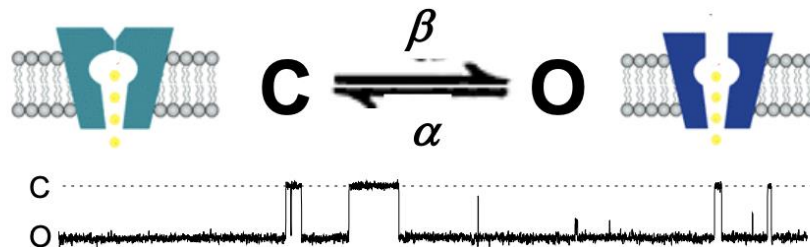


Patch-Clamp Technique



1976

Erwin Neher & Bert Sakmann
Nobel prize for medicine in 1991
for the development of the patch-
clamp technique making possible
the characterization of single ion
channels



1991 Nobel Prize

Patch-Clamp Technique

The Patch Clamp Method

© Sinauer Associates, Inc.

How to study ion channel function?

Molecular approaches

NATURE VOL. 305 27 OCTOBER 1983

Cloning and sequence analysis of calf cDNA and human genomic DNA encoding α -subunit precursor of muscle acetylcholine receptor

Masaharu Noda, Yasuji Furutani, Hideo Takahashi, Mitsuyoshi Toyosato, Tsutomu Tanabe, Shin Shimizu, Sho Kikuyotani, Toshiaki Kayano, Tadaaki Hirose*, Seiichi Inayama* & Shosaku Numa

Department of Medical Chemistry, Kyoto University Faculty of Medicine, Kyoto 606, Japan
* Pharmaceutical Institute, Keio University School of Medicine, Tokyo 160, Japan

Numa

Sakmann



LETTERS TO NATURE

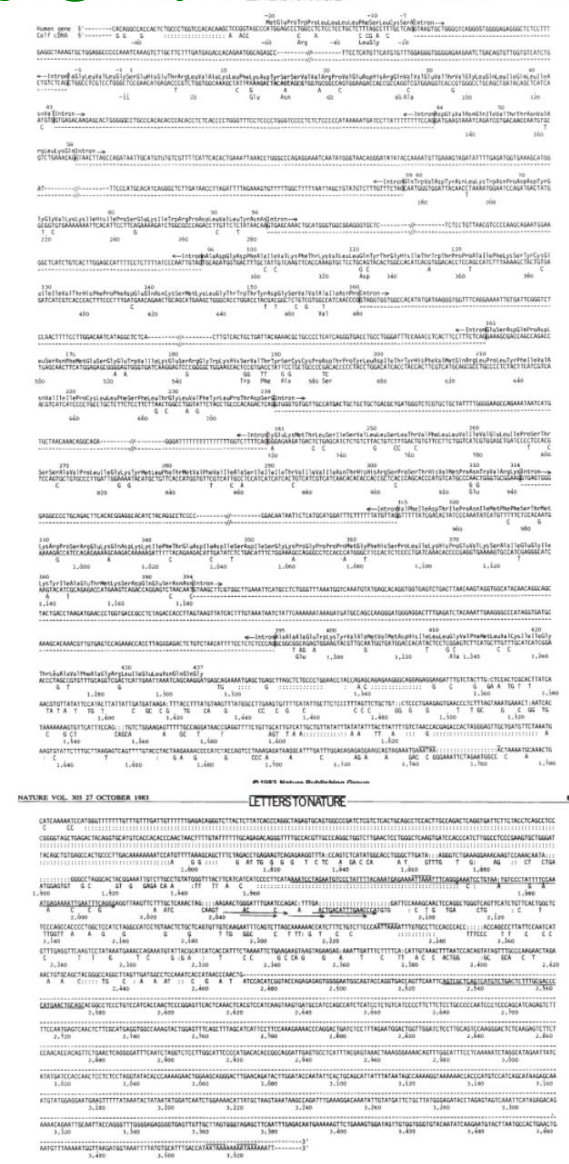


Fig. 1 Nucleotide sequence of the human genomic DNA (upper line) and the calf cDNA (lower line) encoding the AChR α -subunit precursor. The nucleotide sequence of the human genomic DNA, together with the deduced amino acid sequence, is shown. The nucleotide and amino acid differences found in the calf sequence aligned are displayed beneath the human sequence. The absence of a nucleotide or an amino acid in the calf sequence indicates that the human and calf sequences are the same; the presence of a colon in either nucleotide sequence indicates a gap; the hyphens indicate sequences that have not been determined or cloned or that exhibit no evident homology with their counterpart. Amino acid residues are numbered beginning with the amino-terminal residue of the mature α -subunit, and the preceding residues are indicated by negative numbers. Nucleotide residues in the calf cDNA are numbered by vertical lines. The 5'-terminal sequence shown is the first residue of the coding sequence of the mature α -subunit, and the nucleotides on the 5' side of residue 1 are indicated by negative numbers. The non-coding junctions, which are positioned according to the G-C-A-G rule, are indicated by vertical lines. The 3'-terminal sequence shown is not extended to the 3' end of the mRNA or the gene. The 3'-terminal cDNA sequence presented is a poly(A) tail of 66 nucleotides connected with the vector DNA sequence (5'-GGG-3'), thus apparently representing the complete sequence of this region, whereas the 3'-terminal gene sequence shown is probably incomplete (see text). The polyadenylation signal AATAAA (nucleotides 18, 39) present downstream of the transcriptional termination site are overlined. The sequence homologies with the human sequence (complementary to a portion of the consensus sequence given in ref. 2) and the direct repeats present in the 3'-untranslated region are shown by an underline and the horizontal arrows beneath the sequences, respectively.

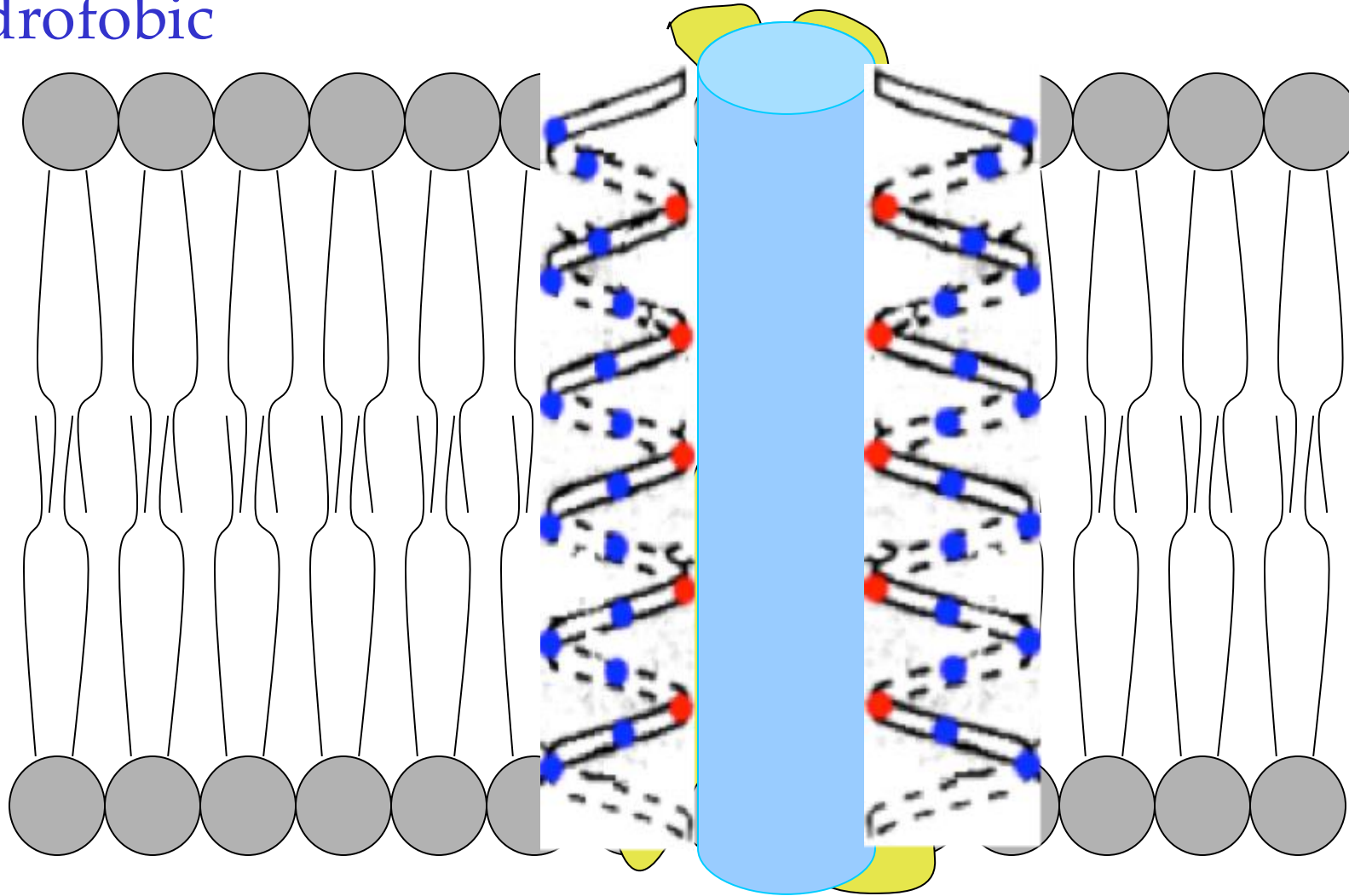
How to study ion channel function?

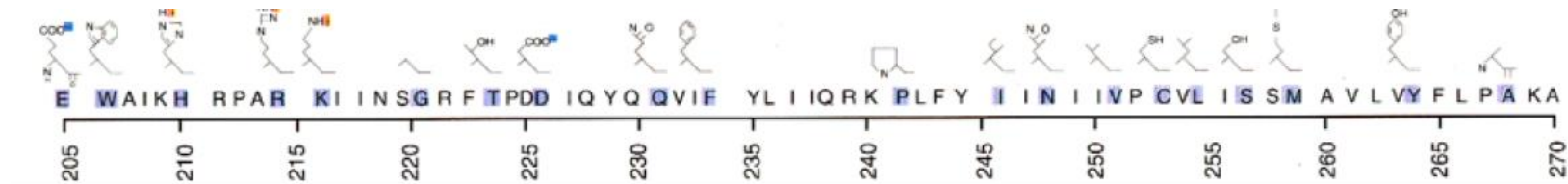
Structural approaches

Hydrophilic
Hydrophobic

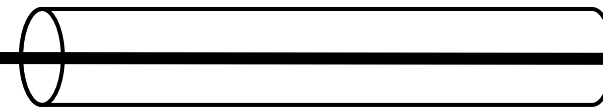
Predictions...

Hydrophilic
Hydrofobic





extracellular



e

Transmembrane segment

(α -elix or β -sheet)

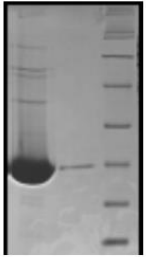


Structure

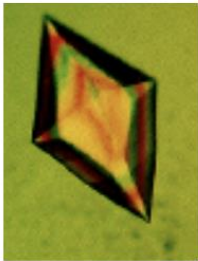
Crystal structure of ion channels

Protein x-ray crystallography

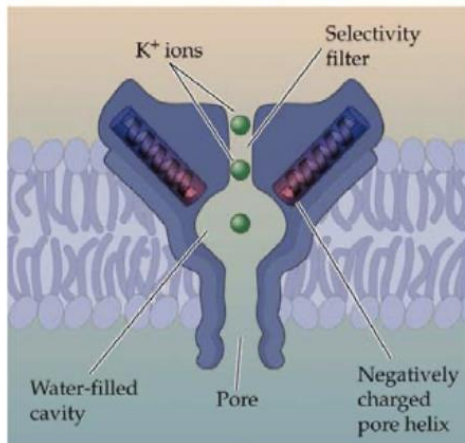
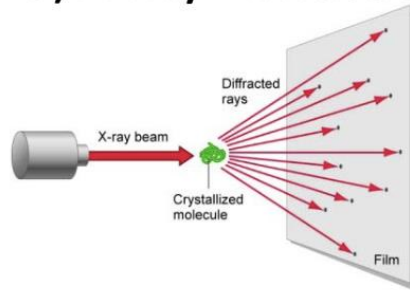
1) Purification



2) Crystallization



3) X-Ray Diffraction



The Nobel Prize in Chemistry 2003
Peter Agre, **Roderick MacKinnon**

Classification of Ion Channels

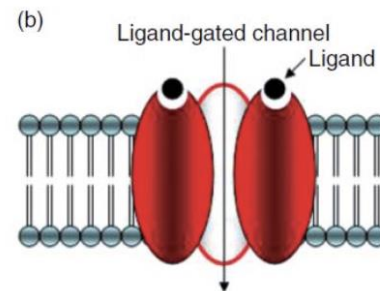
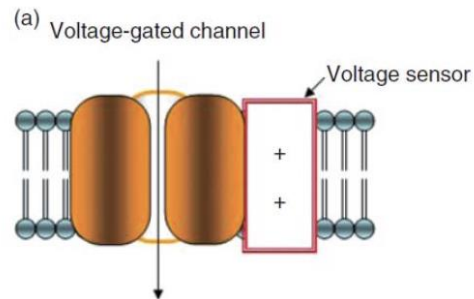
1) Based on ion selectivity:

K^+ , Na^+ , Ca^{2+} , Cl^- channels

2) Based on gating:

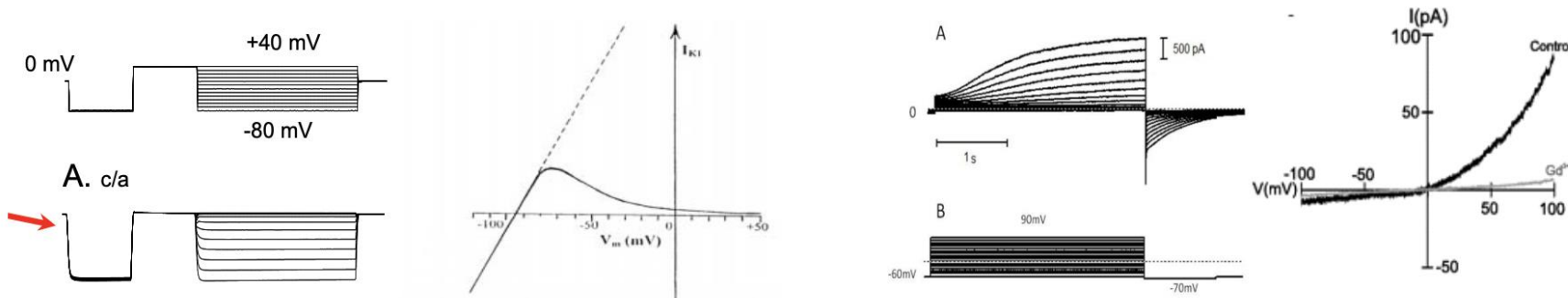
Voltage-gated : ions

Ligand-gated: Glutamate, GABA, ACh, ATP, cAMP



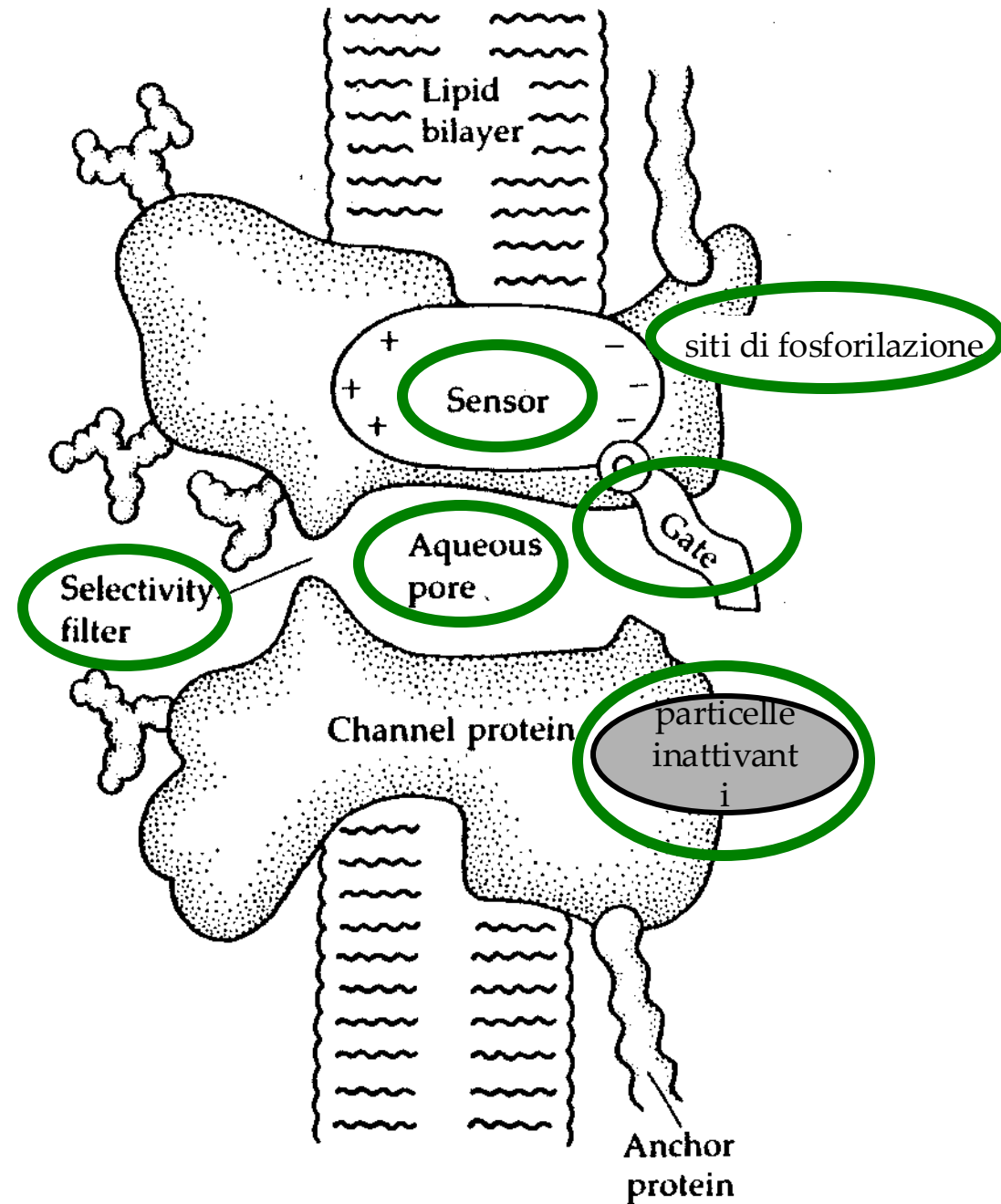
3) Based on rectification:

Inwardly or outwardly rectifying



Structural motifs

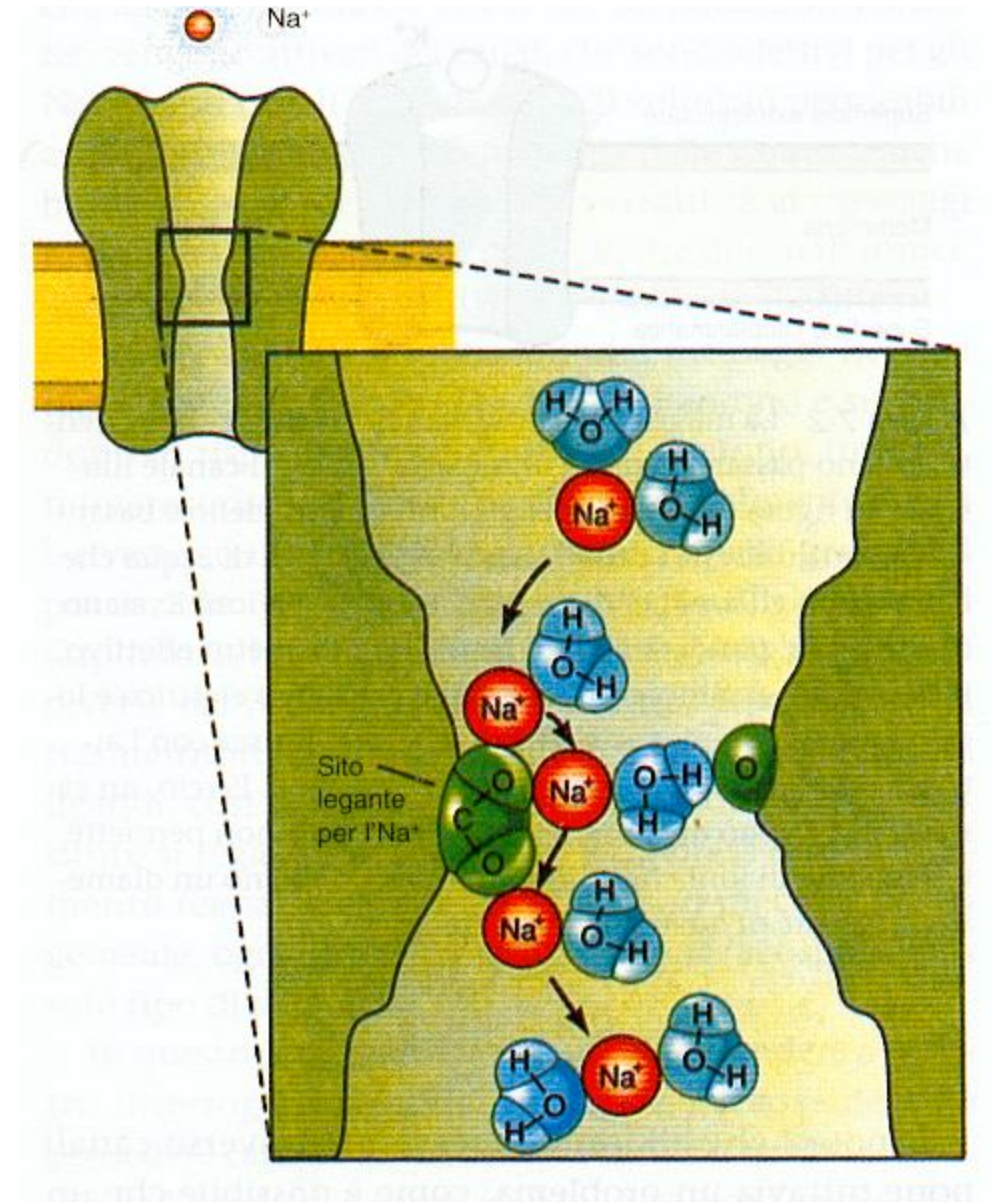
ie: what's in all the channels?



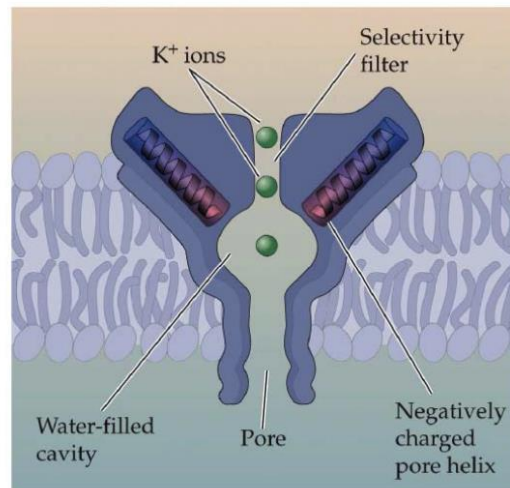
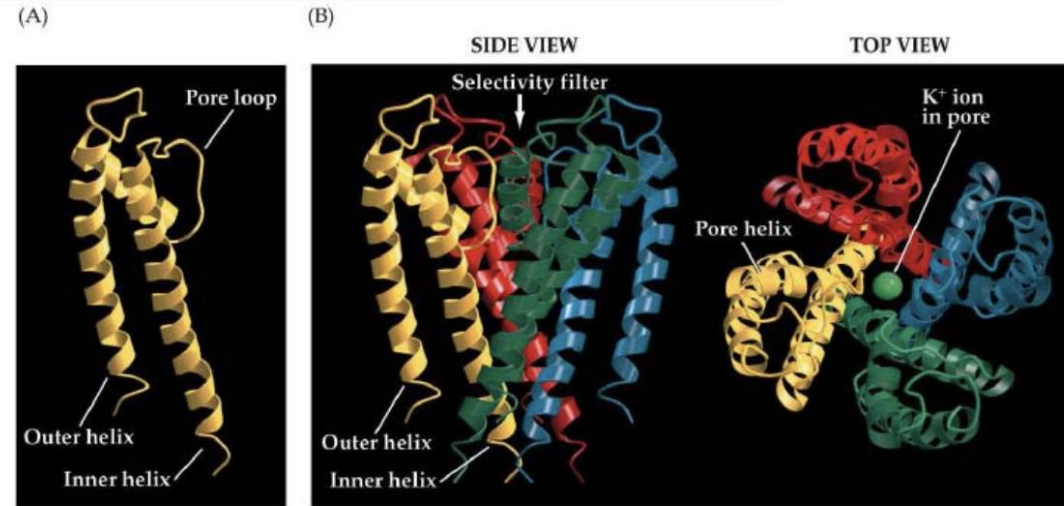
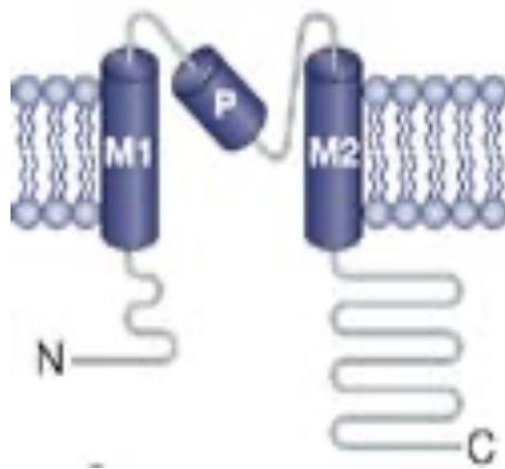
Selectivity filter (3/4 aa)

Ions interact with charged aa

- 1) Each channel excludes ions of dimensions > characteristic value. Es. K $3\text{\AA}$, Ca $5\text{\AA}$...
- 2) Important charged aa are cations binding sites - and exclude ion water hydration molecules
- 3) Due to the reduced size of the filter important steric effects occur, more ions occupy -greater repulsion (2 Ca or 4 Na neutralize 4 + charges)

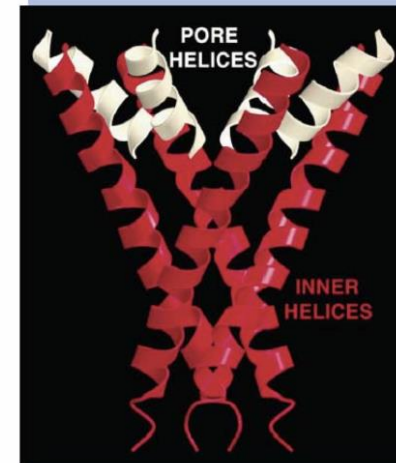


Structure of KCSA Channels: Selectivity Filter and Gating



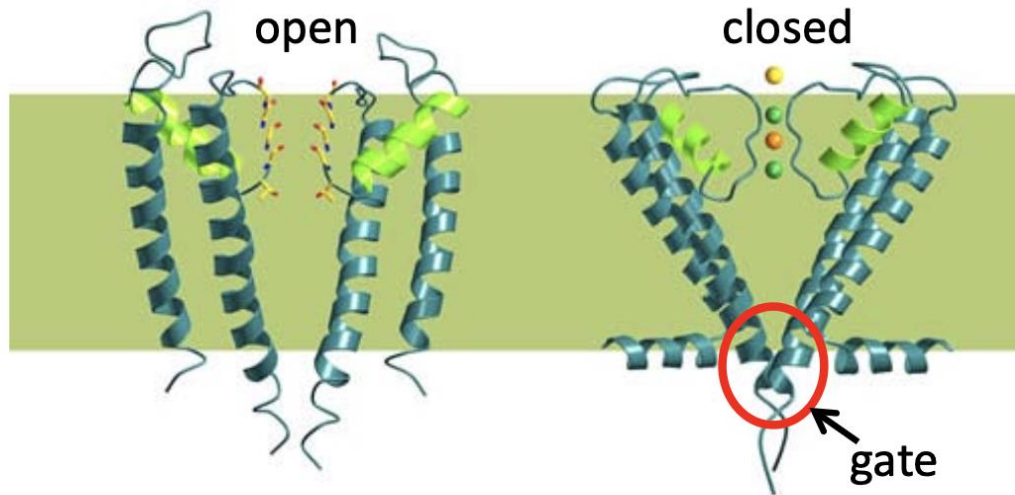
profile

Inner helices form “inverted teepee” structure



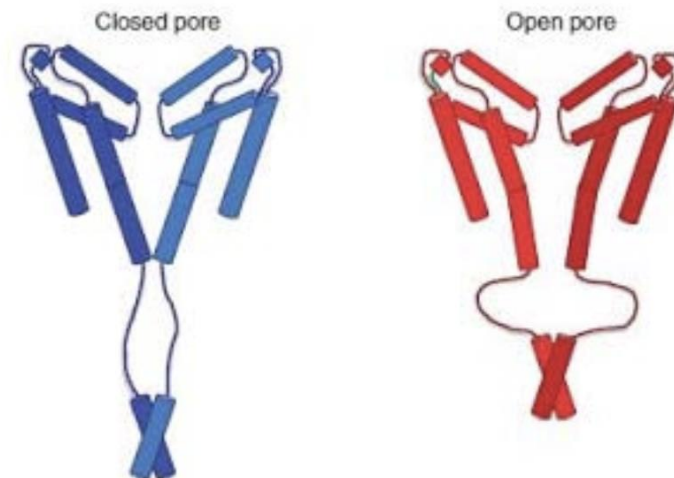
Doyle et al (1998) Science 280, 69

Open-Close Gating



Doyle *et al.* Science 1998;

Bacterial K channel
selective filter: P-loop; **Gating: intracellular side of the pore bundle crossing**

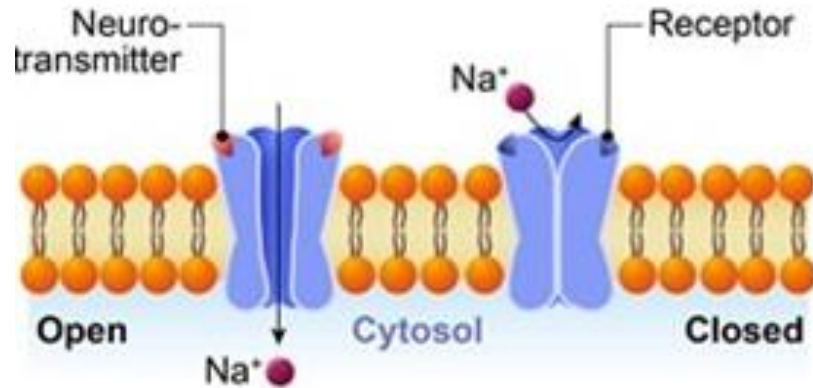


Bacterial Na channel pore in the closed and "open" conformation

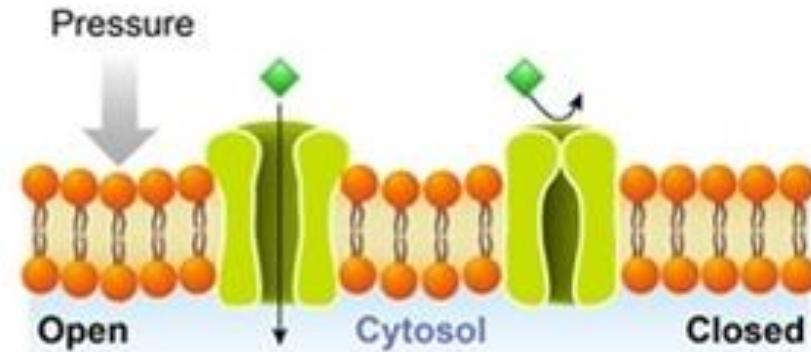
How may we Open/Close a channel?

Different classes of ion channels

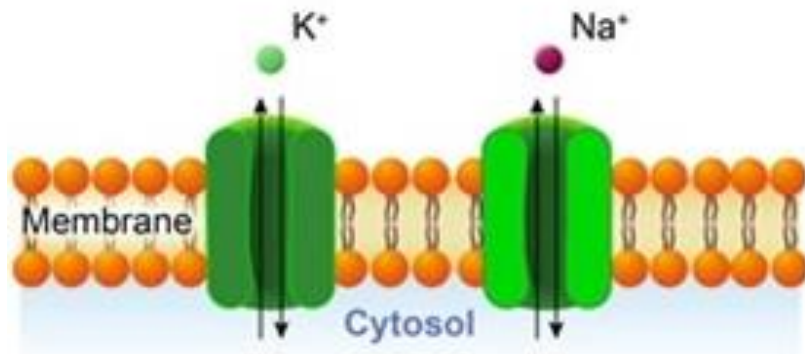
Ligand-gated



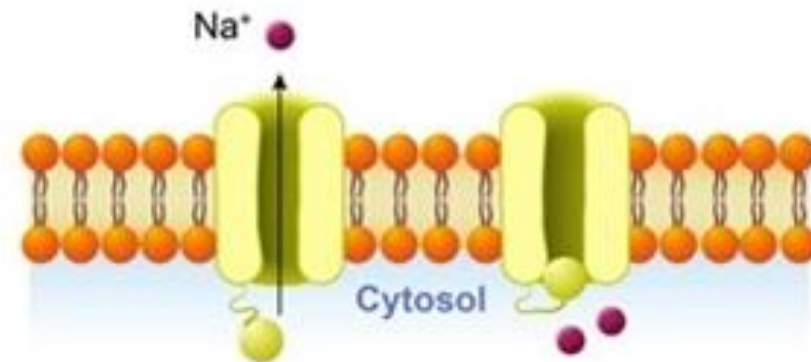
Mechanically-gated



Always open

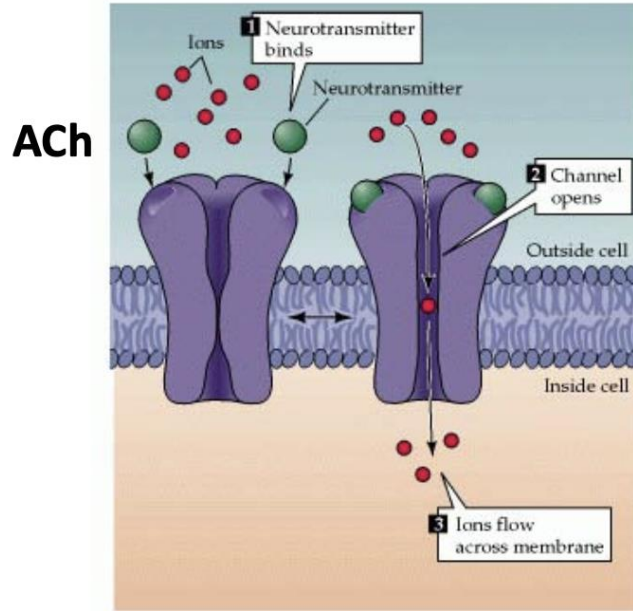


Voltage-gated

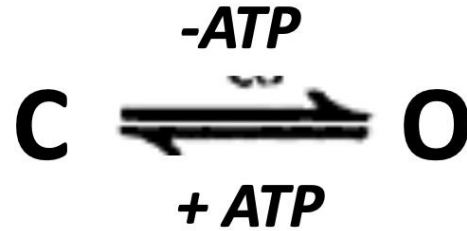


Ligand-Gated Ion Channels

ACh receptor channel



ATP-sensitive K channel

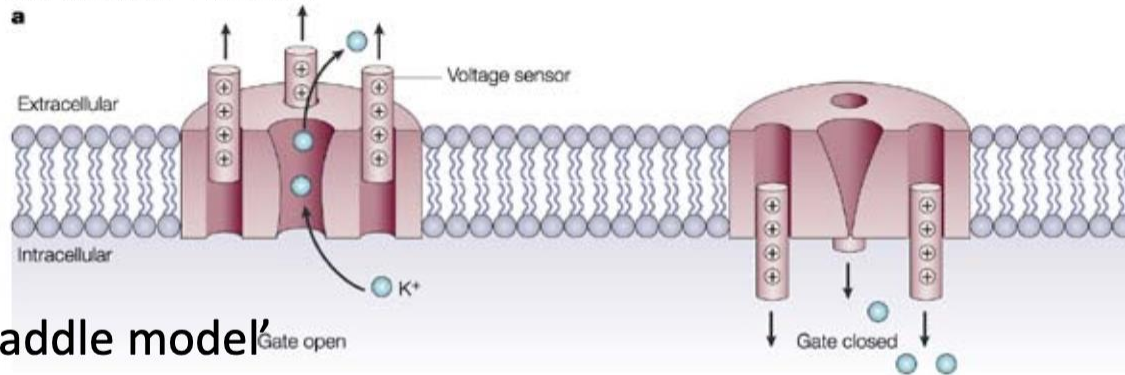


Also a weak inward rectifier

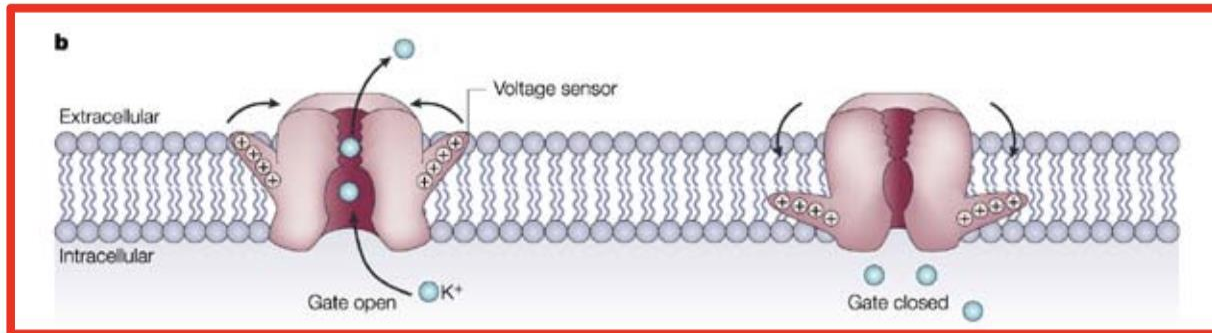
- Open when a signal molecule (ligand) binds to an extracellular receptor region of the channel protein.
- This binding changes the structural arrangements of the channel protein, which then causes the channels to open or close in response to the binding of a ligand such as a neurotransmitter.
- This ligand-gated ion channel, allows specific ions (Na^+ , K^+ , Ca^{2+} , or Cl^-) to flow in and out of the membrane.

Models for Voltage Gating

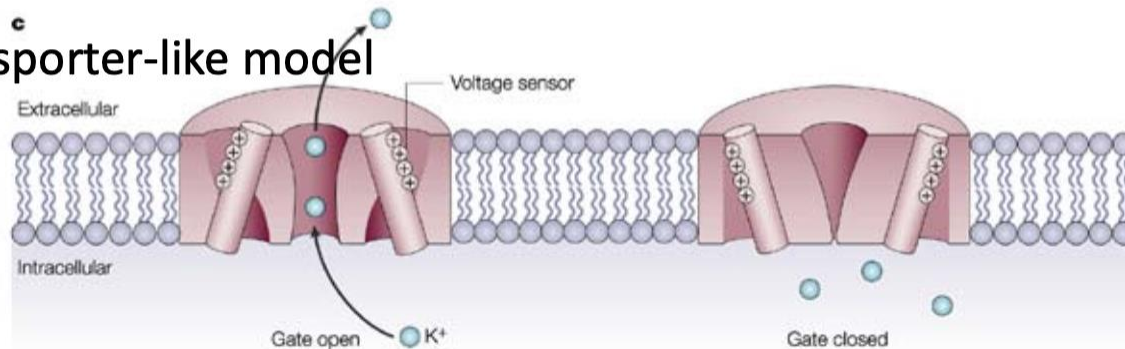
The conventional model



A new 'paddle model'



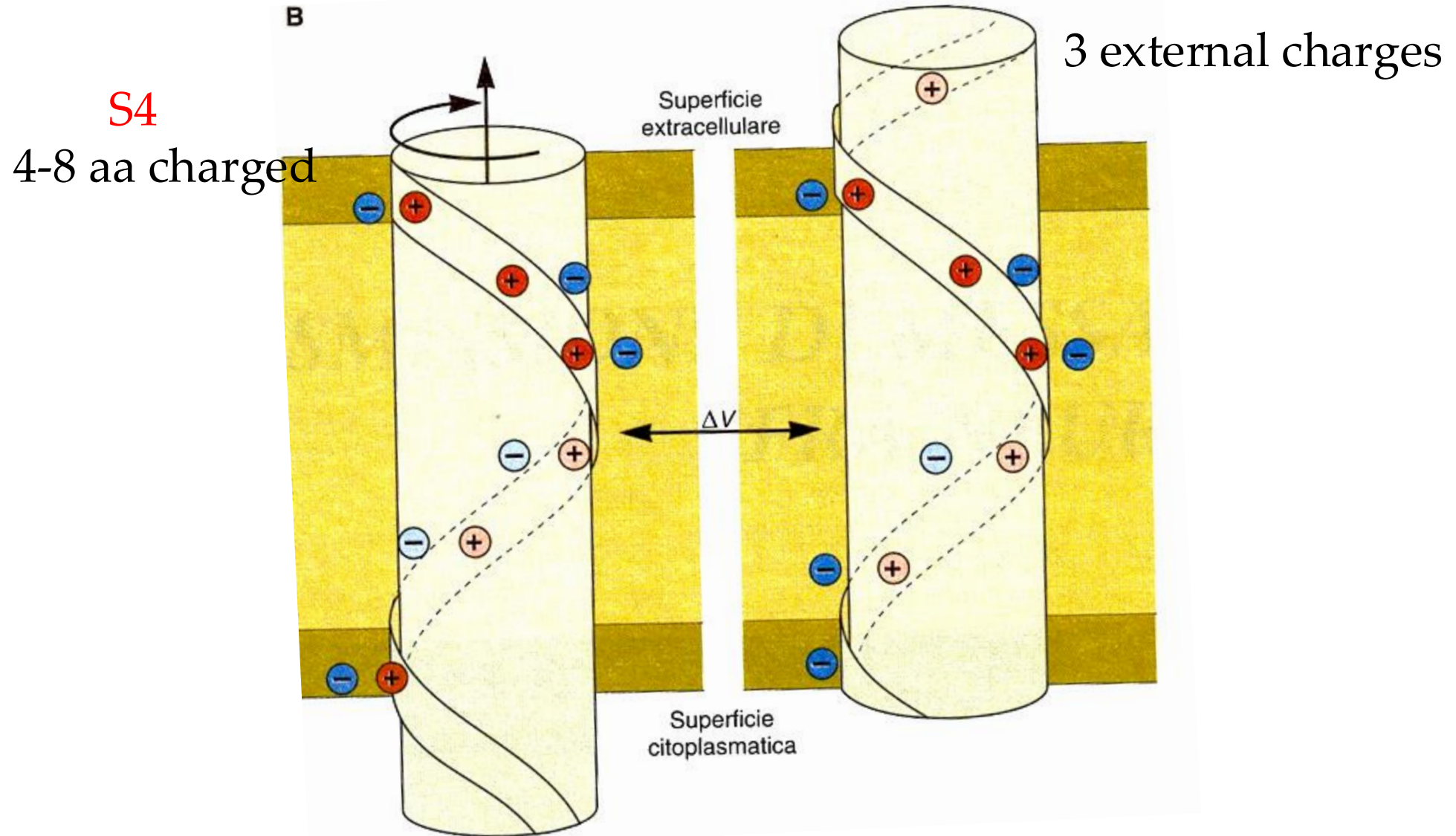
The transporter-like model



the S4 segment is responsible for detecting voltage changes.

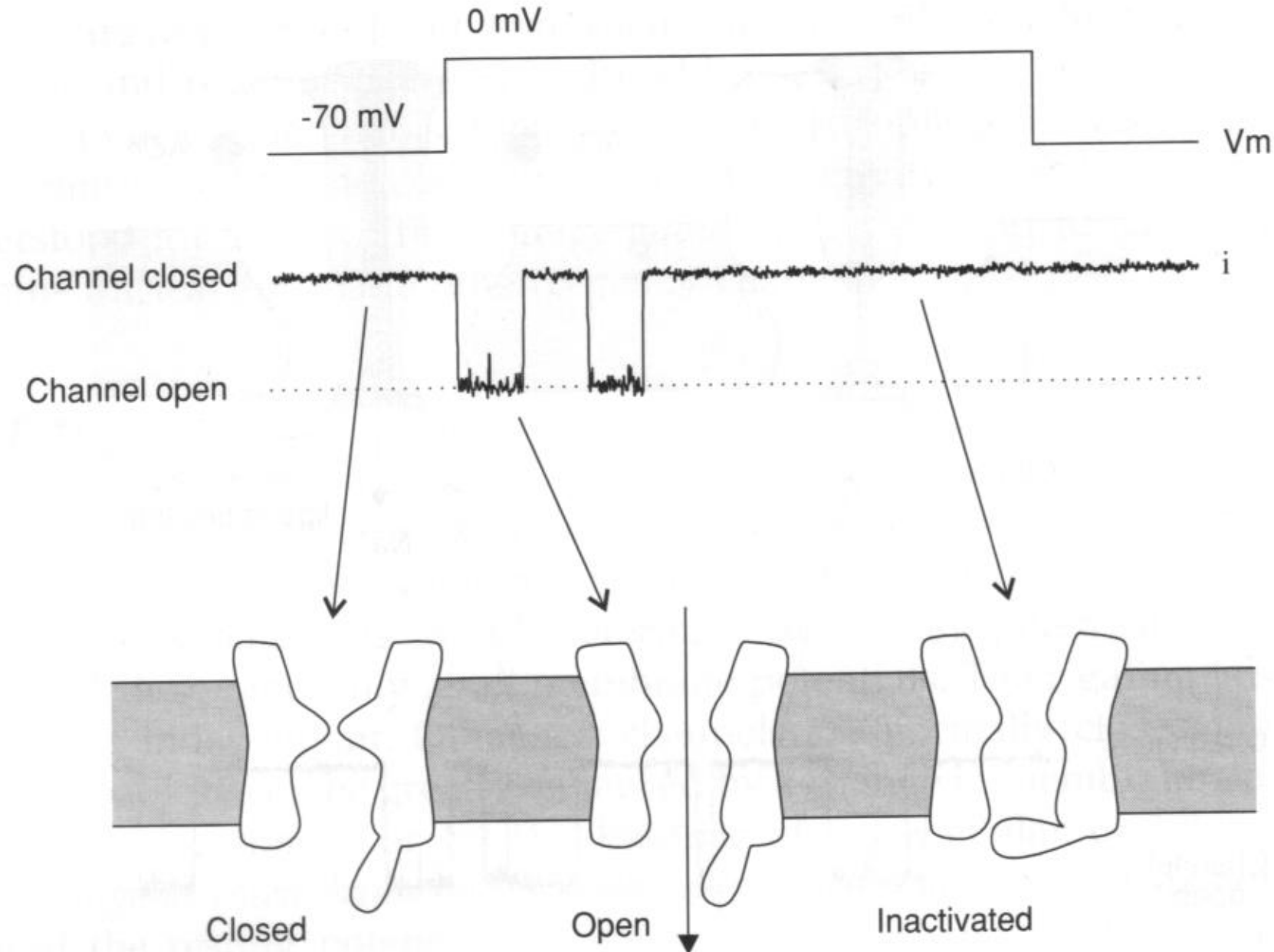
The movement of positively-charged S4 segments within the membrane electric field

Voltage sensor

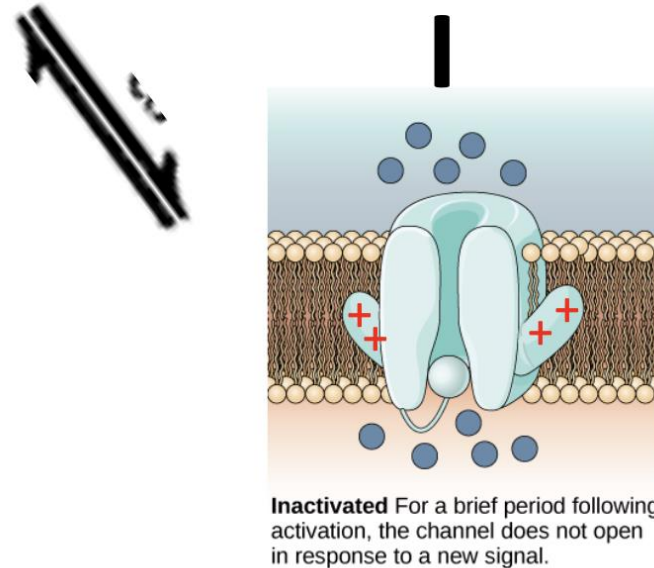
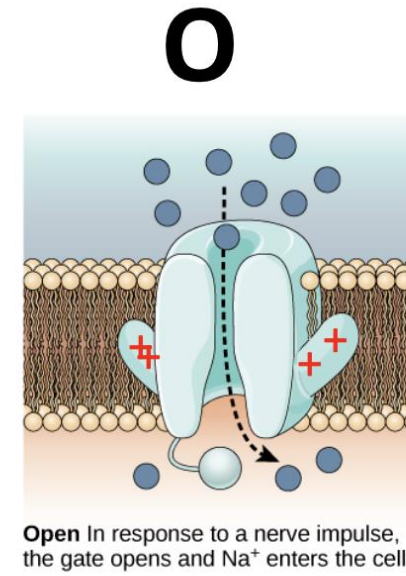
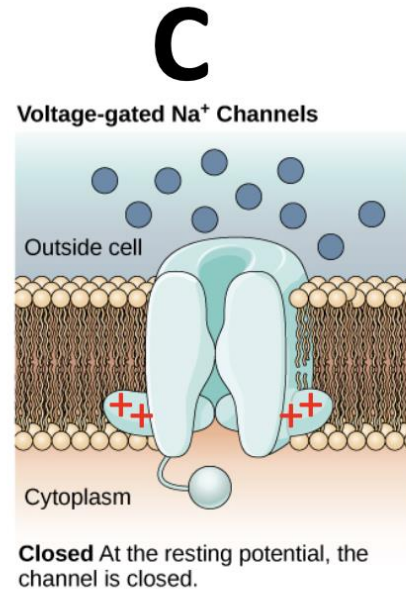


charged groups move in the direction of the electric field
(shown with transient currents)

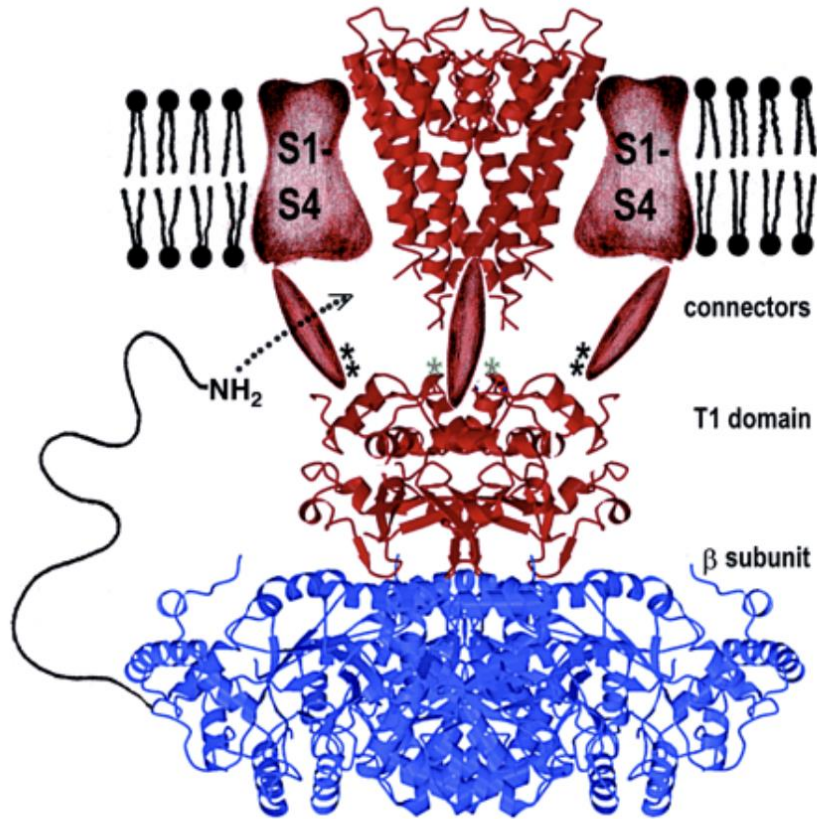
Transition between Closed and Open States... and more..



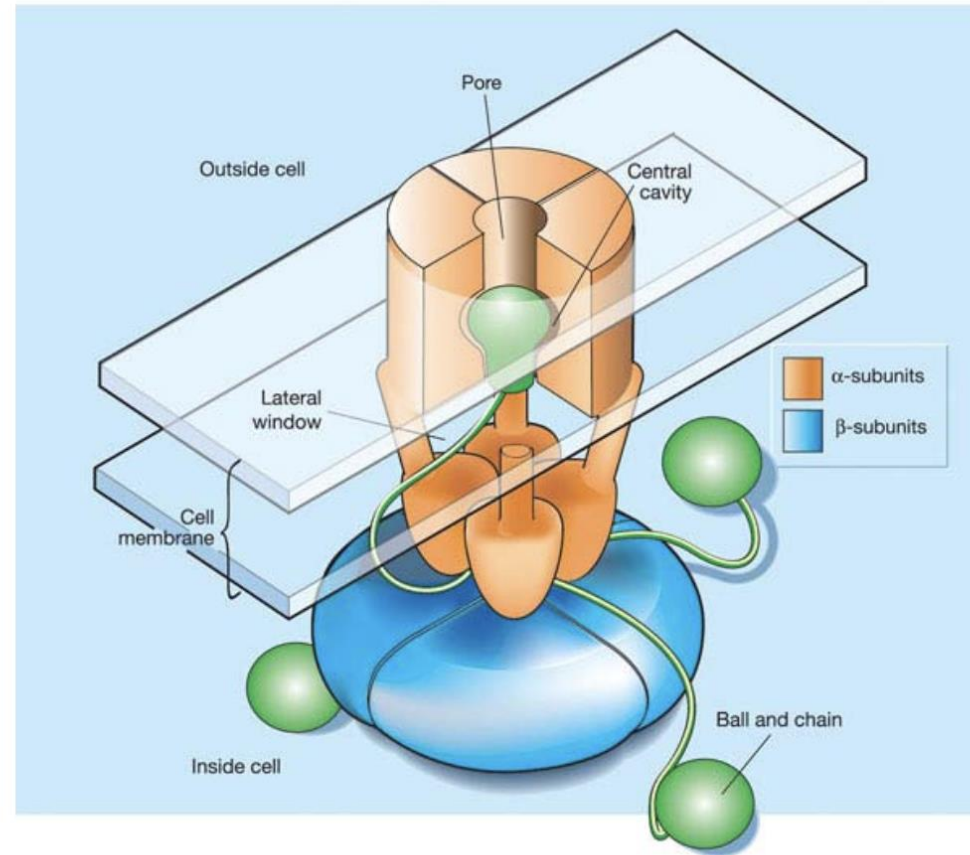
Transition between Closed, Open and Inactivated States



Inactivation Gating of Voltage-Gated Channels -Ball and Chain



(Gulbis et al, Science 2000)



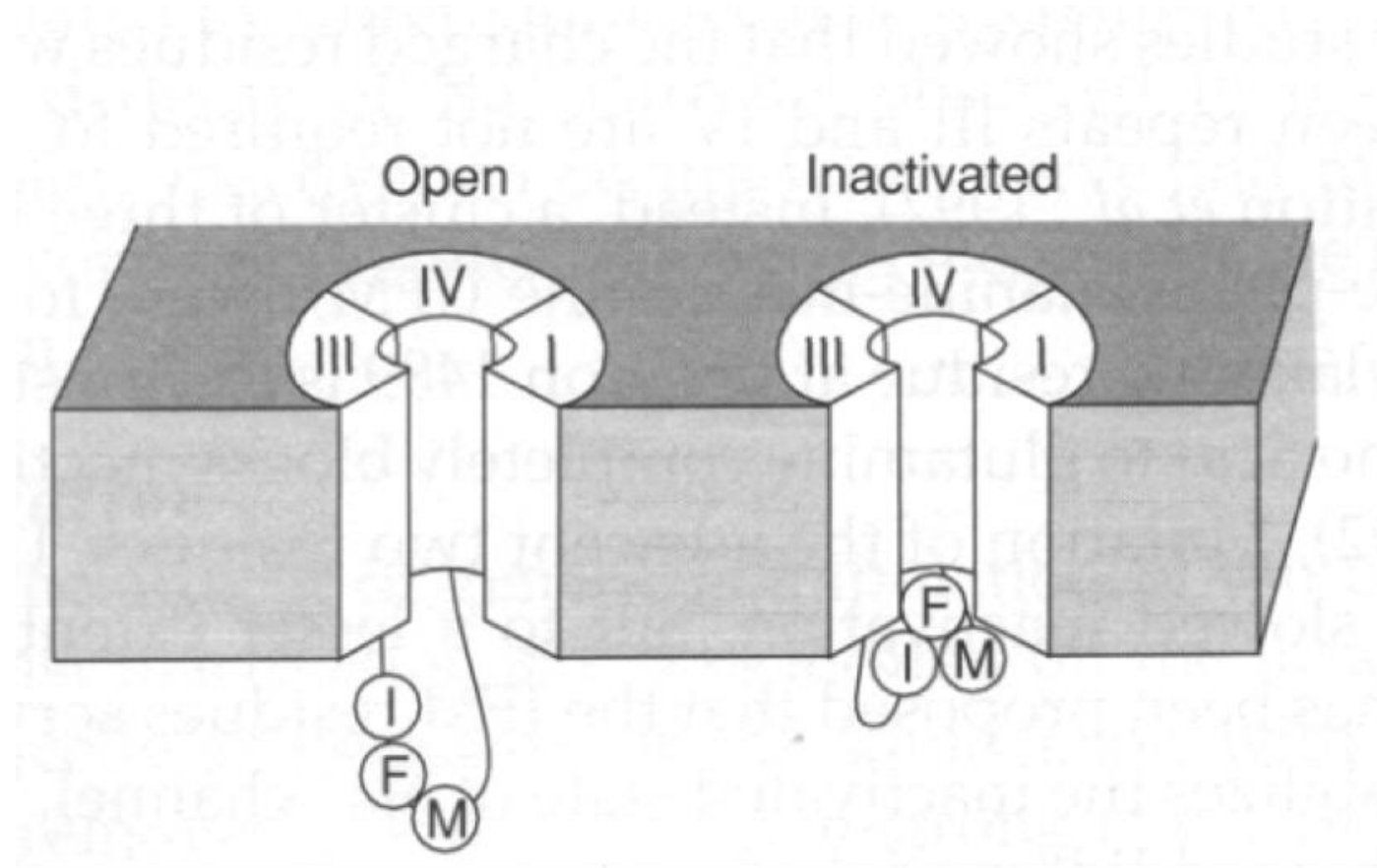
N-terminal inactivation gate

A positively charged inactivation particle (ball) has to pass through one of the lateral windows and bind in the hydrophobic binding pocket of the pore's central cavity.

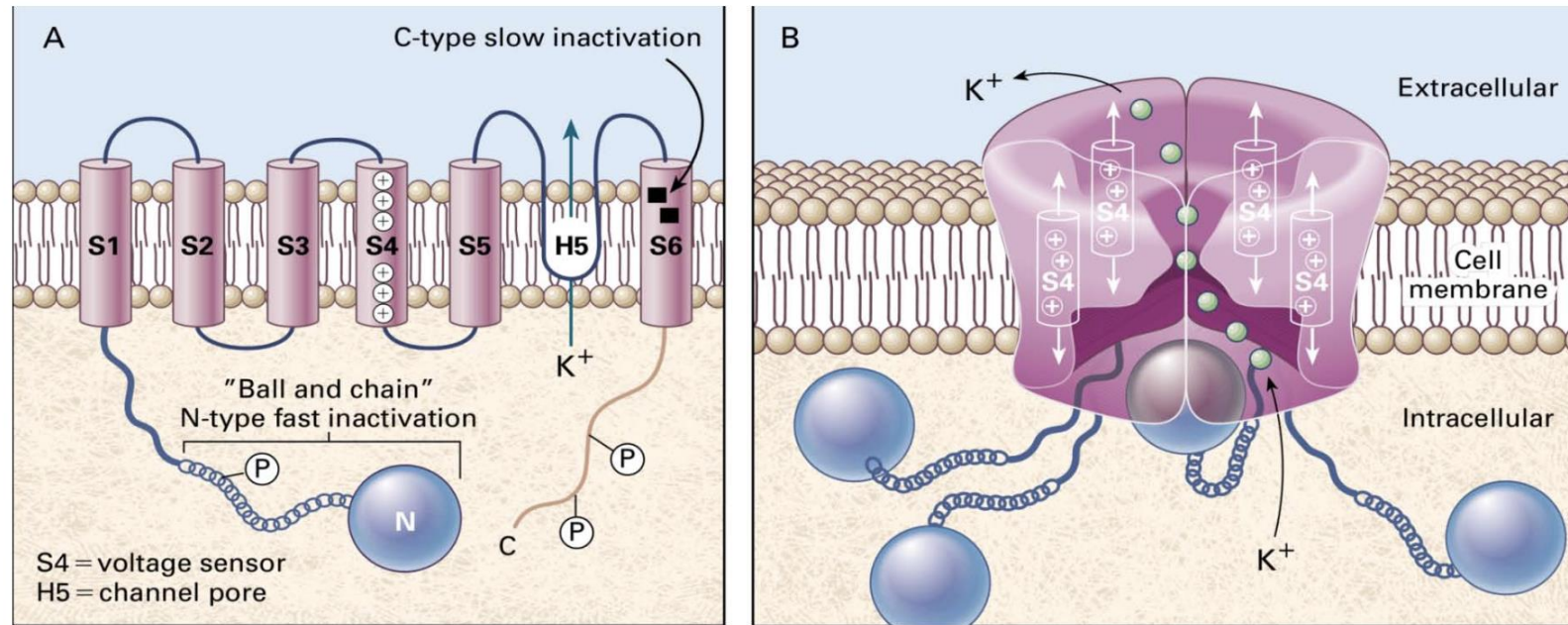
This blocks the flow of ions through the pore.

There are four balls and chains to each channel, but only one is needed for inactivation.

Inactivating particles



Structural Basis of Gating in a Voltage-gated Channel



A: a subunit containing six transmembrane-spanning motifs. S5 and S6 and the pore loop are responsible for ion conduction (channel pore).

S4 is the the *voltage sensor*, which bears positively charged amino acids (Arg) that relocate upon changes in the membrane electric field.

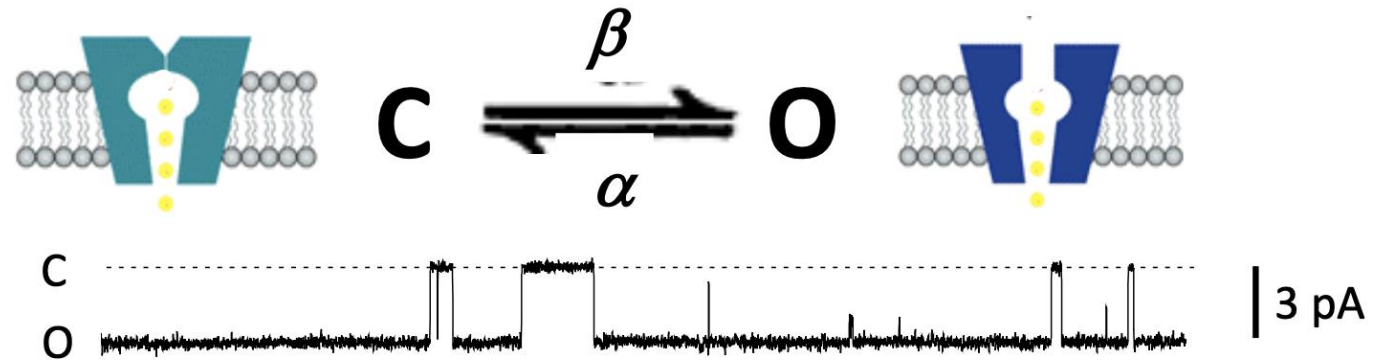
N-terminal ball-and-chain is responsible for inactivation

B: four such subunits assembled to form a potassium channel.

Channel Function: Single Channel and Whole-cell Current

Ion channels are not open continuously but open and close in a stochastic or random fashion.

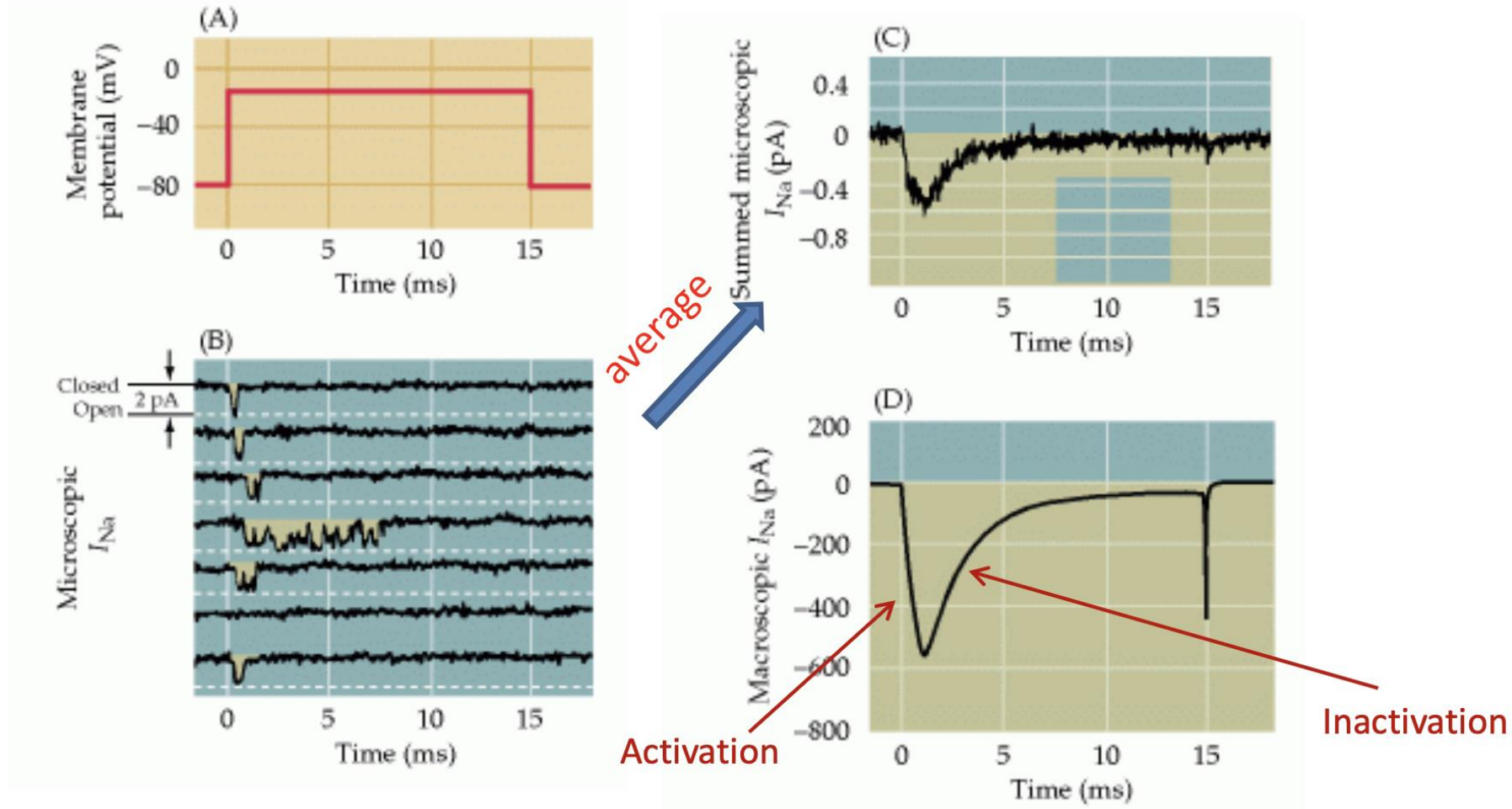
- Ion channel function may be decreased by
 - decreasing the open time (O),
 - increasing the closed time (C),
 - decreasing the single channel current amplitude (i)
 - or decreasing the number of channels (n)



$$I = n * P_o * i$$

$$P_o = \frac{\tau_o}{\tau_o + \tau_c}$$

Channel Function: Single Channel and Whole-cell Current



Depolarizing voltage pulses result in brief openings in the seven successive recordings of membrane current

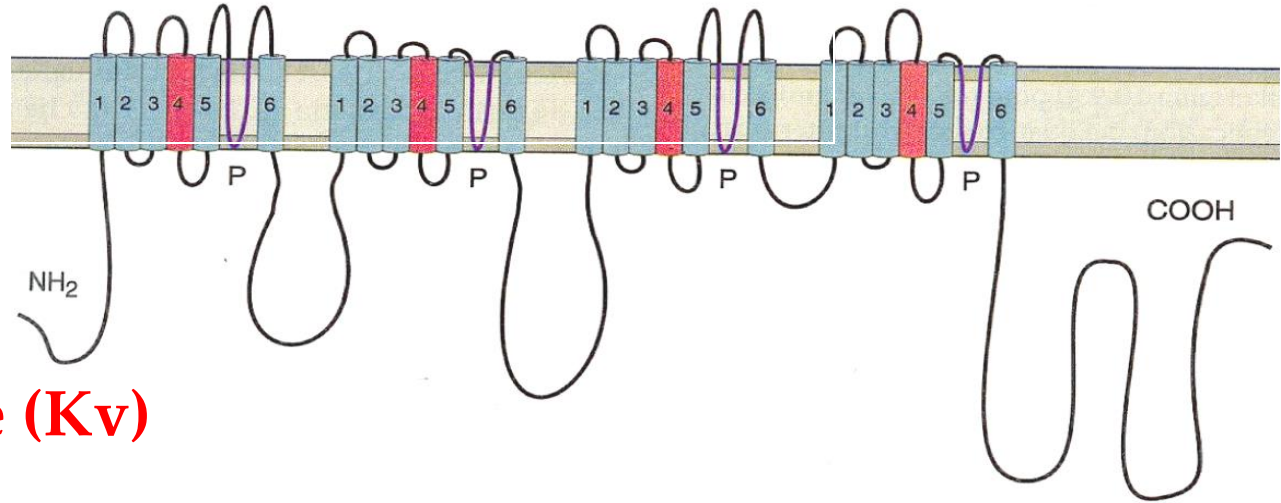
Close correlation between the time courses of microscopic and macroscopic Na^+ currents

Voltage-gated Ion Channels

- **sodium**: I, II, III, $\mu 1$, H1, PN3
- **potassium**: K_A , K_v (1-5), $K_v(r)$, $K_v(s)$, K_{SR} , BK_{Ca} , IK_{Ca} , SK_{Ca} , K_M , K_{ACh}
- **calcium**: L, N, P, Q, T
- **chloride**: ClC-0 - ClC-8

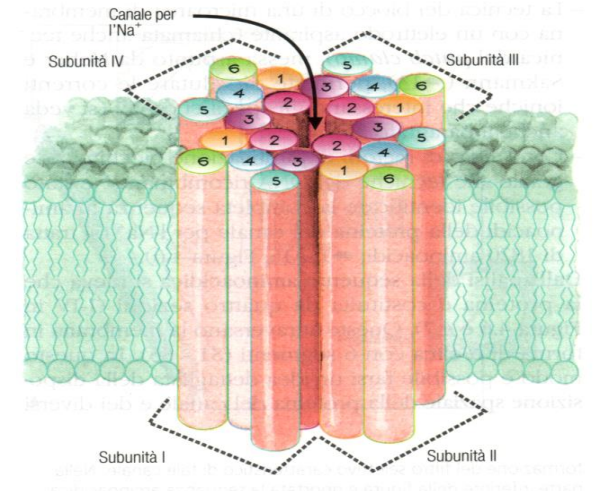
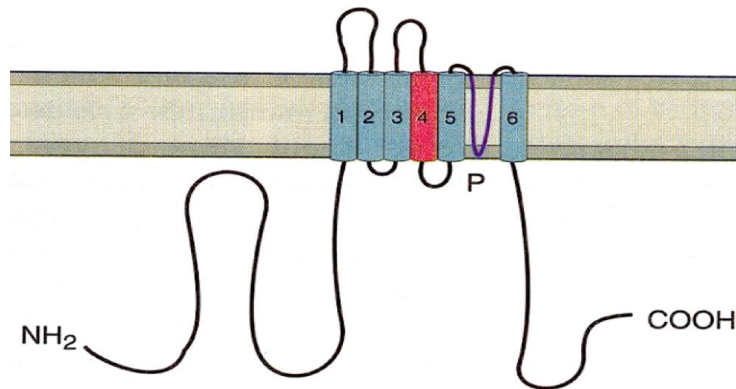
Sodium and calcium channel structure

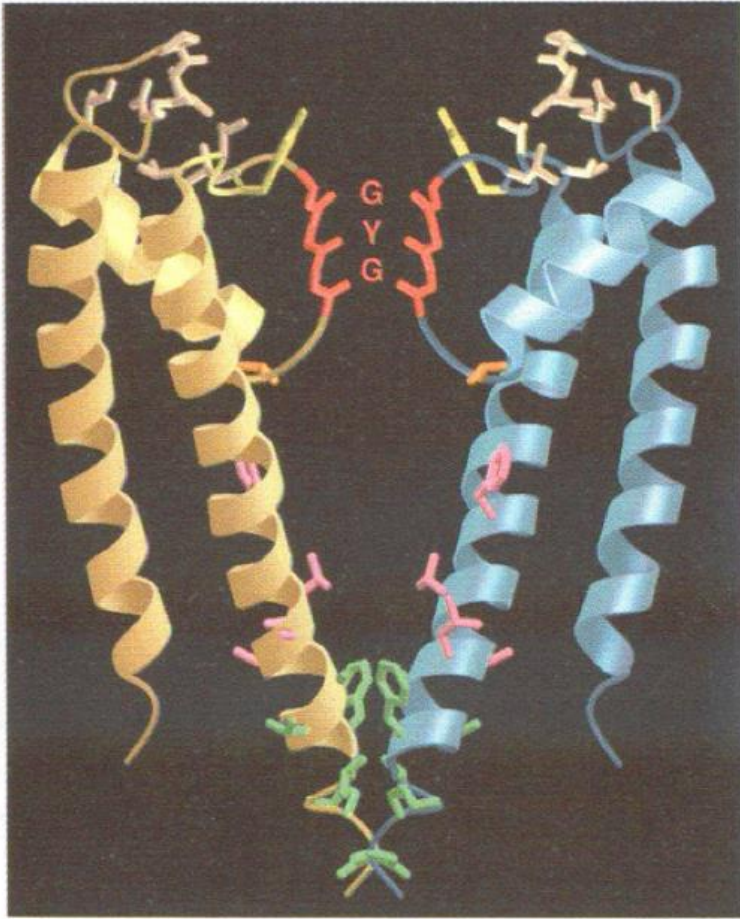
Large protein, α polypeptide chain with 4 homologous domains (I-IV), each with 6 hydrophobic segments (S1-S6), P-handle between S5-S6. S4 aa basic loads = voltage sensor 6TM



Potassium Channel Structure (Kv)

4 polypeptide chains aggregated in tetrameric structure. Each with a similar domain to sodium and potassium 6TM (S1-S6)



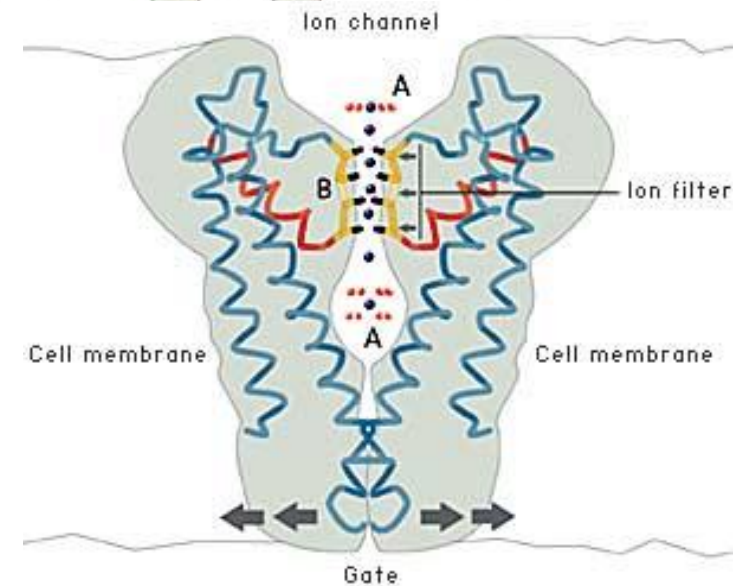
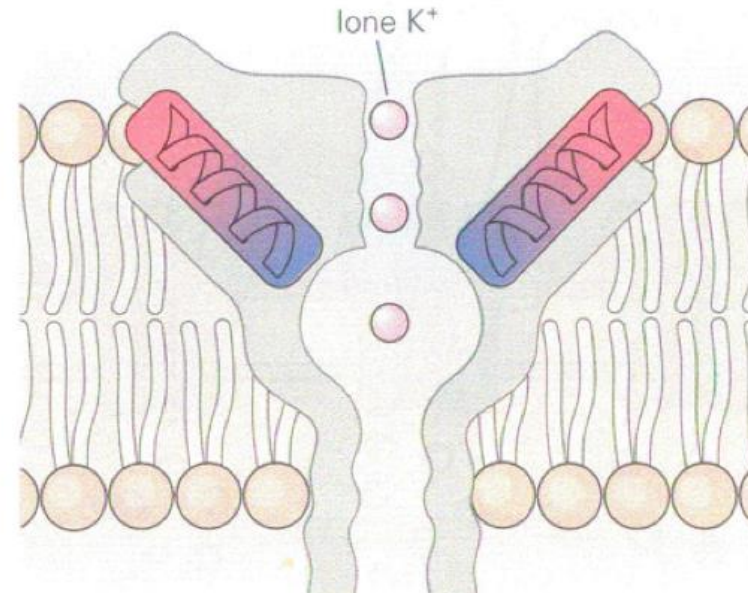


(4 subunits with two transmembrane segments connected by loop P, pore channel)

D

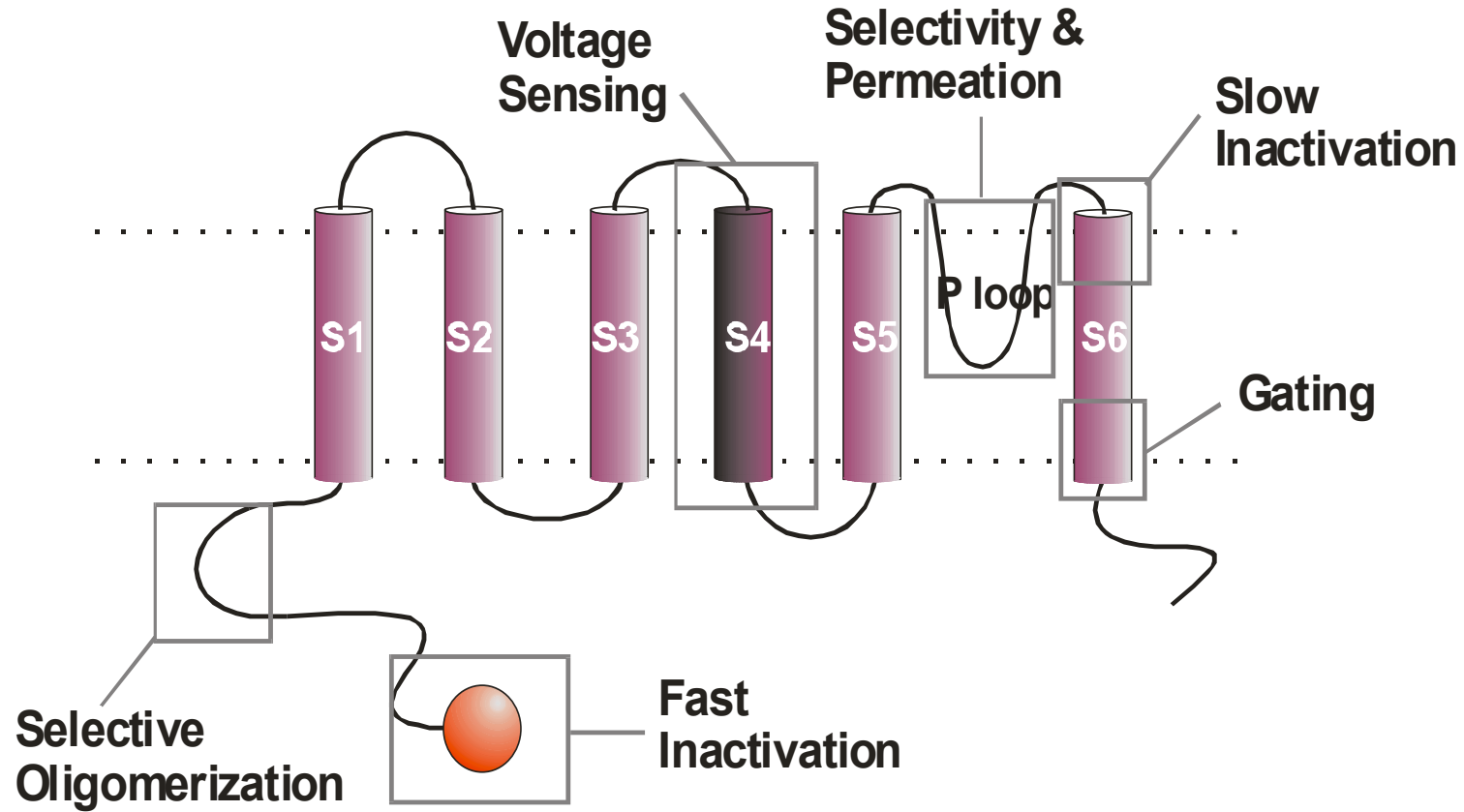
Bacterial K⁺ channel KcsA

MacKinnon, 1998



Voltage-gated Ion Channels

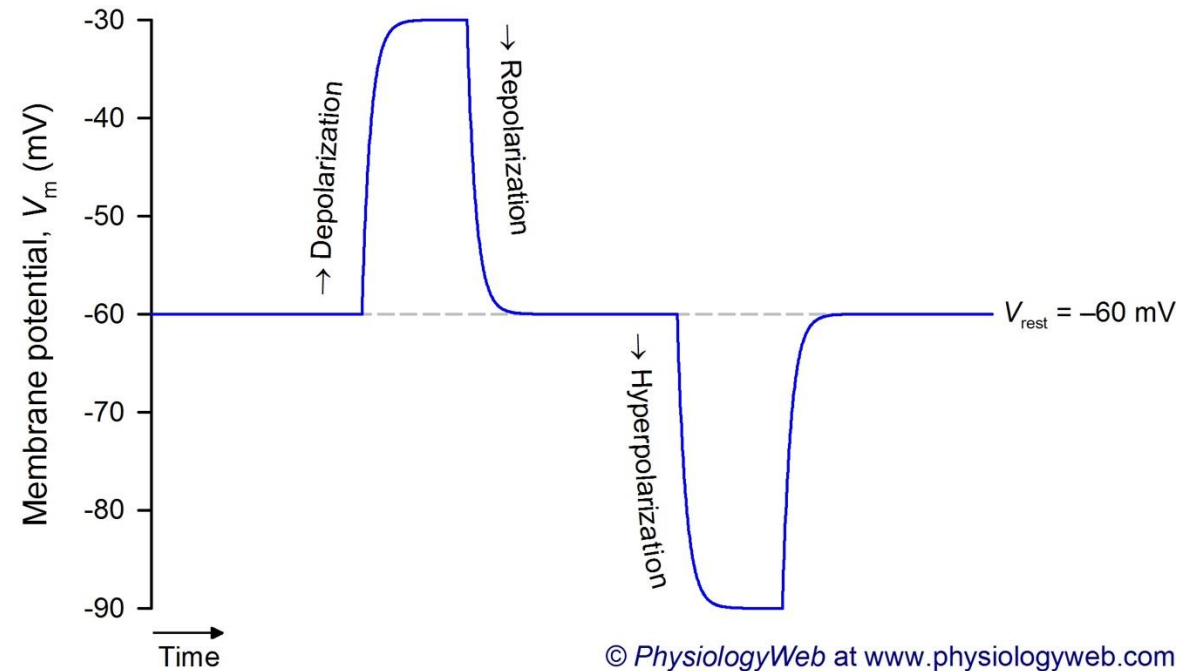
General structure



Two classes of Voltage-dependent Ion Channels

channels open by **depolarization**

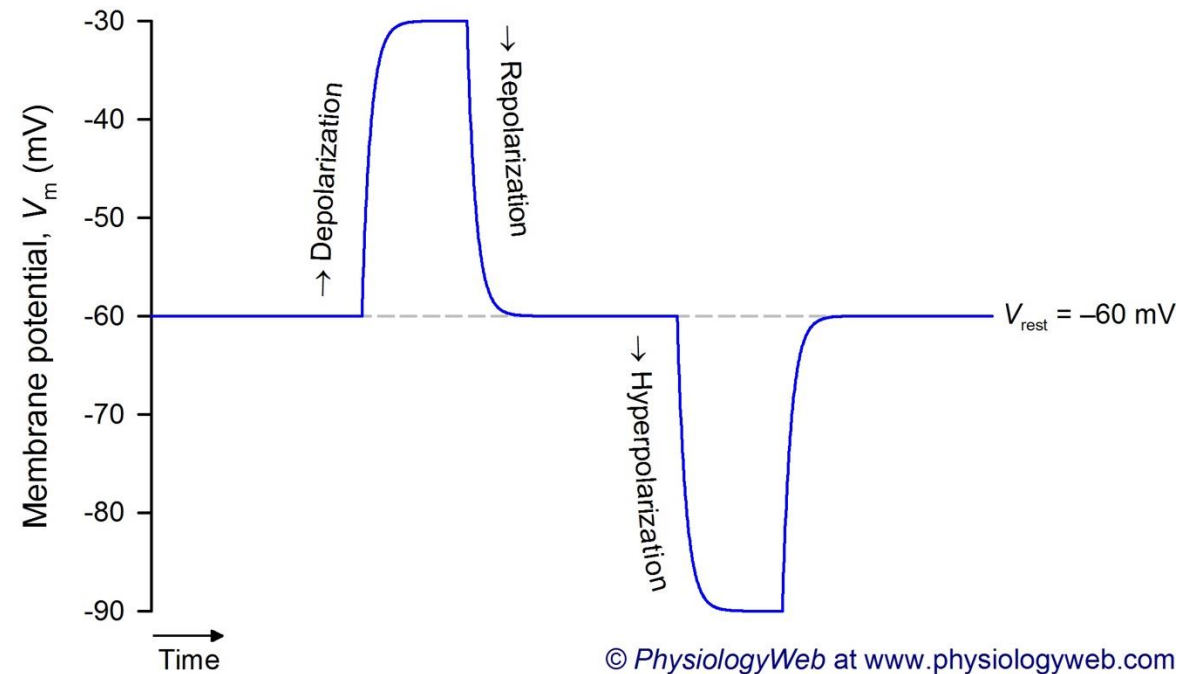
channels open by **hyperpolarization**



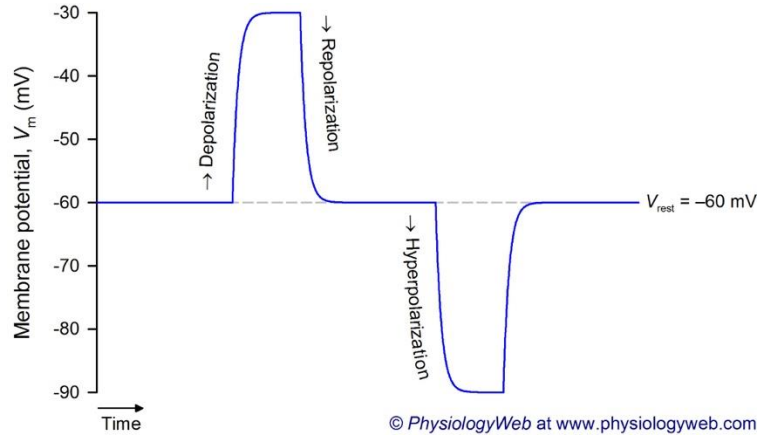
- **Polarization**: a state in which membrane is polarized at rest, negative inside and positive outside.
- **Depolarization**: the membrane potential becomes less negative than the resting potential (close to zero).
- **Hyperpolarization**: the membrane potential is more negative than the resting level.

Voltage-dependent Ion Channels

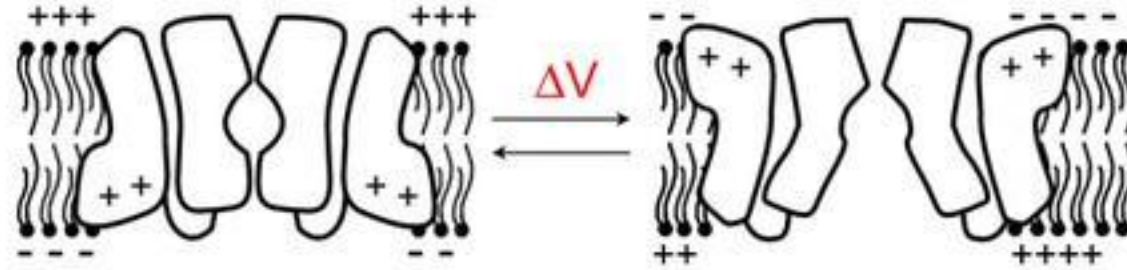
channels open by **depolarization**



Channels Open by Depolarization

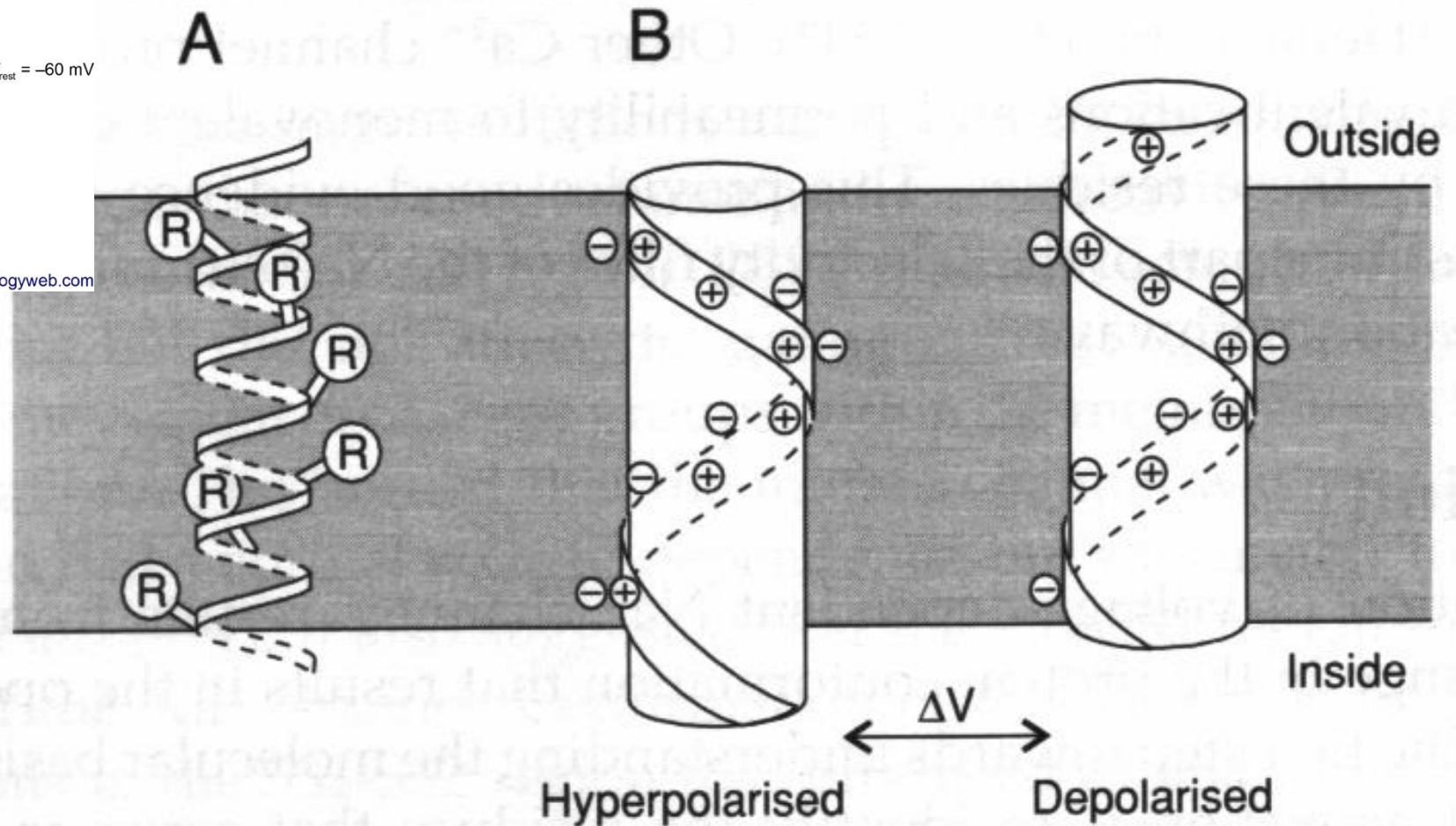
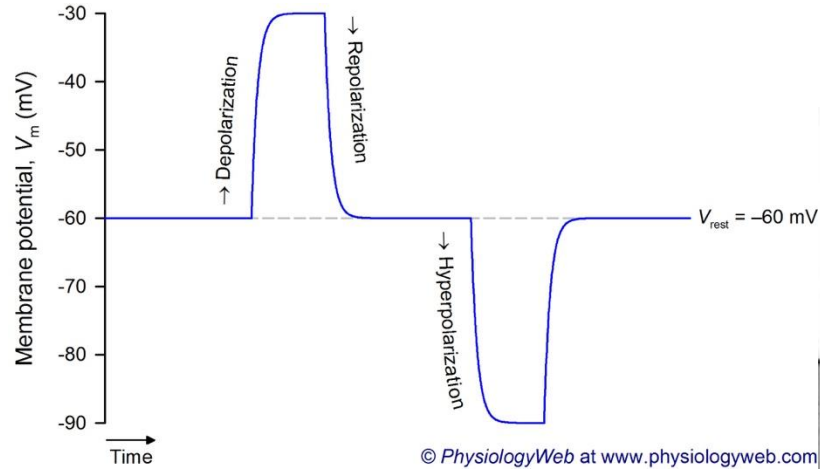


Voltage-dependent channel gating

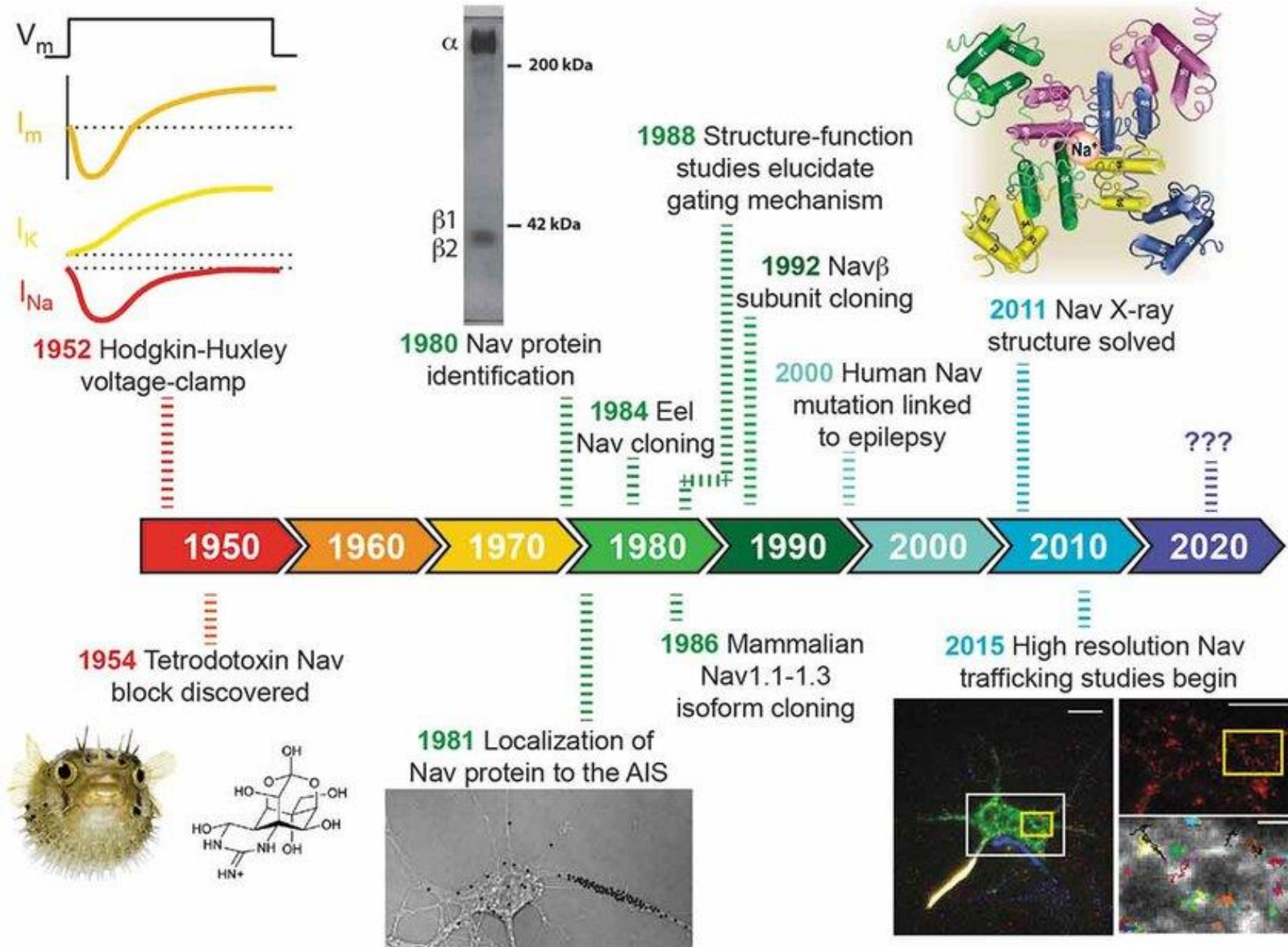


They are a family of selectively permeable channels for Na^+ , K^+ , Ca^{2+}

Channels Open by Depolarization - Gating



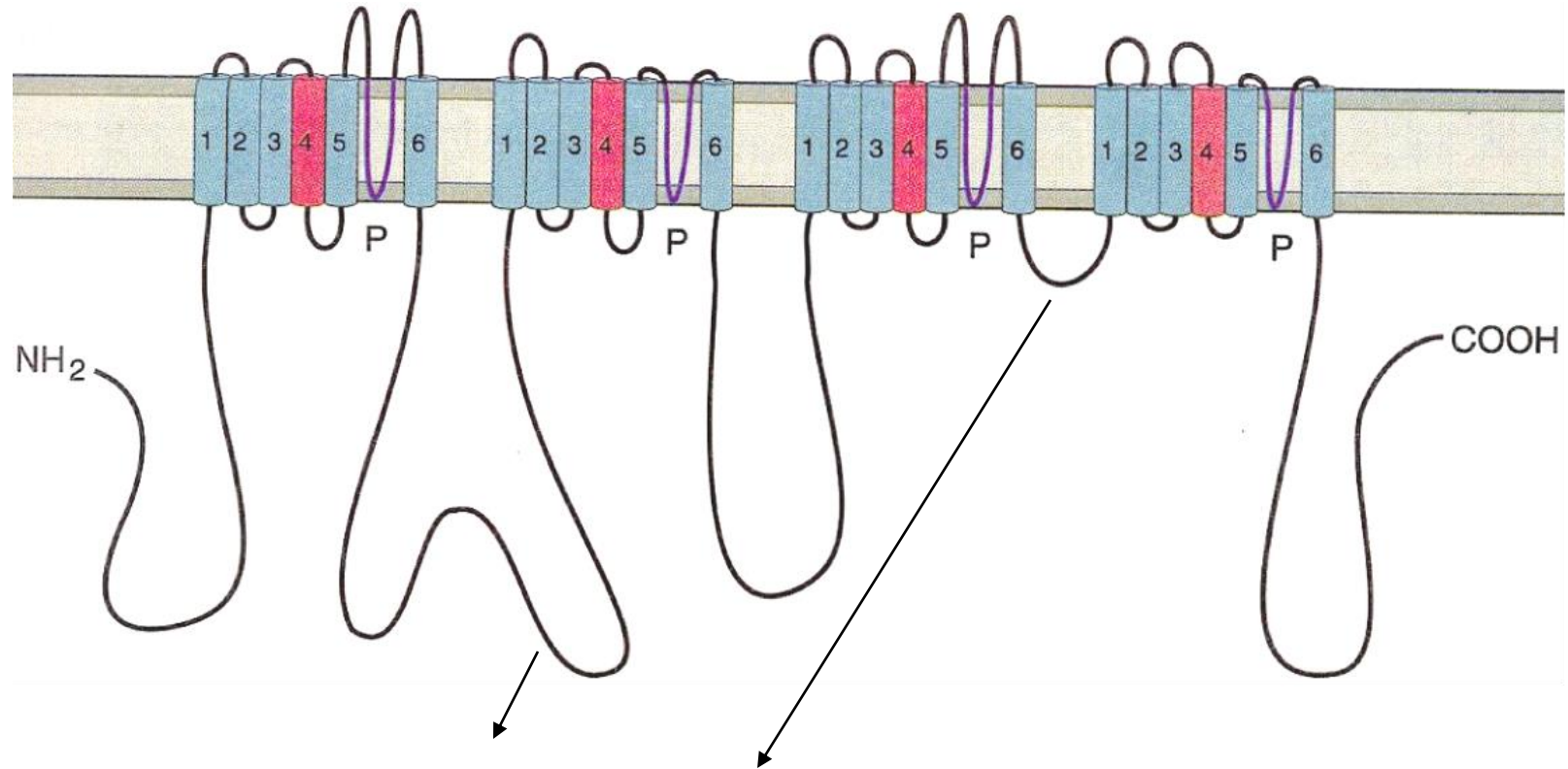
Voltage gated Na⁺ channels



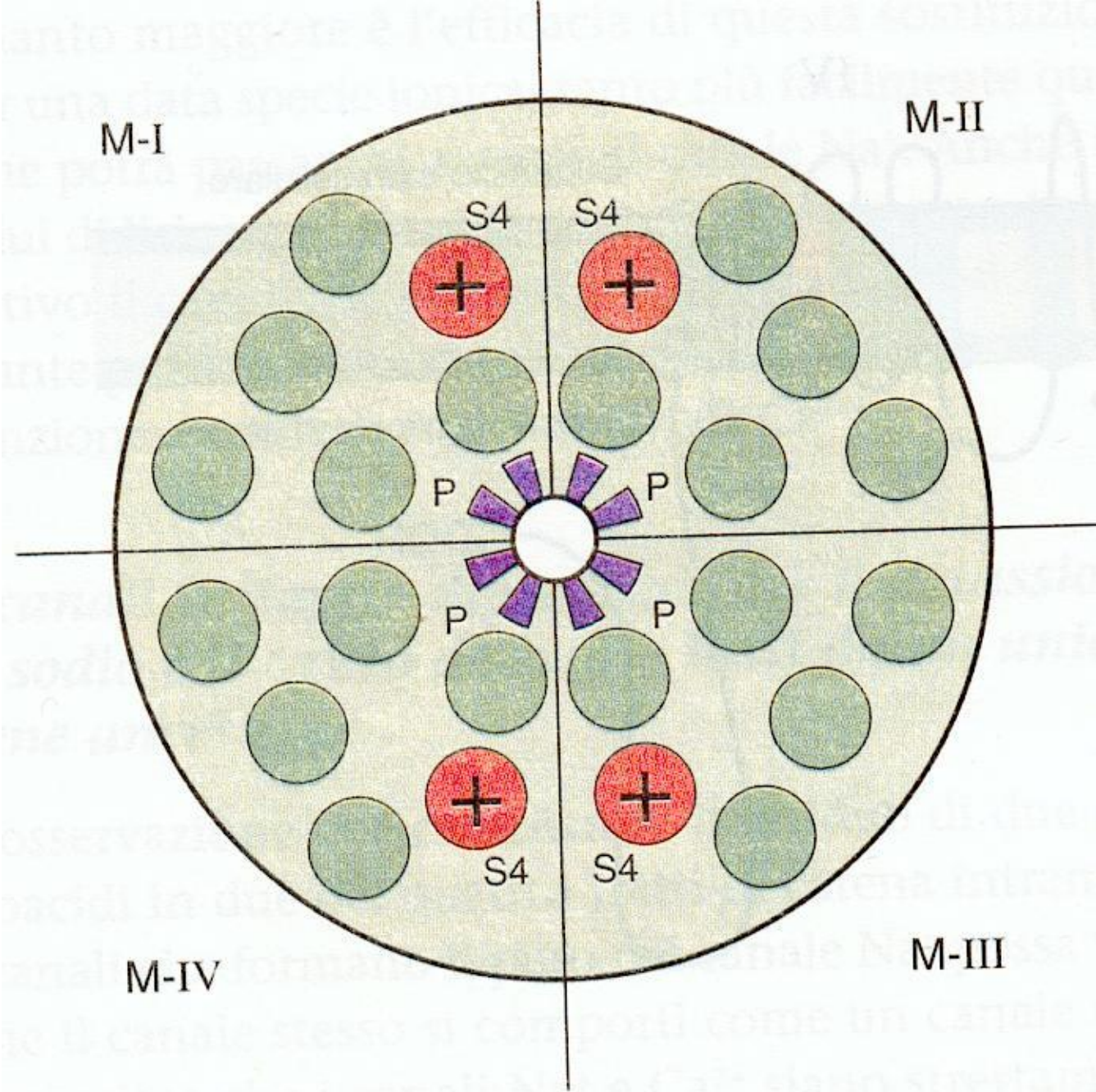
Voltage gated Na⁺ channels

alpha subunit

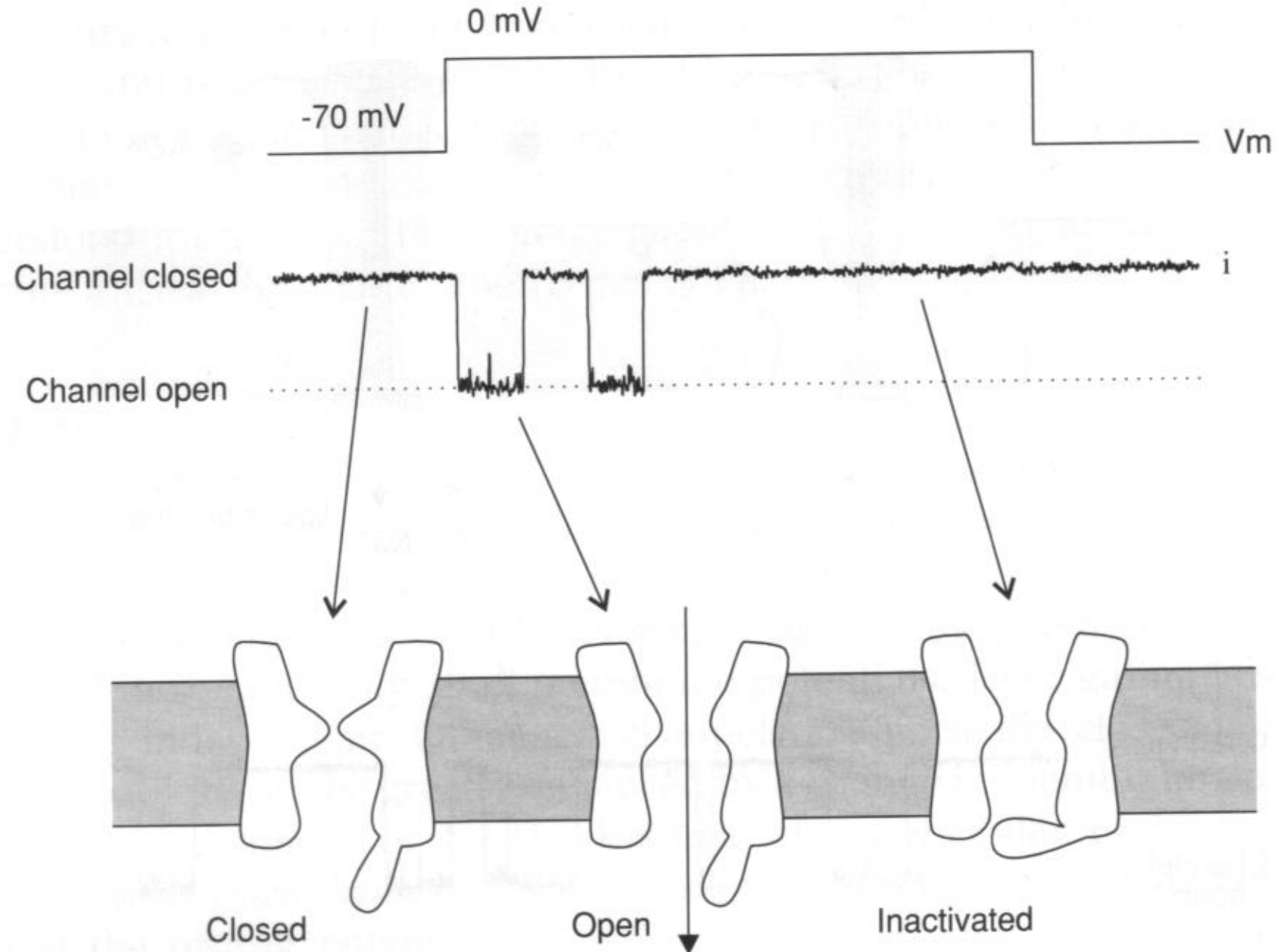
(+ 1 o 2 beta accessorial subunits)



PKA and PKC phosphorylation sites



Channel open = current flow

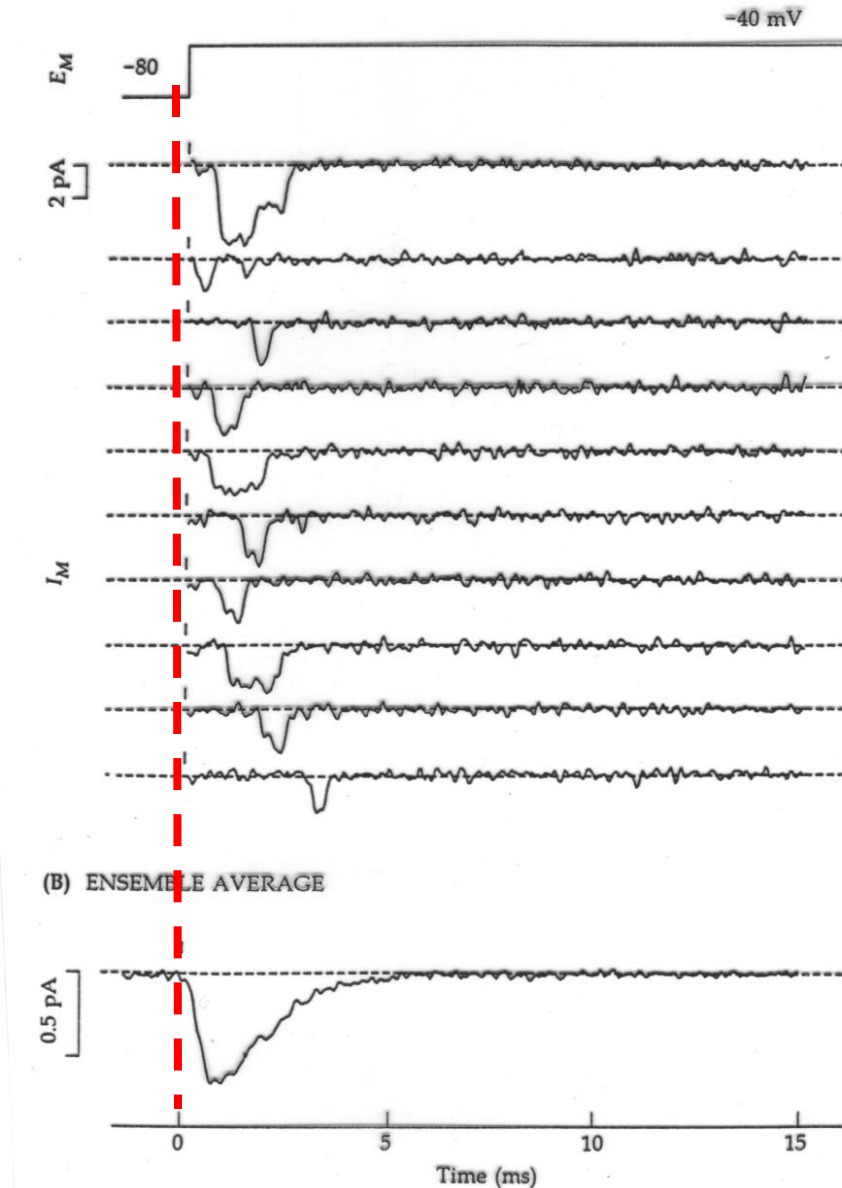


Voltage gated Na^+ channels

Channels selectively permeable to

Na^+

- Rapid activation
- Rapid inactivation

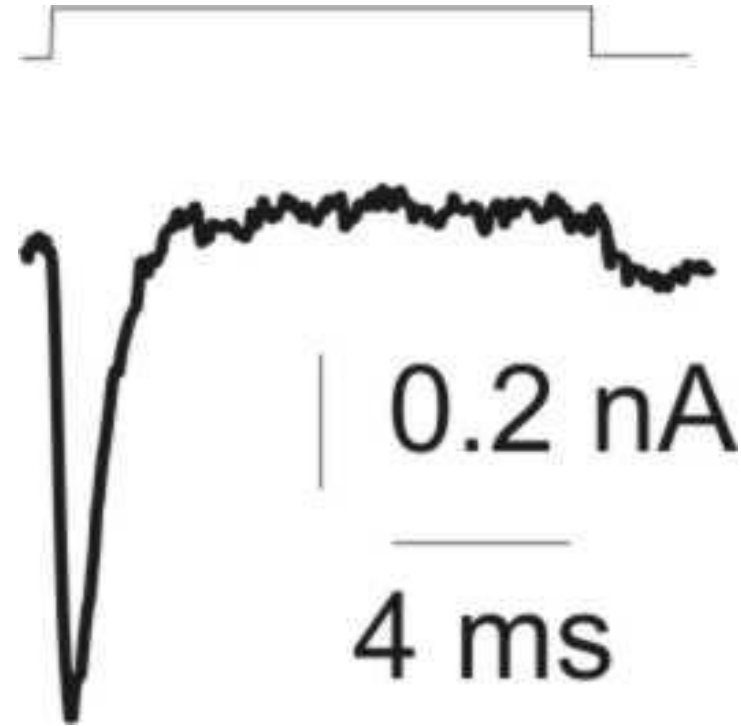


Voltage gated Na^+ channels

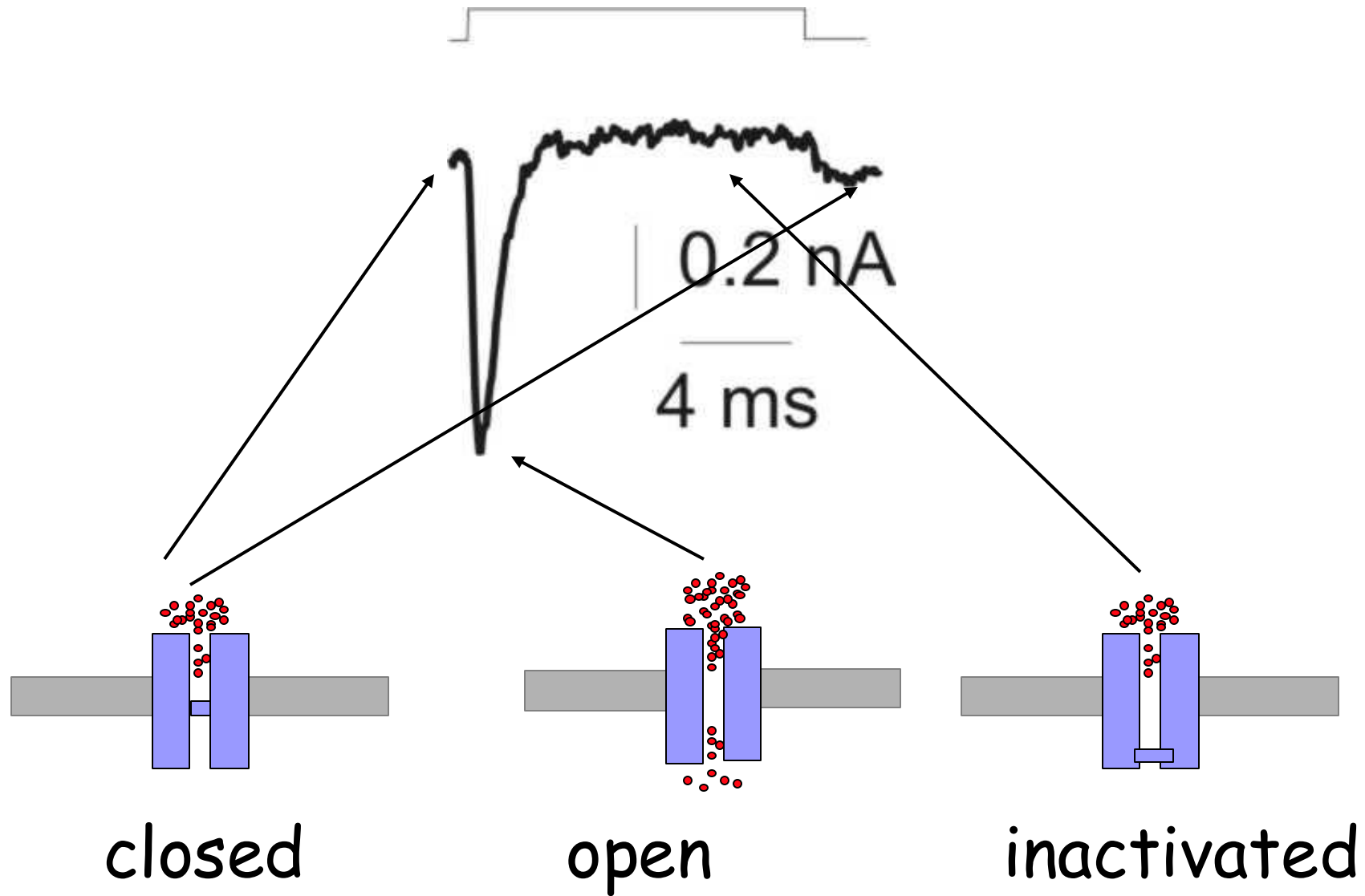
Channels selectively permeable to

Na^+

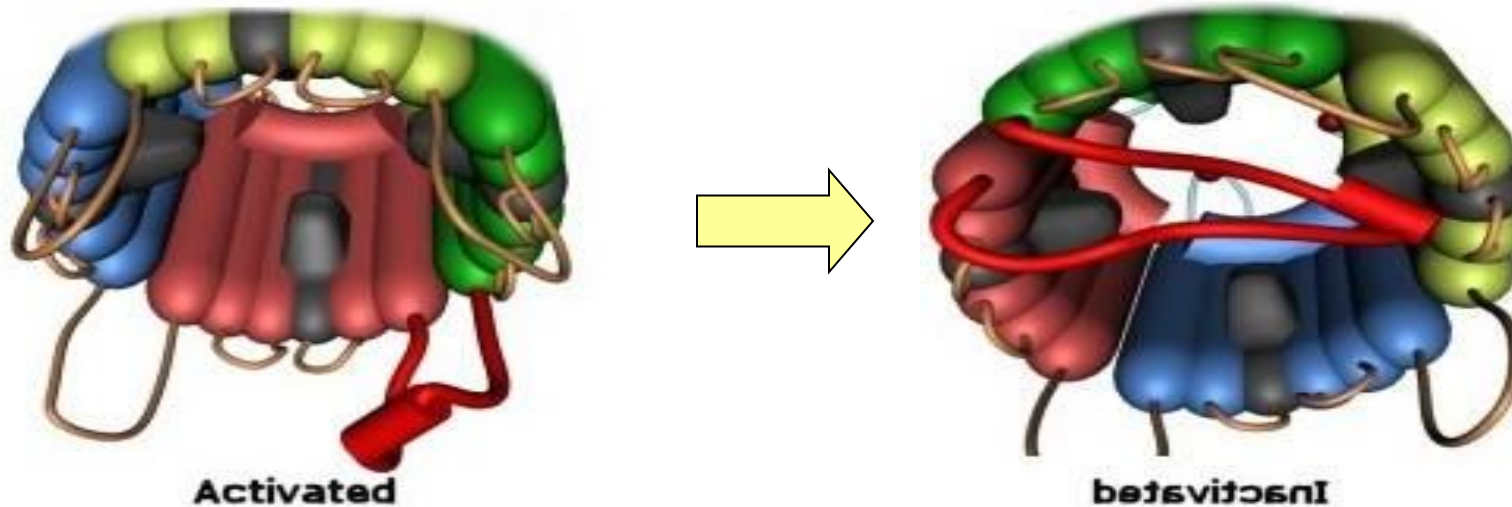
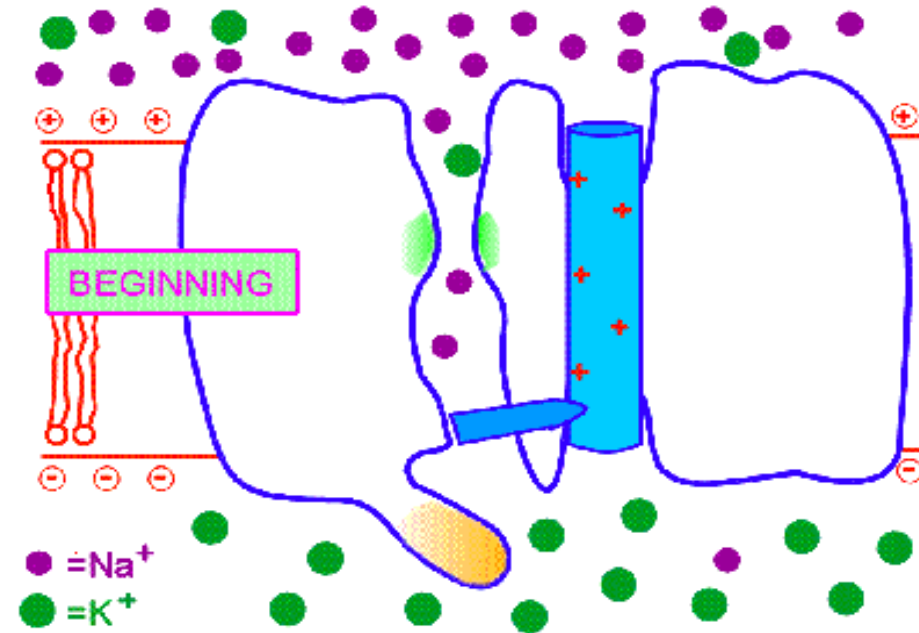
- Rapid activation
- Rapid inactivation



Voltage gated Na^+ channels - Inactivation

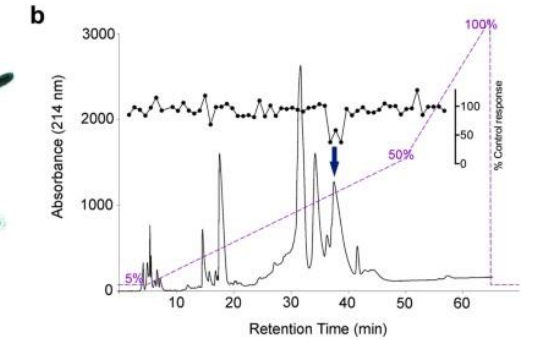
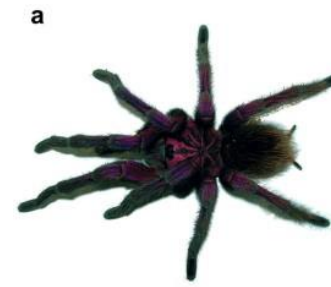
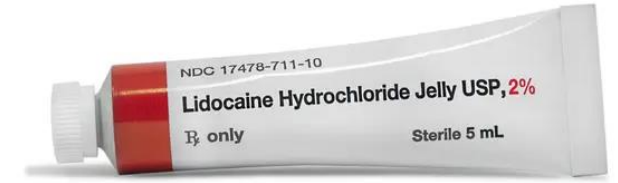


Voltage gated Na^+ channels - Inactivation



Voltage gated Na⁺ channels - Blockers

- Tetrodotoxin (TTX)
- Amioderone
- Lidocaine
- Procainamide
- Mexilitine
- Ketamine
- Many, many others

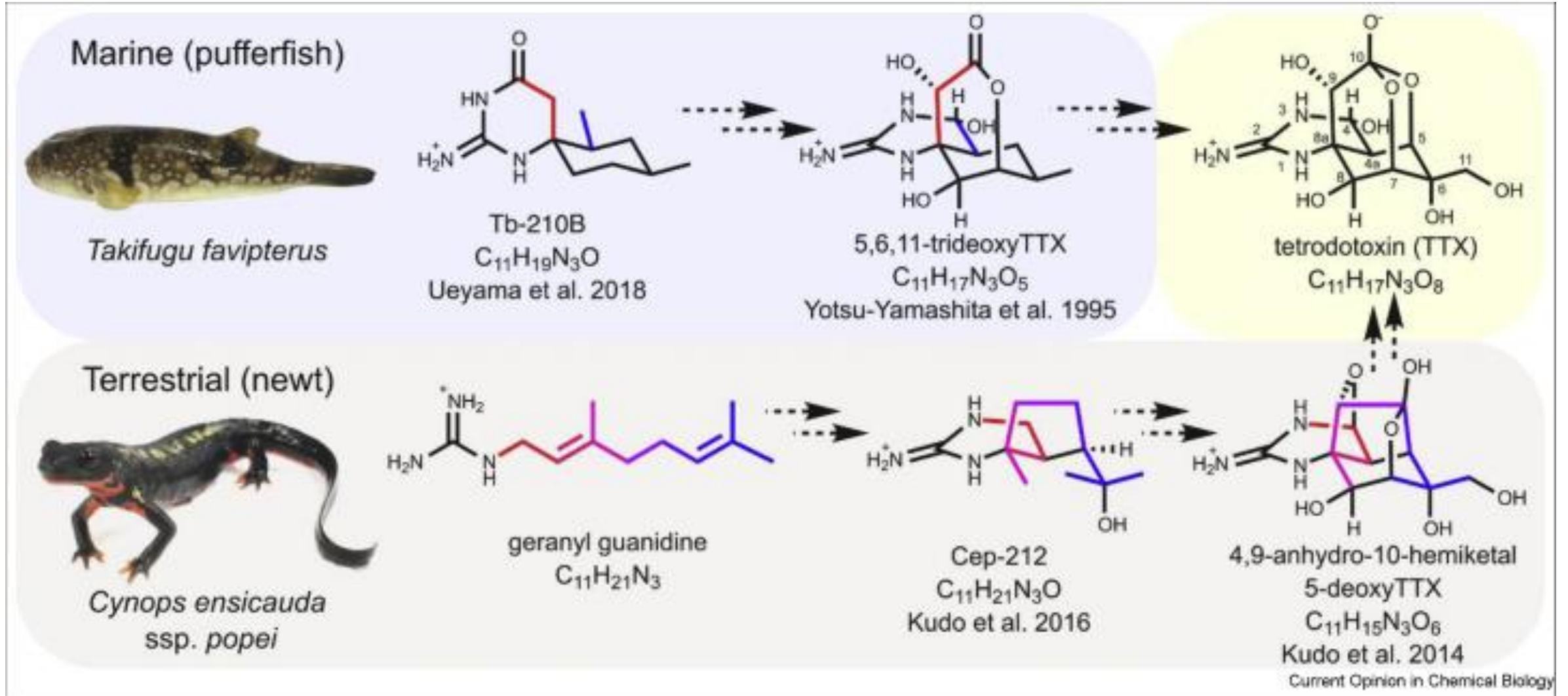


Toxin name	Sequence	Observed Mass (Da)	Calculated Mass (Da)
μ -TRTX-Pn3a	DCRYMFGDCEKDEDCCKHLGCKRKMKYCAWDFTF	4268.5	4268.8
μ -TRTX-Pn3b	GCRYMFGDCEKDEDCCKHLGCKRKMKYCAWDFTF	4210.5	4210.8



Voltage gated Na⁺ channels - Blockers

- Tetrodotoxins (TTX)

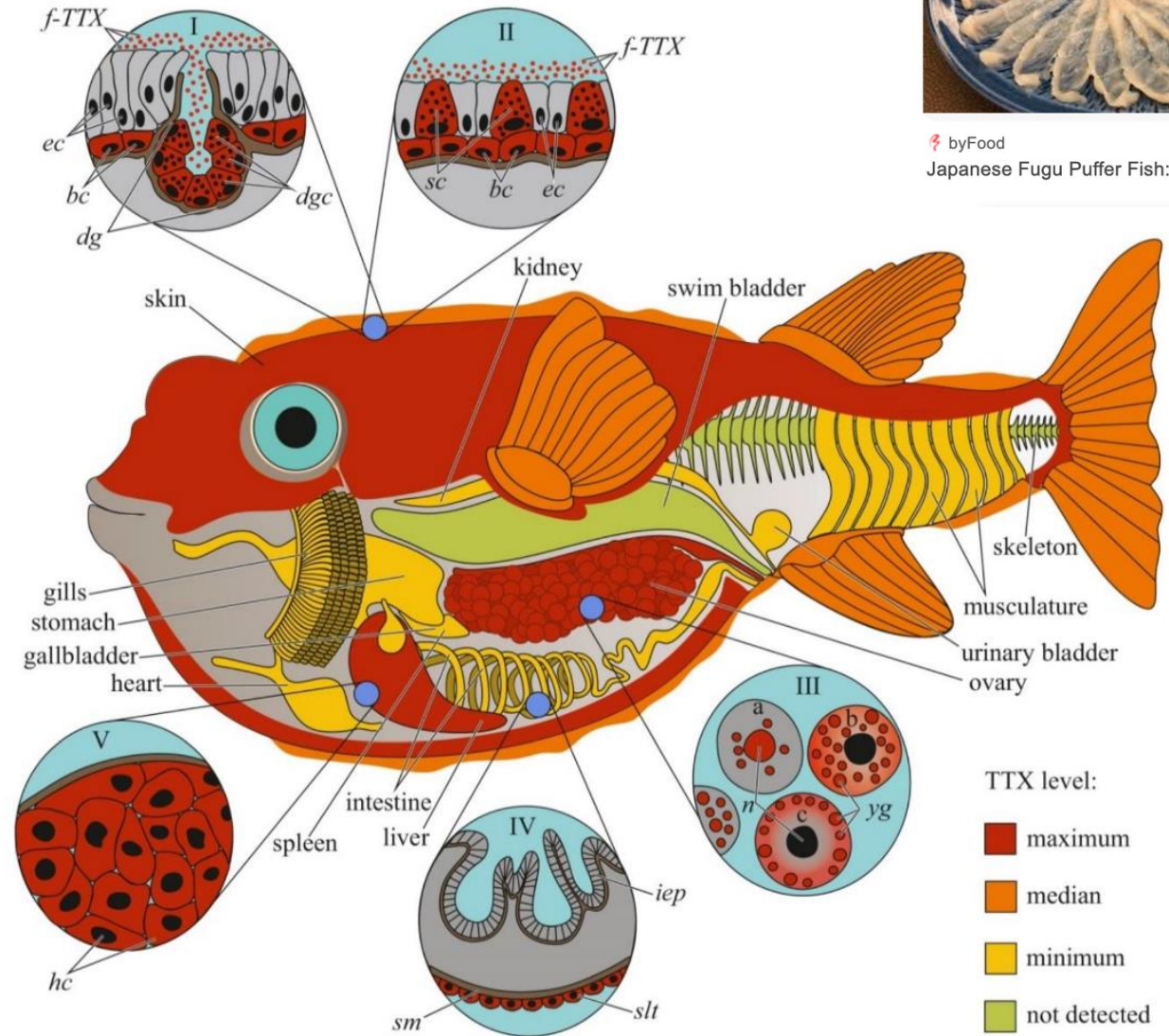
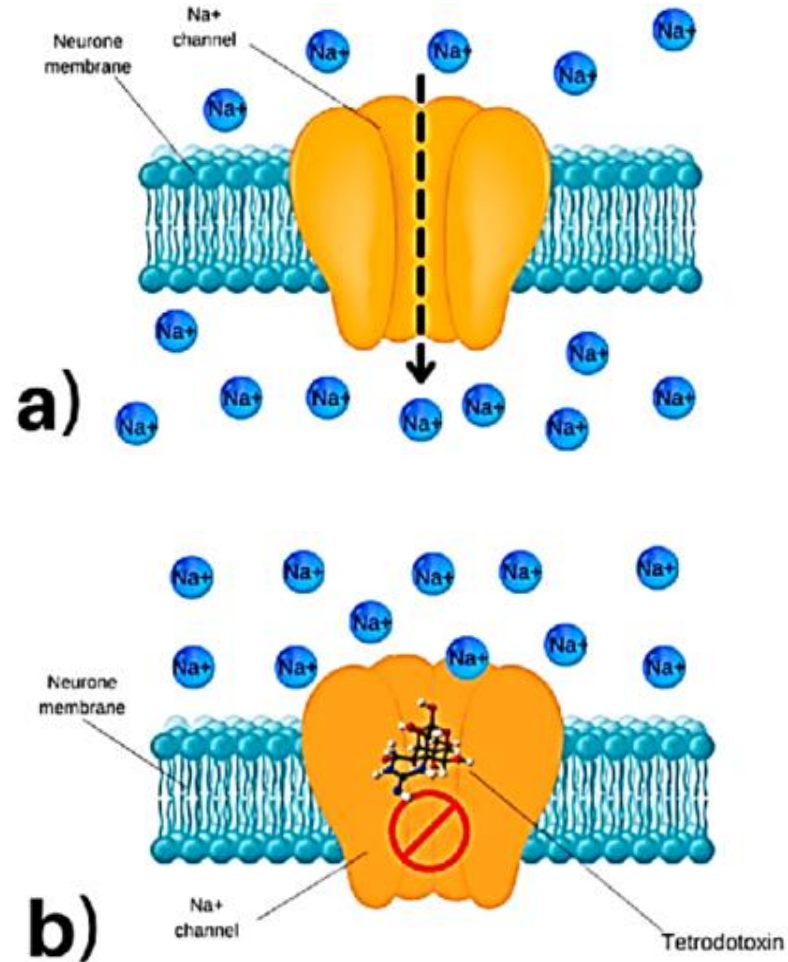


Voltage gated Na^+ channels - Blockers

- Tetrodotoxin (TTX)



byFood
Japanese Fugu Puffer Fish: Dangers, Prep...



Voltage gated Na^+ channels - Functions

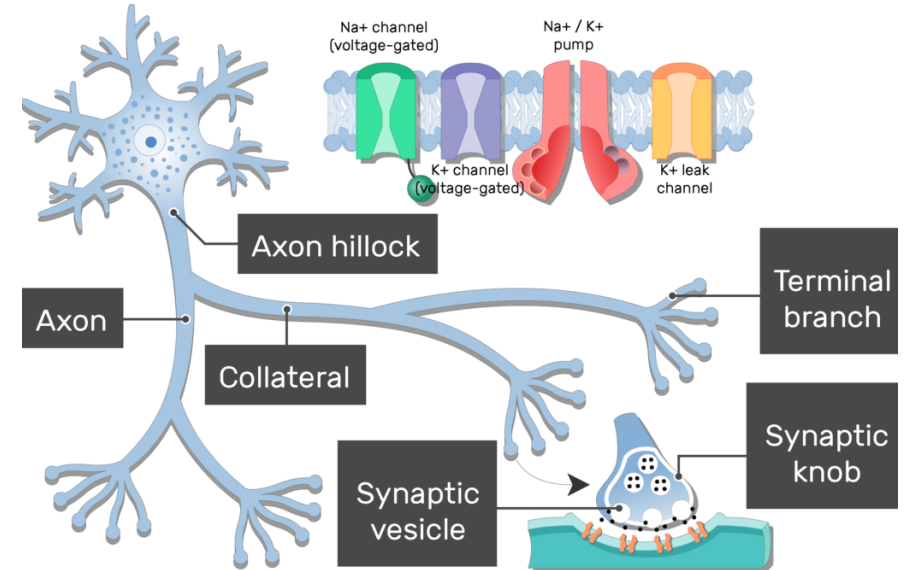
Opening of voltage gated Na^+ channels @ **depolarization**
($E_{\text{Na}} = +55 \text{ mV}$)

Na^+ channels in:

- skeletal muscle
- working myocardium
- Axons

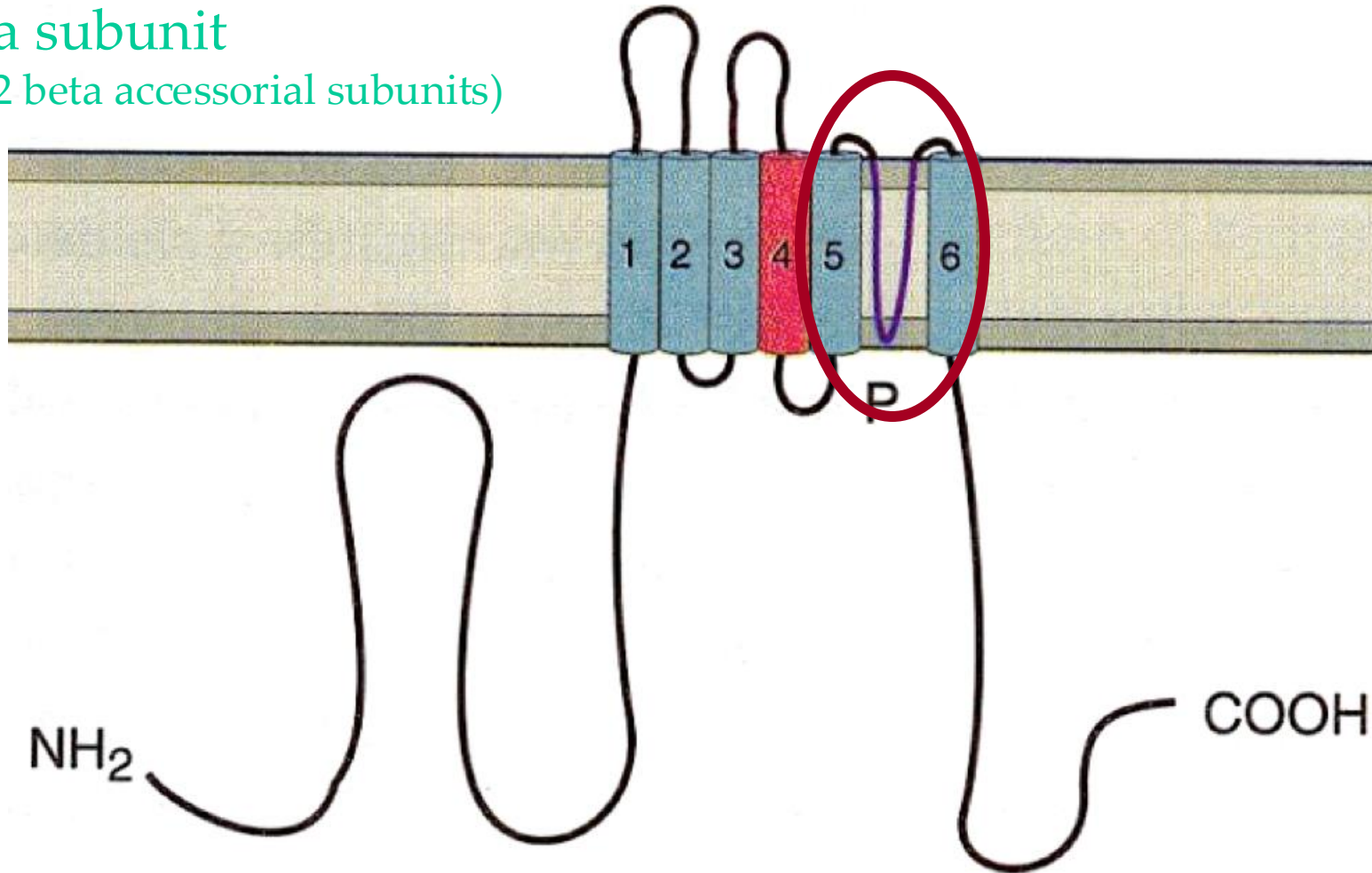
Important in

- 1) Generation and propagation of action potential
- 2) Depolarization post action potential
- 3) Discharge frequency regulation



Voltage gated K⁺ channels

alpha subunit
(+ 1 o 2 beta accessorial subunits)



Voltage gated K⁺ channels

Selectively permeable to K⁺

There are various channel families for K⁺
they are found in all cell types

Channel opening K⁺ @
repolarization ($E_K = -90 \text{ mV}$)

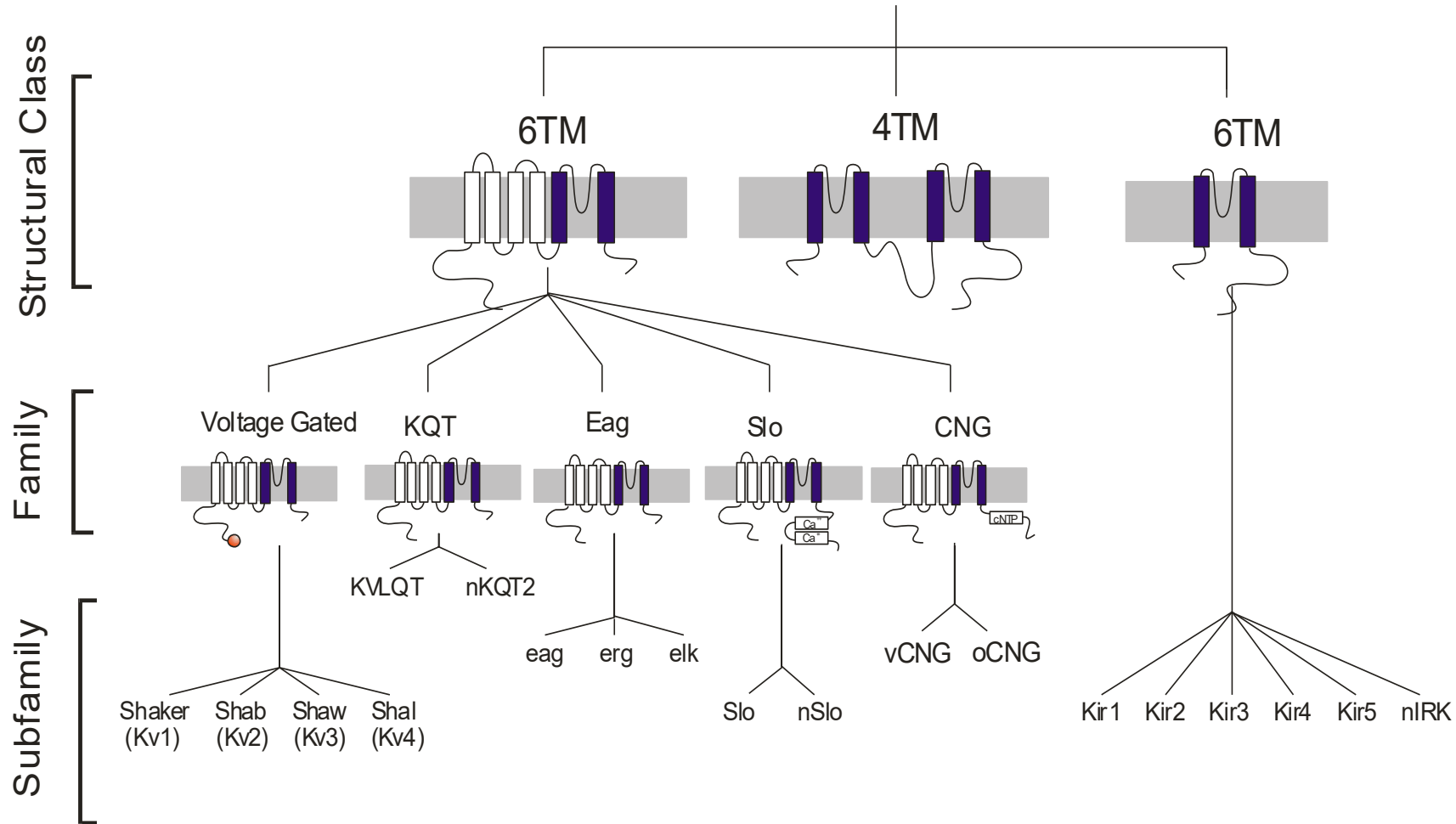
Important in excitable cells for:

- 1) They contribute to the resting potential
- 2) They support the repolarizing phase of the action potential
- 3) Inhibitively contribute to synaptic integration

Voltage gated K⁺ channels

+

more than 100 genes!!!

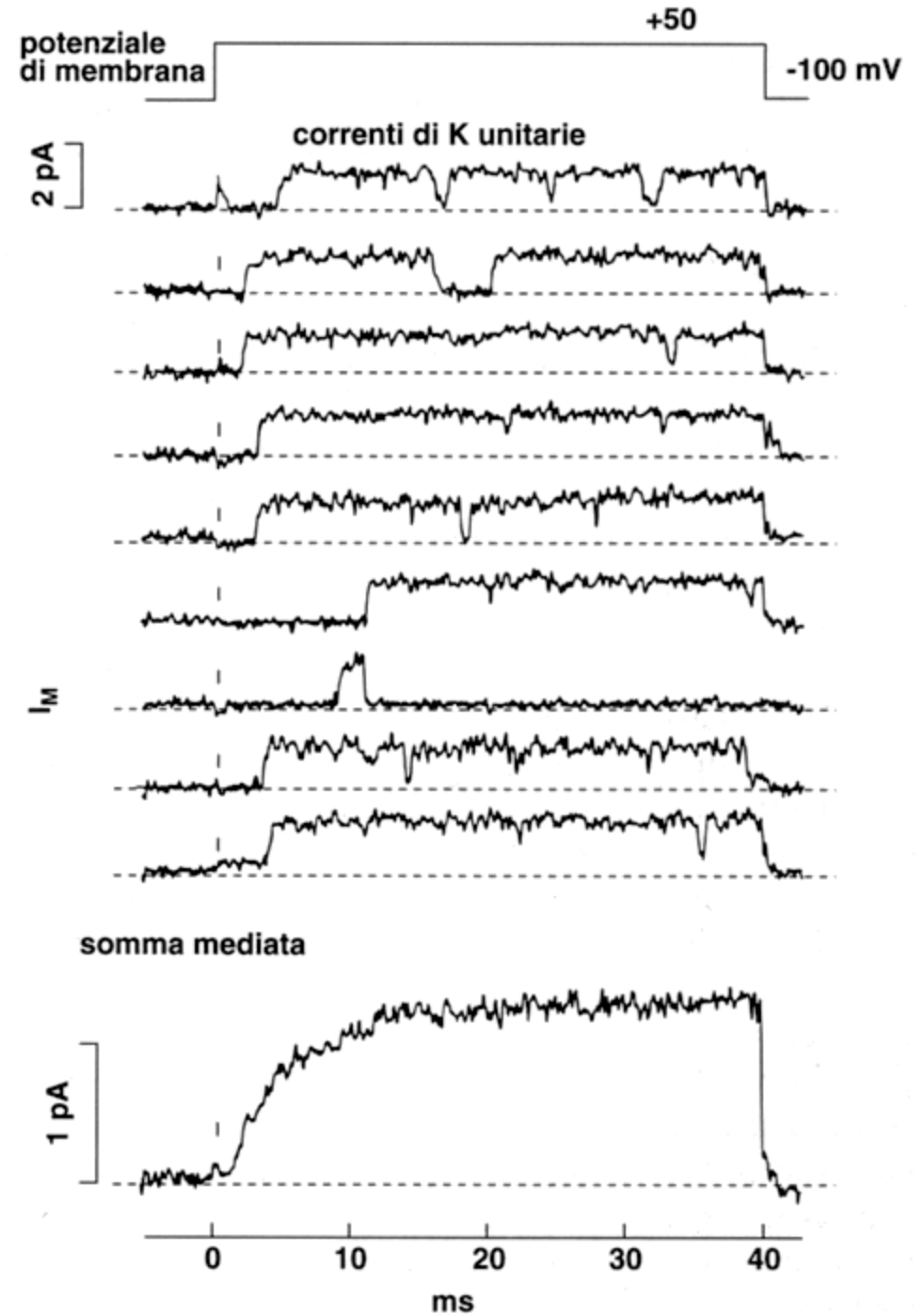


Voltage gated K⁺ channels

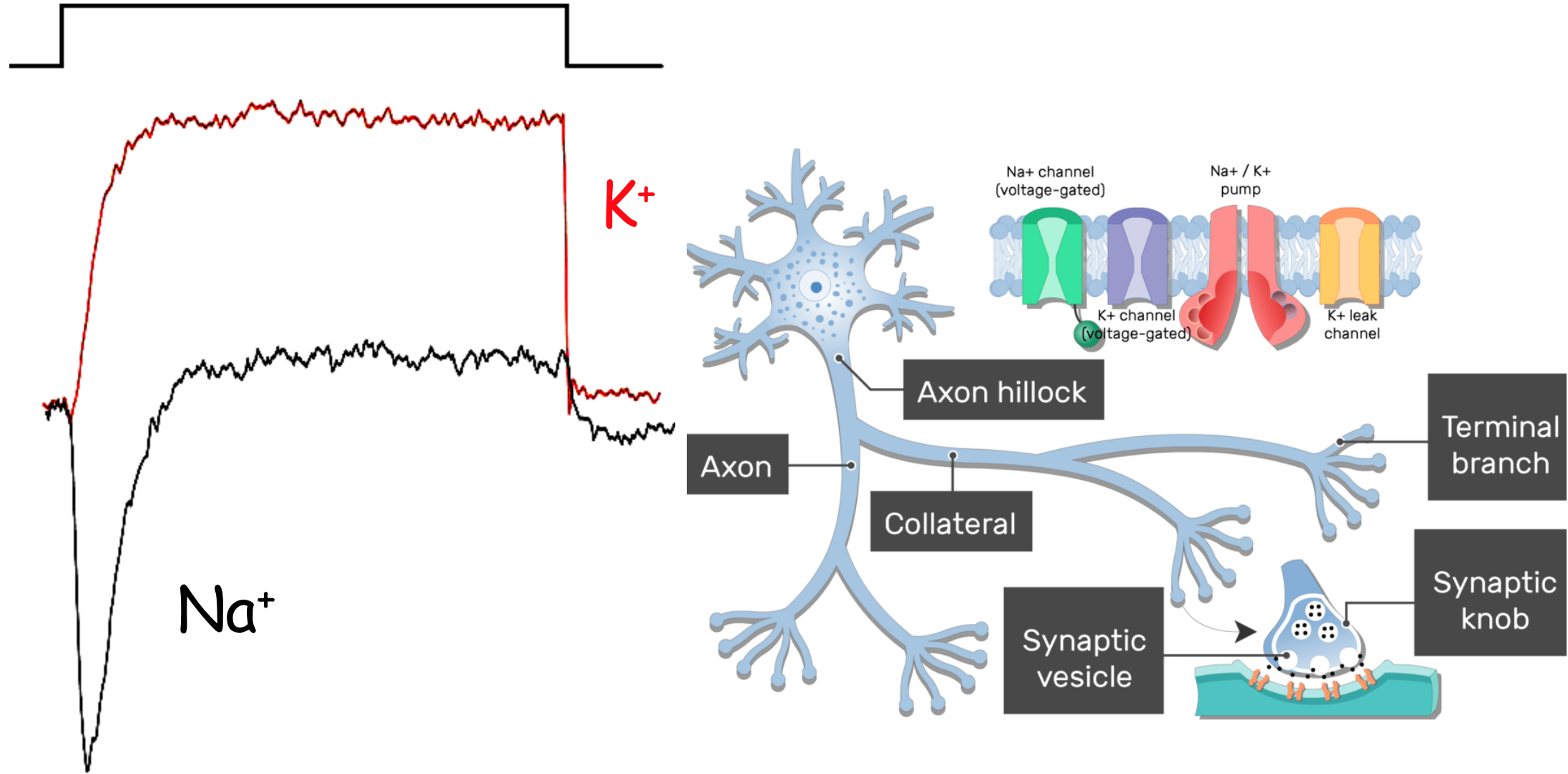
K⁺ Channels in the axons

Channels selectively permeable to
K⁺

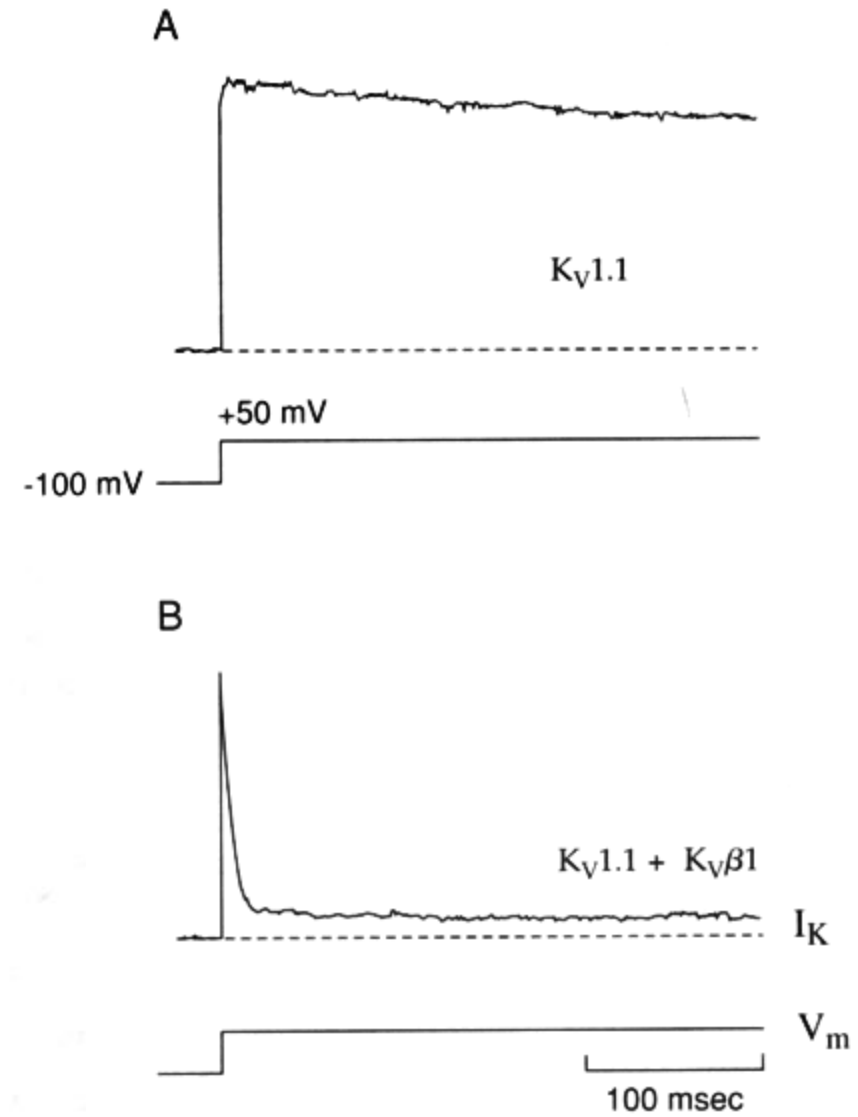
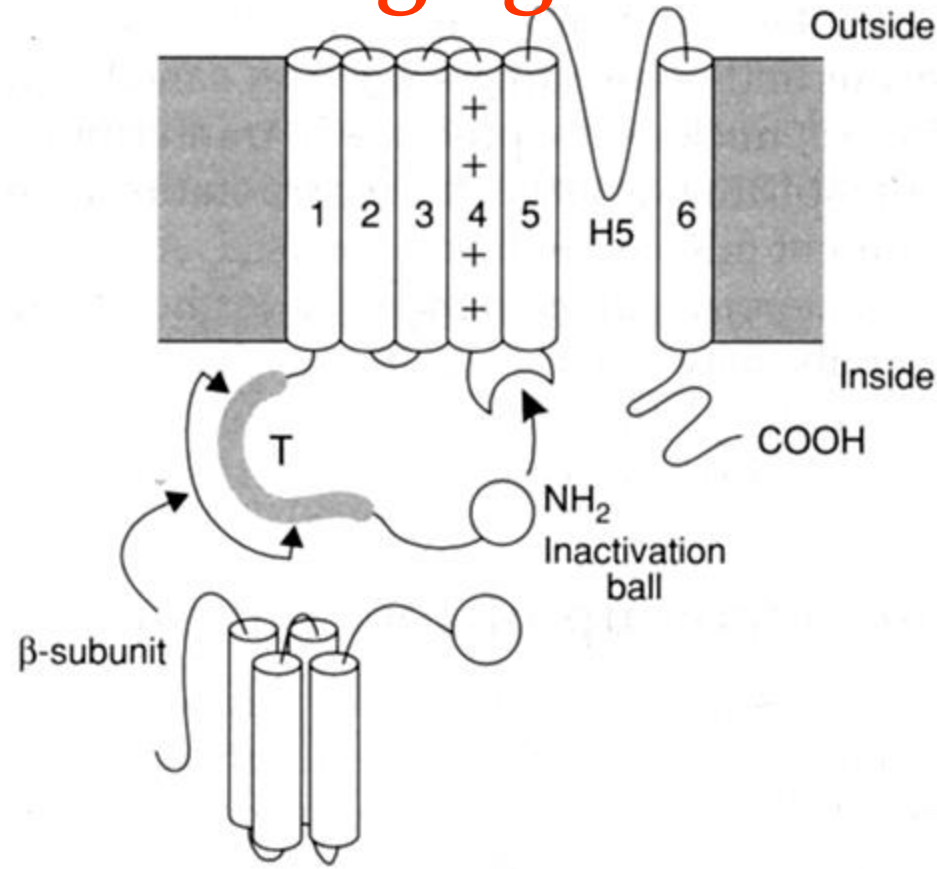
- Slow activation
- No inactivation



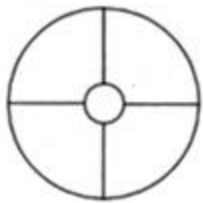
Voltage gated K^+ channels vs Voltage gated Na^+ channels



Different Voltage gated K⁺ channels



B



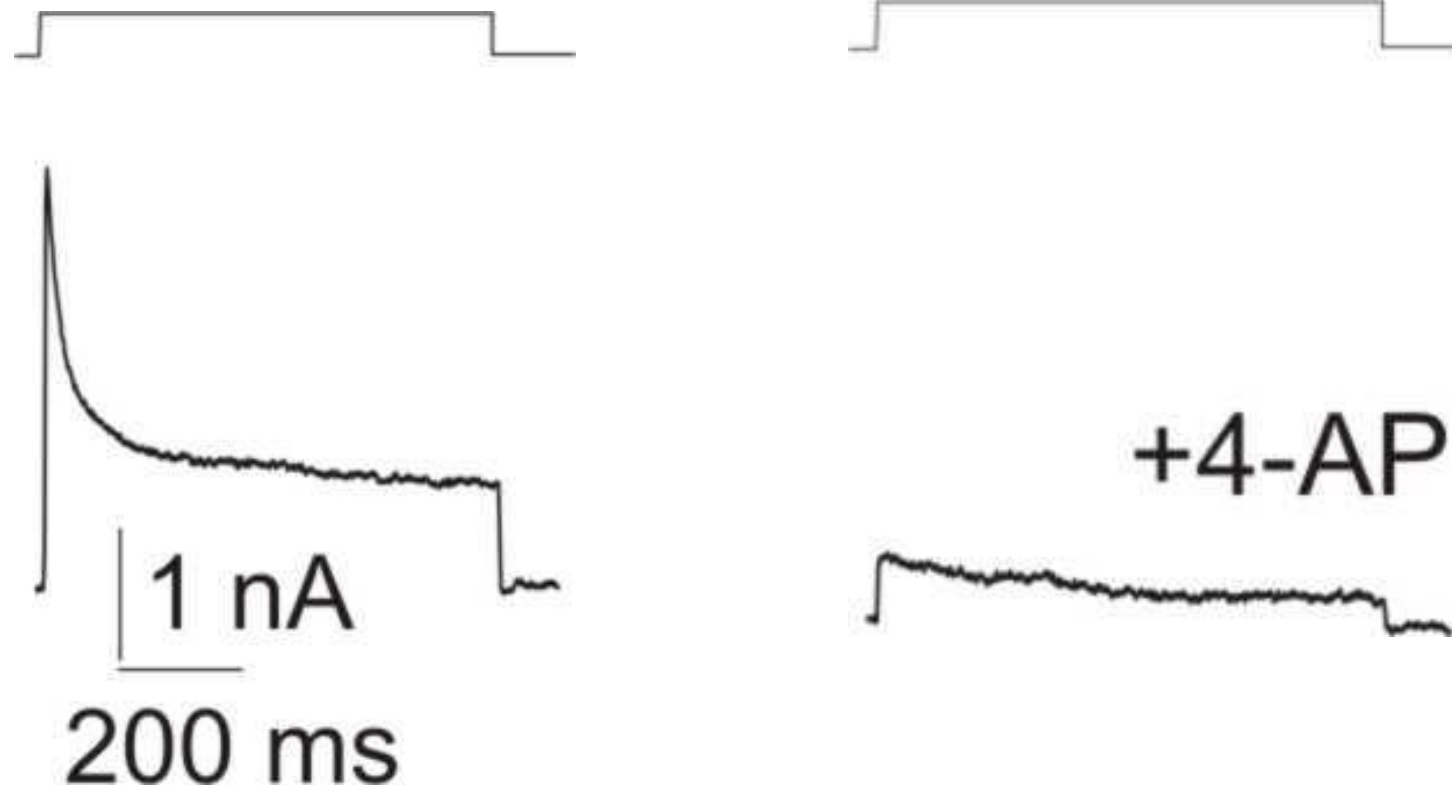
Homomeric



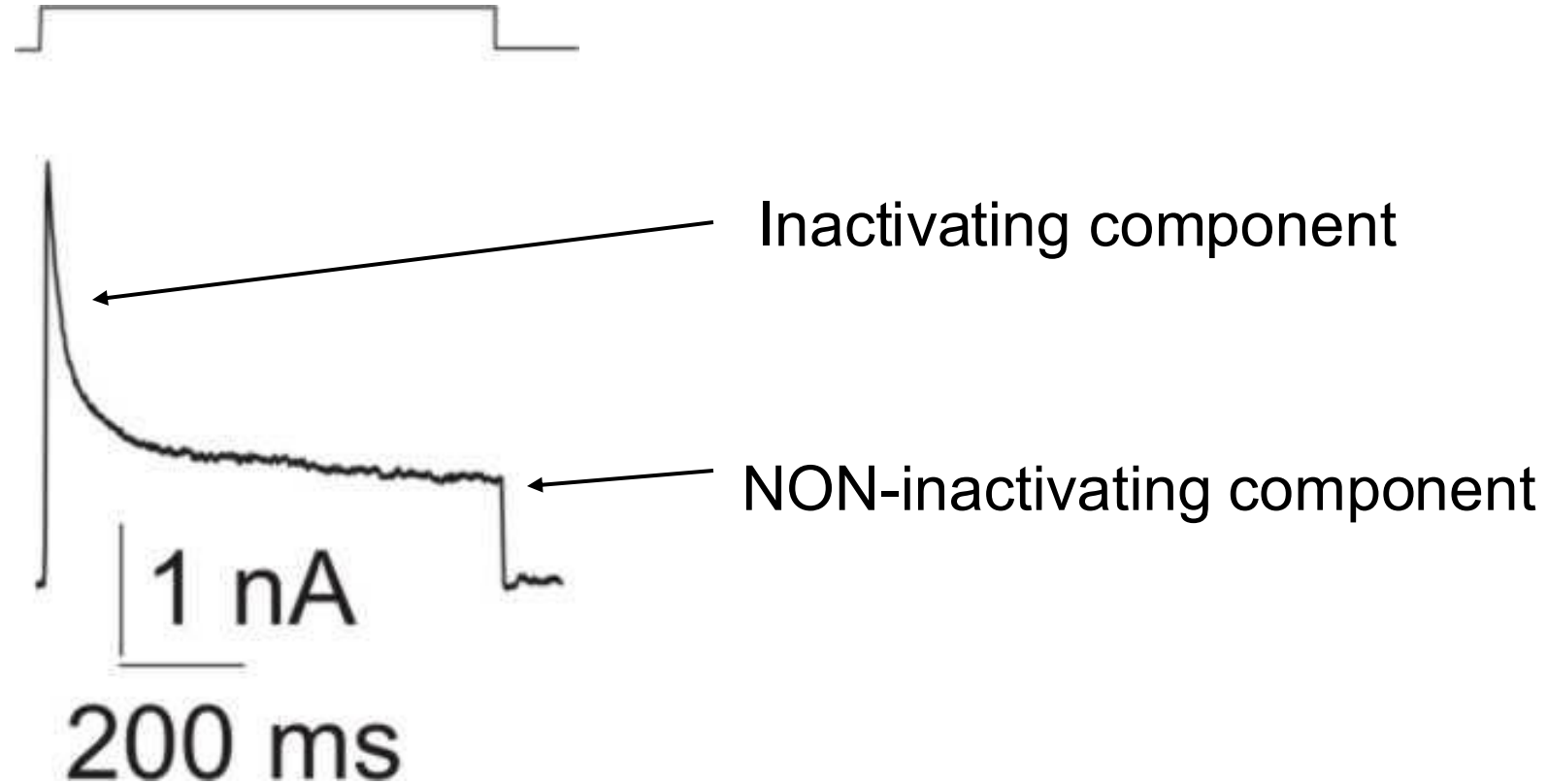
Heteromeric

Different Pharmacology of Voltage gated K⁺ channels

Many types of K⁺ channels are blocked by 4 aminopyridine

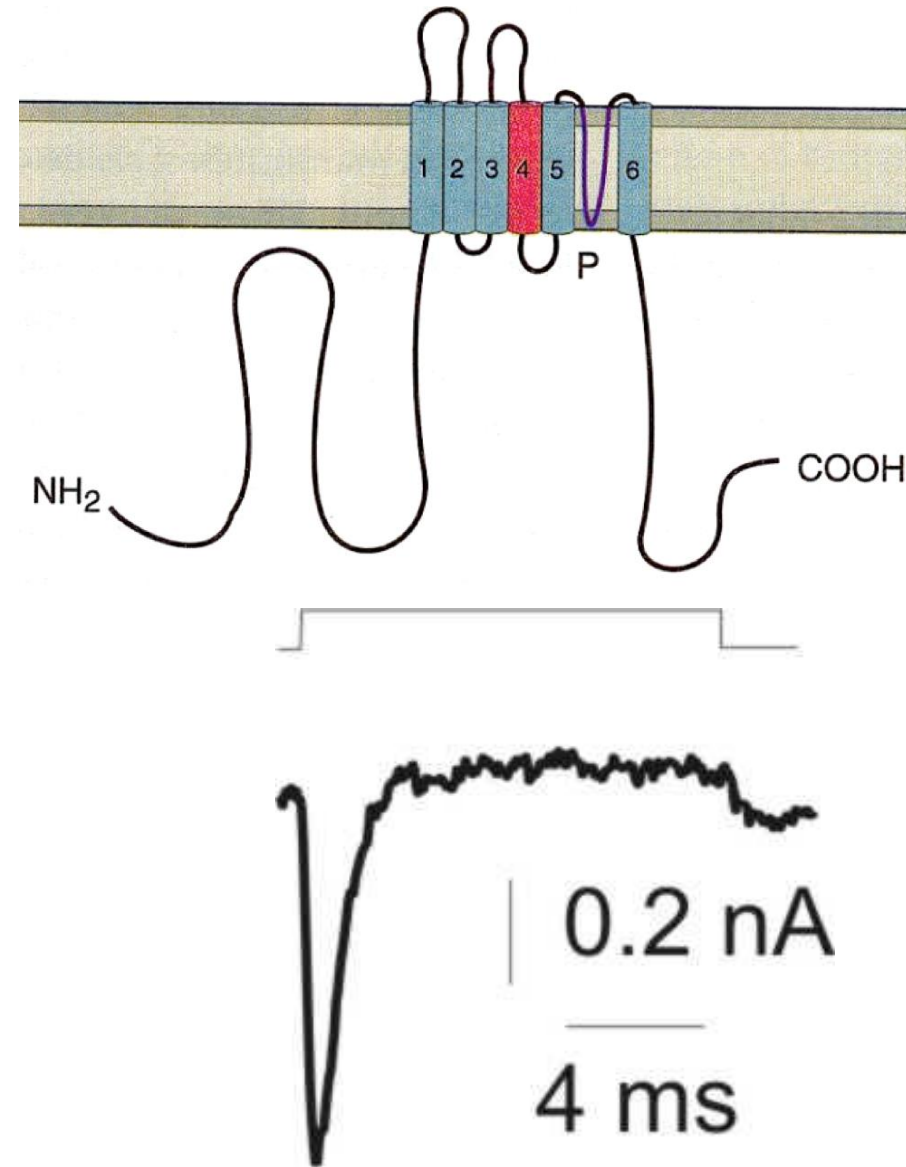
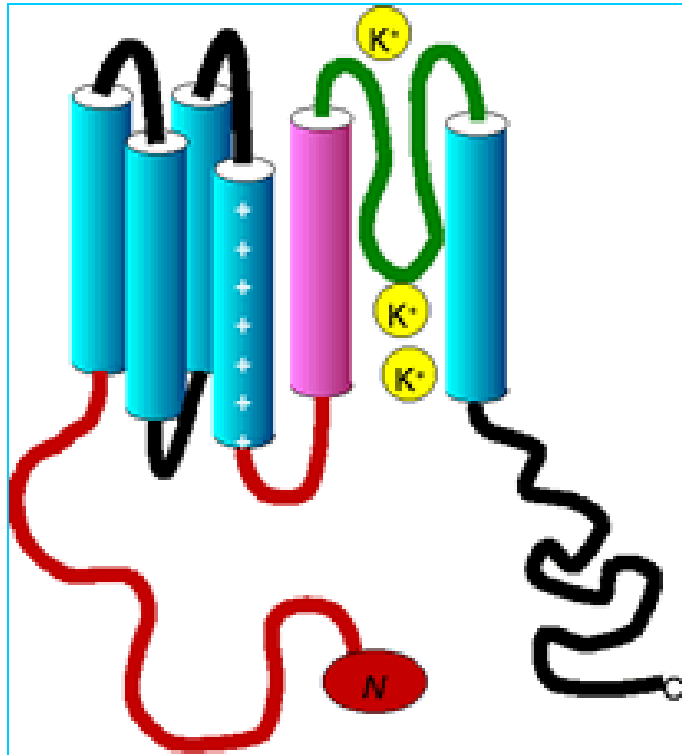


Different Inactivation of Voltage gated K⁺ channels



Depends on the different combinations of subunits

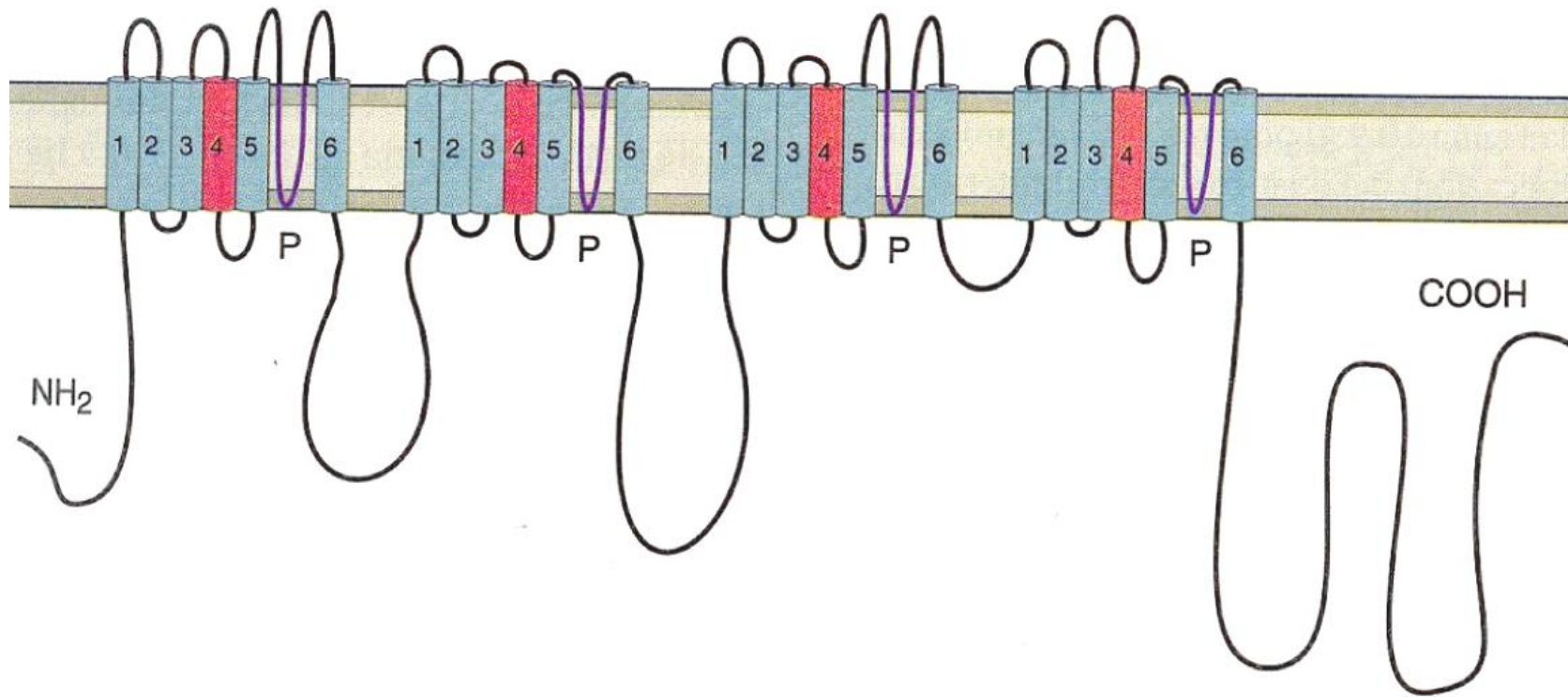
Inactivating Voltage gated K⁺ channels



Voltage gated Ca^{2+} channels

Alpha Subunit

(+ 1 beta, alpha2, delta and gamma)



They transduce electrical signals into metabolic signals

Intracellular and extracellular Ca^{2+} concentrations

Intracellular

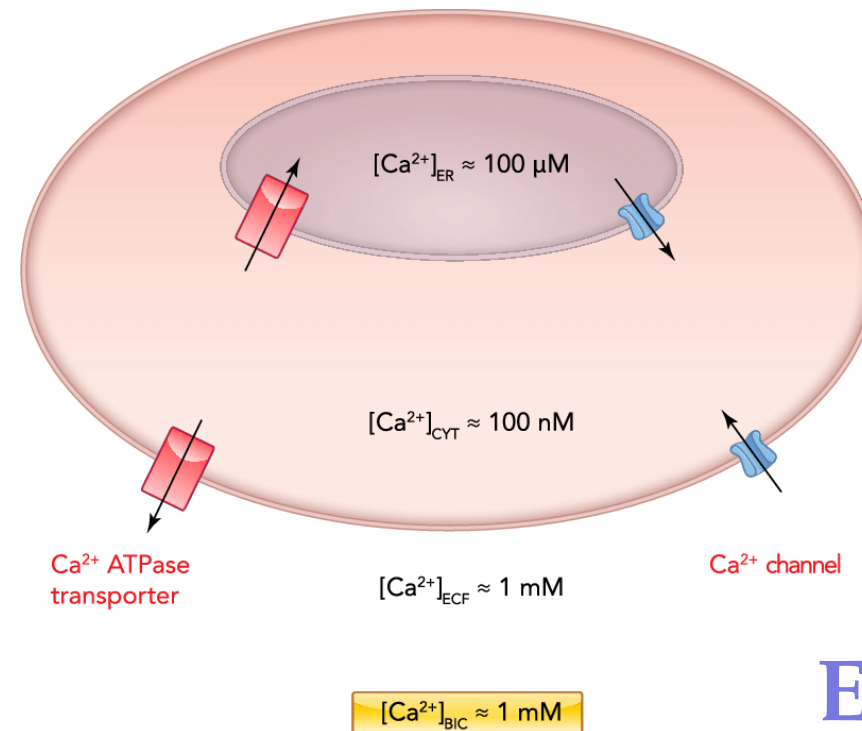
$$[\text{Ca}^{2+}]_{\text{rest}} = 100 \text{ nM}$$

$$[\text{Ca}^{2+}]_{\text{stim}} = 1000 \text{ nM}$$

(1 μM)

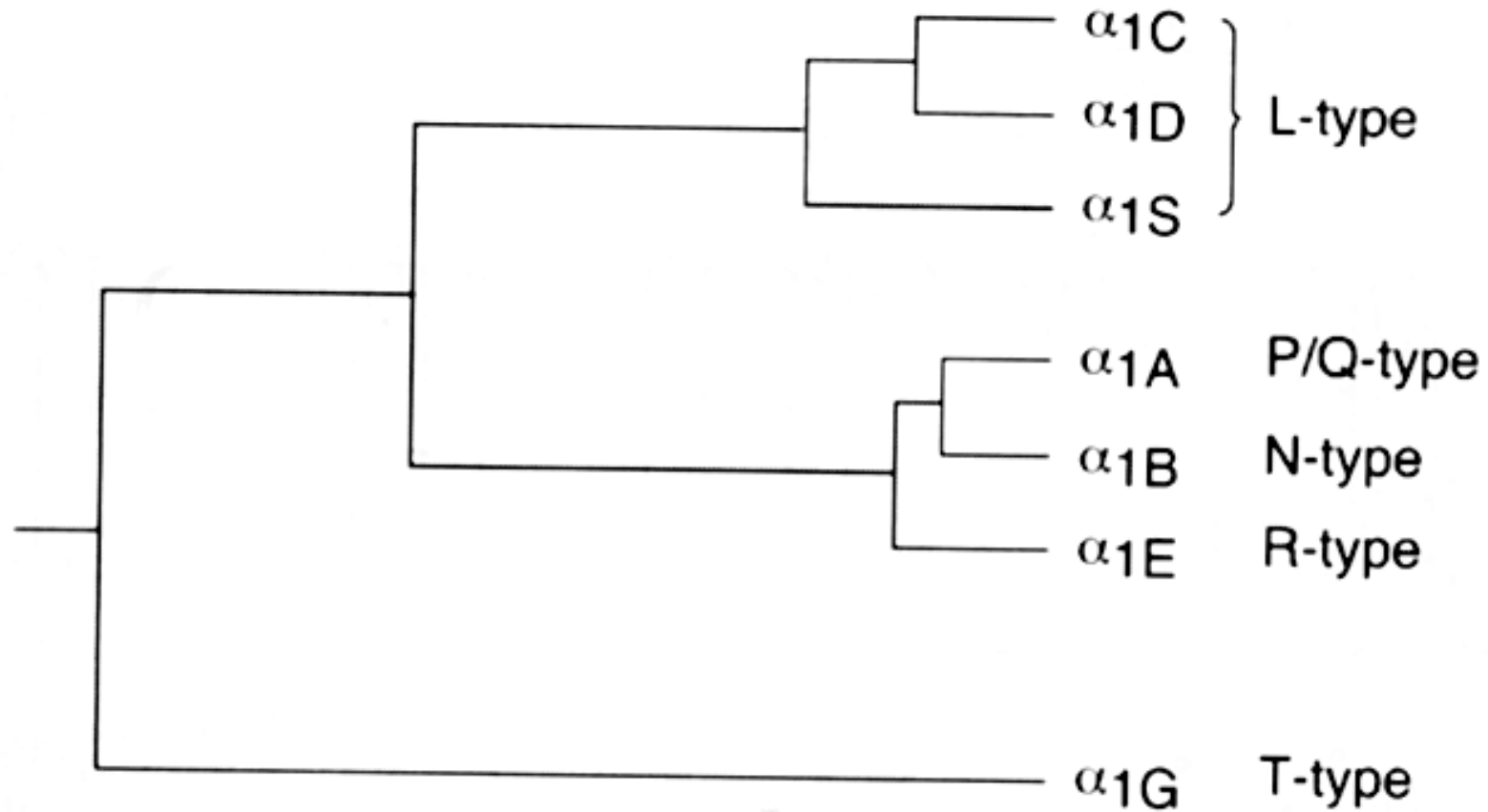
Extracellular

$$[\text{Ca}^{2+}] = 1.2 \text{ mM}$$



$$E_{\text{Ca}} = +110 \text{ mV}$$

Voltage gated Ca^{2+} channels



Voltage gated Ca^{2+} channels

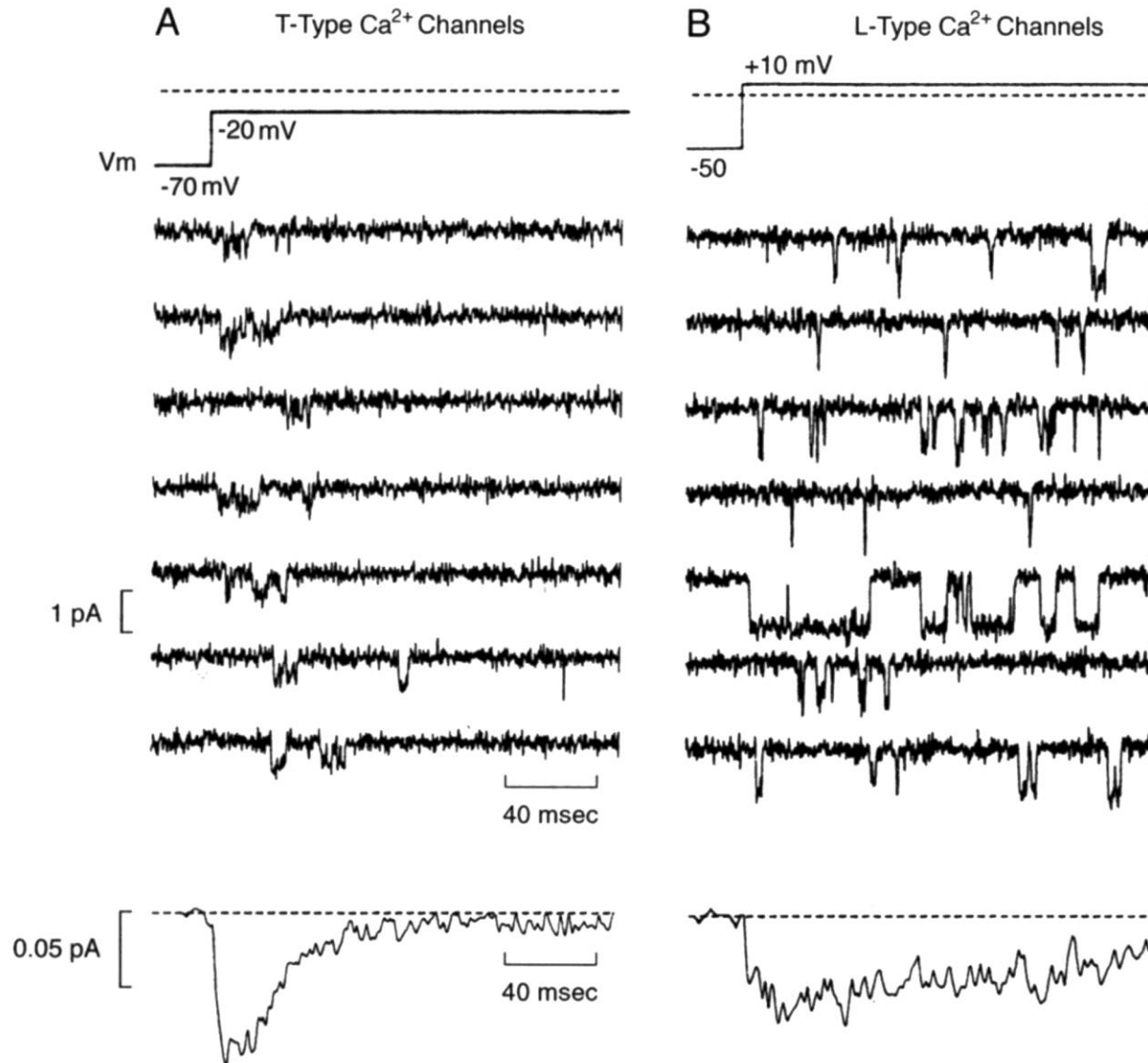
There are 2 types of voltage-dependent channels for Ca^{2+} :

HIGH threshold
(open for large depolarizations)

LOW threshold
(open with slight depolarizations)

Low threshold and High threshold Voltage gated Ca^{2+} channels

$$E_{\text{Ca}} = +110 \text{ mV}$$



Low threshold and High threshold Voltage gated Ca^{2+} channels

Low threshold

In posthumous hyperpolarization
Repeated discharge control
Generation of action potentials independent of Na

Inhibite: amiloride, low nickel conc

High threshold

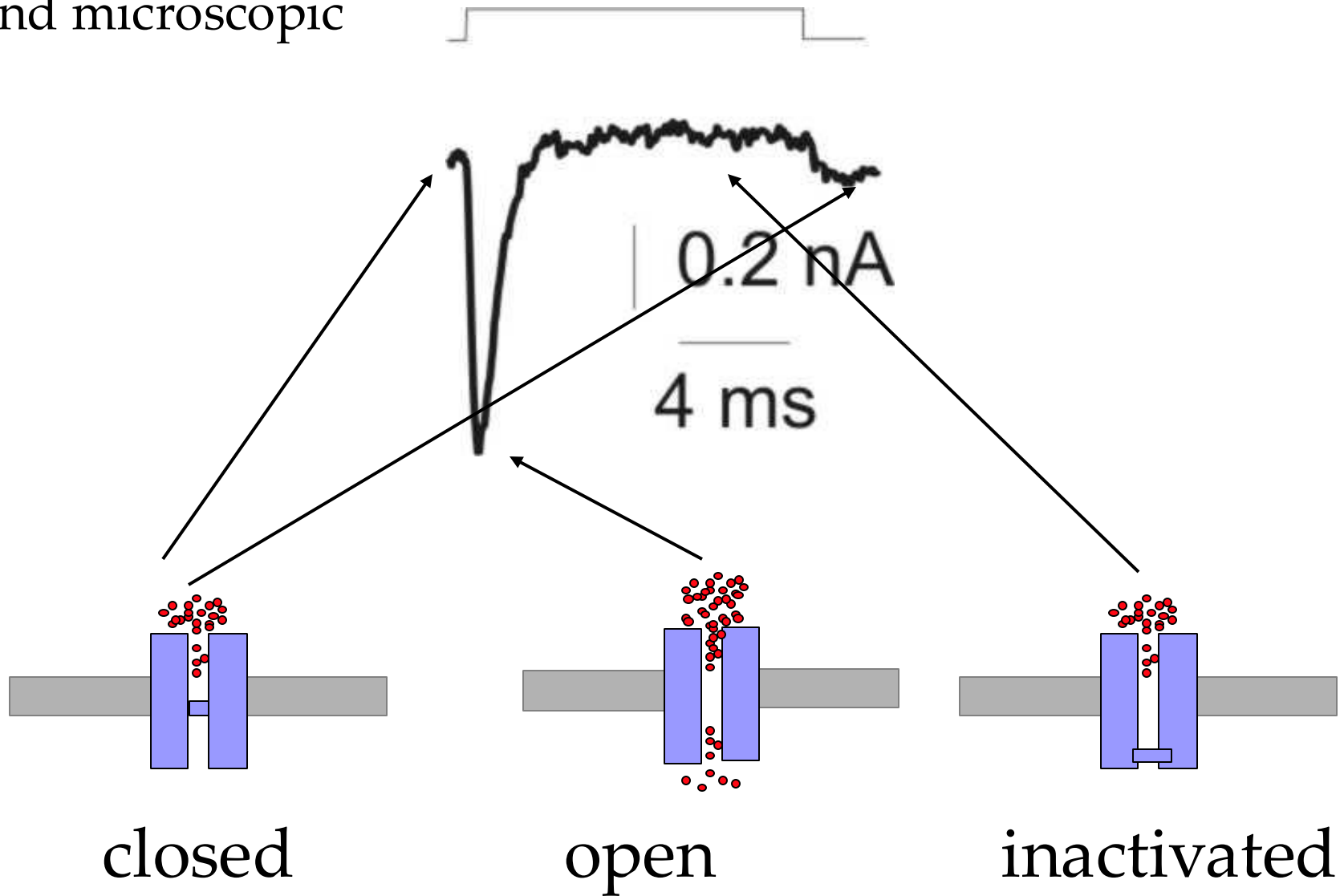
Activate Ca-dip potassium channels
Vesicular exocytosis

Dihydropyridine inhibitors: L channels
Not inhibited by dihydropyridines: non-L channels (N, R, P/Q)

Parameters that characterize Ion channels

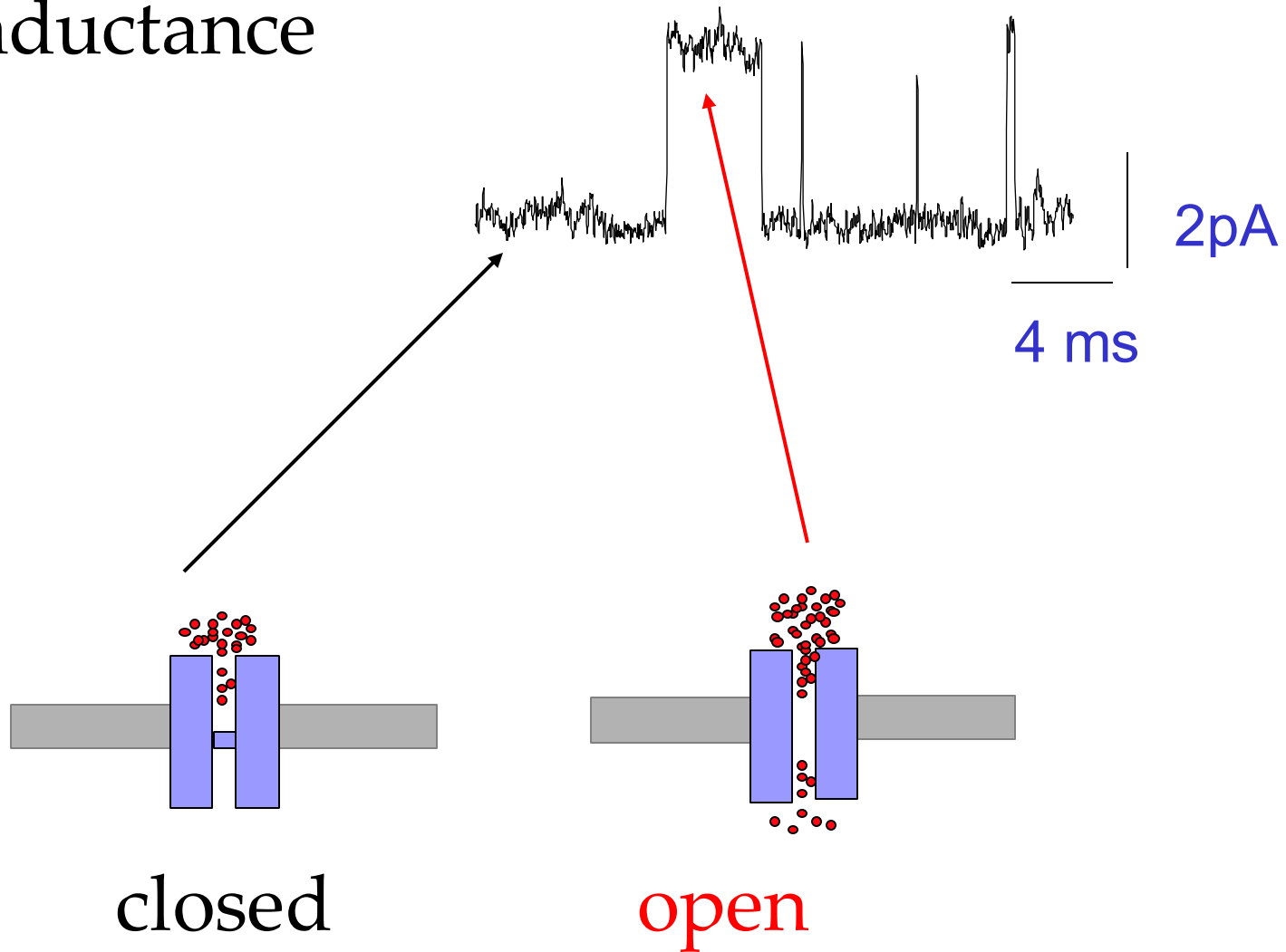
Single Channel and Whole-cell Current

Macroscopic and microscopic



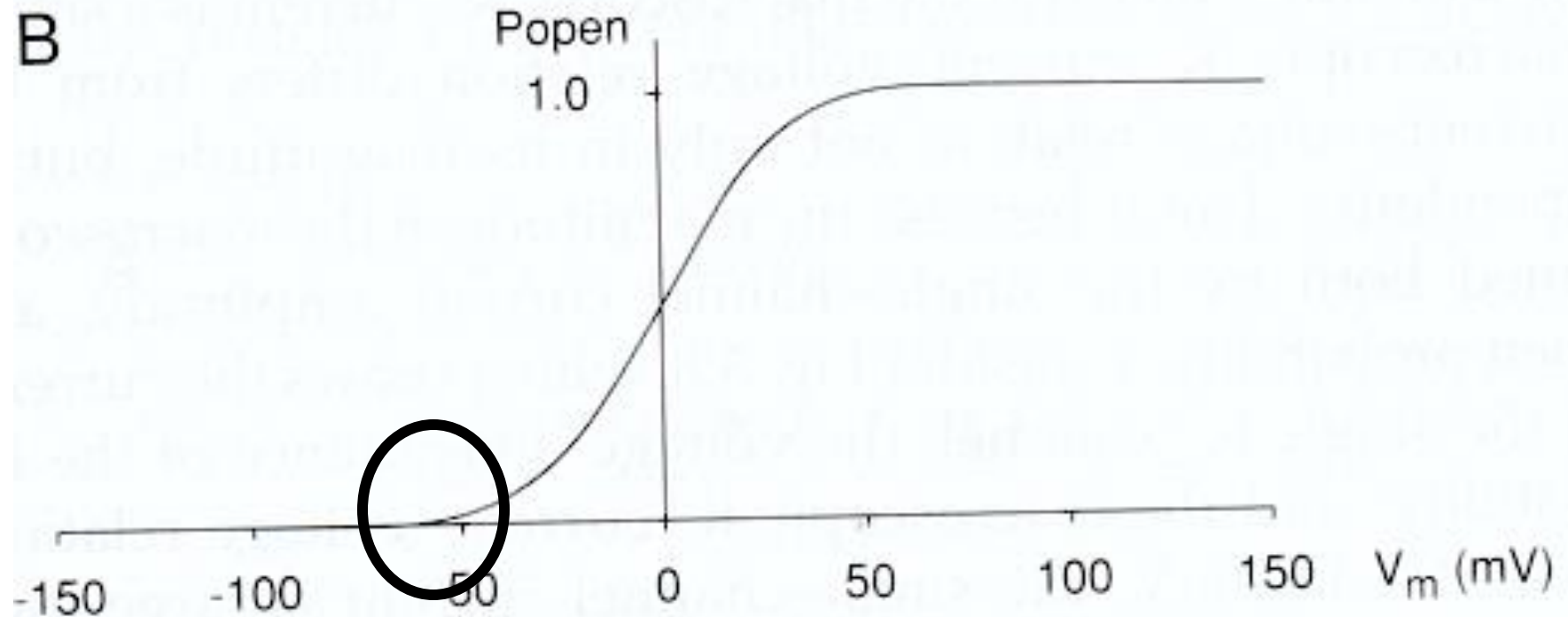
Single Channel Current

Conductance



Single Channel and Whole-cell Current

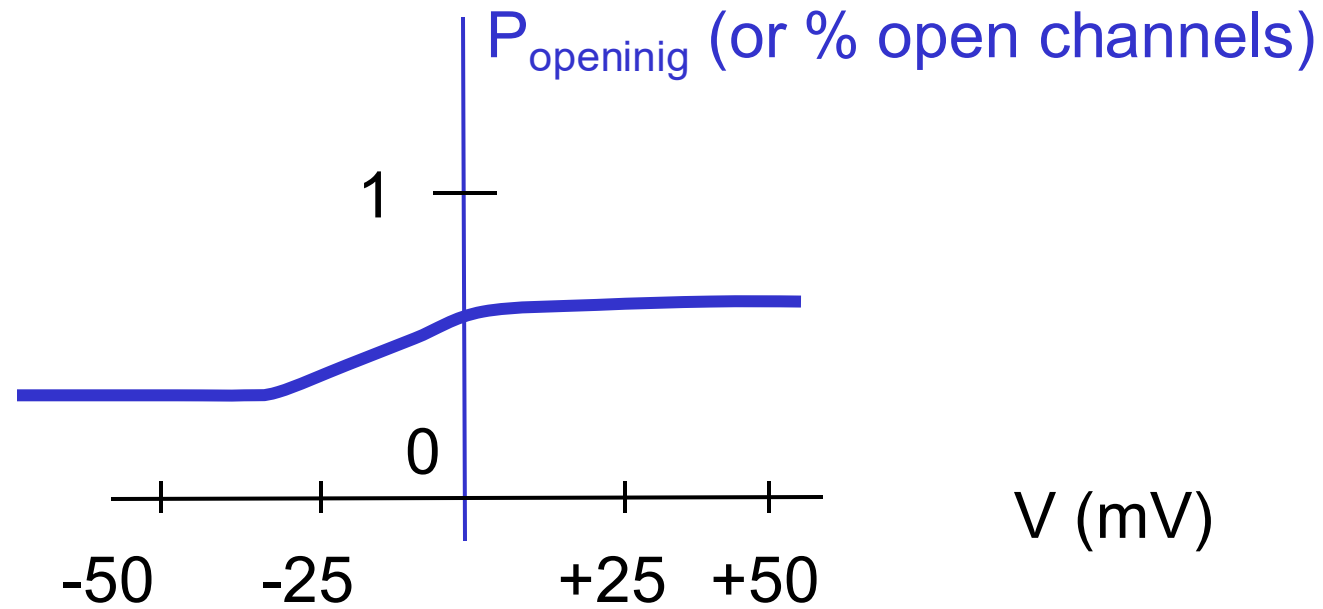
"threshold"
potential



Current-voltage relationship

Single Channel and Whole-cell Current

Opening probability



Single Channel and Whole-cell Current

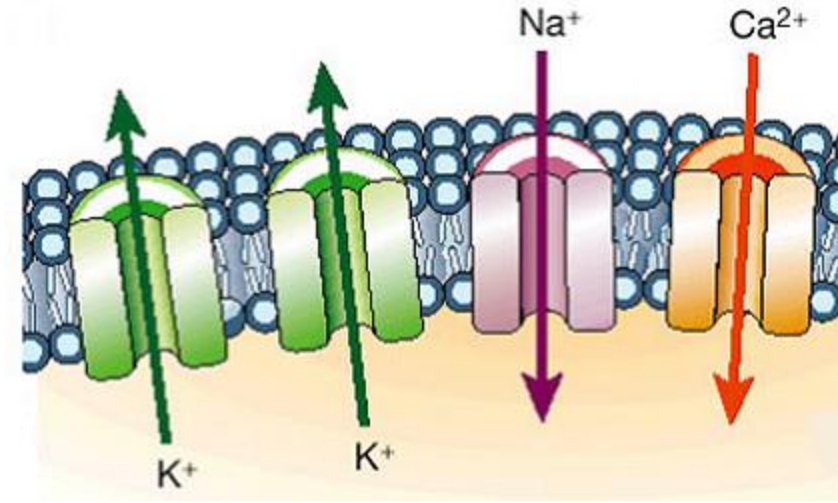
Electrochemical gradient

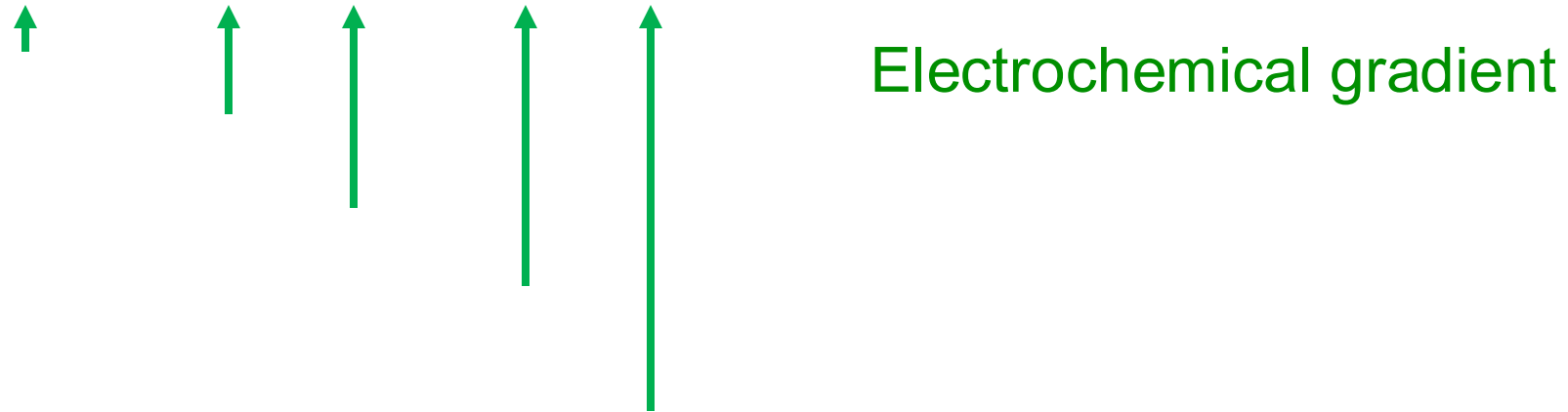
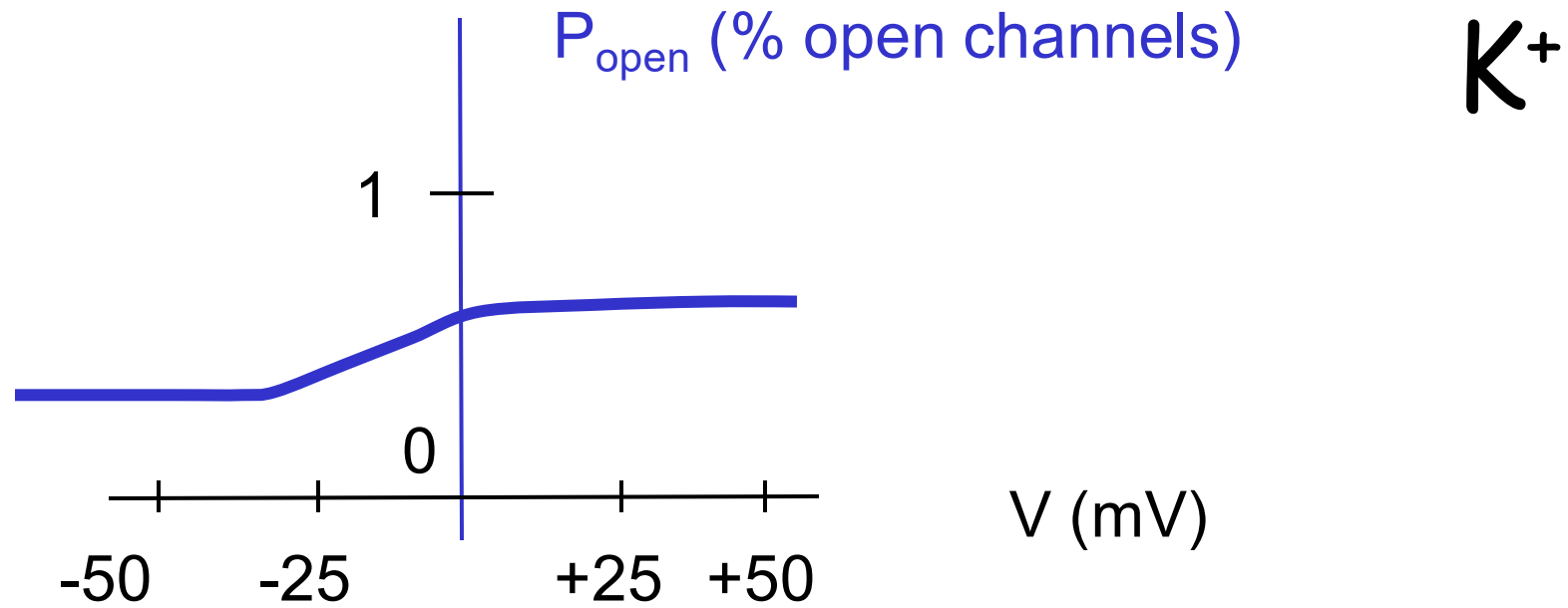
$$I = 1/R \times V$$

$$I = \text{conductance} \times df$$

$$\text{current} = \text{No. of open channels} \times (\text{channel conductance}) \times df$$

$$\text{current} = \text{No. of open channels} \times (\text{channel conductance}) \times (V_m - E_{Na})$$





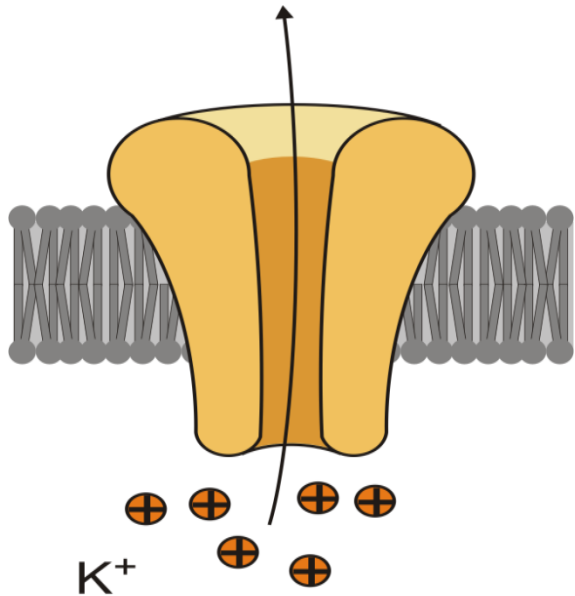
$$\text{Current} = N. \text{ open channels} \times (V_m - E_K) \times (\text{conduttanza canale})$$

The outward flux of K⁺

$$DF = V_m - E_K$$

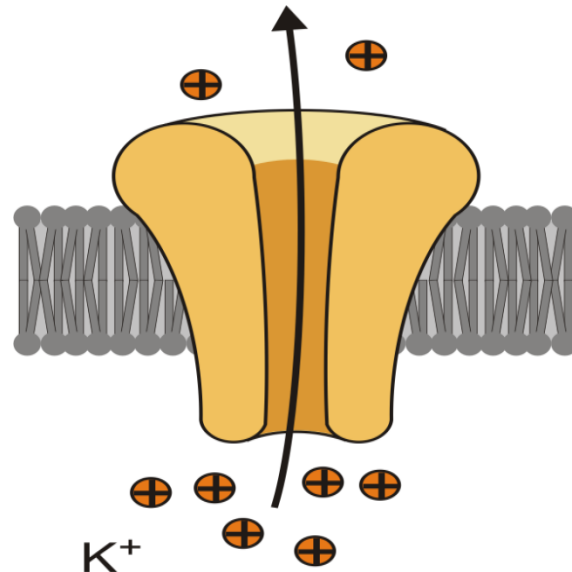
$$E_K = -80 \text{ mV}$$

The total current of K⁺ which leaves the cell depends not only on f.e.m. also from the number of channels open to a certain potential, or from the conductance (g_K):



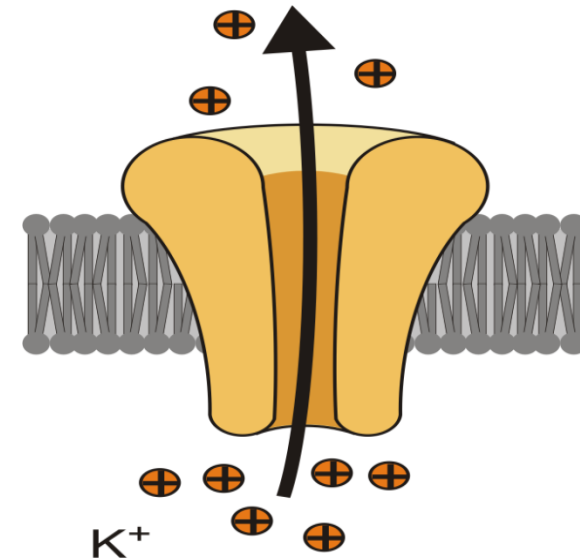
-35 mV

4% open channels
Small Driving Force
Small K⁺ **flux**
Total I_K low



0 mV

70% open channels
Average Driving Force
Average K⁺ **flux**
Total I_K totale **average**



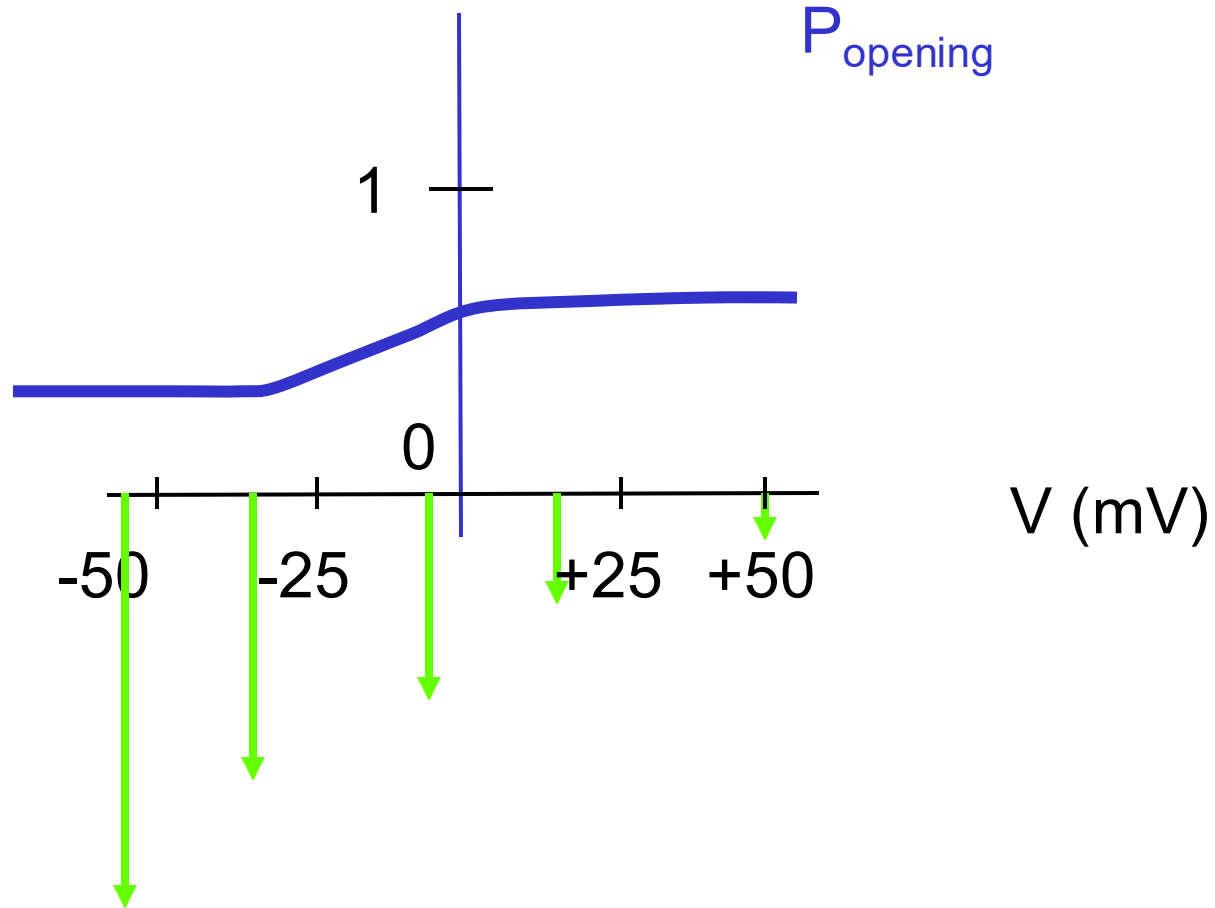
+63 mV

100% open channels
Huge Driving Force
Big K⁺ **flux**
Total I_K totale **Big**

Single Channel and Whole-cell Current

Electrochemical gradient

Na⁺

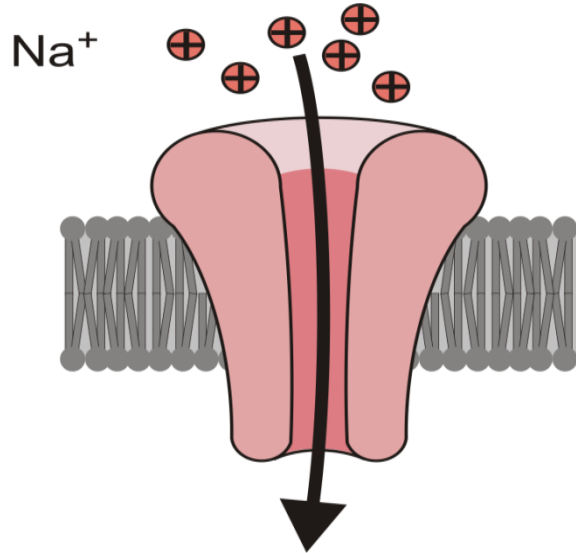


The inward flux of Na⁺

$$DF = V_m - E_{Na}$$

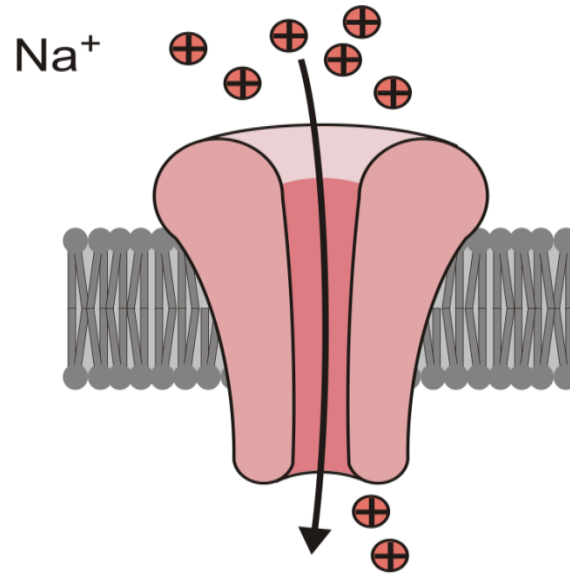
$$E_{Na} = +63\text{mV}$$

The total current of Na⁺ which enters the cell depends not only on f.e.m. also from the number of channels open to a certain potential, or from the conductance (g_{Na}):



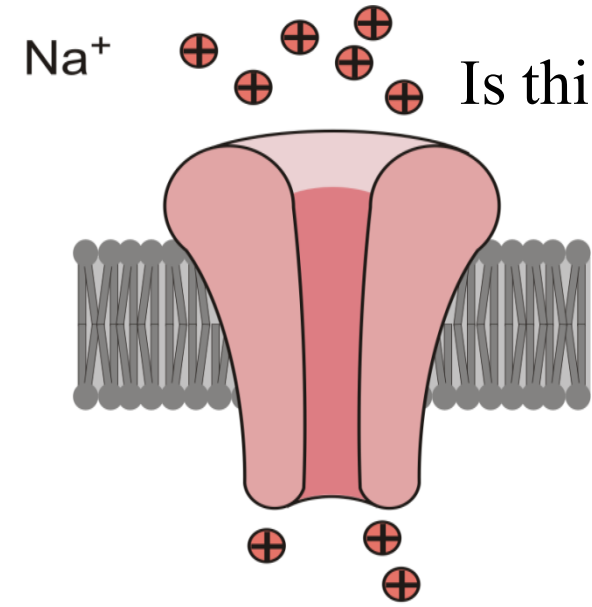
-35 mV

10% open channels
Huge driving force
Big Na⁺ flux
Total I_{Na} **small**



0 mV

80% open channels
Average Driving force
Average Na⁺ flux
Total I_{Na} **Highest**



+63 mV

100% open channels?
Zero Driving force
No Na⁺ Flux
Total I_{Na} **zero**

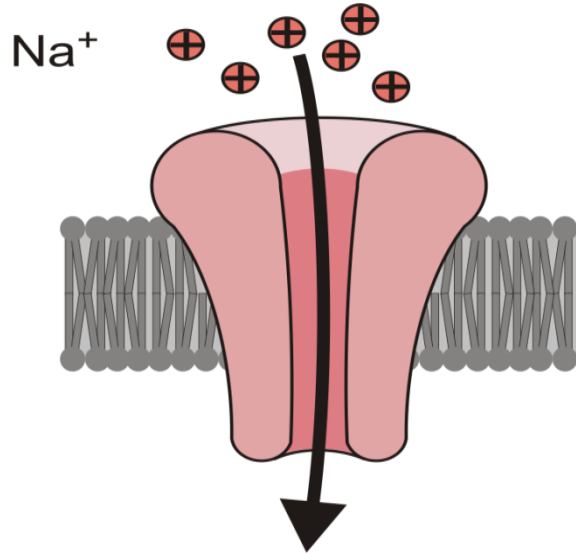
Is this correct?

The inward flux of Na⁺

$$DF = V_m - E_{Na}$$

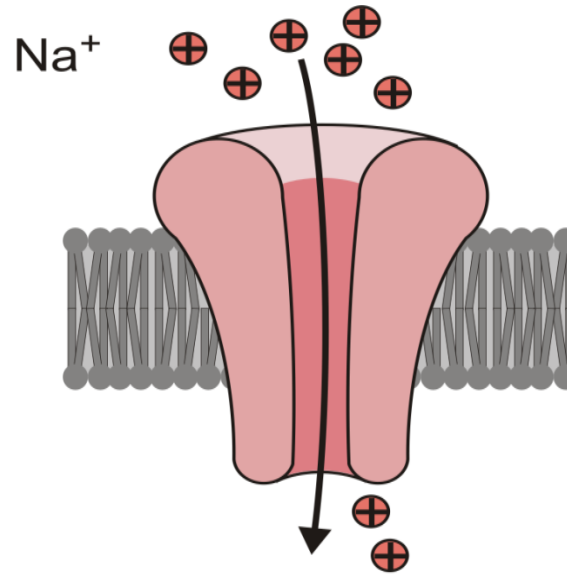
$$E_{Na} = +63\text{mV}$$

The total current of Na⁺ which enters the cell depends not only on f.e.m. also from the number of channels open to a certain potential, or from the conductance (g_{Na}):



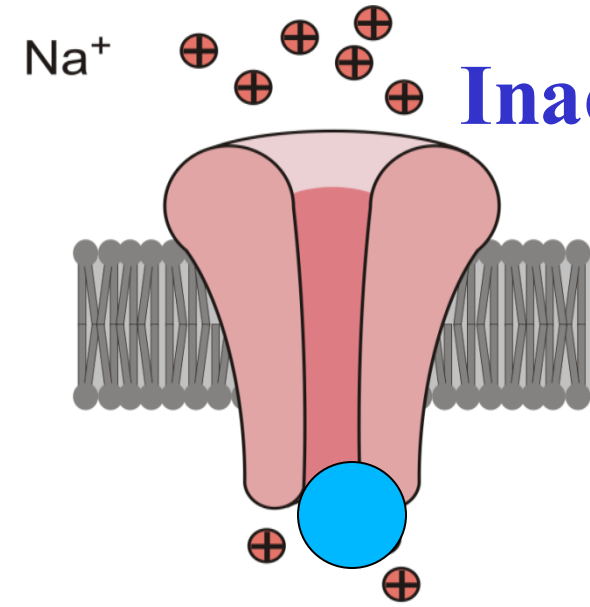
-35 mV

10% open channels
Huge driving force
Big Na⁺ flux
Total I_{Na} **small**



0 mV

80% open channels
Average Driving force
Average Na⁺ flux
Total I_{Na} **Highest**



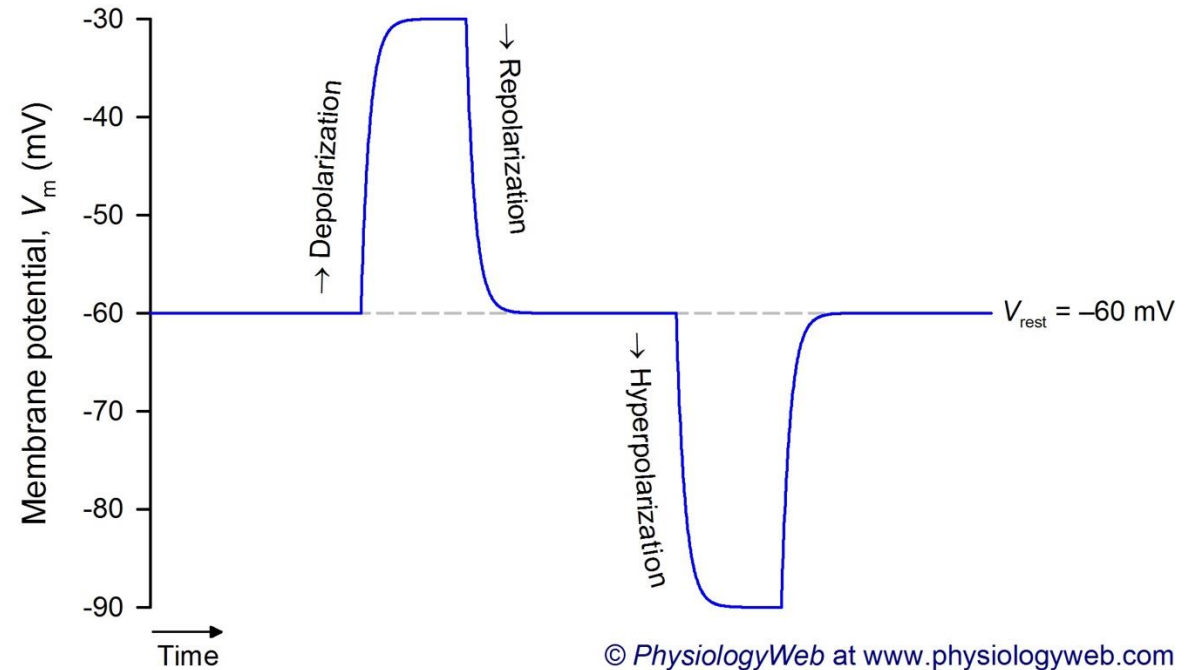
+63 mV

100% Inactivated
channels
No Na⁺ Flux
Total I_{Na} **zero**

channels open by hyperpolarization

Voltage-dependent Ion Channels

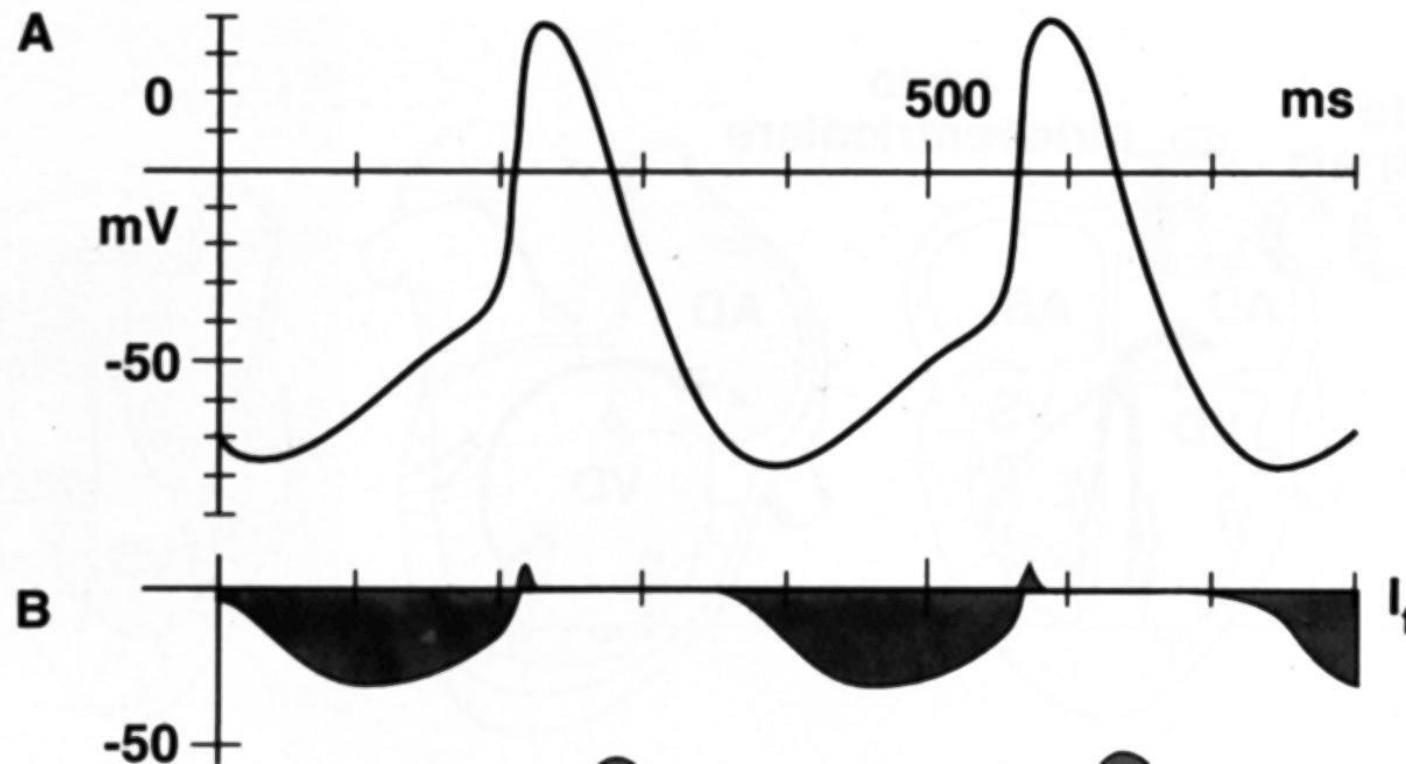
channels open by **hyperpolarization**



I_{funny}

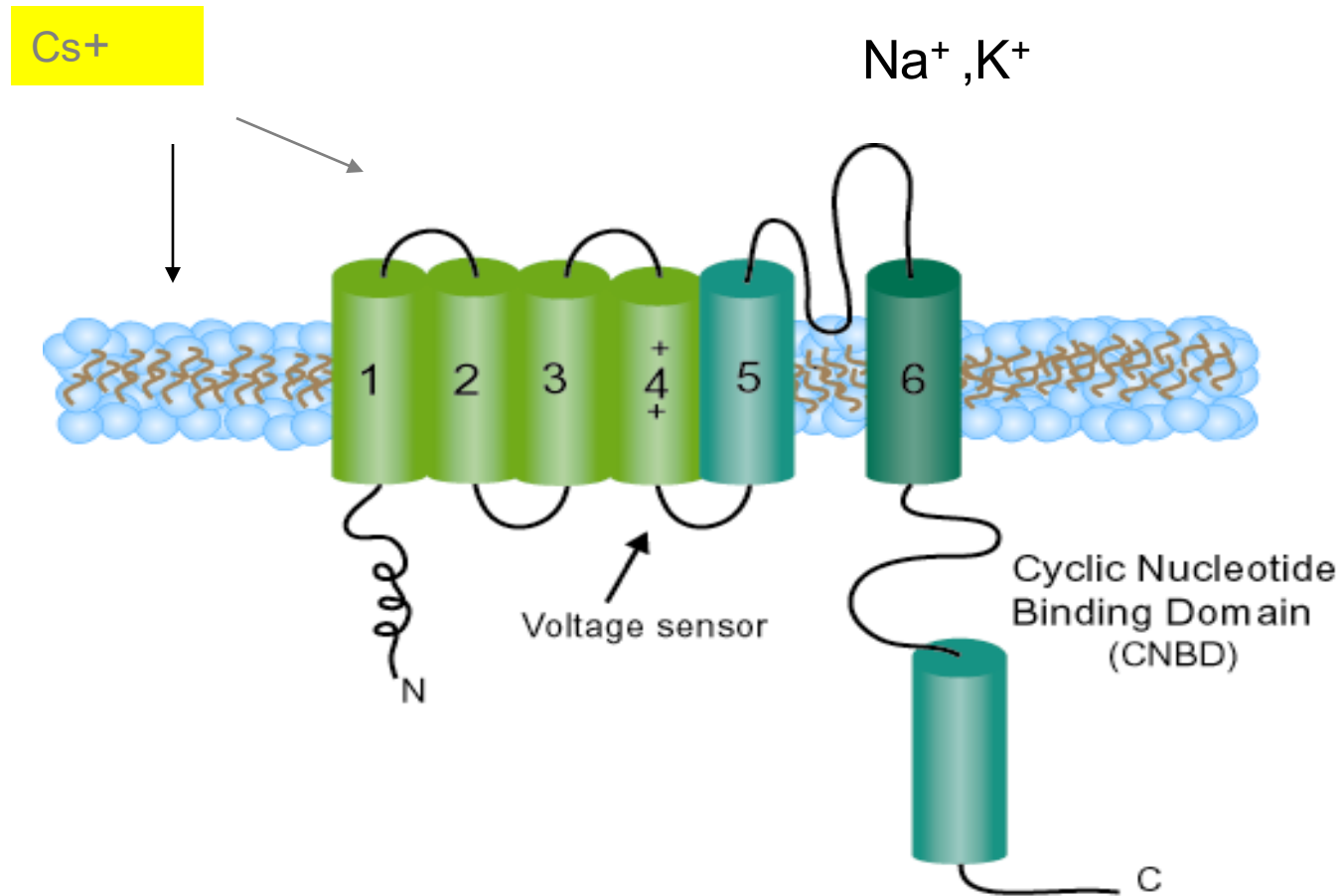
They are cationic channels permeable to Na^+ and K^+

They determine rhythmic variations of V_m



Structure of HCN channels

similar to V gated K channels



The current I_f has singular characteristics

- 1) It is a voltage dependent current that is activated in hyperpolarization: the channel opens when the others close
- 2) The opening of the channels induce a very slow current, entering "inward" that begins at the end of the action potential after the cell has reached its negative potential
- 3) Both sodium and potassium flow in the channel, it has a mixed conductance
- 4) The channel is modulated directly by the cyclic nucleotides and in particular by cAMP and blocked by Cesium

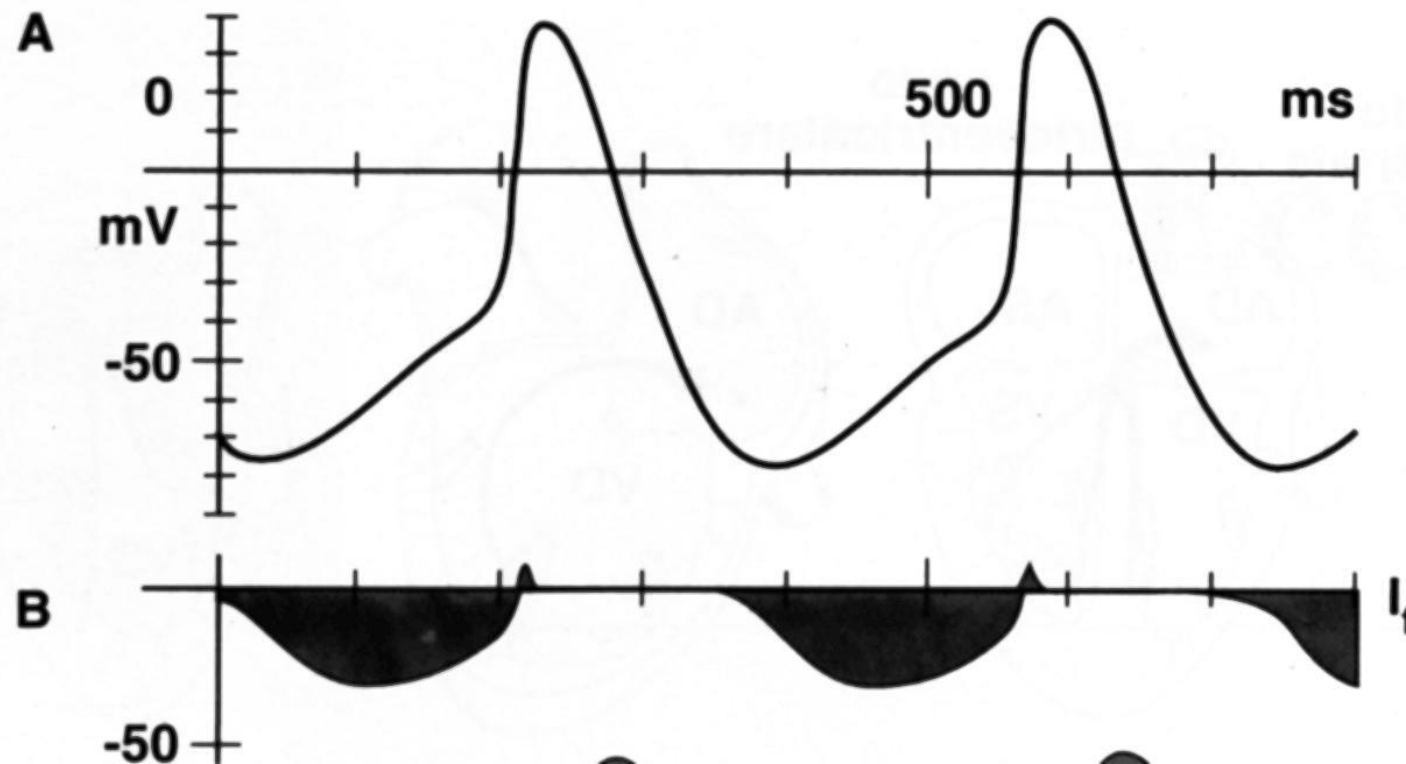
If has helped to found a new family of channels whose exact definition is "activated by hyperpolarization and modulated by nucleotides (HCN)" whose recognition occurred in 1998 with the identification of the genes coding for these proteins.

To this family belong similar currents to If discovered in neurons and photoreceptors.

I_{funny}

They are cationic channels permeable to Na^+ and K^+

They determine rhythmic variations of V_m



Action potential in the Sino-atrial node

- 1) Starter tissue cells (pacemakers) have a low membrane potential of -60 mV
- 2) To this V_m is active the cationic current depolarizing I_f permeable to Na^+ and to K^+ .
- 3) After a first depolarization due to I_f follows a current of voltage-dependent Ca^{2+} which at a certain point becomes strongly regenerative. Depolarization inhibits I_f .
- 4) Depolarization activates a current to K^+ that repolarizes the cell.
- 5) Repolarization closes the channels to the Ca^{2+} and activates the I_f

Action potential in the Sino-atrial node

- I_f : activated by repolarization
- $I_{Ca^{2+}}$: activated by depolarization, transient current
- I_{K^+} : activated by depolarization

