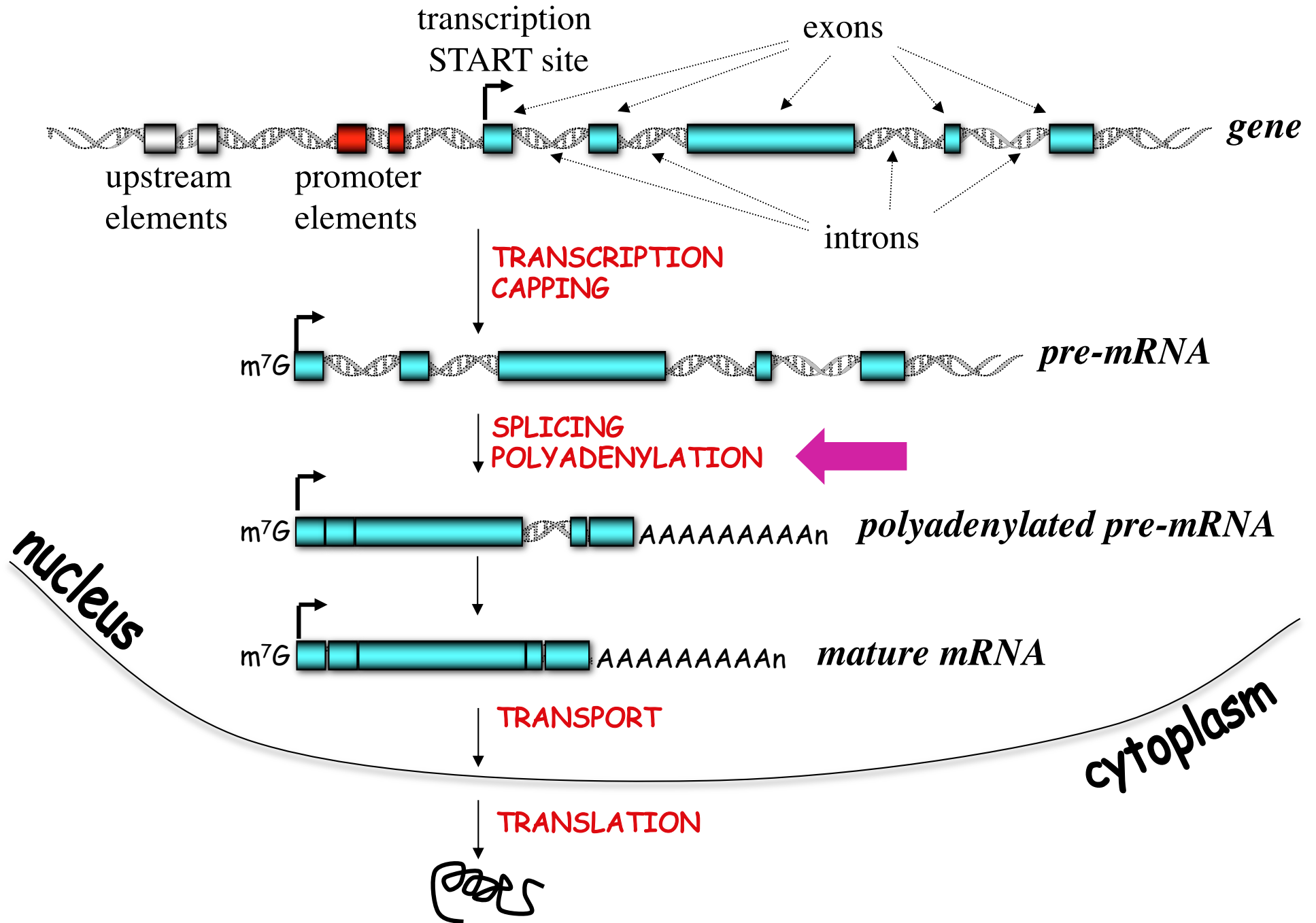


**3'-end formation**

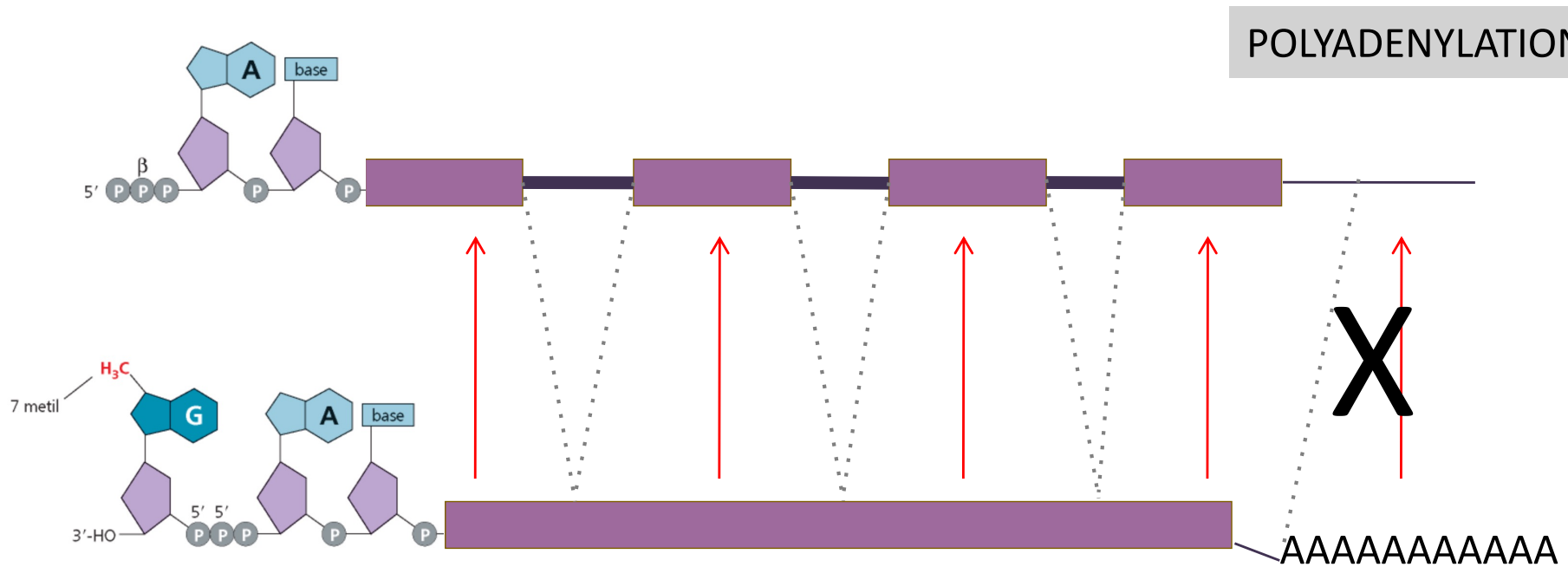
# Eukaryotic gene expression



# What is the function of the PAP-dependent polyA tail?

- 1) Increases RNA stability
- 2) Favours the mRNA transport to the cytoplasm
- 3) Increases translation efficiency by favouring the loading of ribosomal 40S subunit
- 4) Stimulates mRNA transcription termination

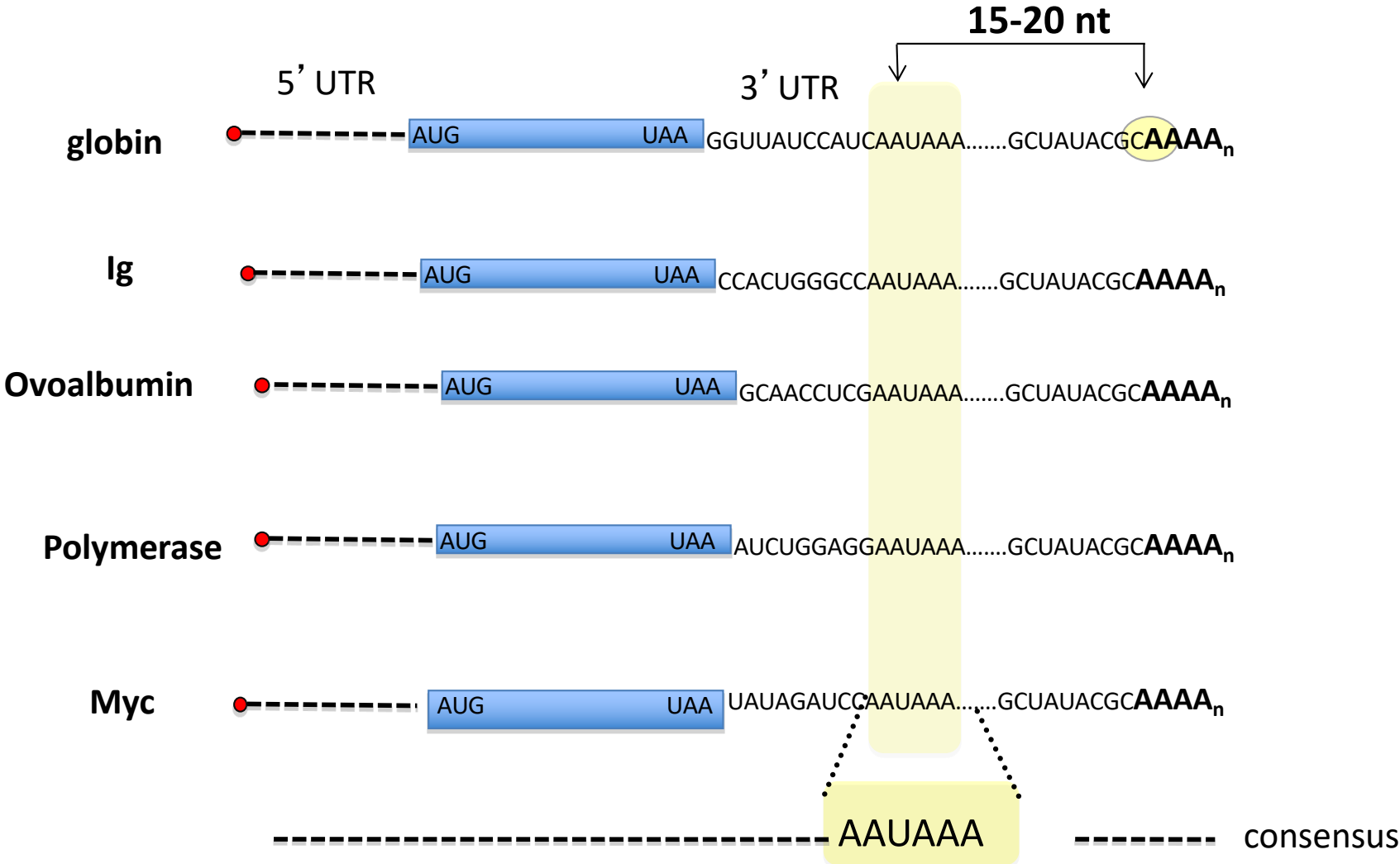
# mRNA maturation



The polyA tail is not transcribed from the DNA but it is added to the mRNA during a process named polyadenylation.

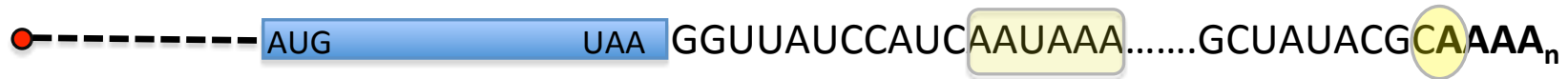
# Looking for consensus sequences in mammals

## SEQUENCE ALLINEAMENT OF cDNAs STARTING FROM THE POLYA TAIL



# LOOKING FOR CONSENSUS SEQUENCES IMPORTANT FOR POLYADENYLATION

**mRNA**

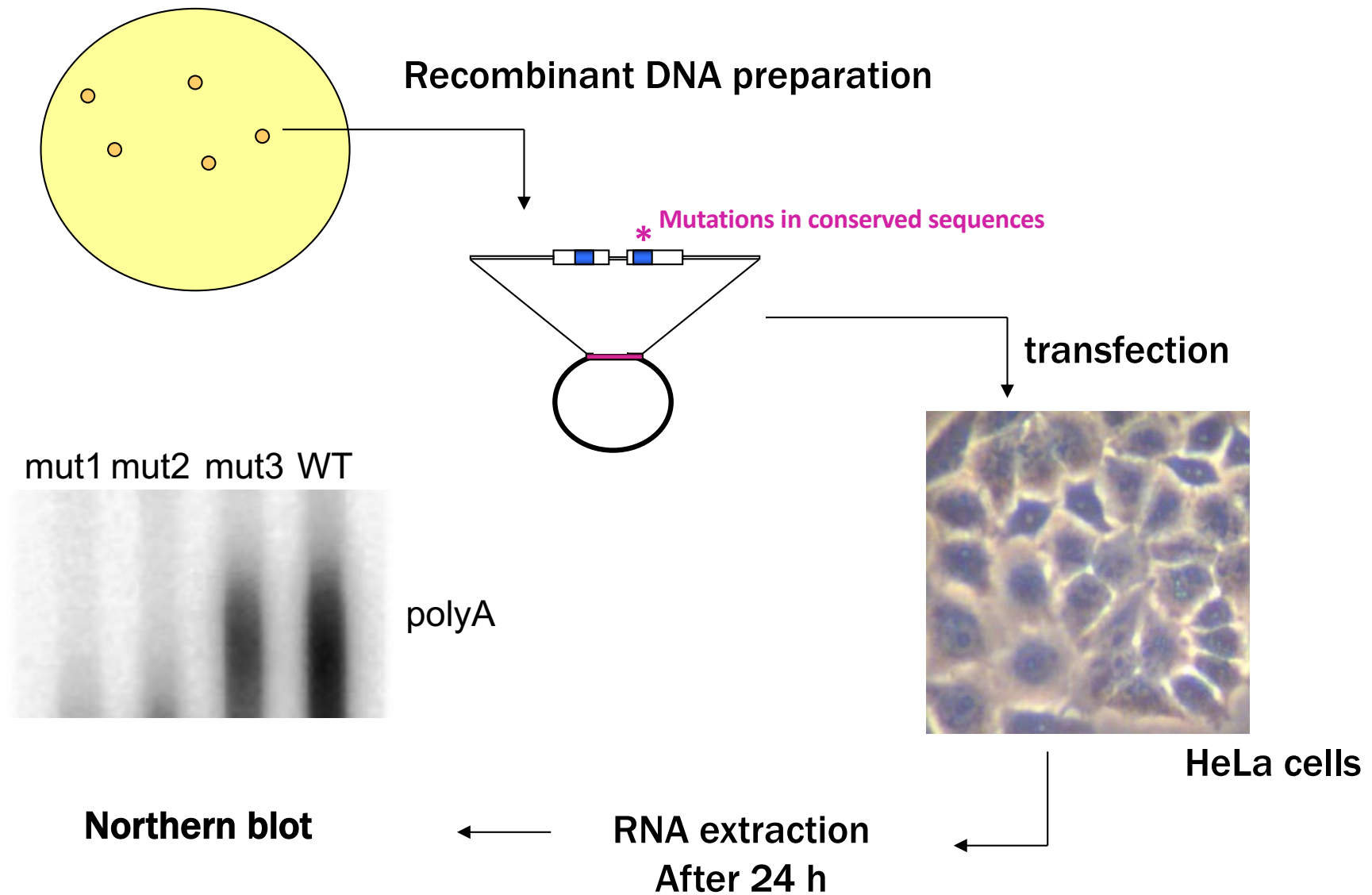


**DNA**



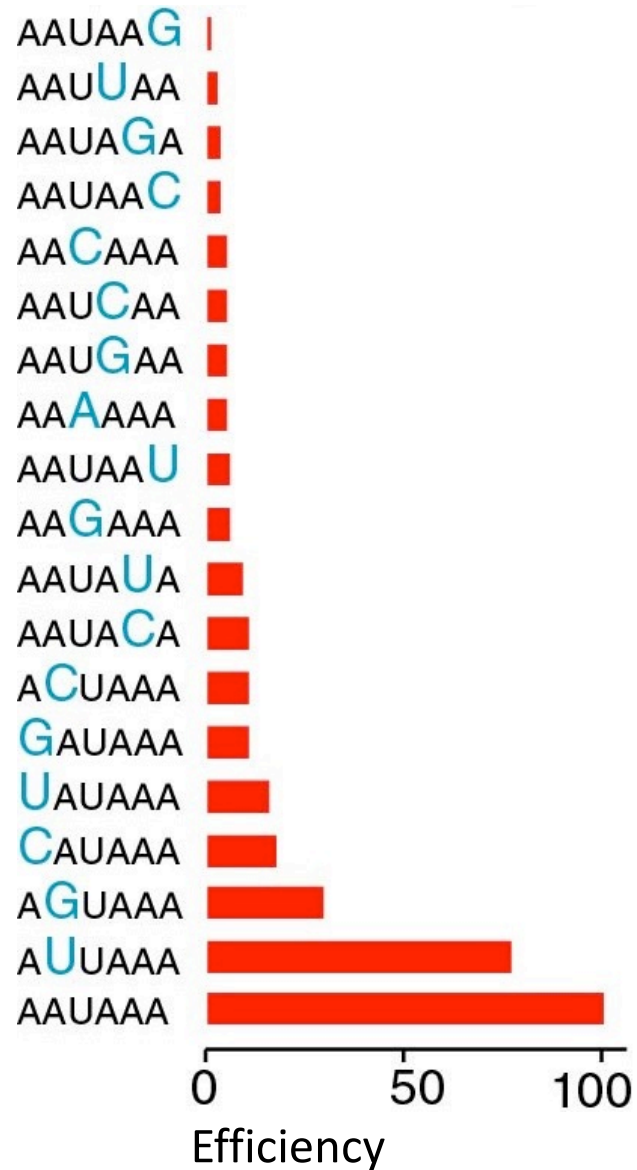
↑  
GU rich

# *In vivo* polyadenylation analysis



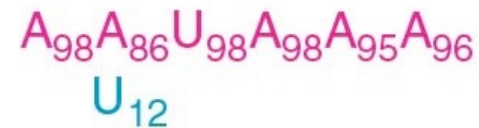
# *In vivo* polyadenylation analysis

Polyadenylation efficiency



*mammals*

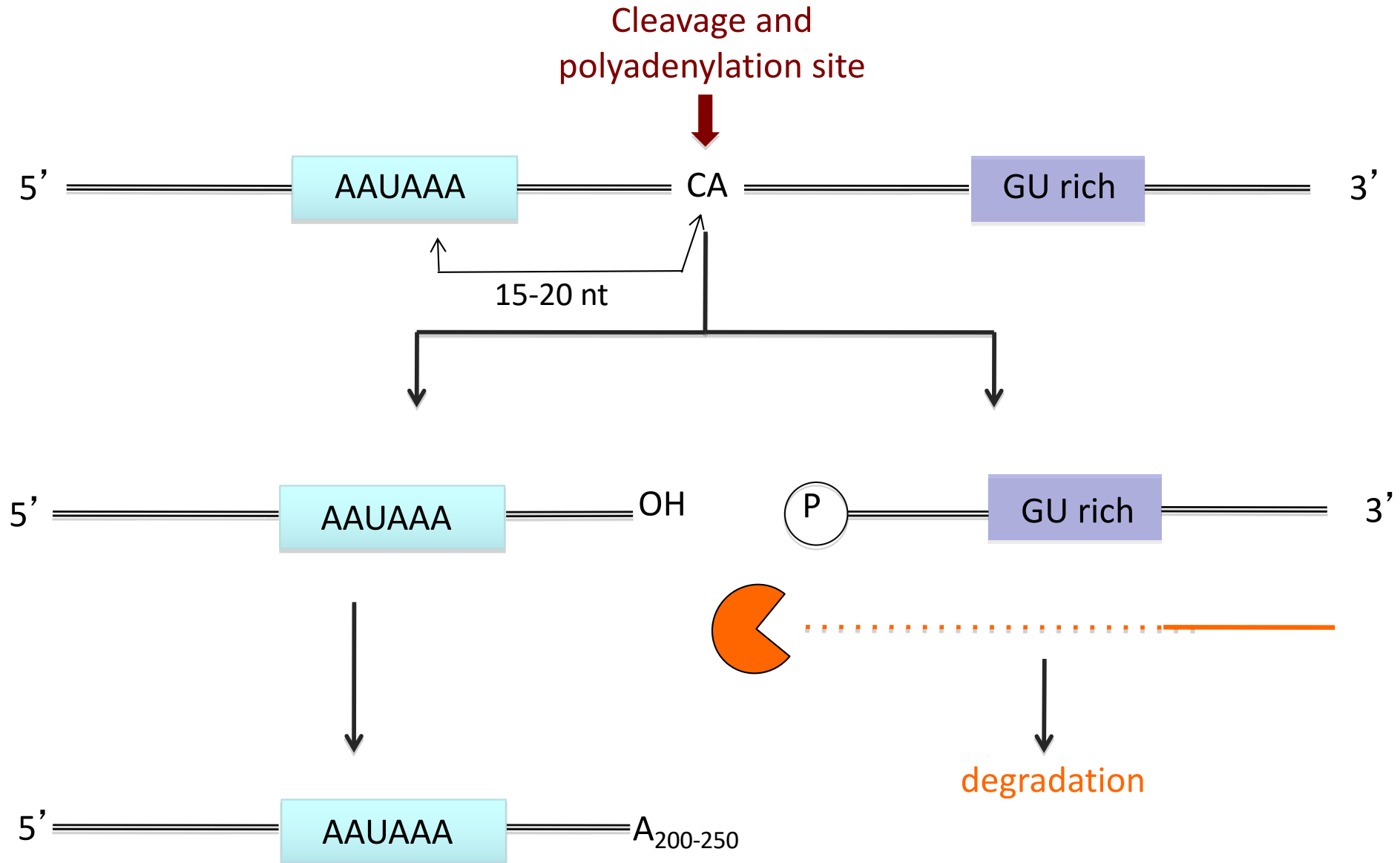
consensus

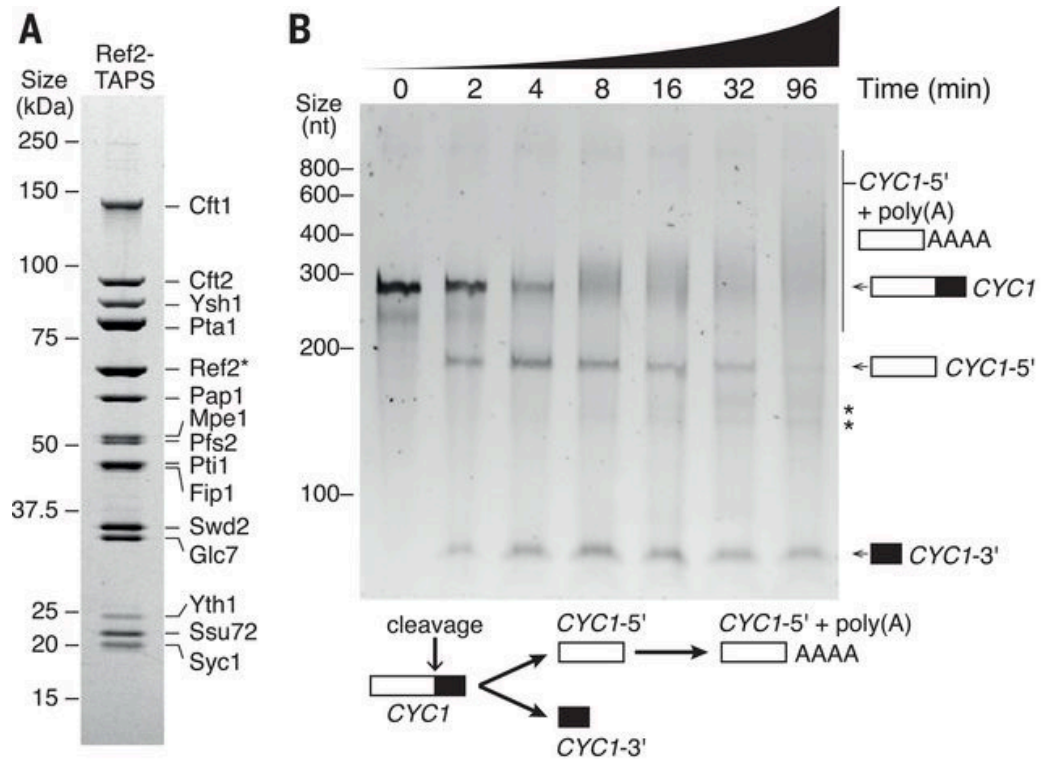
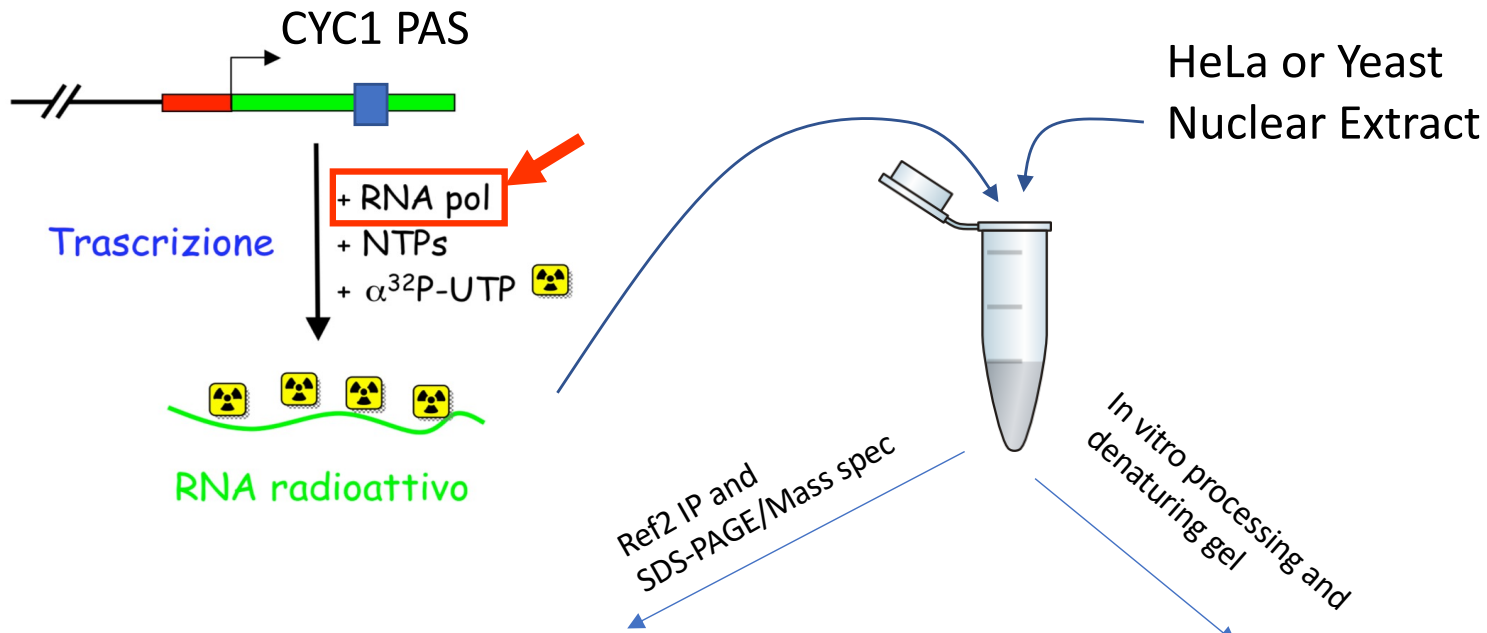


- AAUAAA in mammals  
Located ~20-30 bp from the polyA site
- Other hexamers less efficient but are used
- Mutagenesis and *in vivo* expression studies reveal also the importance of the GU-rich downstream of AAUAAA sequence.



# 3'-end formation in mammalian cells





# PAS sequences are not conserved in evolution

*Cis* elements around the PAS in simpler species differ from those in mammals:

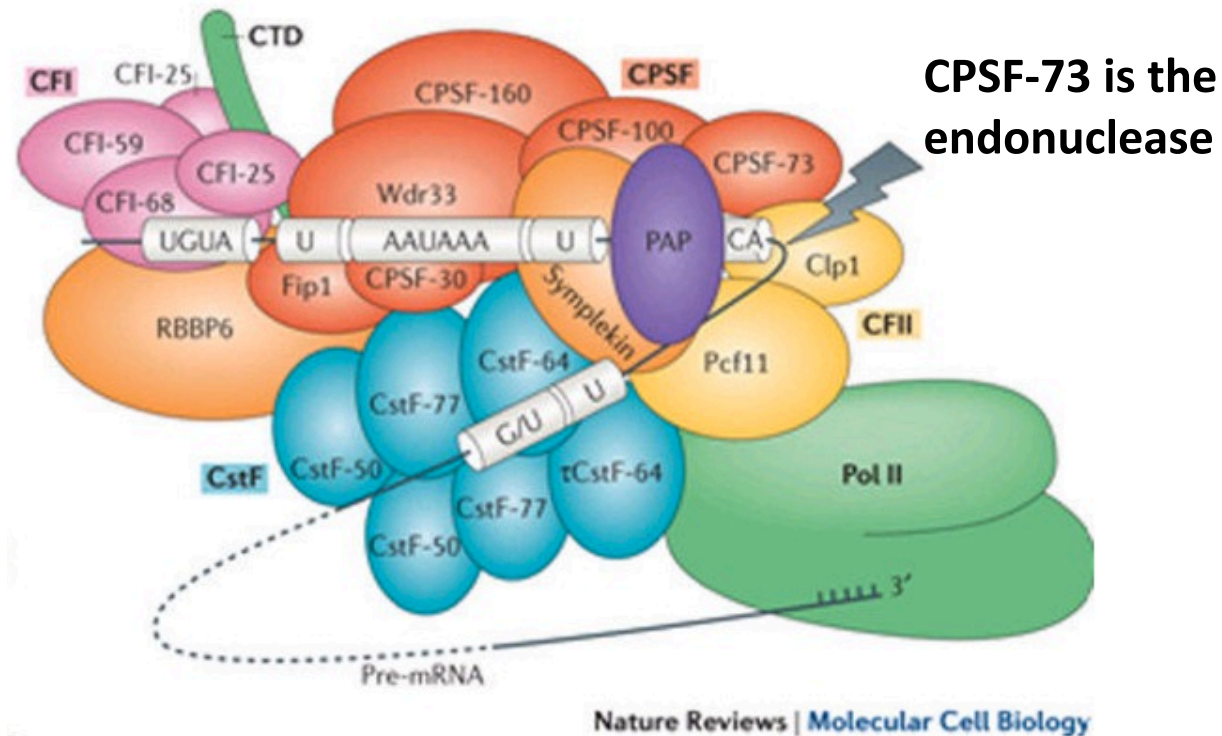
- downstream GU-rich elements are not present in nematode PASs.
- AAUAAA is prominent in *Schizosaccharomyces pombe*, while a general A-rich sequence is present in the same region in *Saccharomyces cerevisiae*. Yeast PASs do not have downstream GU-rich elements. However, UAUA elements are highly enriched in the upstream region.
- a typical plant PAS is more similar to that of yeast than of metazoan, with a loosely defined upstream AAUAAA element flanked by U-rich sequences.

# The polyadenylation machinery in metazoans

The polyadenylation machinery in metazoans is composed of ~20 core proteins, including four protein complexes and several single proteins:

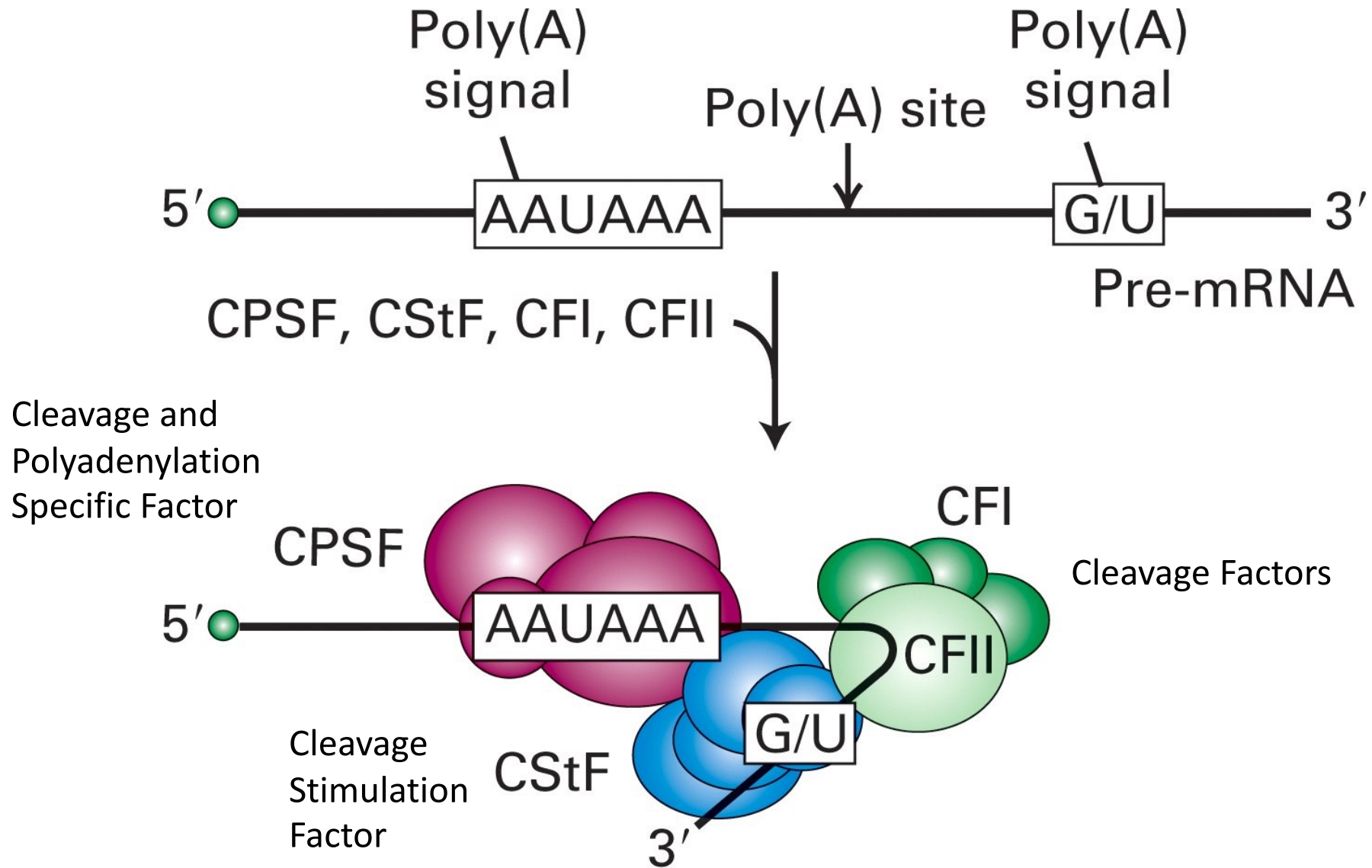
- **cleavage and polyadenylation specificity factor (CPSF)**, which contains **CPSF160** (also known as CPSF1), **CPSF100** (also known as CPSF2), **CPSF73** (also known as CPSF3), **CPSF30** (also known as CPSF4), **FIP1** (factor interacting with PAP) and **WDR33**
- **cleavage stimulation factor (CSTF)**, which contains **CSTF 77**, **CSTF50** (also known as CSTF1) and **CSTF64**
- **cleavage factor I (CFI)**, which contains **CFI25** and either **CFI68** or **CFI59**;
- **cleavage factor II (CFII)**, which contains **PCF11** and **CLP1**
- Single proteins include **symplexin**, **poly(A) polymerase (PAP)**, **retinoblastoma-binding protein 6 (RBBP6)**.

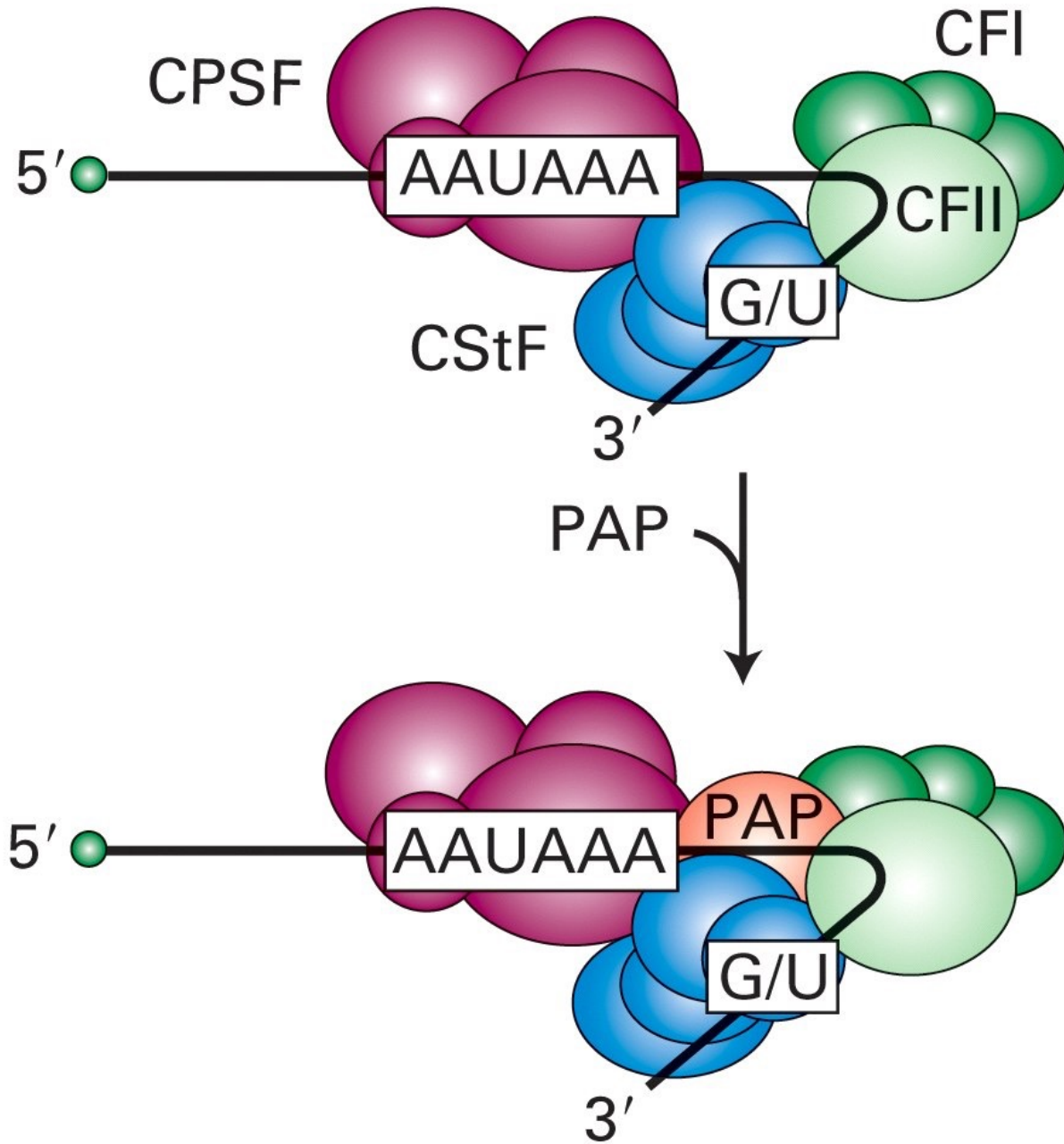
# Specific complexes are involved in mRNA 3' end-processing

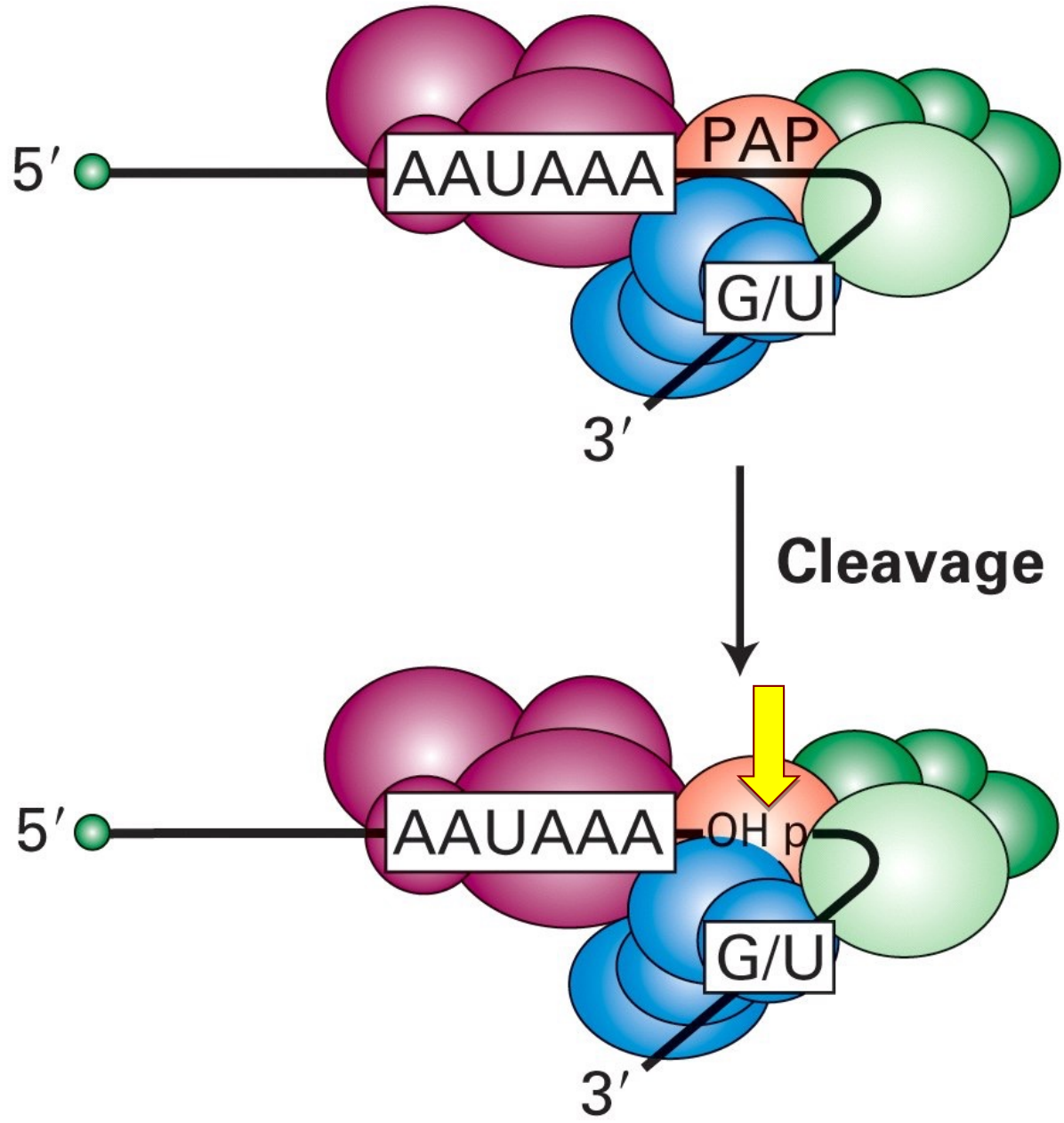


Despite considerable divergence between yeast and mammals in the core RNA sequences that constitute the PAS, nearly all mammalian polyadenylation factors have homologues in yeast, with the exception of the CFI proteins and CSTF50. Moreover, the yeast polyadenylation factor Hrp1p, which interacts with UA-rich elements, is missing from metazoans. The polyadenylation machinery in plants is similar to that in metazoans, but with substantial gene (and thus protein) duplications.

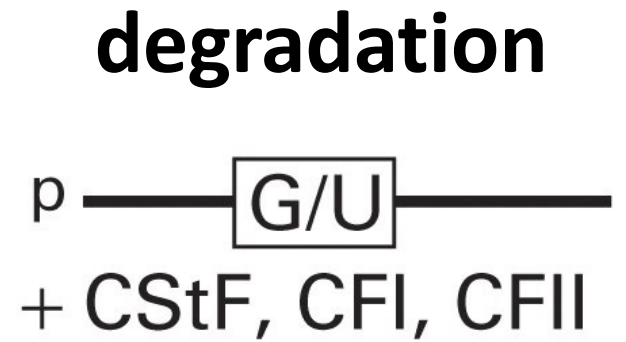
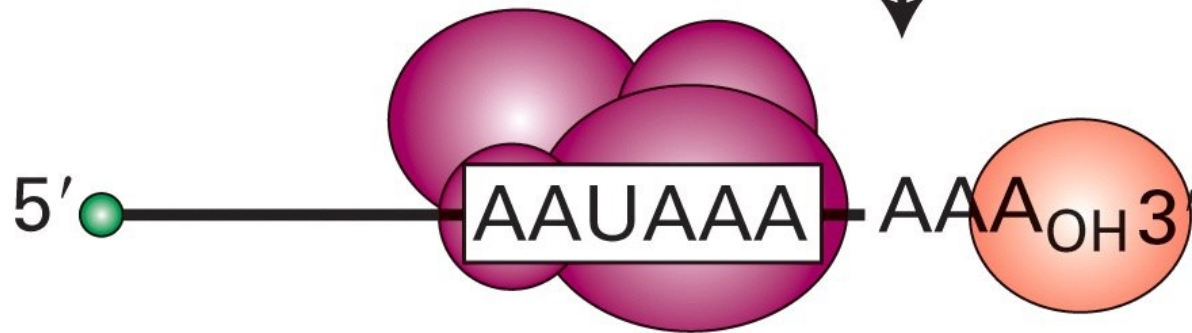
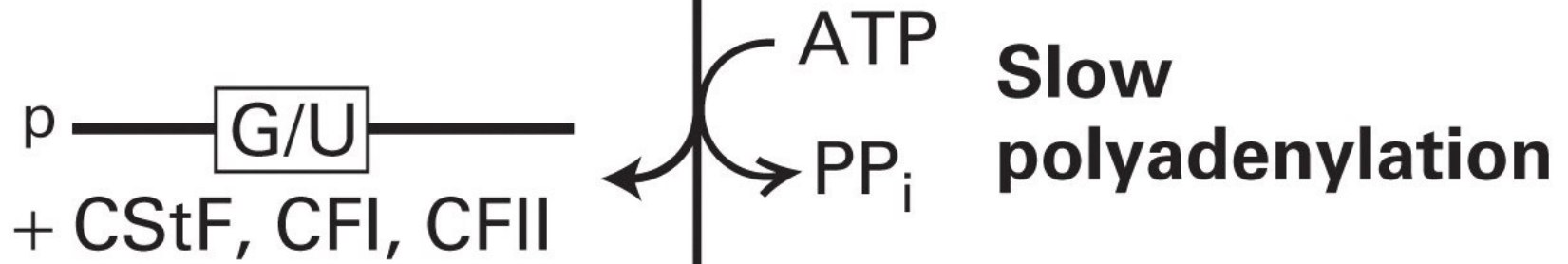
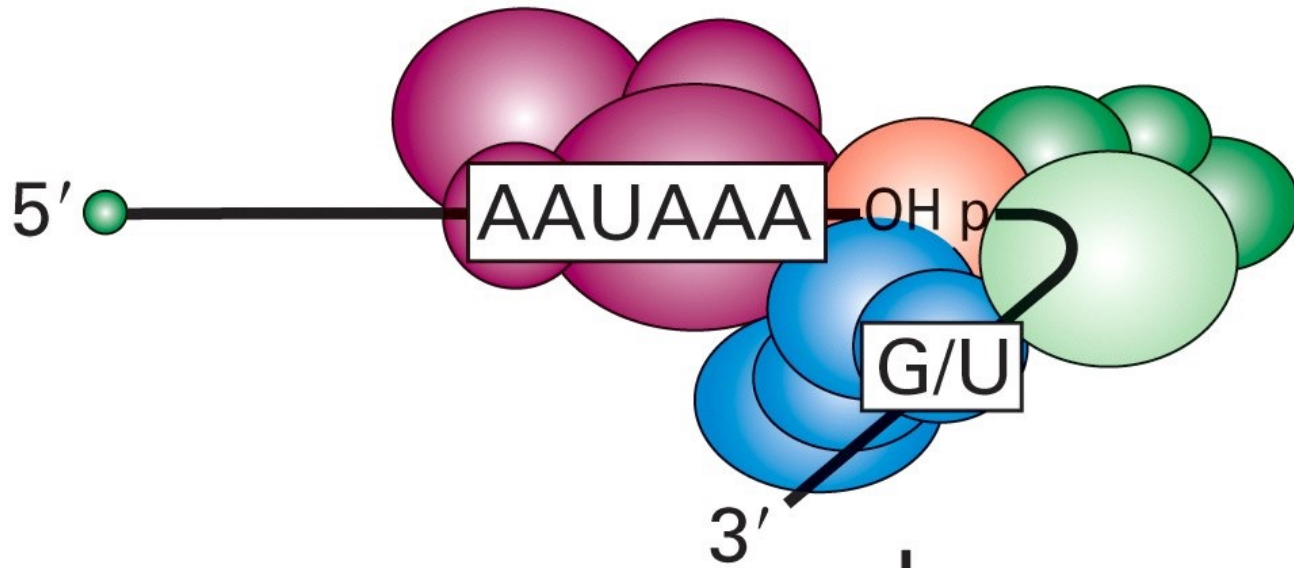
# 3'-end formation: RNA Processing

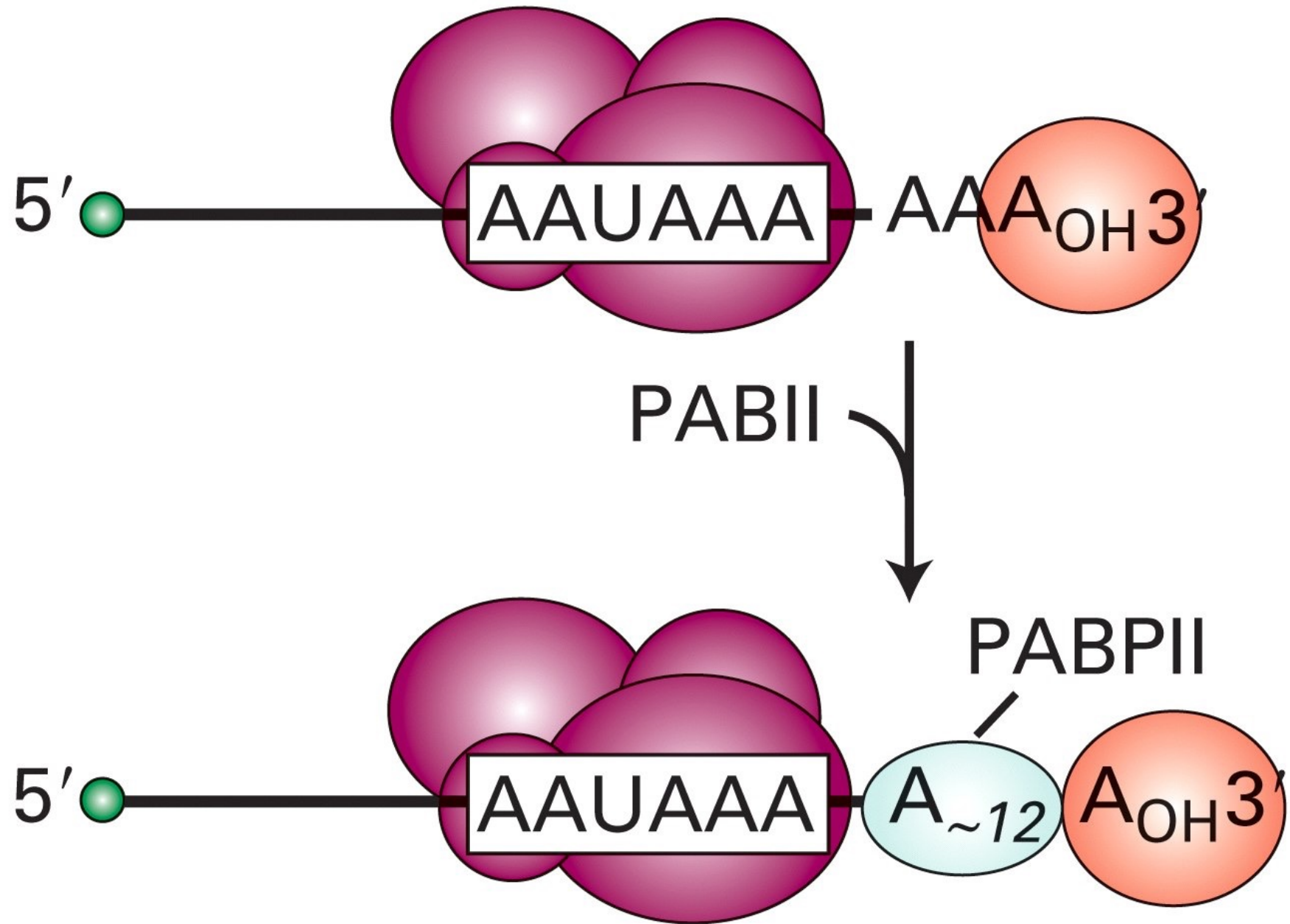


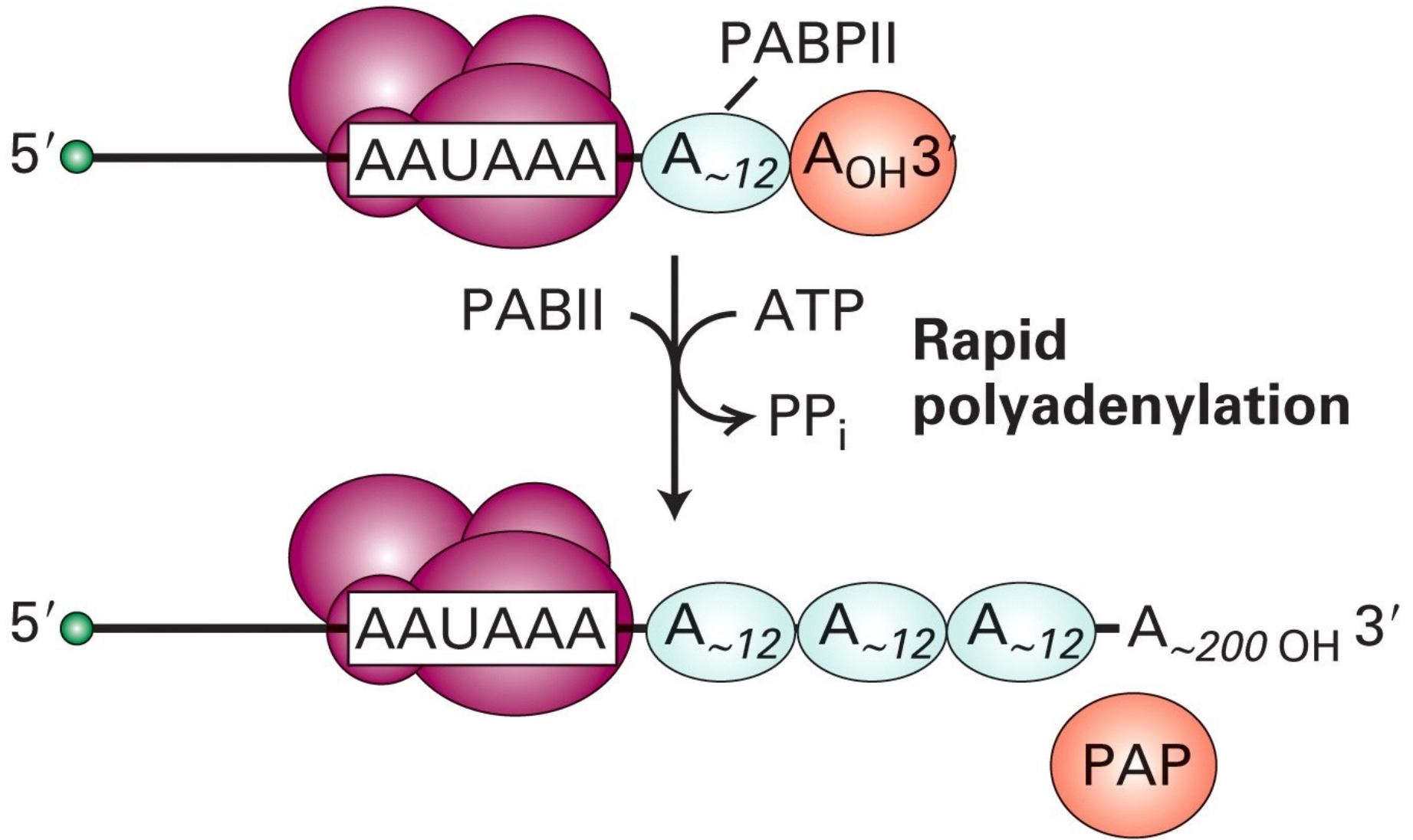




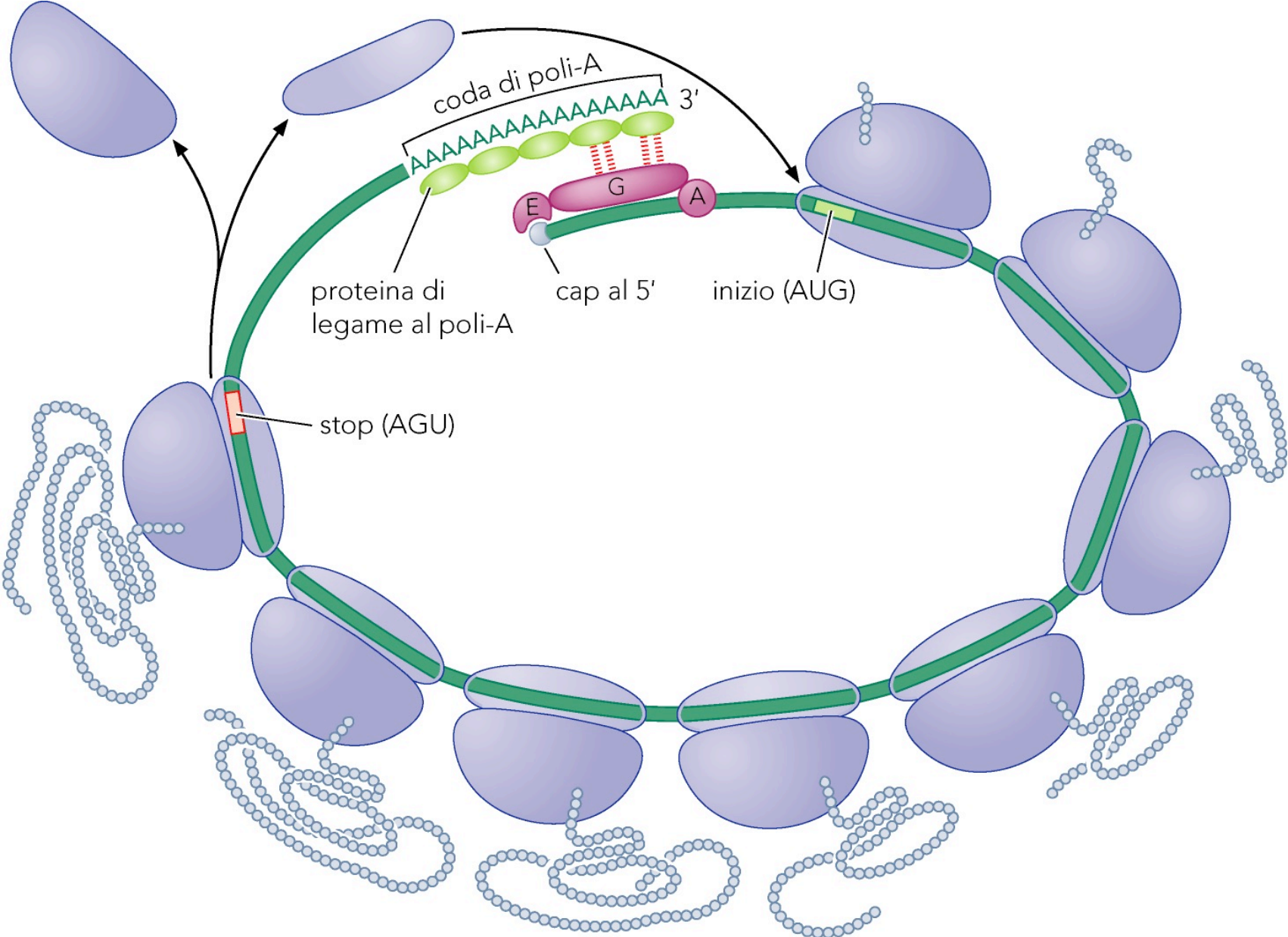






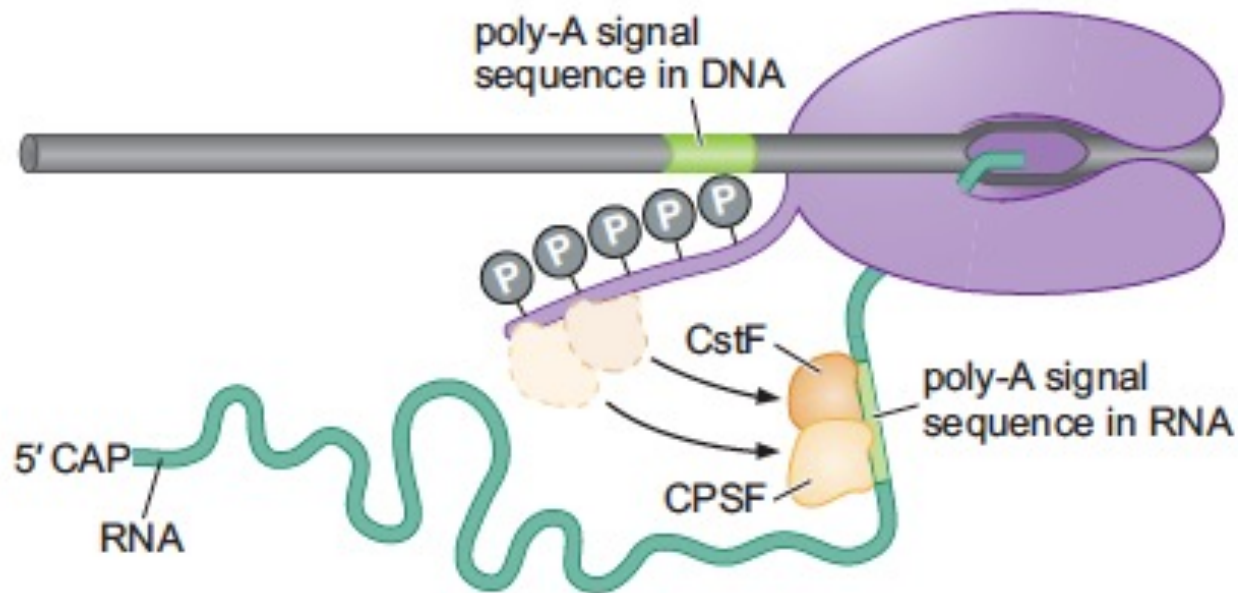


# CAP and polyA tail promote mRNA translation

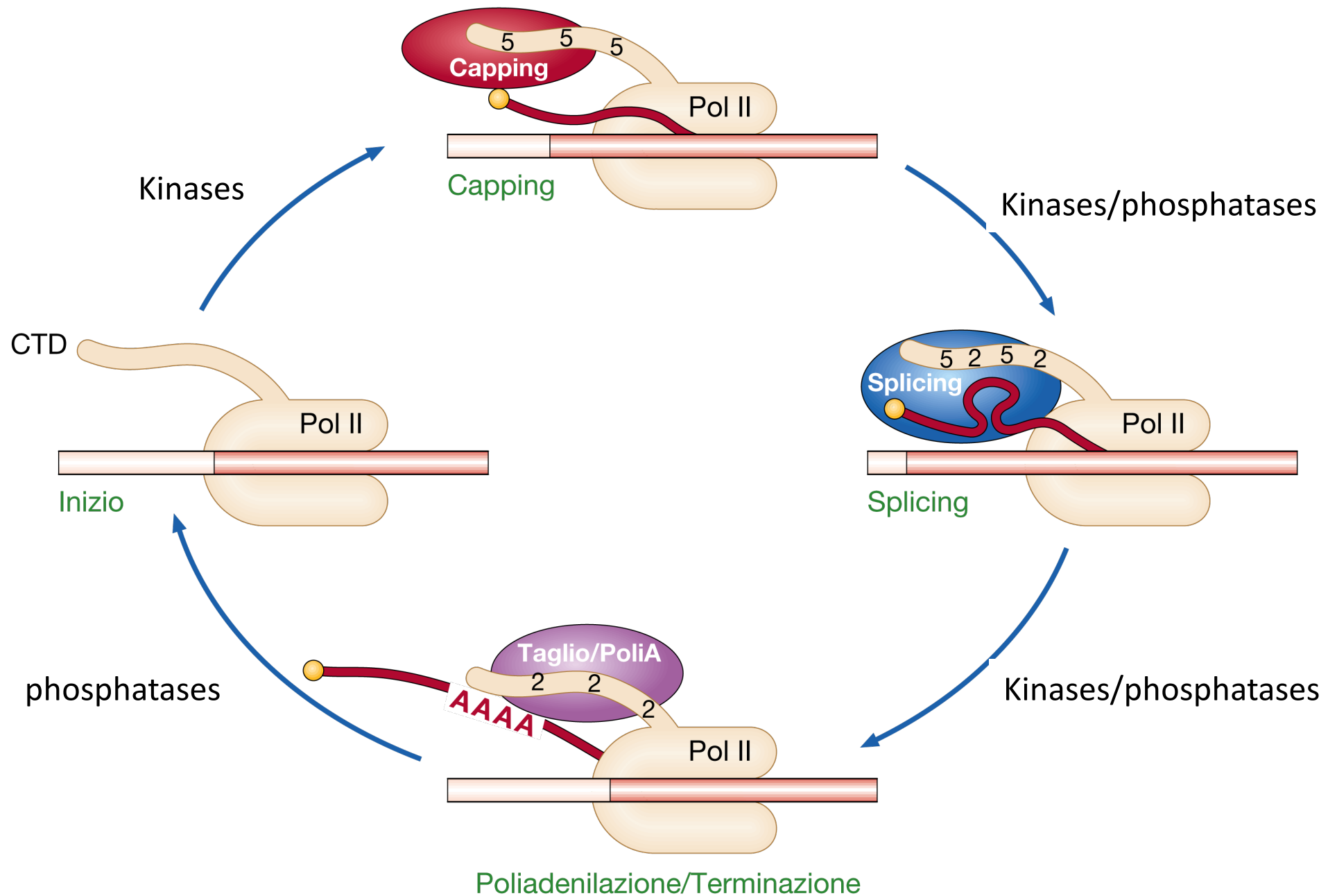


# 3'-end formation occurs during transcription

- TFIID associates with CPSF in the preinitiation complex (PIC)
- After transcription initiation CPSF dissociates from the PIC and associates with the CTD of elongating RNA Pol II together with CstF



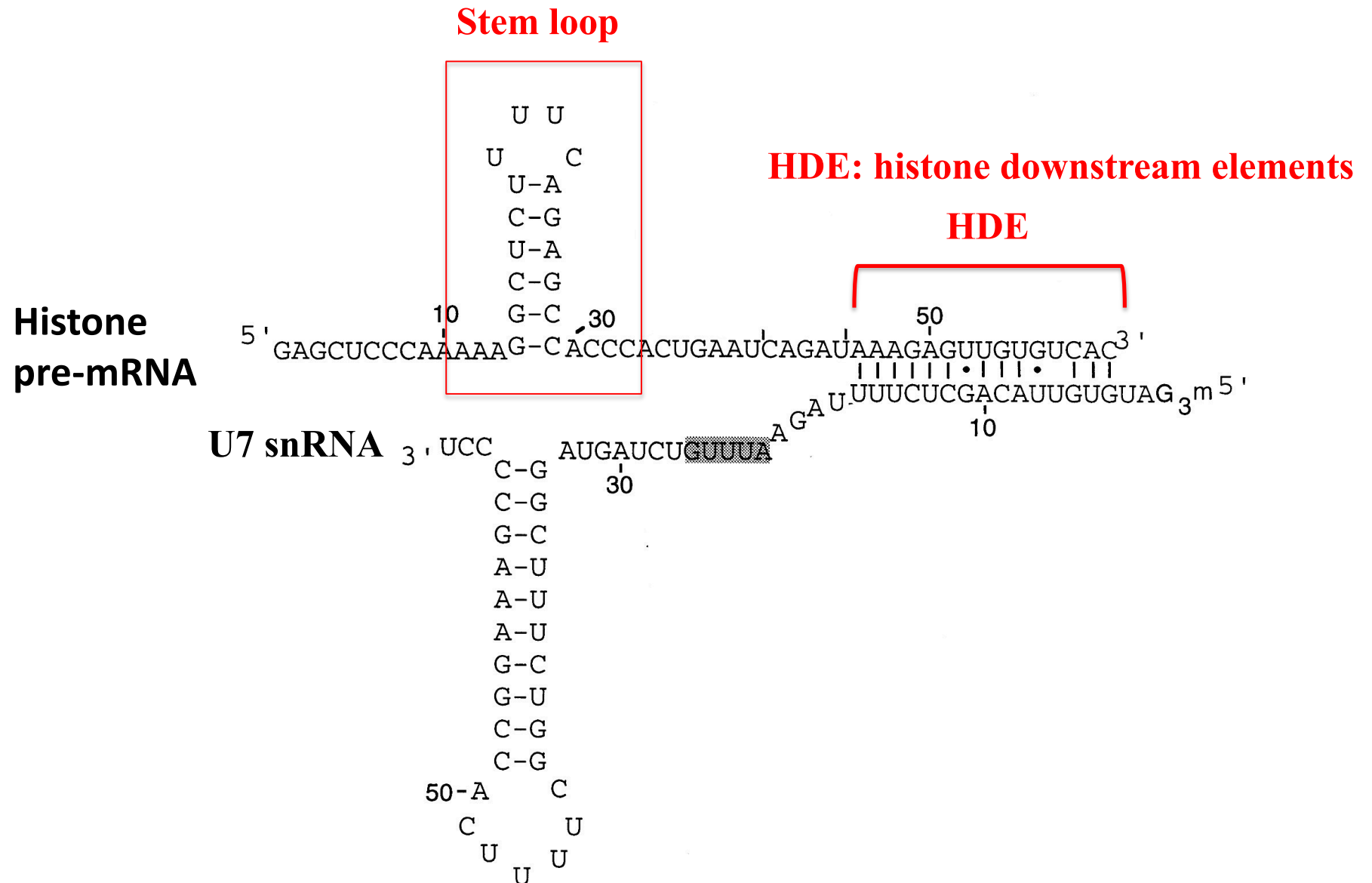
# capping, splicing and polyadenylation processes are tightly coupled with to transcription



# Characteristics of Histone Messenger RNA

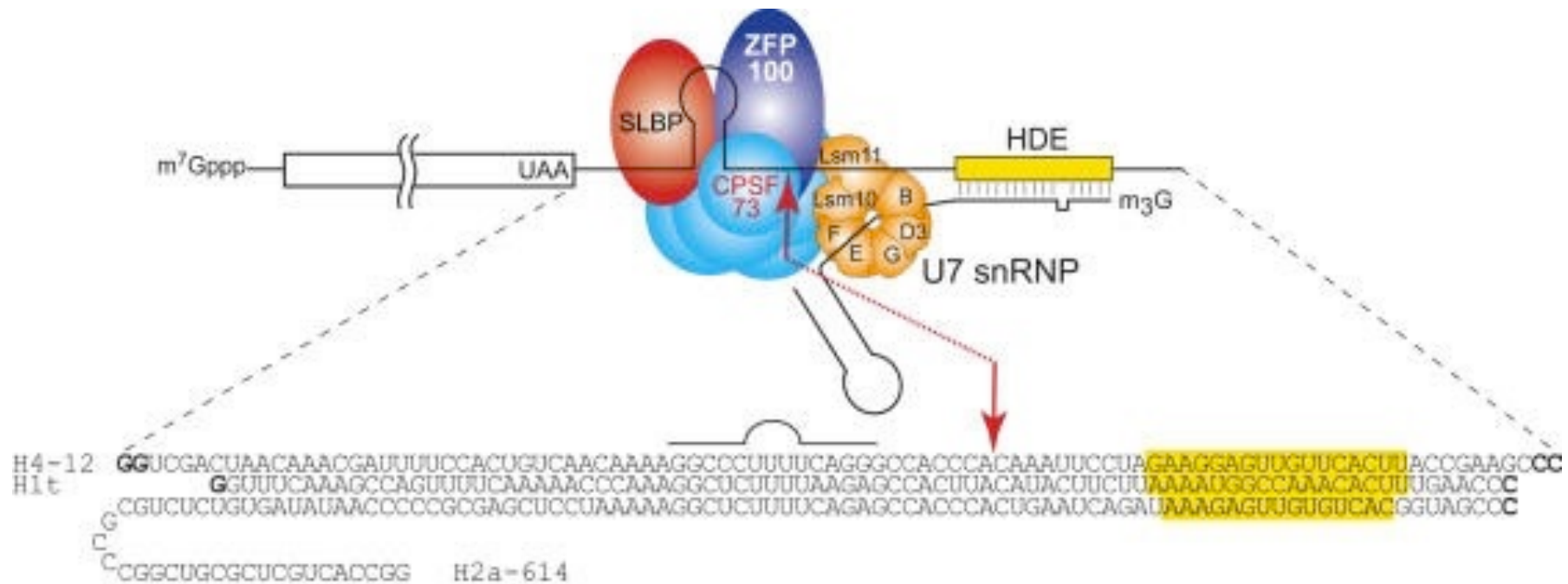
- Lacks introns: No splicing
- Lacks a polyA tail: cleavage not followed by polyadenylation
- Export to the cytoplasm is independent of the polyA tail
- Degradation occurs via uridylation
- Regulation of expression is dependent on the cell cycle

# The maturation of histone mRNA requires cis elements and trans factors, including U7 snRNA



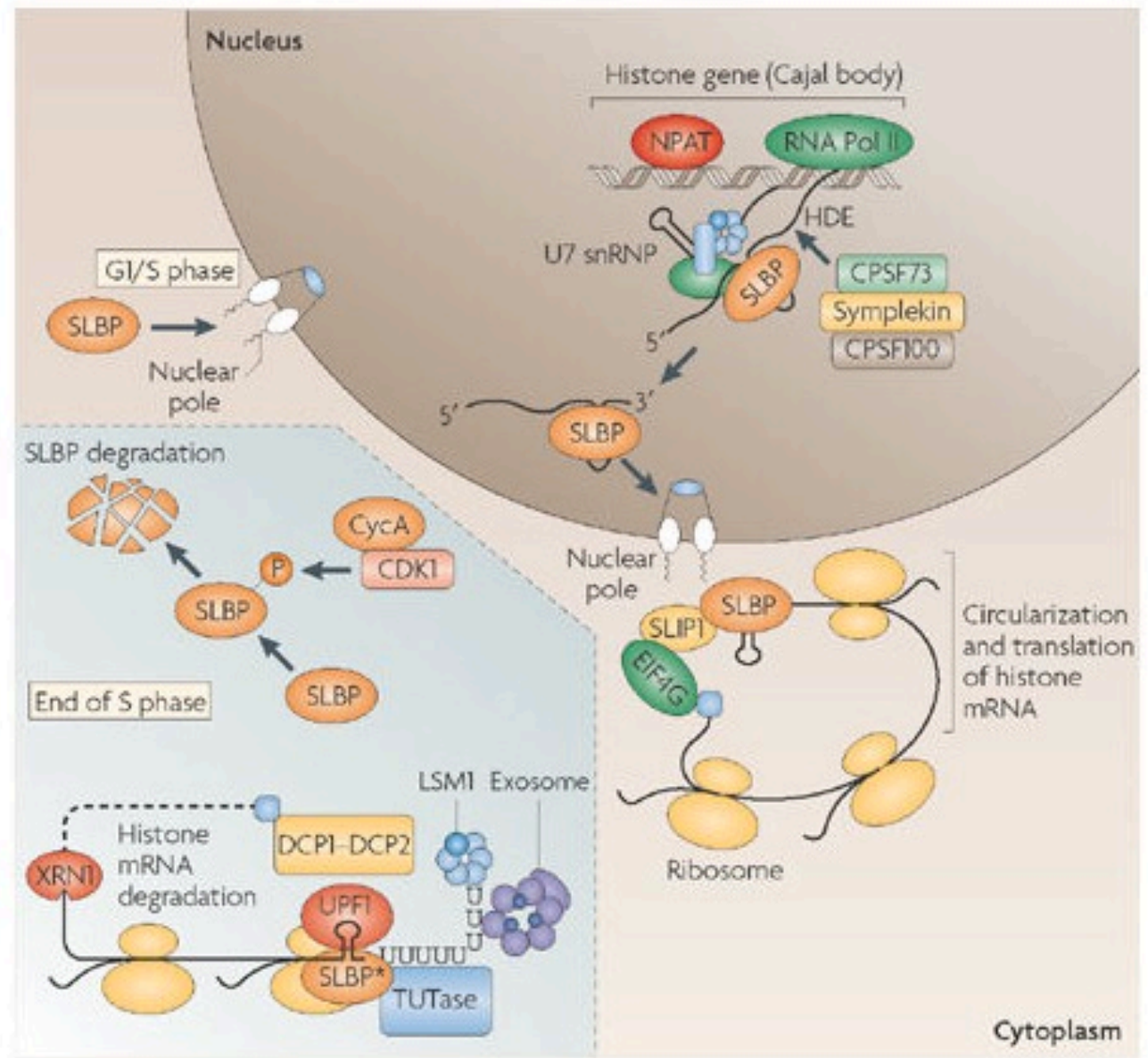


# Histone mRNA processing is polyadenylation independent



The U7 snRNP and stem-loop binding protein (SLBP), and the cleavage complex are responsible for the cleavage of the pre-mRNA from the DNA template, forming the mature histone mRNA

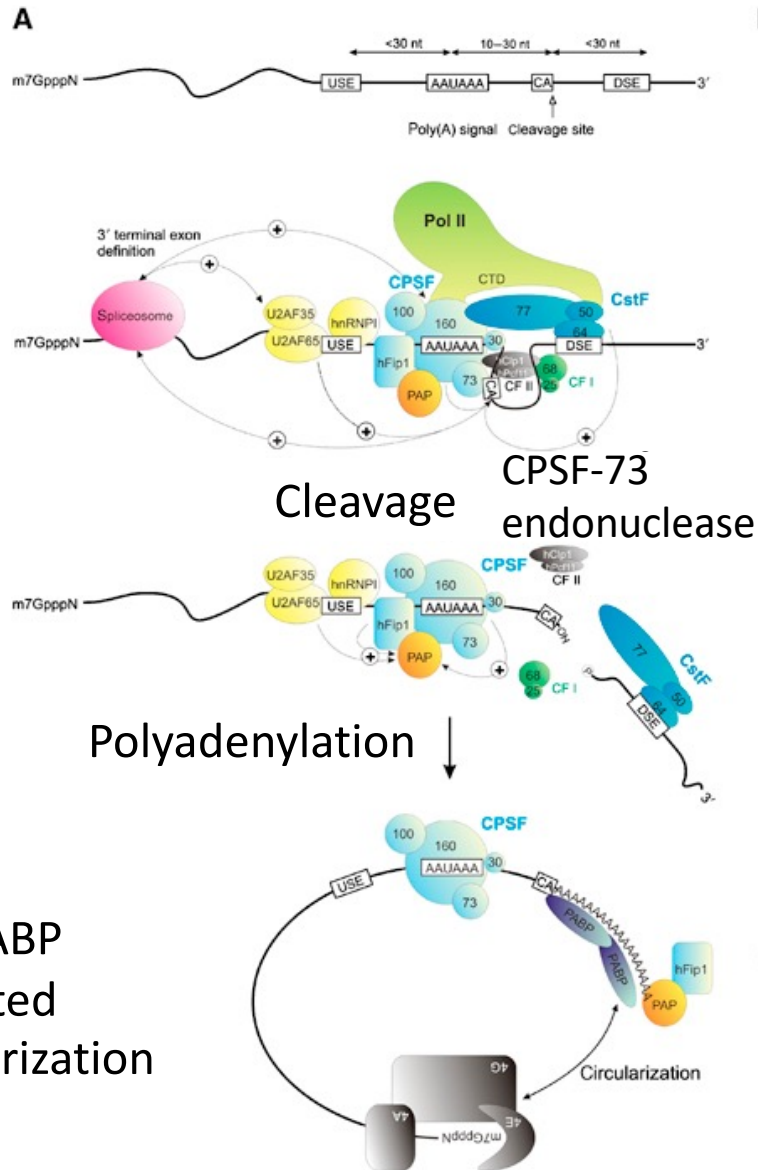
# Histone mRNAs: life without a poly(A) tail



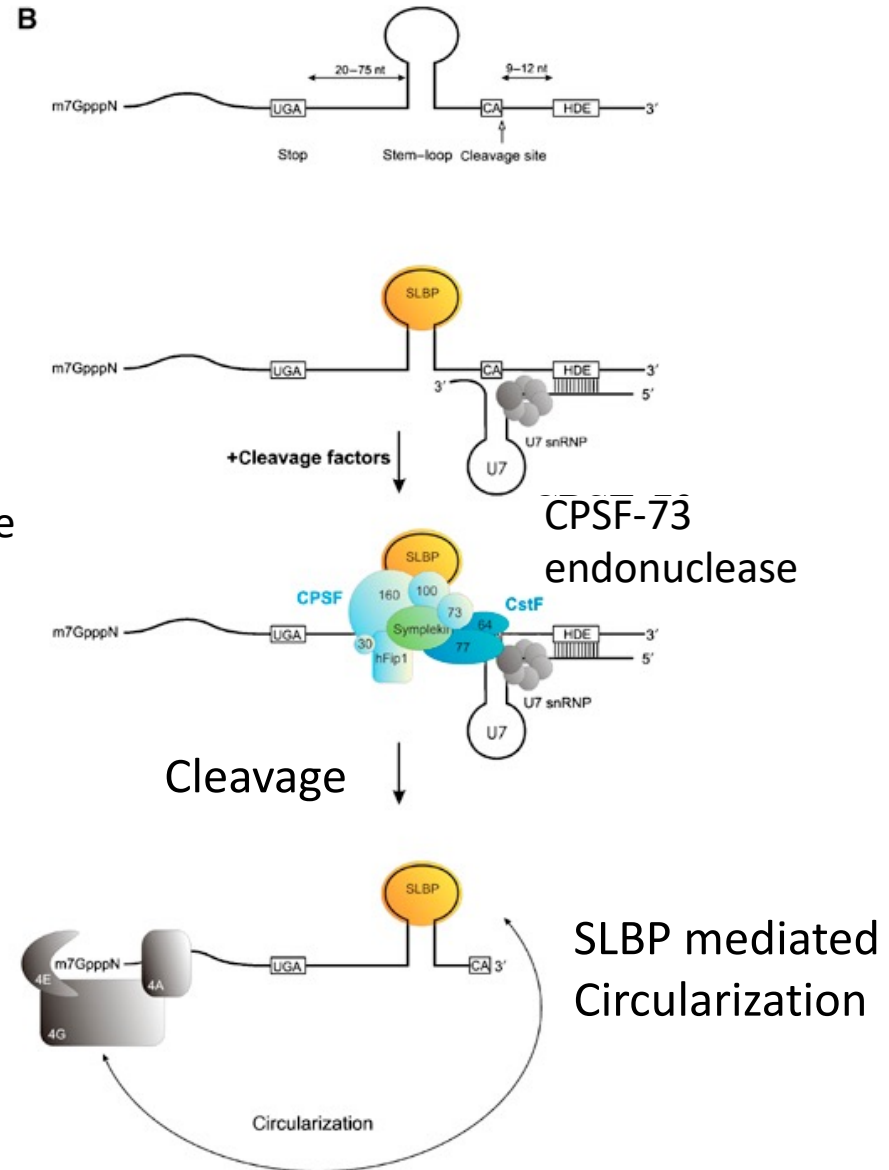
SLBP remains bound to the histone mRNA as it goes to the cytoplasm, where histone mRNA is circularized through a complex of proteins mediating translation of histone mRNA. At the end of S phase, a short U tail is added to histone mRNA in the cytoplasm. The LSM1–7 ring binds the oligo(U) to cooperate in the recruitment of the decapping complex and the exosome to degrade the mRNA. In addition, cyclin A (CycA)–CDK1 (cyclin-dependent kinase 1) phosphorylates SLBP to trigger its degradation at the end of S phase, preventing further histone mRNA synthesis.

# Processing and translation of mRNAs

## Poly(A)<sup>+</sup> mRNAs



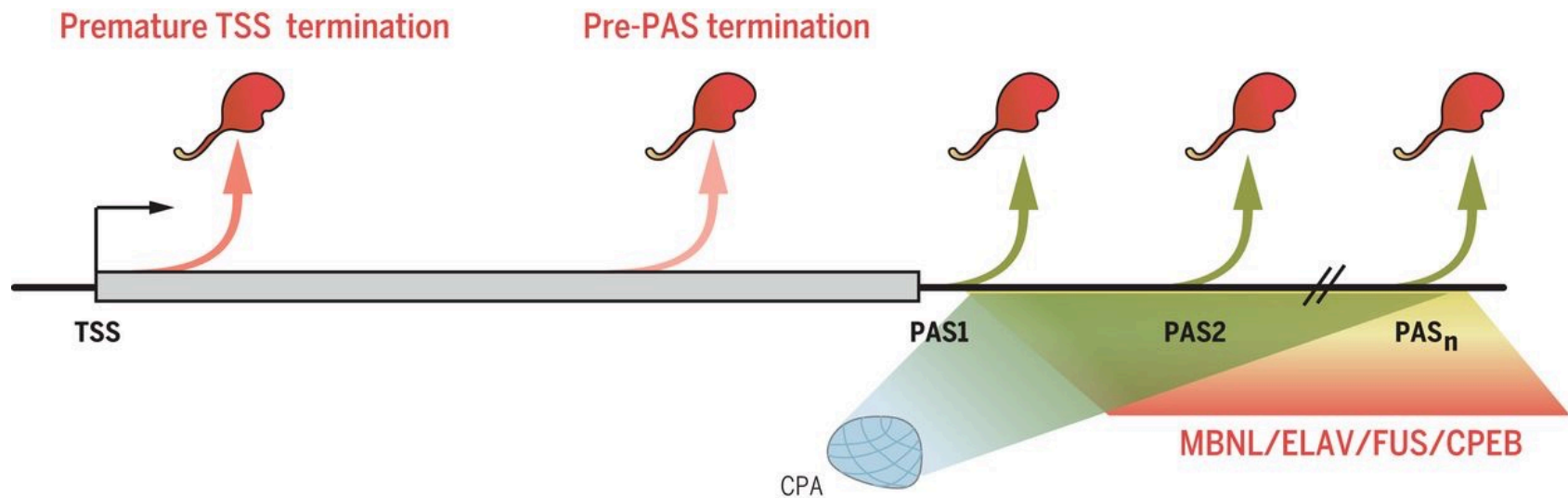
## Histone mRNAs



# RNA Pol II alternative 3'-end formation.

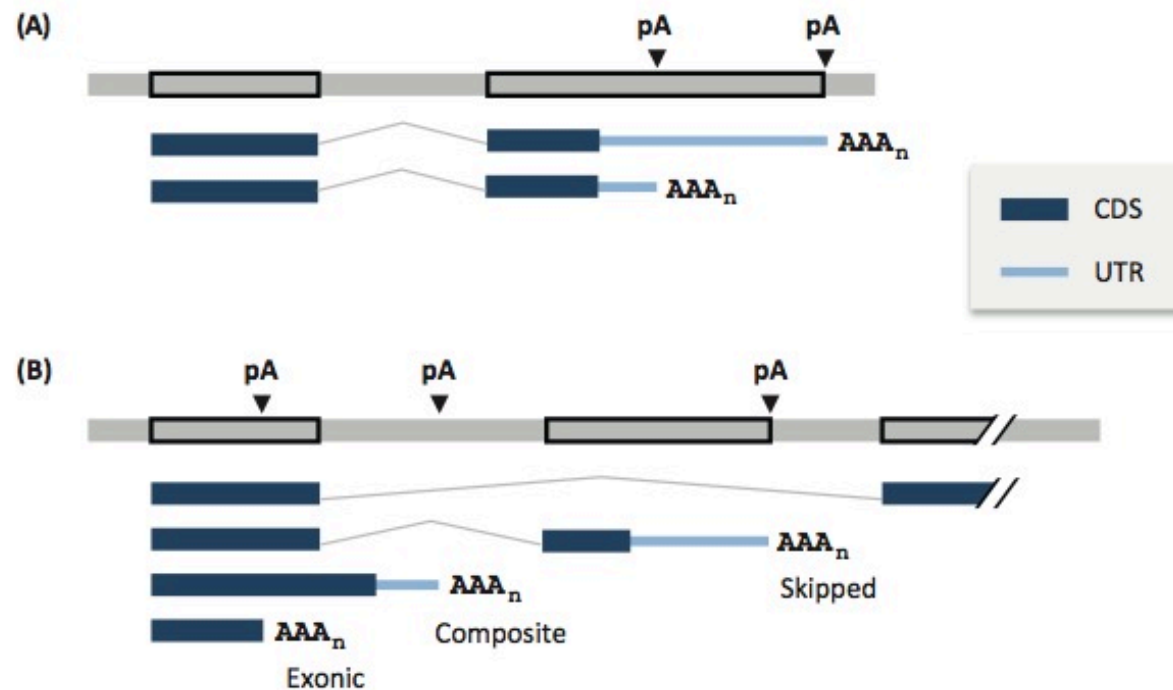
Termination at gene 3'-ends is also subject to intense regulation. Many mRNAs possess variable lengths of 3'-untranslated sequence defined by the selective usage of different PASs. Because mRNA 3'UTRs define mRNA cytoplasmic functions, the use of alternative poly(A) sites (APA) can constitute a key regulatory process in gene expression.

Polymerase speed and in trans elements can affect APA choice



# Alternative polyadenylation

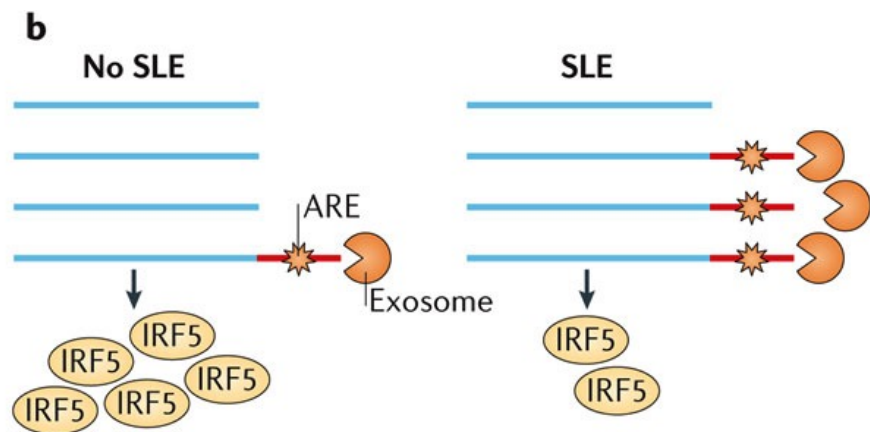
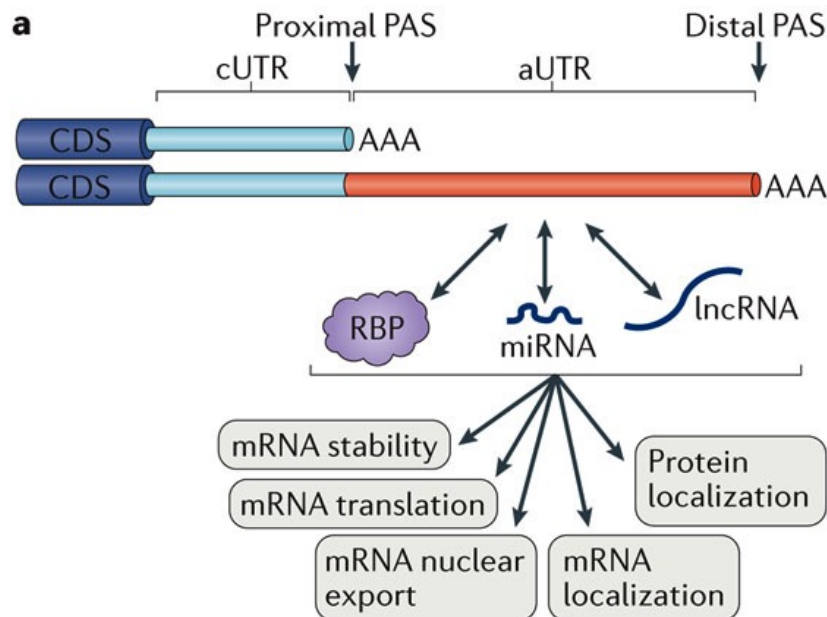
- 70–80% of mammalian genes have been reported to display alternative cleavage and polyadenylation site (APA) ,
- alternative pAs in the 3'-most exon (A) typically leads to variable 3' UTRs, whereas pAs in upstream introns and exons (B) cause both coding sequence (CDS) and 3' UTR changes,
- APA is dynamic under different biological conditions (cell growth and development, cancer)





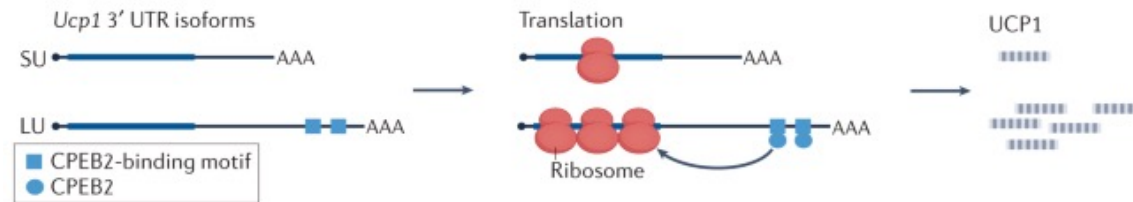
# Alternative polyadenylation

Most alternative polyadenylation (APA) sites are located in 3' UTRs. As 3' UTRs contain *cis* elements that are involved in various aspects of mRNA metabolism, 3' UTR-APA can considerably affect post-transcriptional gene regulation in various ways, including through the modulation of mRNA stability, translation, nuclear export and cellular localization, and even through effects on the localization of the encoded protein.

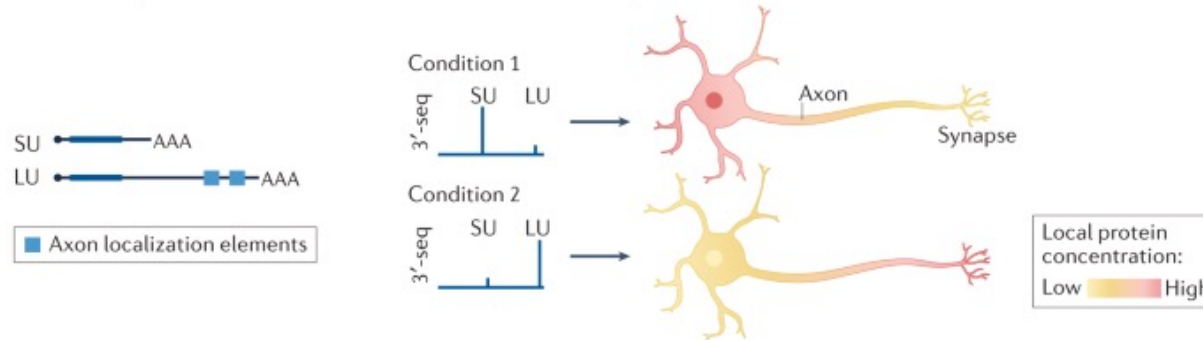


In patients with systemic lupus erythematosus (SLE), a single nucleotide polymorphism reducing the use of the proximal PAS leads to the production of long isoforms at the expense of short isoforms, which results in reduced IFN-regulatory factor 5 (IRF5) levels.

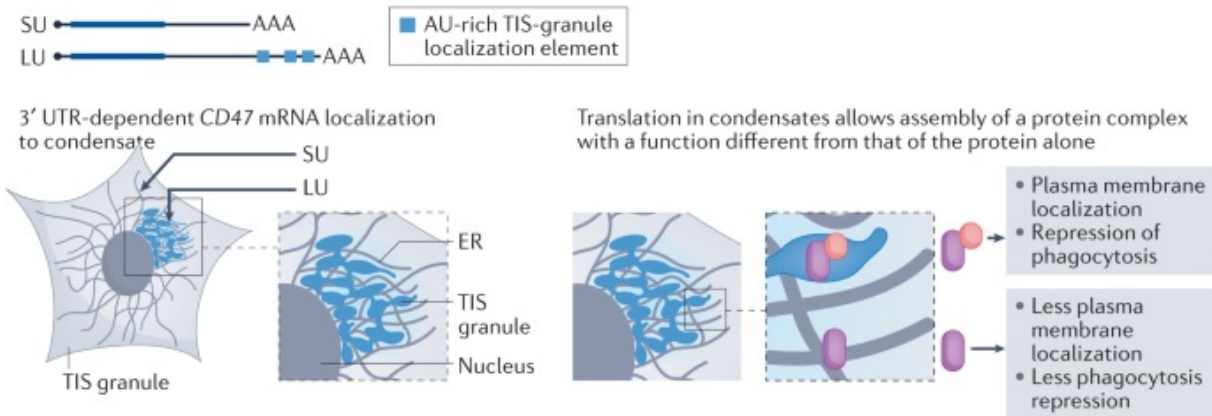
**a 3' UTR-dependent regulation of protein abundance**



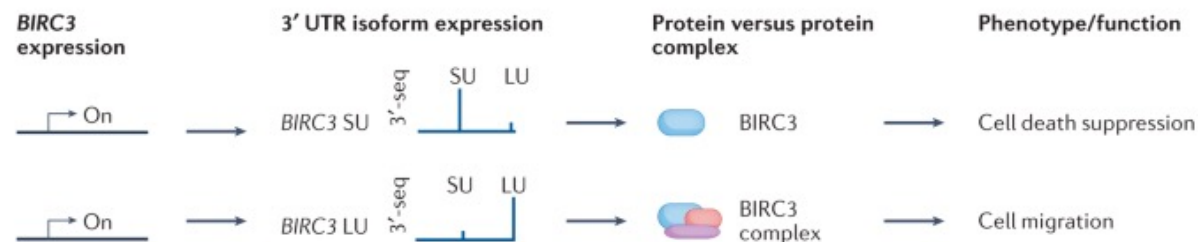
**b 3' UTR-dependent mRNA localization allows translation of proteins at their final destination**



**c 3' UTR-dependent protein complex assembly in condensates**



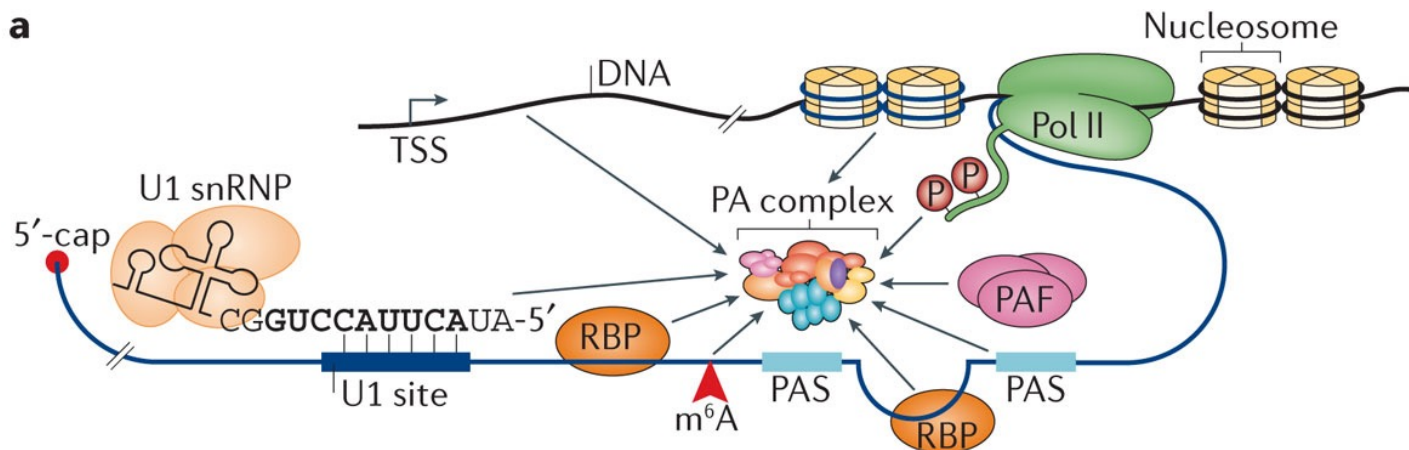
**d 3' UTR-dependent protein complex formation**





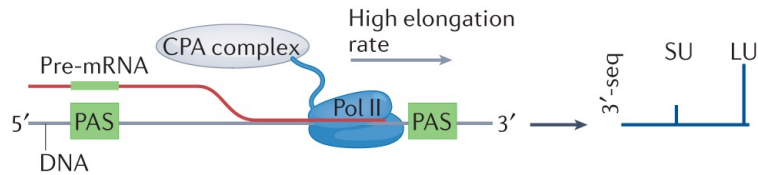
# Regulation of Alternative cleavage and polyadenylation

1. U1 can exert a negative role in pA usage
2. RNA binding proteins may inhibit pA usage by occluding the binding of core C/P factors, and some enhance pA usage by recruiting core factors.
3. Transcription activity impacts pA choice by:
  - **cis elements** that cause pausing of RNA polymerase (RNAP) II, such as G-rich elements, facilitate pA usage,
  - **factor recruitment** at the promoter can influence downstream pA usage.

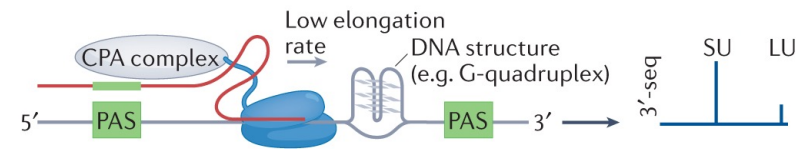


# Co-transcriptional regulators of alternative polyadenylation

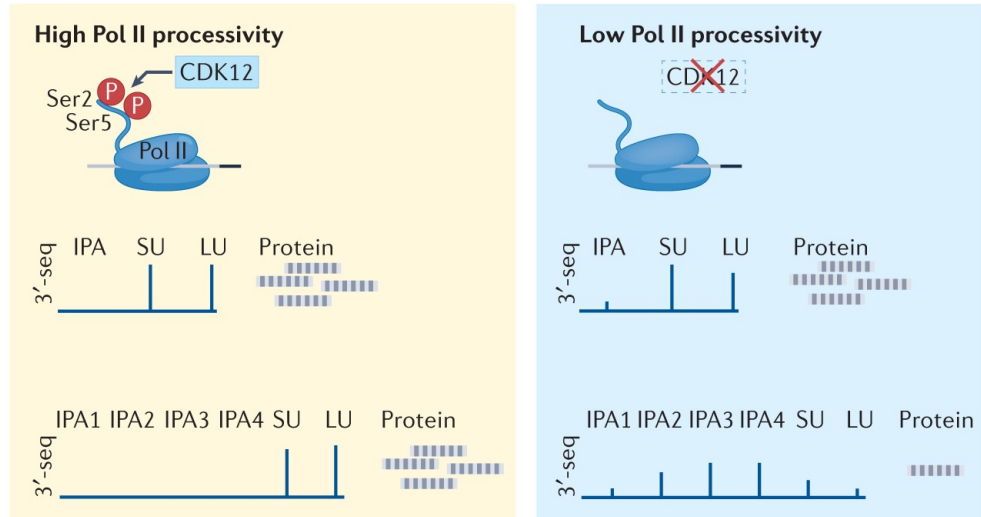
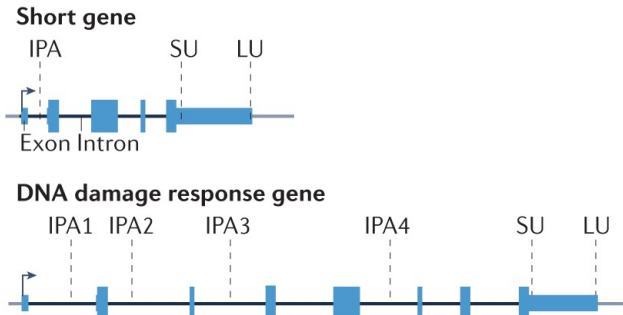
**a High transcript elongation rates promote full-length mRNA expression**



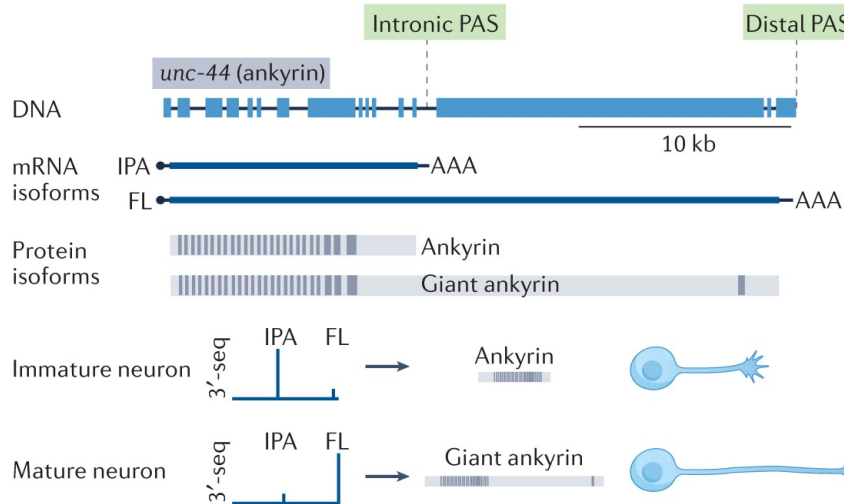
**Structured DNA templates create roadblocks that induce proximal PAS usage**



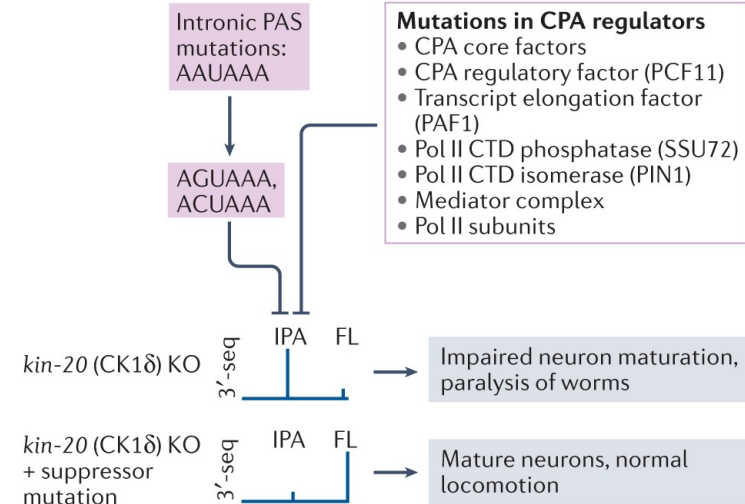
**b**



**c Physiological regulation of ankyrin APA**



**IPA suppressor screen**

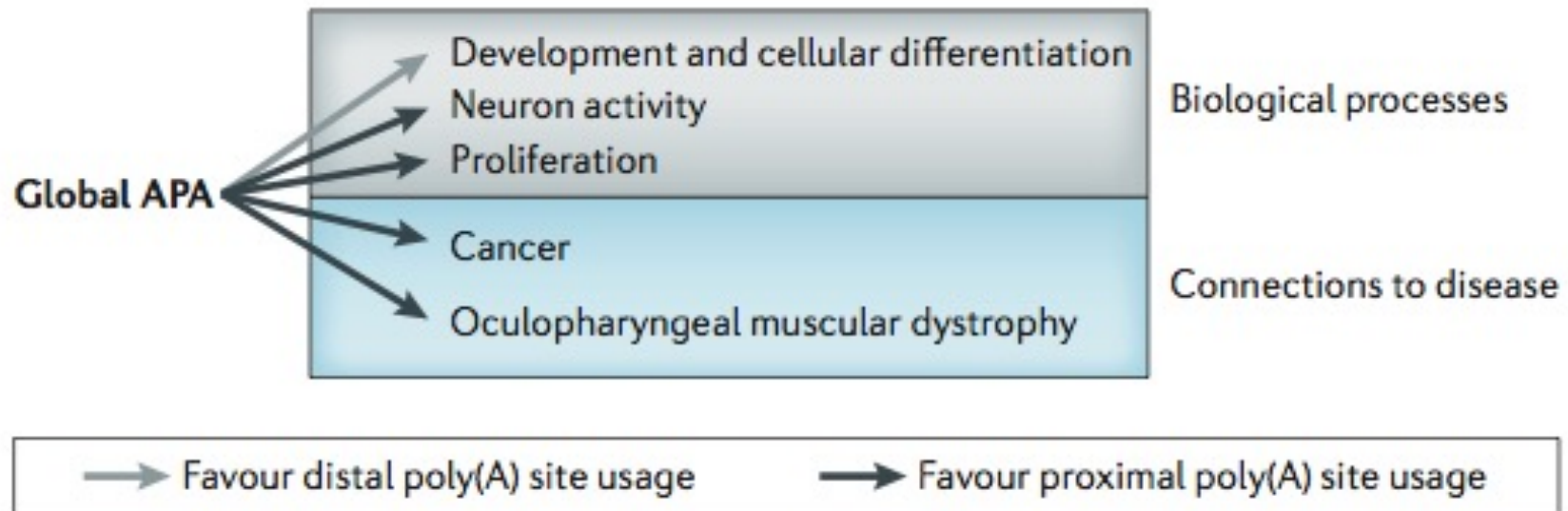


# Mutations in poly(A) *cis*-elements that cause or contribute to human diseases

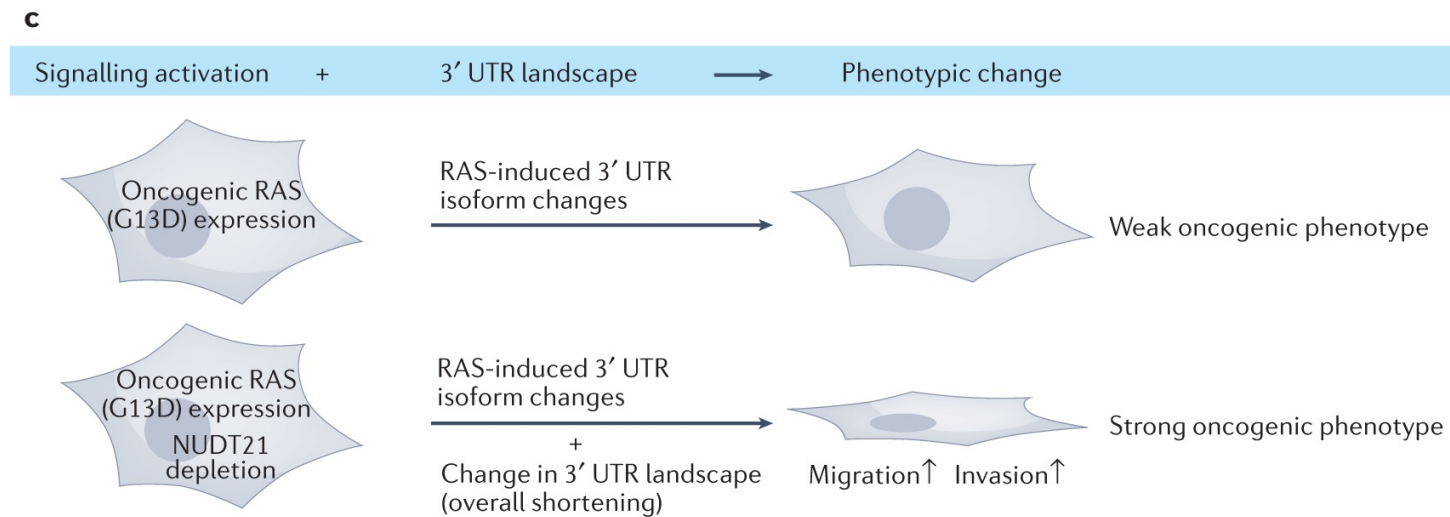
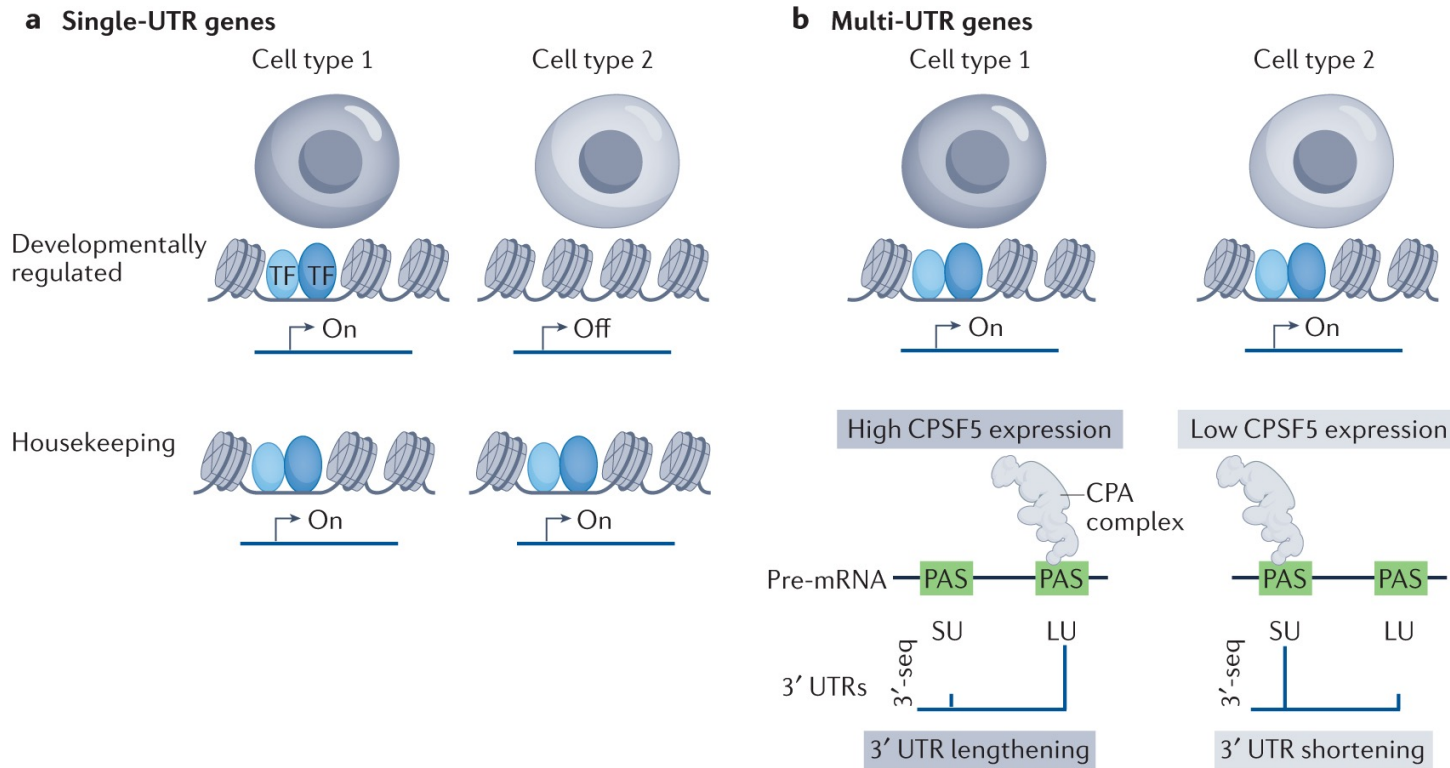
Disease	Disease description	Poly(A) site mutation	Affected gene	Comments	Refs
$\alpha$ -Thalassaemia	Thalassaemias are a group of common human genetic diseases that result from defects in haemoglobin production	AATAAA to AATAAG	<i>HBA2</i>		84
$\beta$ -Thalassaemia		AATAAA to AACAAA	<i>HBB</i>	The mutation results in generation of an unstable transcript that is ~900nt longer	85
Metachromatic leukodystrophy	A neurodegenerative disorder caused by null mutations in <i>ARSA</i>	AATAAC to AGTAAC	<i>ARSA</i>	Carriers of this mutation show reduced mRNA levels and enzyme activity but do not develop the disease symptoms; this condition is termed 'pseudodeficiency'	86,87
IPEX	A rare multifaceted and fatal disease	AATAAA to AATGAA	<i>FOXP3</i>		88
Fabry's disease	A rare and severe X-linked lysosomal storage disease caused by mutations in <i>GLA</i>	An AA dinucleotide deletion within the poly(A) site	<i>GLA</i>	<i>GLA</i> is one of the unusual mammalian genes that lack a 3'UTR and has its PAS within the CDS. The mutation in the poly(A) site results in aberrant 3' end formation and multiple unstable transcripts	89

*ARSA*, arylsulfatase A; CDS, coding DNA sequence; *FOXP3*, forkhead box P3; *GLA*, alpha-galactosidase; *HBA2*, haemoglobin, alpha 2 (also known as alpha-2-globin); *HBB*, haemoglobin, beta (also known as beta-globin); IPEX, immune dysfunction, polyendocrinopathy, enteropathy, X linked; PAS, poly(A) signal; UTR, untranslated region. Only selected examples are discussed here. For a more thorough discussion, see REF. 64.

# Biological processes that have been linked with broad APA modulation

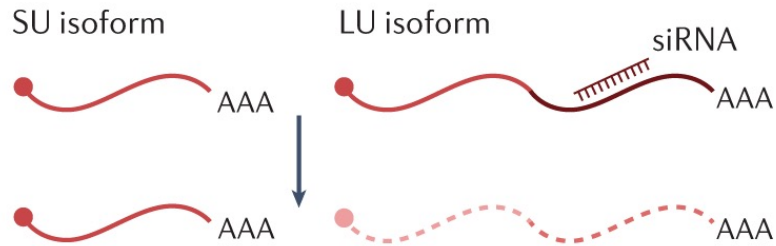


# Biological processes that have been linked with broad APA modulation

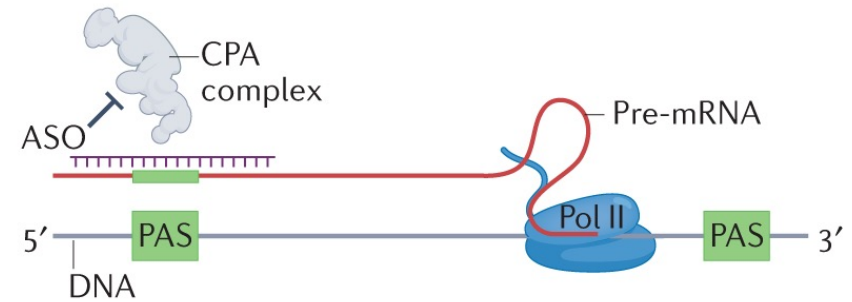


# Manipulation of alternative 3' UTR expression.

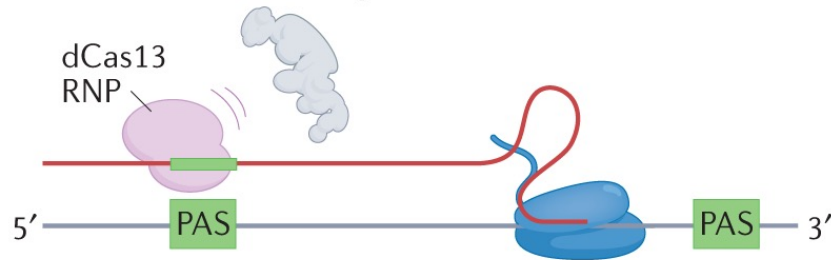
**a Selective siRNAs: degradation of LU isoforms**



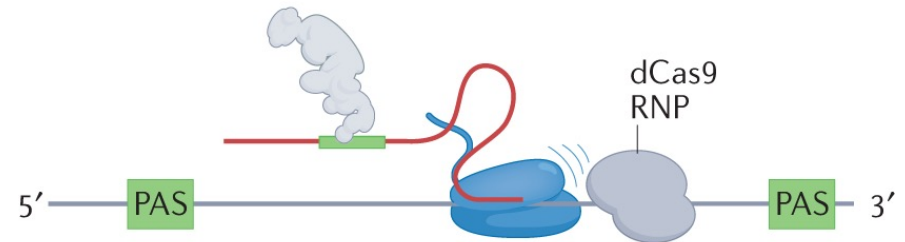
**b ASOs: masking of proximal PAS**



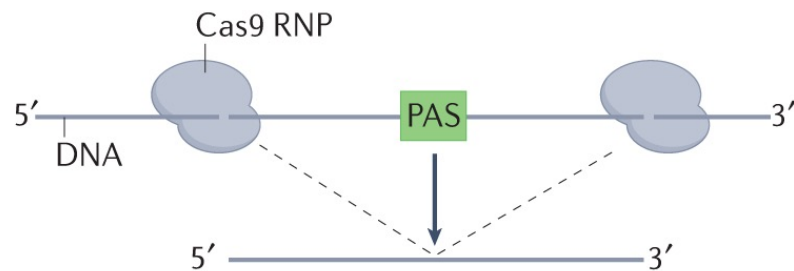
**c CRISPR-iPAS: masking of PAS**



**d CRISPRpas: roadblock for Pol II**



**e KO of 3' UTR isoforms through CRISPR-mediated PAS deletion**



**f KO of 3' UTR isoforms through CRISPR-mediated PAS mutation**

