

# Molecular Dynamics Simulations

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# 1 Introduction

## *Experimental techniques*

***X-ray, NMR***  
(dynamics and structure)

***Light/X-ray/neutron scattering***  
(dynamics and structure)

***Imaging/Cryo-EM***  
(dynamics and structure)

***Calorimetry, pKas,  
thermodynamics,  
physical measurements***

**Development of new  
theories and models  
to rationalize and  
predict experimental  
observations**

## *Theoretical methods*

***Development of  
mathematical  
models  $v(r)$***

***Development of  
methods to  
explore models***

***Exploration of model  
phenomenology and  
properties***

***Understanding biomolecular  
structure, dynamics and function***

## 2 Force field

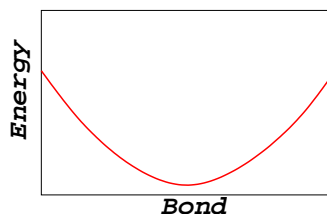
- A force field is an empirical approximation for expressing structure-energy relationships in biopolymers
- It is a compromise between speed and accuracy
- Common form (CHARMM):

$$\begin{aligned} E(\mathbf{r}_1, \mathbf{r}_2, \dots, \mathbf{r}_N) &= \sum_{bonds} \frac{1}{2} k_i^b (d_i - d_i^0)^2 \\ &+ \sum_{angles} \frac{1}{2} k_i^\theta (\theta_i - \theta_i^0)^2 \\ &+ \sum_{torsions} k_i^\phi [1 + \cos(n_i \phi_i - \delta_i)] \\ &+ \frac{1}{2} \sum_{nonbond} \left( \epsilon_{ij}^{min} \left[ \left( \frac{d_{ij}^{min}}{d_{ij}} \right)^{12} - 2 \left( \frac{d_{ij}^{min}}{d_{ij}} \right)^6 \right] + \frac{q_i q_j}{\epsilon d_{ij}} \right) \end{aligned}$$

## 2.1 Energy terms

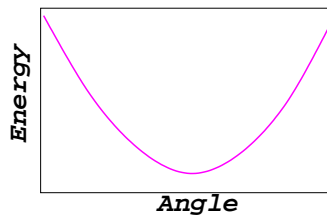
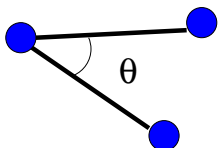
From spectroscopy, IR, NMR

**Bonds**



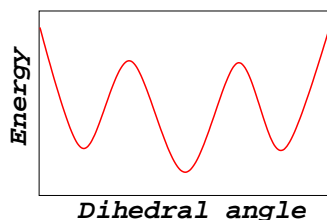
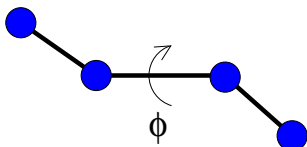
$$v_{bond} = \frac{1}{2}k_i^b(d_i - d_i^0)^2$$

**Angles**



$$v_{angle} = \frac{1}{2}k_i^\theta(\theta_i - \theta_i^0)^2$$

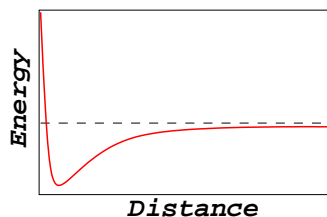
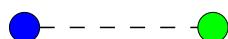
**Dihedrals**



$$v_{dihedral} = k_i^\phi[1 + \cos(3\phi_i)] + k_i^{\phi'}[1 - \cos(\phi_i - \pi)]$$

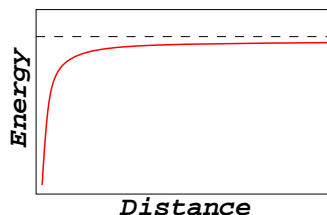
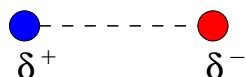
From quantum chemistry, thermodynamics

**Van der Waals**



$$v_{vdW} = \epsilon_{ij}^{min} \left[ \left( \frac{d_{ij}^{min}}{d_{ij}} \right)^{12} - 2 \left( \frac{d_{ij}^{min}}{d_{ij}} \right)^6 \right]$$

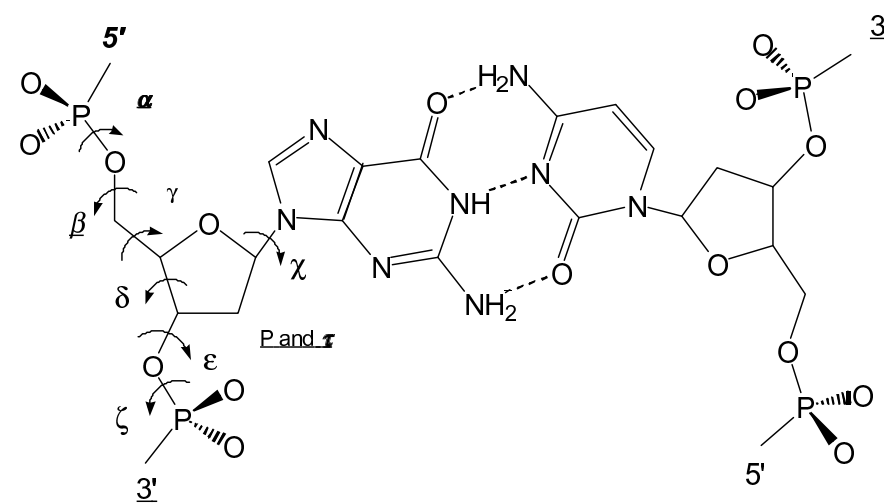
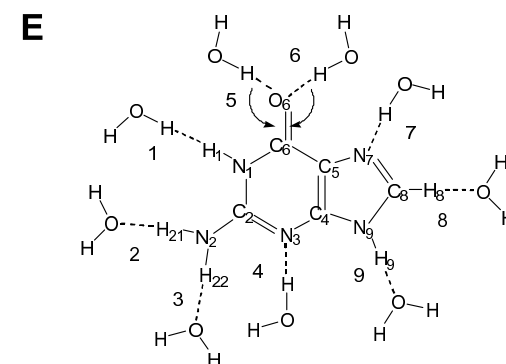
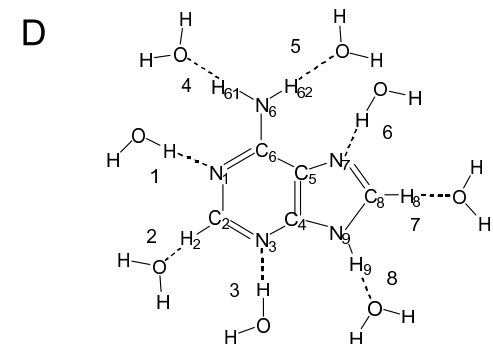
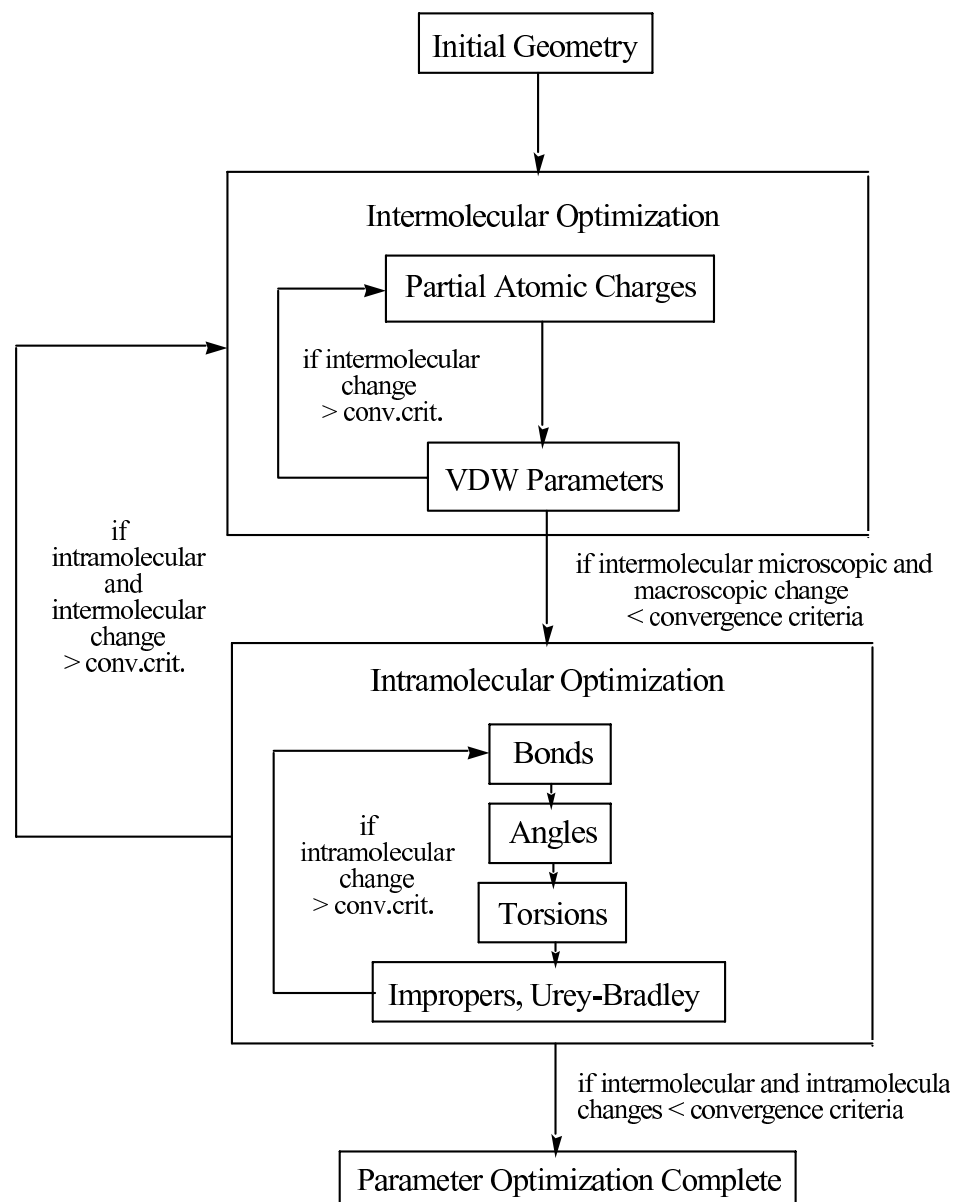
**Electrostatics**



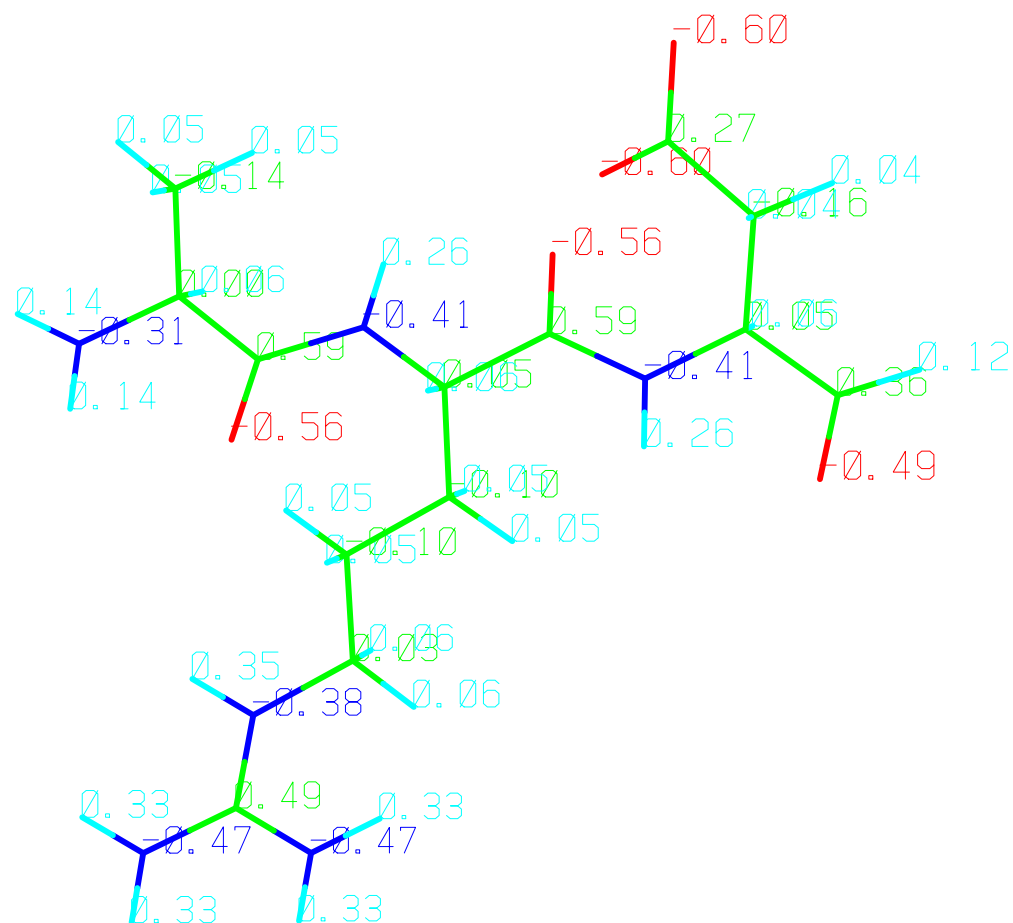
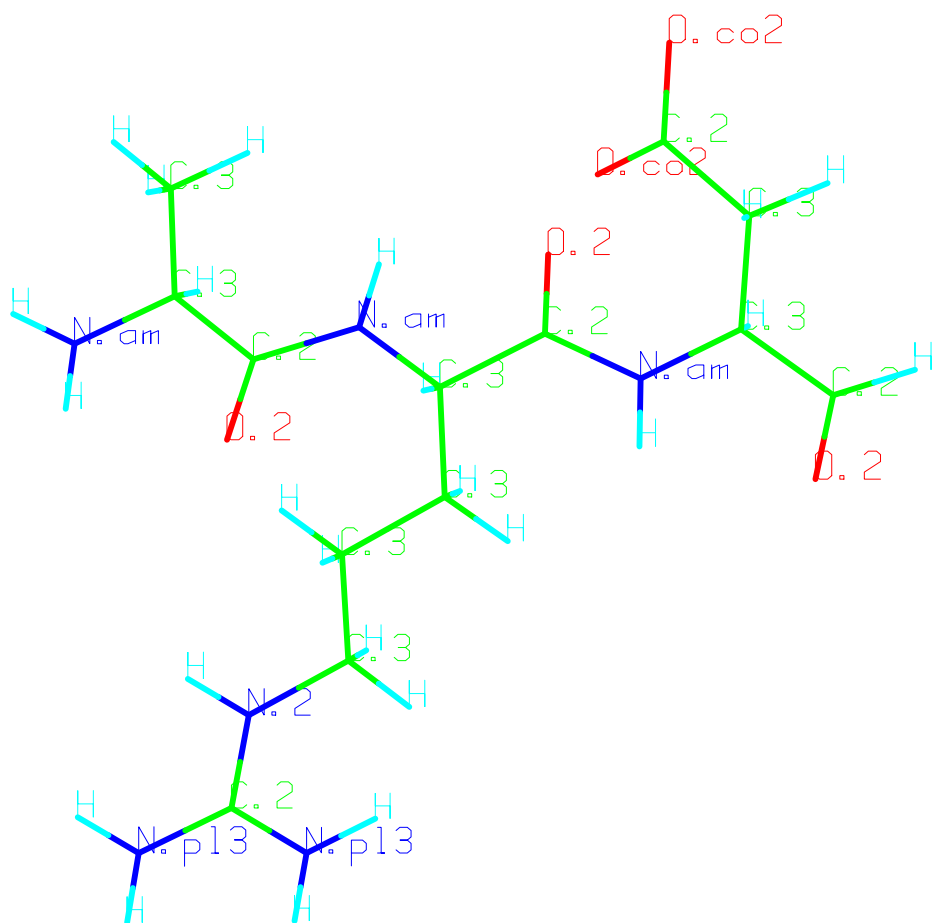
$$v_{Coulomb} = \frac{q_i q_j}{\epsilon d_{ij}}$$

## 2.2 Parametrization

MacKerell, Parameter Notes



### 2.3 Atom types and partial charges

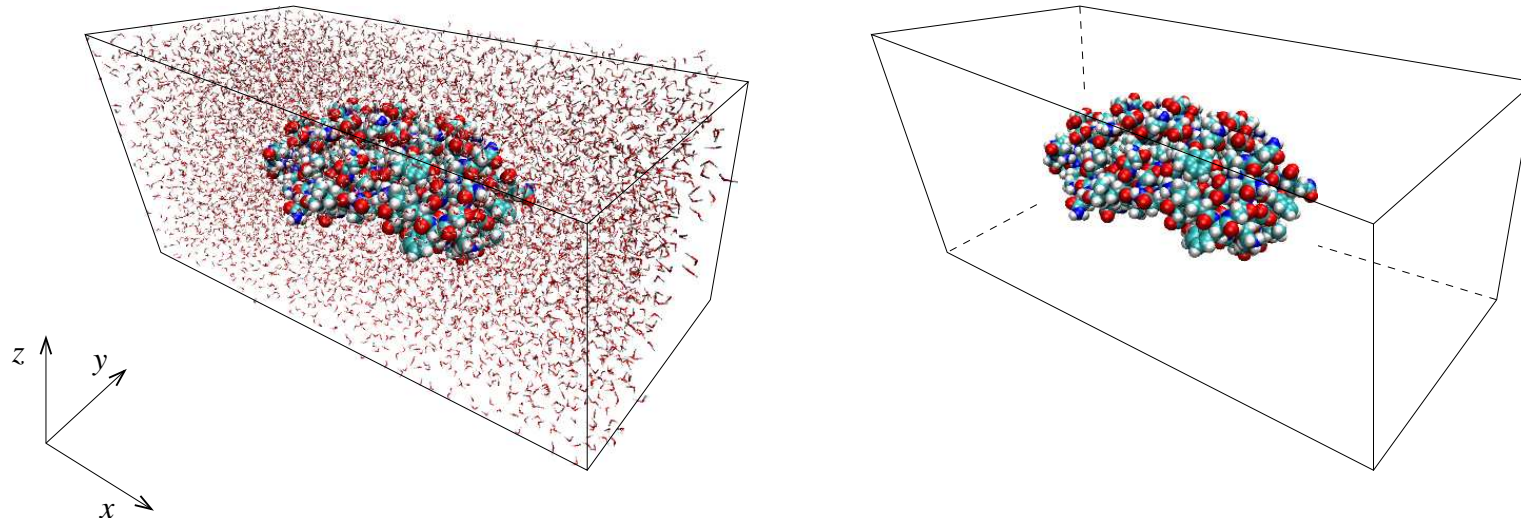


## 2.4 Models for simulation studies

- How similar is the (macro)molecular system being studied to systems that were employed in the development of the model?
- The generality (i.e., range of possible applications) of a given model can only be established by comparison to experimental data for a wider and wider variety of (macro)molecular systems.
- Example: protein force field parameters are developed using small compounds which represent the peptide bond (e.g., N-methyl-acetamide) and the functional groups of the side chains of the natural amino acids (e.g., propane for Val, benzene for Phe, imidazole for His, indole for Trp, etc). Hence, it is straightforward to determine the parameters for a nor-leucine side chain ( $-\text{C}_\beta-\text{C}_\gamma-\text{C}_\delta-\text{C}_\epsilon$ ) but not for a phosphotyrosine because a protein force field does not usually have parameters for phosphate.
- Some properties are more amenable to accurate computation than others. Example: Energy minimization to obtain refined structures is much simpler than calculating free energy differences.

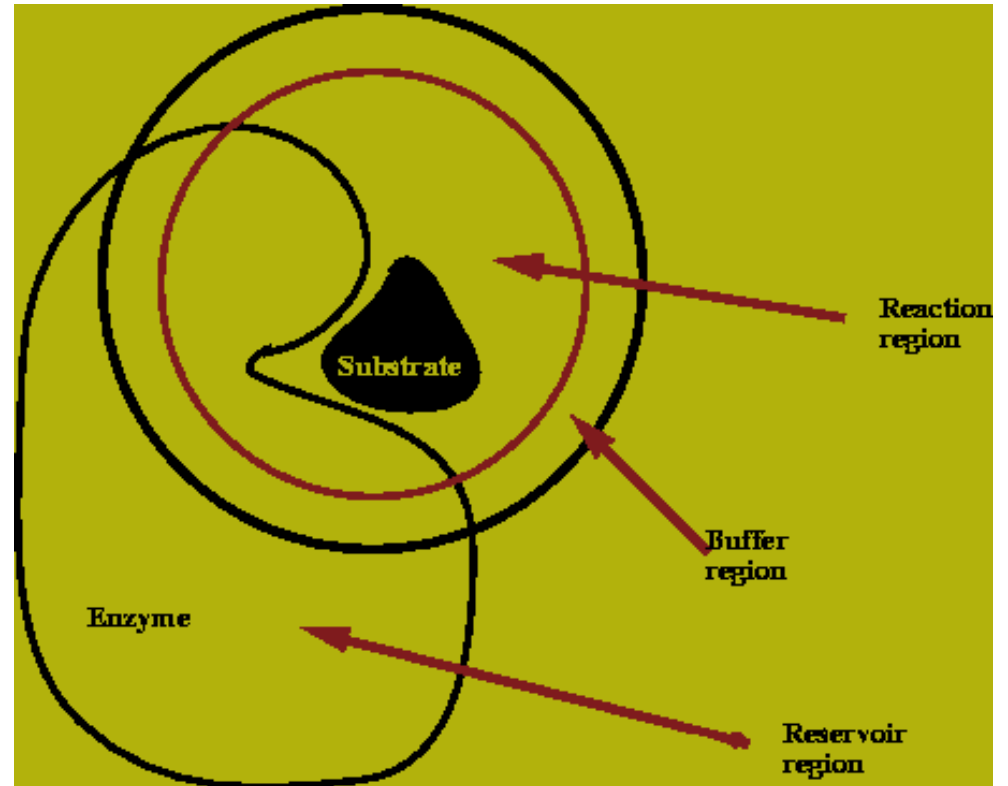
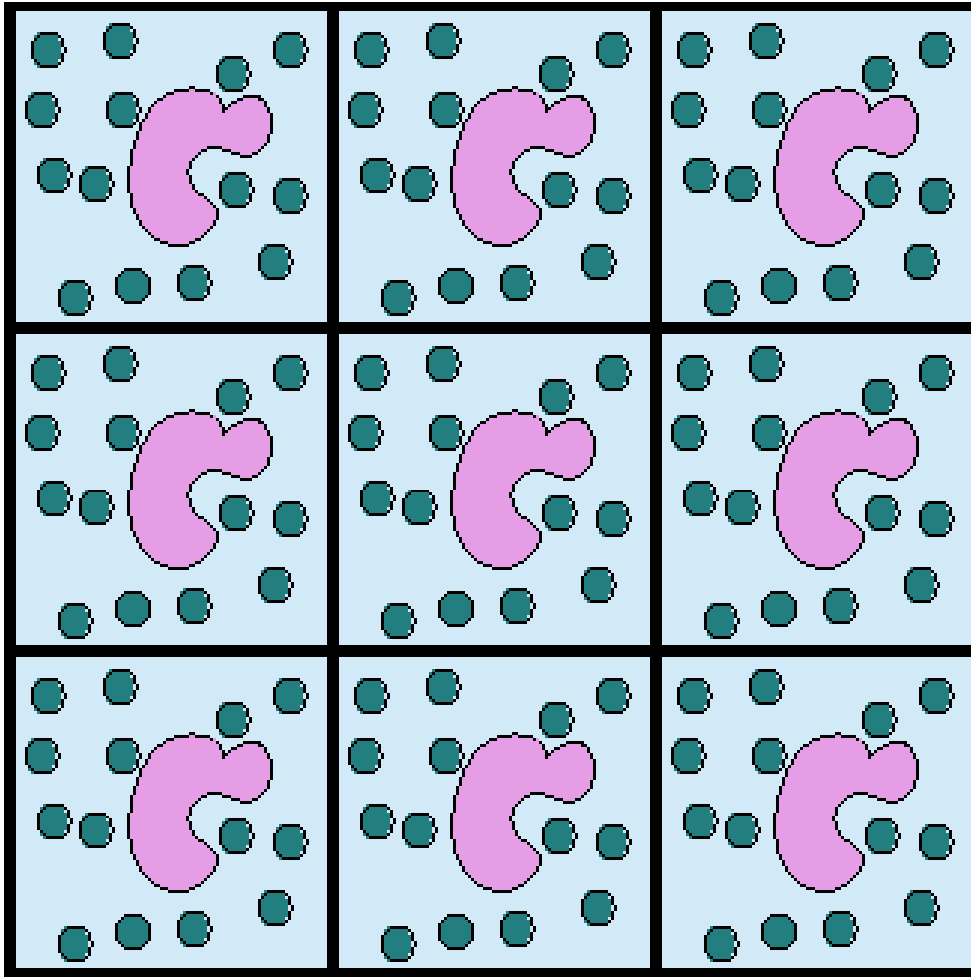


### 3 Solvation models

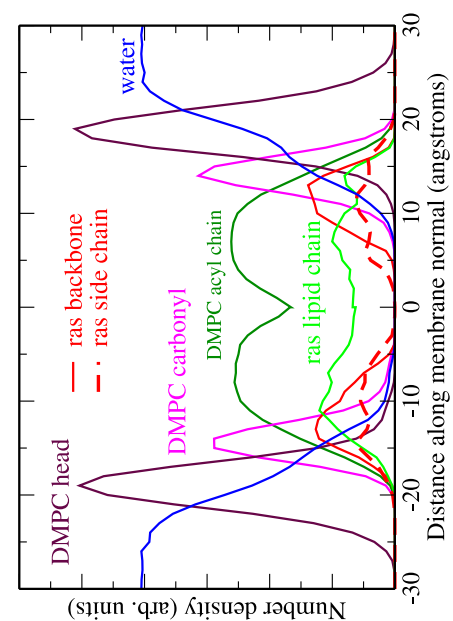
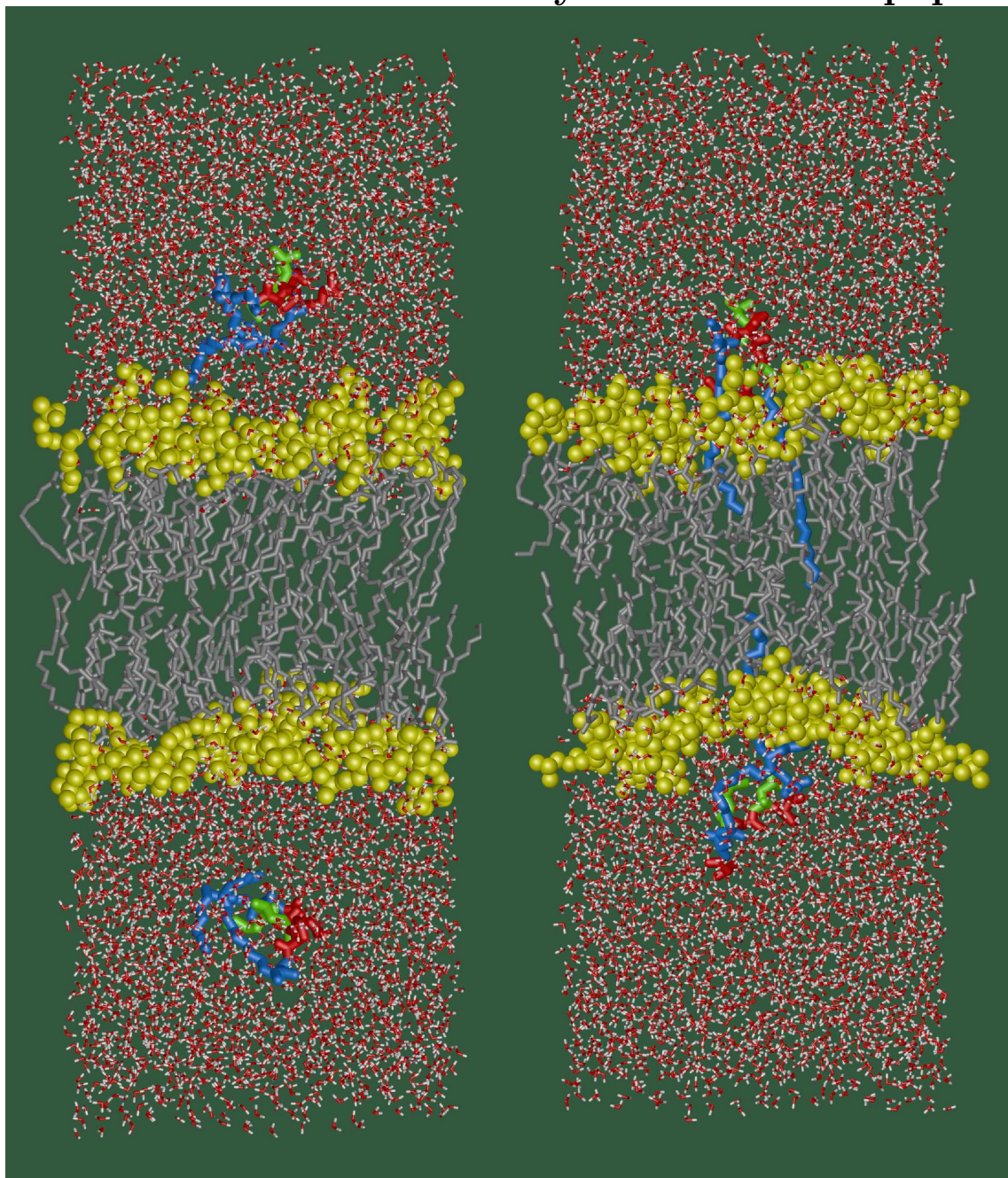


- Simulations *in vacuo* suffer from serious artifacts.
- Implicit solvation models reduce computational costs by removal of H<sub>2</sub>O interaction centers and degrees of freedom.
- Solvation term based on the solvent accessible surface area (Ferrara et al., *Proteins* **46**, 24, 2002).

### 3.1 Periodic boundary conditions (PBC) vs. spherical boundary



## 3.2 Periodic boundary conditions for peptide/membrane system



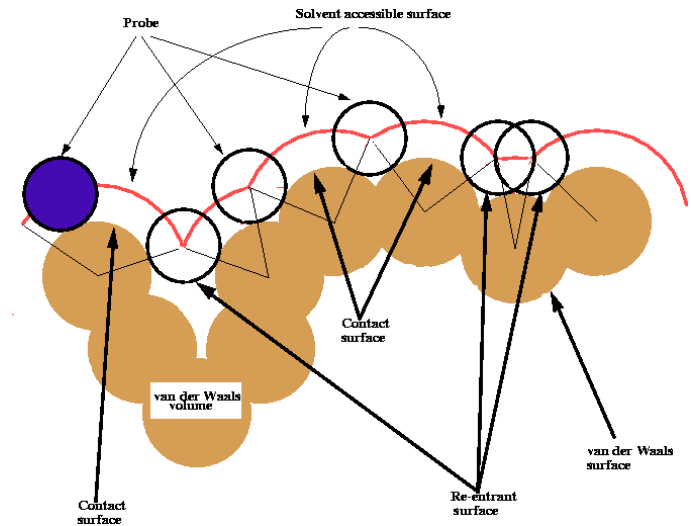
Gorfe et al., Membrane localization and flexibility of a lipidated ras peptide studied by molecular dynamics simulations. J. Amer. Chem. Soc. 126, 15277-15286, 2004.



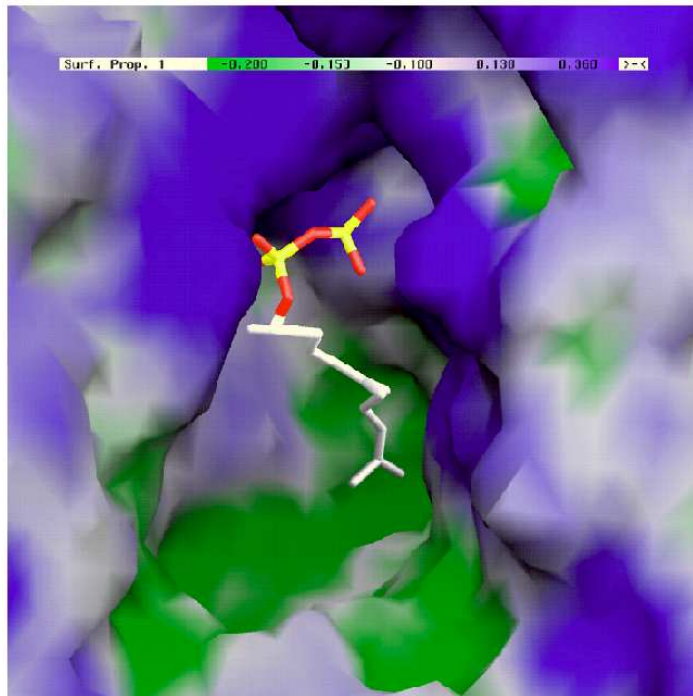
### 3.3 Solvent accessible surface area

$$W(\mathbf{r}) = \mathbf{E}(\mathbf{r}) + \mathbf{V}_{\text{solvation}}(\mathbf{r})$$

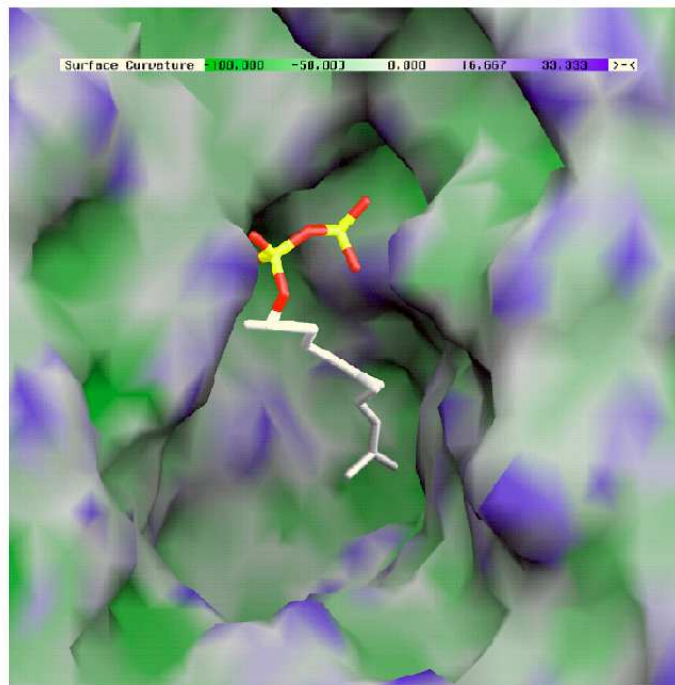
$$V_{\text{solvation}}(\mathbf{r}) = \sum_{i=1}^{N_{\text{polar}}} \sigma_{\text{N,O}} A_i + \sum_{i=1}^{N_{\text{apolar}}} \sigma_{\text{C,S}} A_i$$



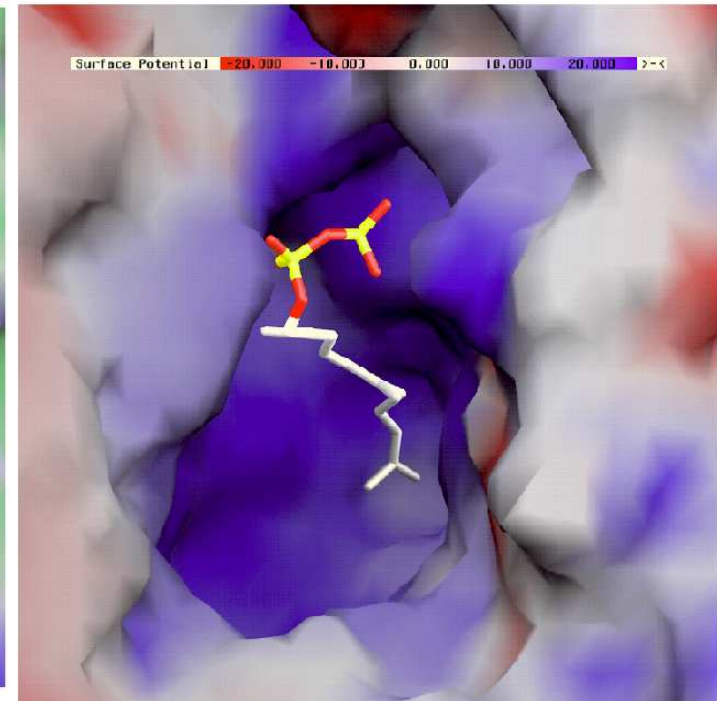
Farnesyltransferase-farnesylpyrophosphate complex



hydrophilic/hydrophobic



concave/convex

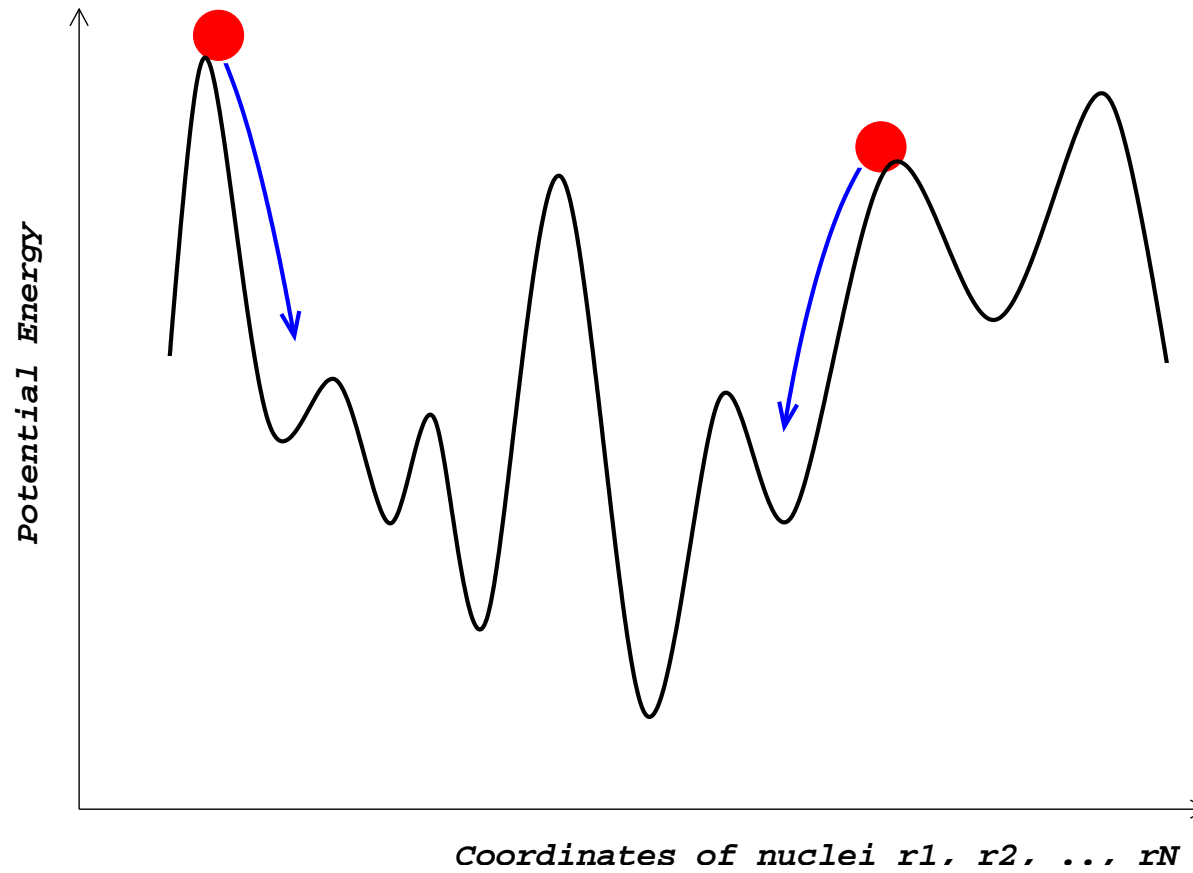


electrostatic potential

Scarsi et al., *Proteins* **37**, 565, 1999

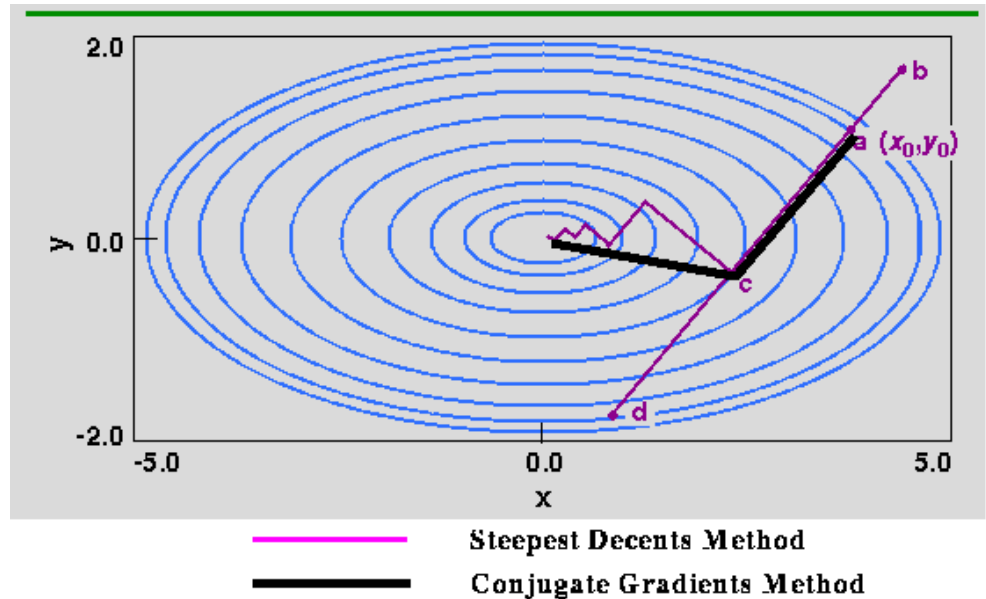
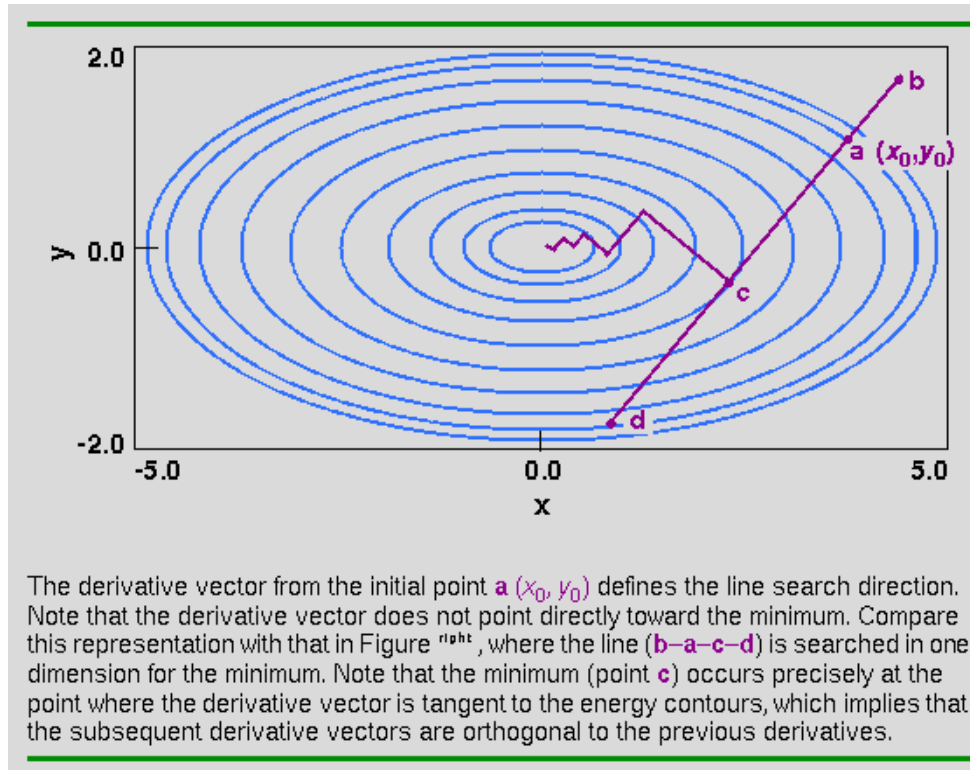
## 4 Sampling techniques

### 4.1 Energy minimization



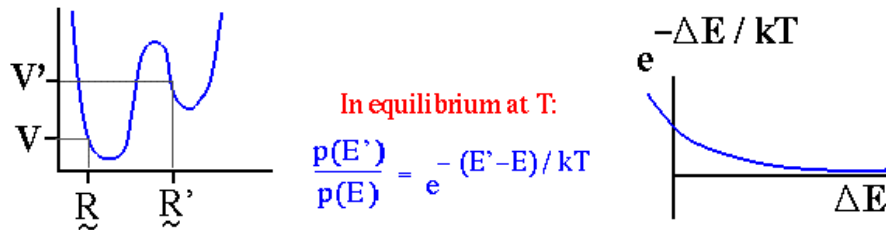
- Minimization follows gradient
- Reaches the nearest **local** minimum
- Steepest descent, conjugate gradient

## 4.2 Steepest descent vs. conjugate gradient

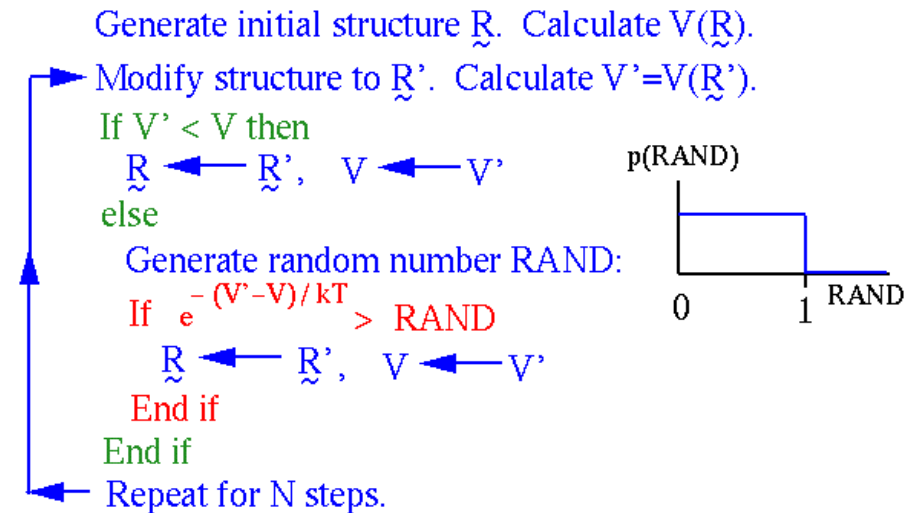


### 4.3 Metropolis Monte Carlo (Boltzmann statistics)

MC compares energies. No forces calculated.

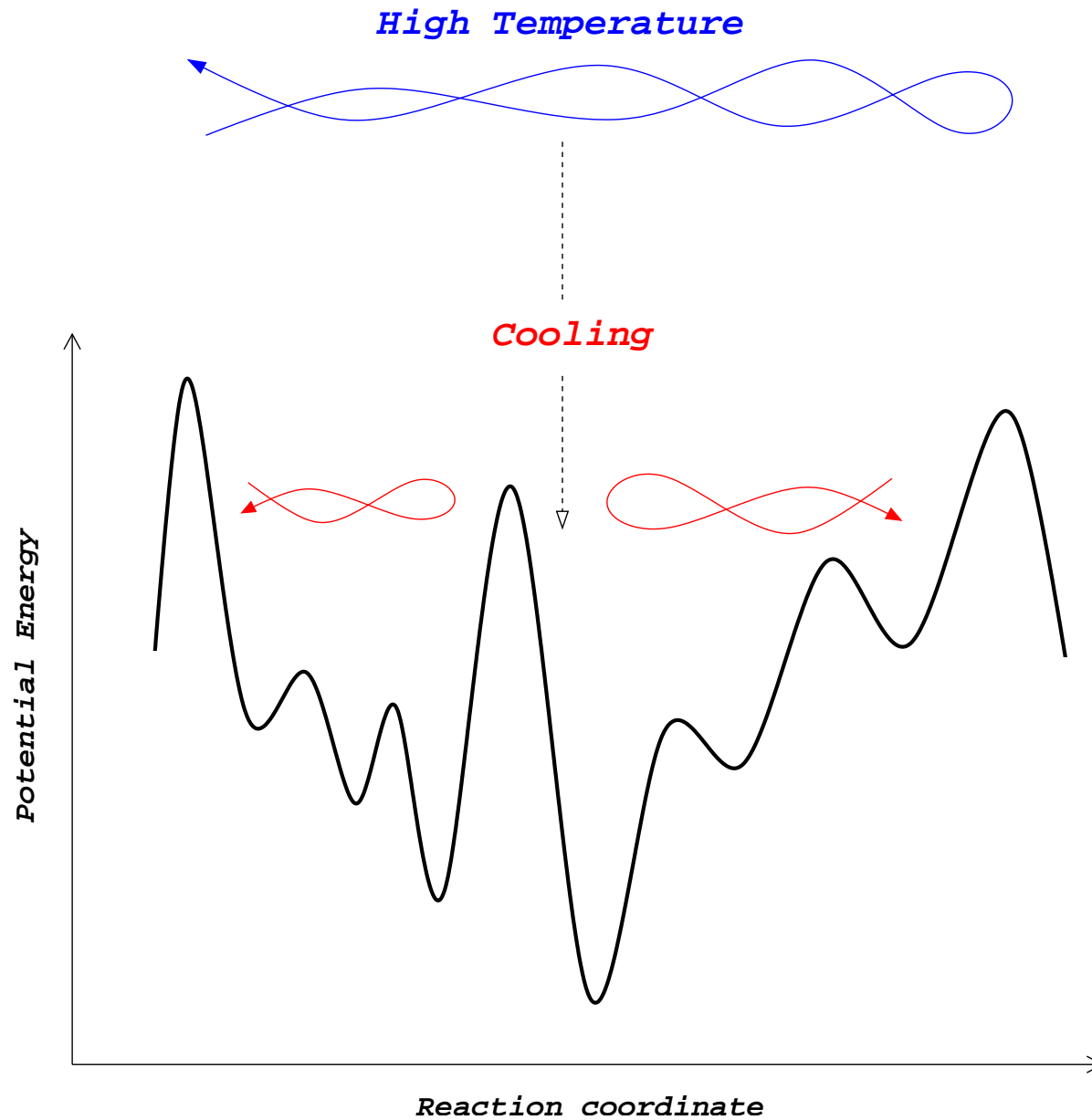


Monte Carlo Algorithm



- Metropolis Monte Carlo yields an ensemble (Boltzmann statistics).
- Ergodicity: every accessible point in configuration space should be reached in a finite number of Monte Carlo steps from any other point.
- Kinetics are usually not meaningful.

#### 4.4 Simulated annealing (good for sampling but no ensemble)





## 4.5 Parallel tempering (equilibrium Monte Carlo scheme)

- $M$  *non-interacting* copies of the system at different  $T_m$
- A *state* is defined by

$$X = \left( \overbrace{x_{m(1)}^1, \dots, x_{m(M)}^M}^{T_1, T_2, \dots, T_M} \right), \quad x_m^i \equiv (q^i, p^i)_m$$

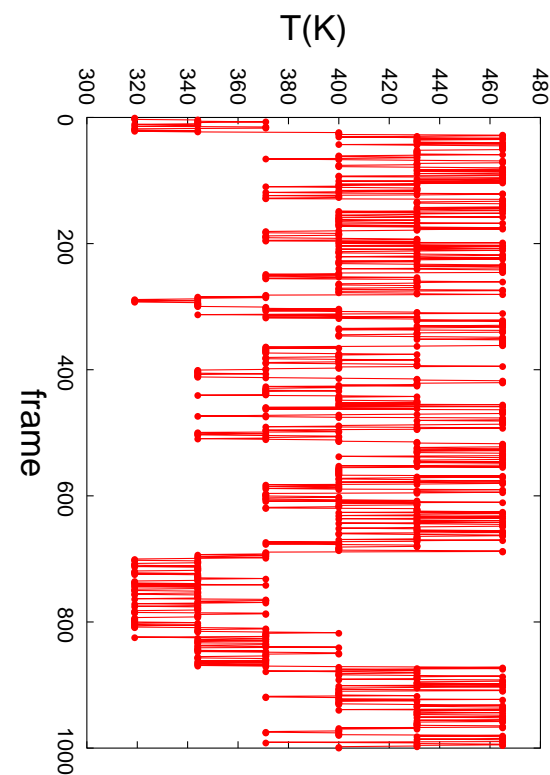
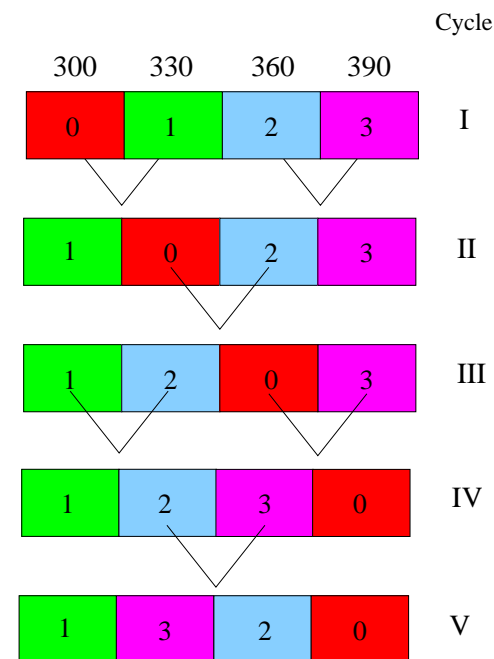
- In order to converge toward equilibrium the *detailed balance* should be satisfied. Therefore:

$$w(X \rightarrow X') = \begin{cases} 1, & \Delta \leq 0, \\ \exp(-\Delta), & \Delta > 0. \end{cases}$$

where  $\Delta \equiv [\beta_n - \beta_m] (\mathcal{E}(x_m^i) - \mathcal{E}(x_n^j))$ .

<i>High <math>T</math></i>	replicas jump from basin to basin	( <i>inter-basin</i> )
<i>Low <math>T</math></i>	replicas explore a single valley	( <i>intra-basin</i> )

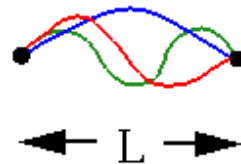
Rao and Caflisch, *J. Chem. Phys.* **119**, 4035, 2003



## 5 Normal mode analysis

### Examples of Normal Modes

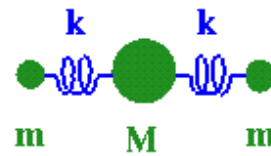
Guitar string  
fixed at both ends



$$\lambda = 2L / n$$

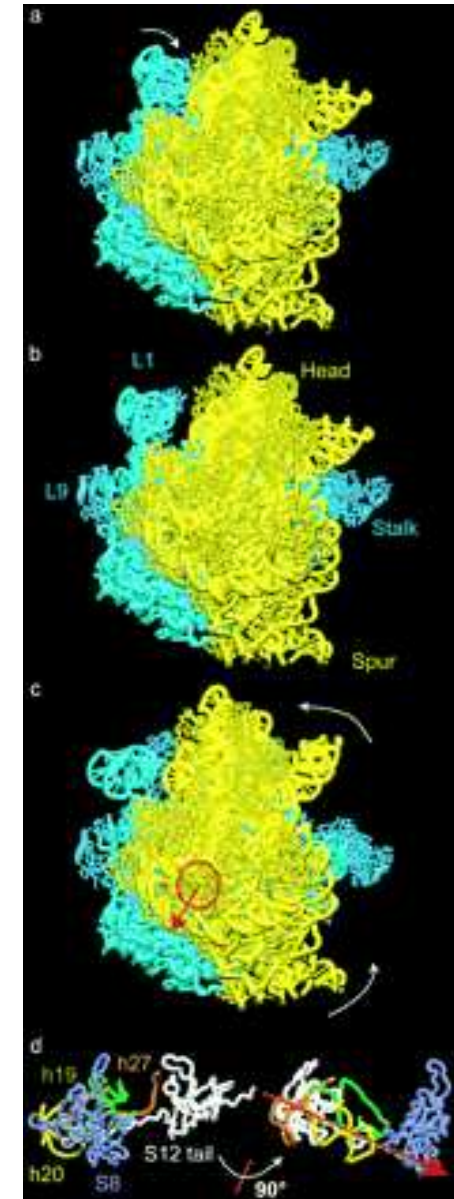
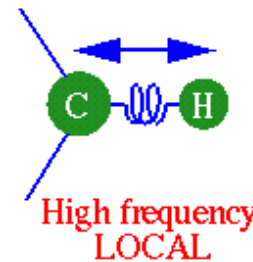
$$n = 1, 2, 3$$

Balls & Springs  
in 1D



$\omega$	mode
0	→ → → (Pure Translation)
$(k/m)^{1/2}$	← • → (Center at Rest)
$[(k/m)(1+2m/M)]^{1/2}$	← → ←

Proteins

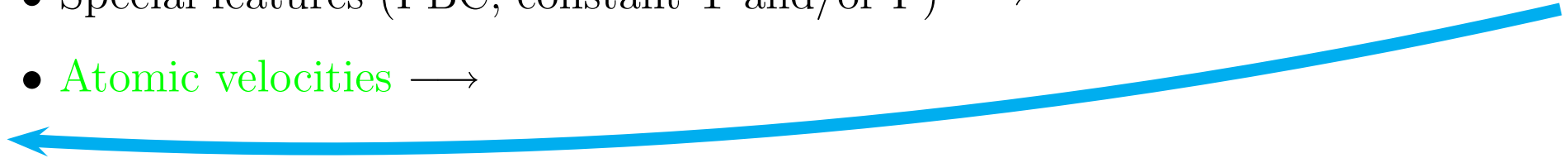


Tama et al., PNAS 100, 9319, 2003

## 6 Molecular dynamics

### 6.1 Basics

- Atomic positions (coordinate file) →
  - Covalent structure (topology file) →
  - Potential energy function (parameter file) →
  - Additional atoms (solvent, counterions) →
  - Special features (PBC, constant T and/or P) →
  - Atomic velocities →
- Effective temperature (through kinetic energy)
  - Forces on each atom



## 6.2 Equations

- $\mathbf{F}_i = m_i \mathbf{a}_i \quad \mathbf{F}_i = -\mathbf{grad}_i E \quad E = E_{bonding} + E_{non-bonding}$

1. solve for  $a_i$  at  $t$   $-\frac{dE}{dr_i} = F_i = m_i a_i(t)$
2. update  $v_i$  at  $t + \Delta t/2$   $v_i(t + \Delta t/2) = v_i(t - \Delta t/2) + a_i(t)\Delta t$
3. update  $r_i$  at  $t + \Delta t$   $r_i(t + \Delta t) = r_i(t) + v_i(t + \Delta t/2)\Delta t$
4. go to 1.

- Timestep controls accuracy of numerical solution.
- Fundamental timestep is determined by high frequency vibrations (covalent bonds  $\longrightarrow \Delta t = 10^{-15}$  sec).
- Highest frequency motions, i.e., hydrogen atom vibrations, can be removed with holonomic constraints.

### 6.3 Thermodynamic variables $T$ and $P$

- Statistical ensembles connect *microscopic* to *macroscopic*

Microcanonical (NVE, entropy)

Canonical (NVT, Helmholtz free-energy)

$$\cdot T = \sum m \langle v^2 \rangle / (3k_b)$$

Isothermal-isobaric (NPT, Gibbs free-energy)

$$\cdot P = \text{kinetic} + \text{virial contributions}$$

- Thermostats, barostats allow to choose the appropriate ensemble.
  - Andersen, Nosé, Hoover.

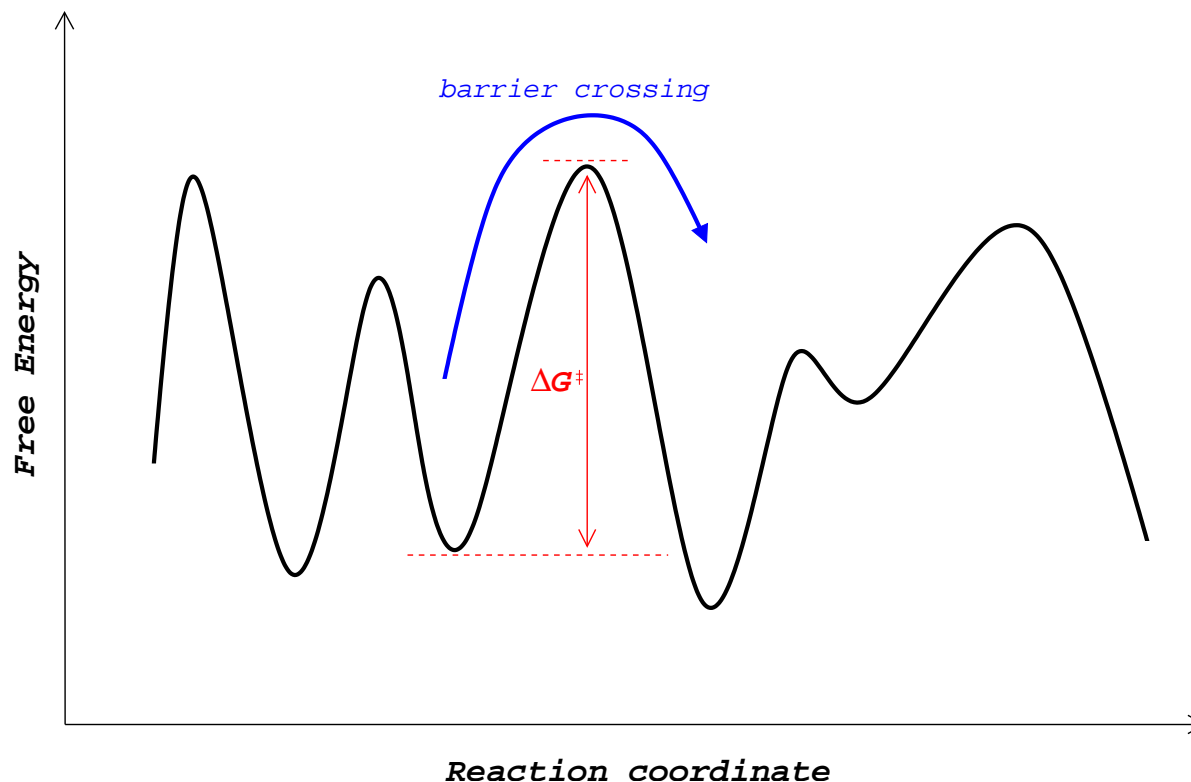
## 7 Applications of molecular dynamics

1. Sampling configuration space,  
e.g., simulated annealing to determine or refine structures.
2. Obtaining a description of the system at equilibrium,  
i.e., sampling with appropriate Boltzmann factor:
  - structural and motional properties
  - values of thermodynamic parameters
3. Obtaining actual dynamics and kinetics,  
i.e., sampling with appropriate Boltzmann factor and  
correct representation of the development of the system over time.

For (1) and (2) Monte Carlo simulations can be also used.

From Karplus and McCammon, Nature Structural Biology **9**, 646, 2002.

## 8 Free-energy barriers and timescales



- To cross a free-energy barrier  $\tau = \tau_0 \exp(\Delta G^\ddagger / k_B T)$  with  $\tau_0 \sim 10^{-12}$  s:  
1 kcal/mol :  $\sim ps$ , 5 kcal/mol :  $\sim ns$ , 10 kcal/mol :  $\mu s$  or longer
- Sampling should exceed timescales of interest by  $\sim 10$ -fold.
- System size and complexity increase required timescales (equilibration of ions, complex landscapes, multiple minima)

## 9 Approximations in molecular dynamics

- Approximations inherent to the force field ( $E$ )  $\longrightarrow$

### **Systematic error:**

Calculations of free energy differences is still very difficult.

- Time scale and sampling problem  $\longrightarrow$

### **Statistical error:**

Conformational transitions that require  $> 0.1 - 1\mu\text{s}$  cannot be simulated (yet) by conventional molecular dynamics techniques.

- Other simulation approaches:
  - MD with implicit solvent (approximate)
  - Brownian dynamics
  - Monte Carlo (move definitions are difficult for macromolecules)



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