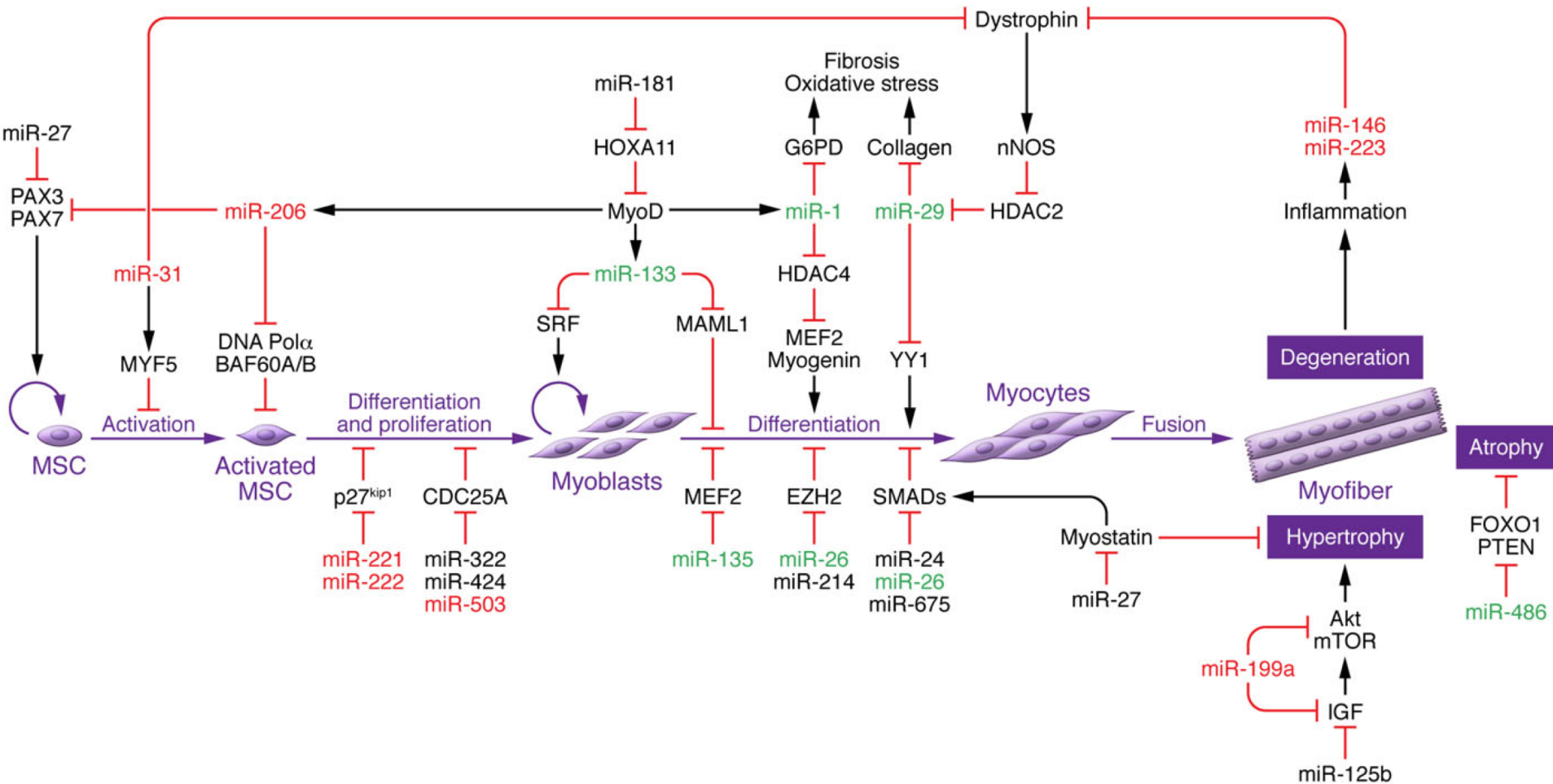


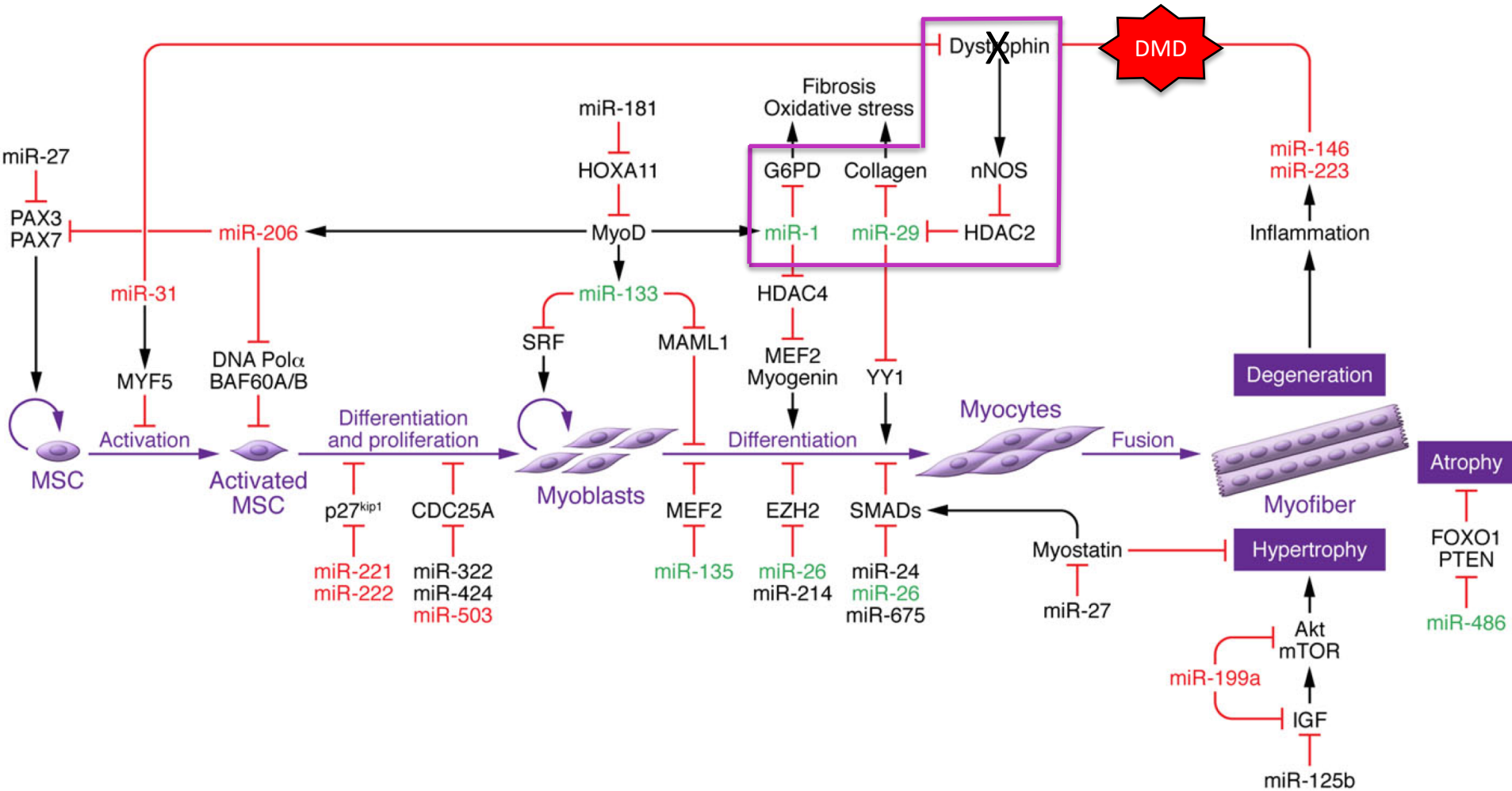
# lncRNA and miRNA in muscle differentiation

# miRNA in myogenesis

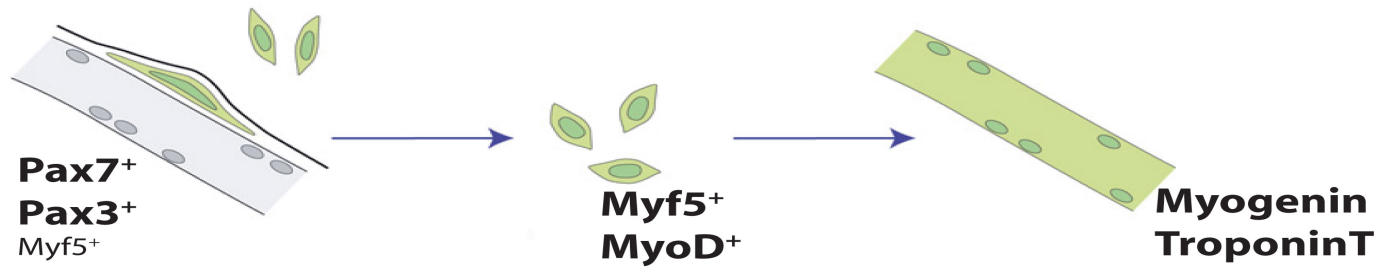




# miRNA in myogenesis

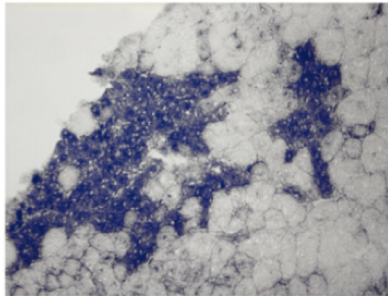


# *miR-206 and miR-1 are differentially activated during myogenesis*

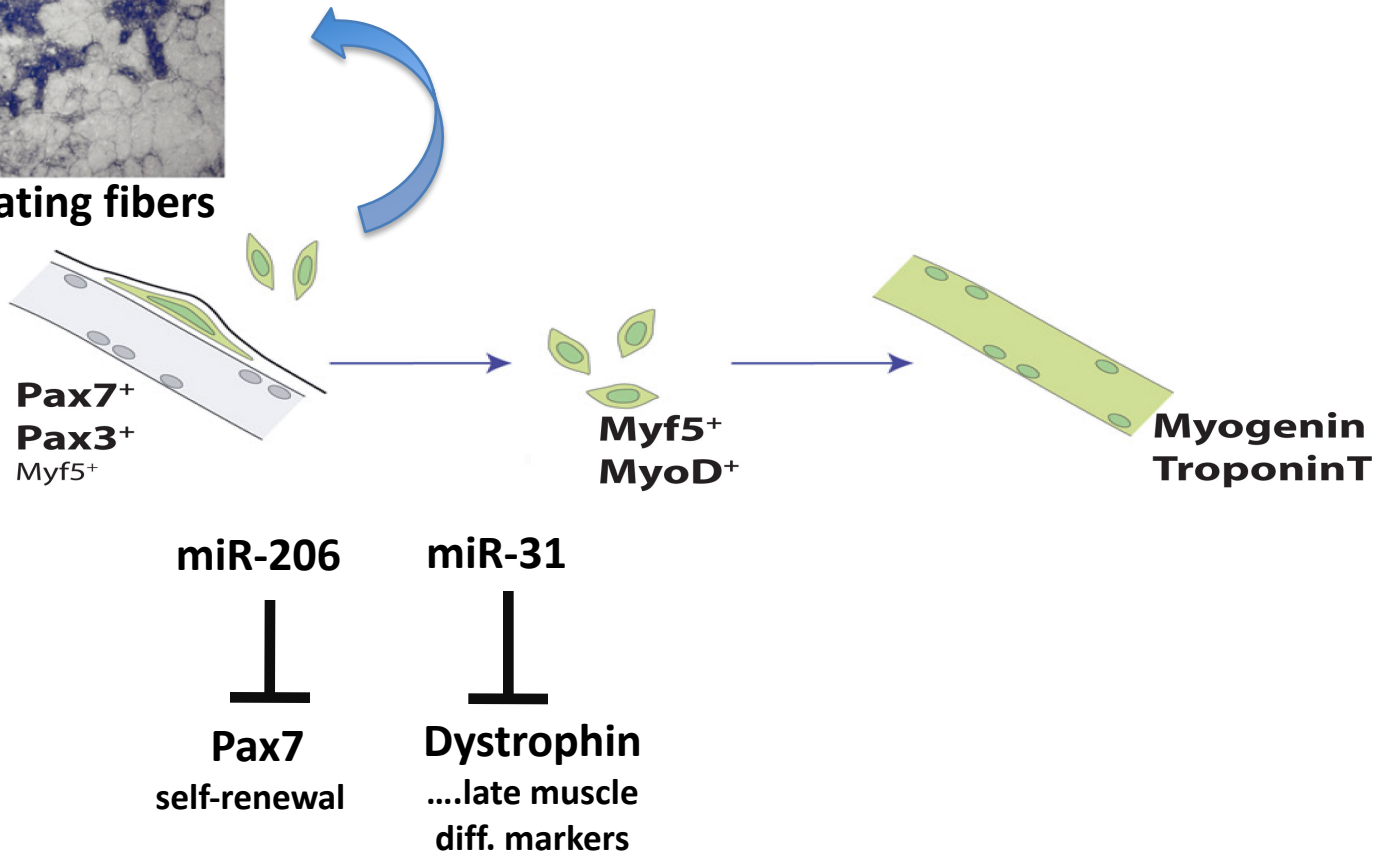


# *miR-206 and miR-31 are expressed in early phases of myogenesis*

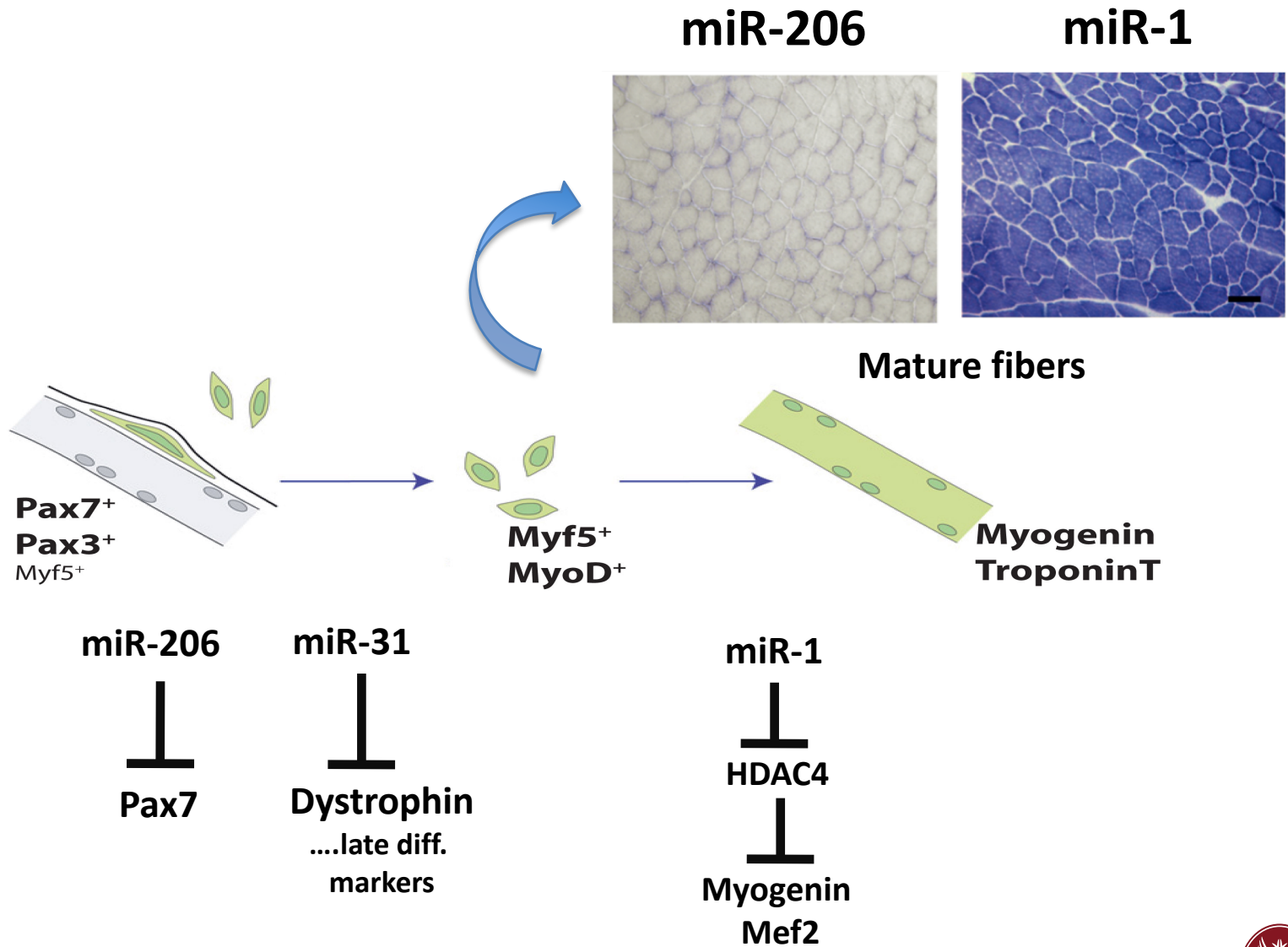
## miR-206



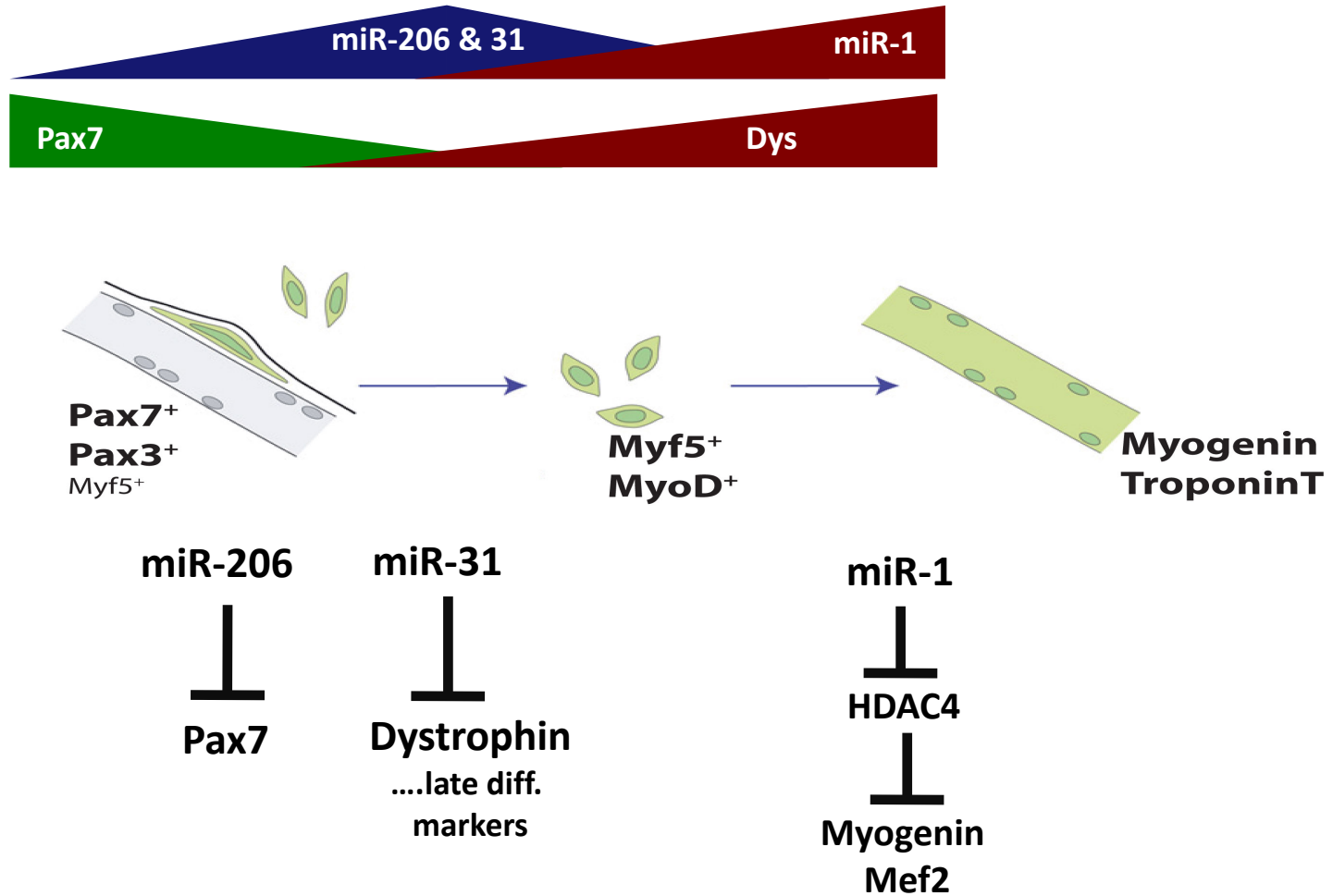
regenerating fibers



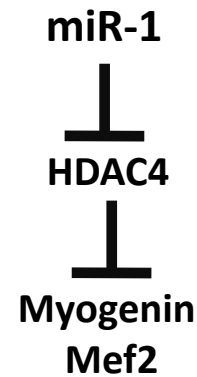
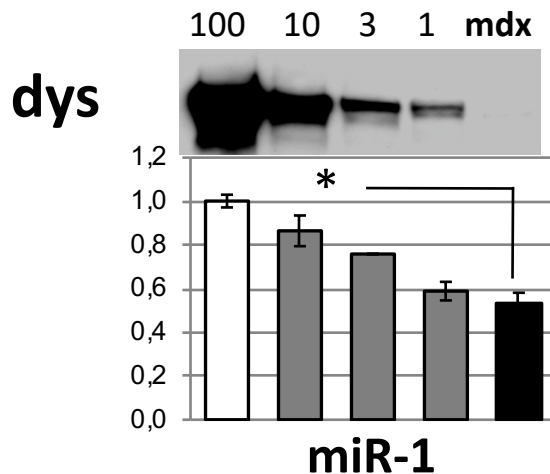
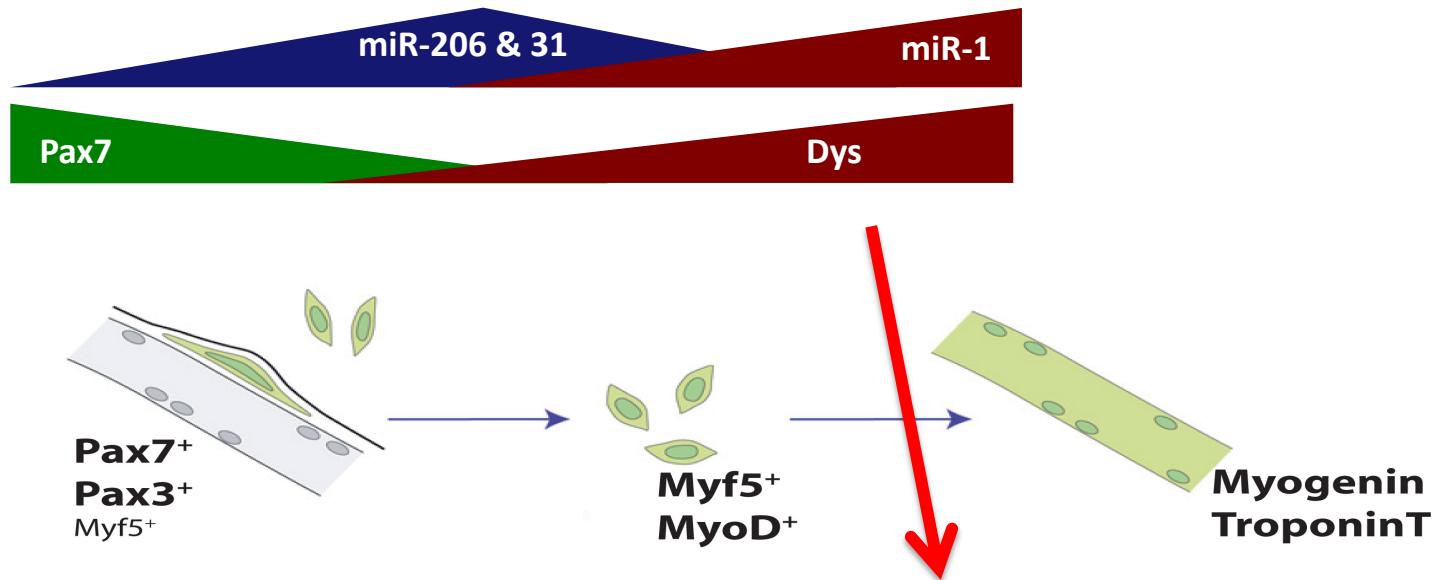
# *miR-1 is expressed at later stages of myogenesis*



# *miR-206 and miR-1 are differentially activated during myogenesis*



# miR-1 levels correlate with dystrophin expression



## miR-206 is *present only in vertebrates*

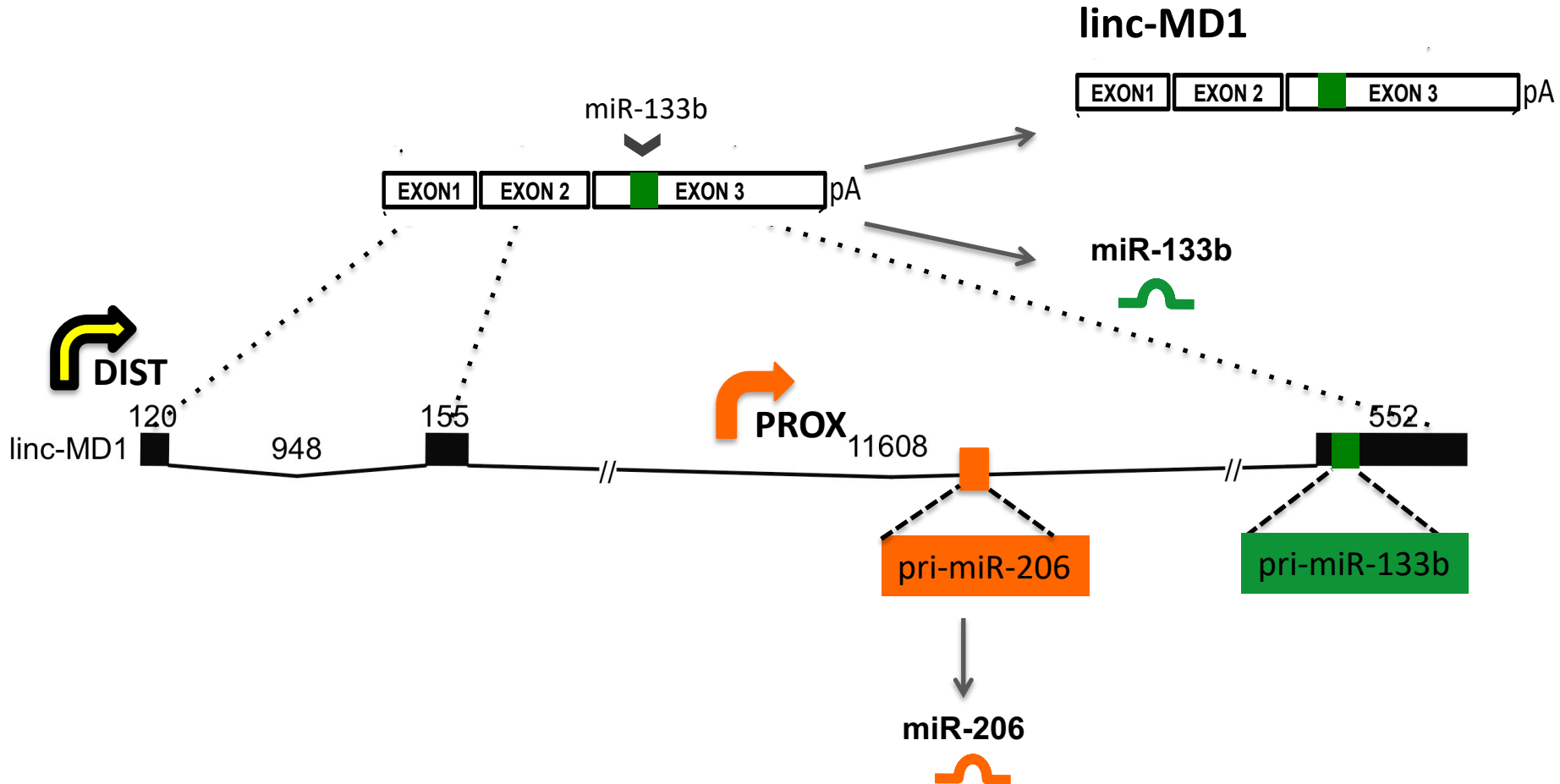
The origin of the miR-206/miR-133b locus correlates, indeed, with the increase in complexity of vertebrate skeletal muscle: in fact, fly and worm do not have satellite cells nor do they possess different fiber types such as slow-twitch and fast twitch fibers as found in vertebrate skeletal muscle

*miR-206 – in activated precursor cells*  
*miR1 – in differentiated myoblasts*

**Mouse genomic localization**

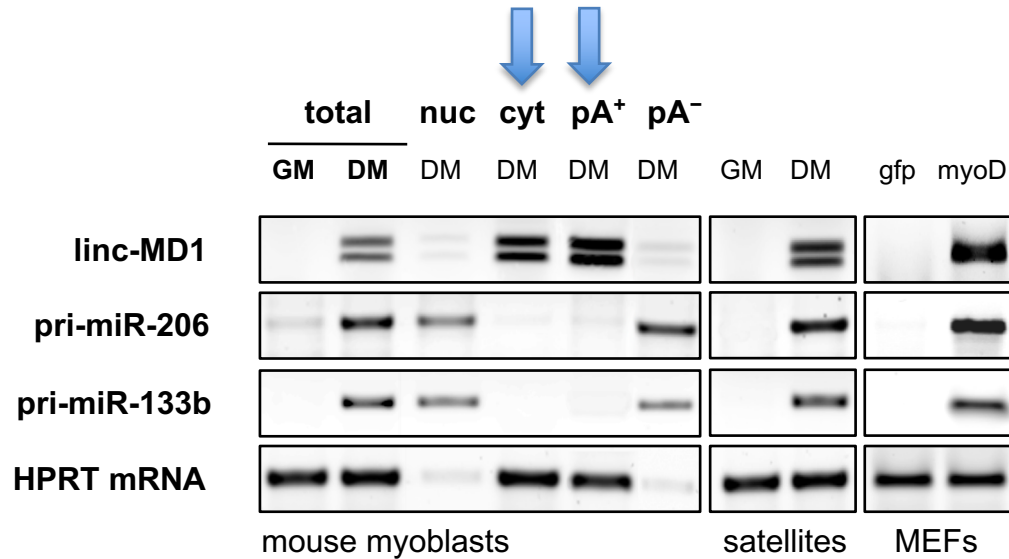
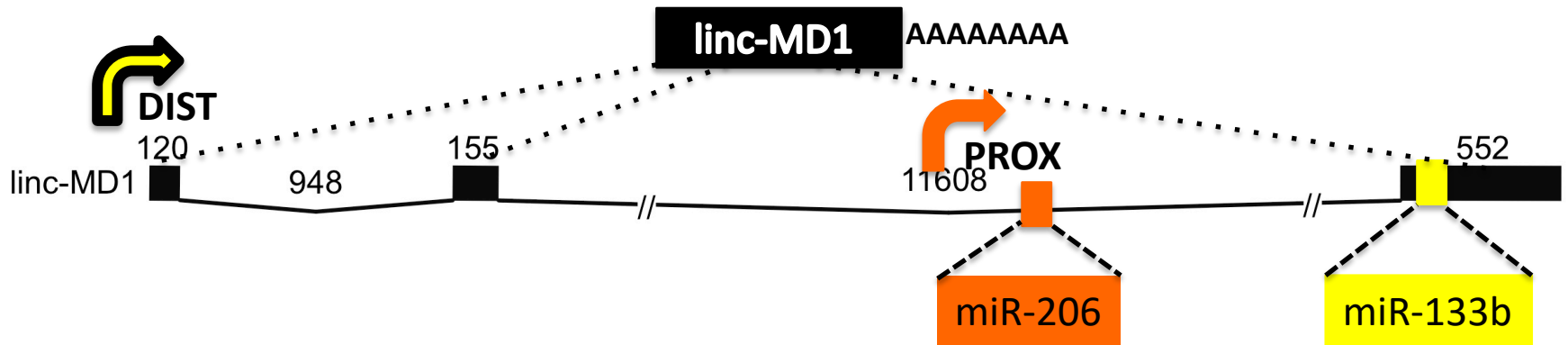


# Different types of transcripts originate from mir-206/mir-133b locus

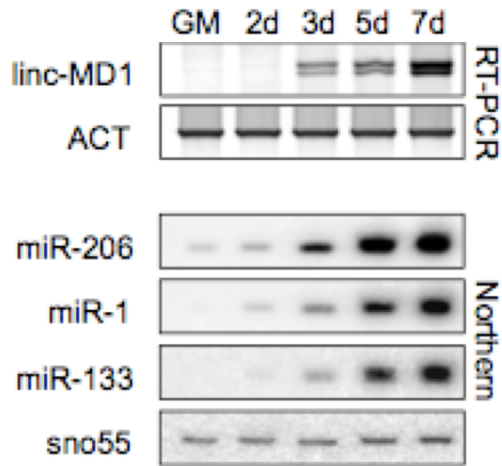
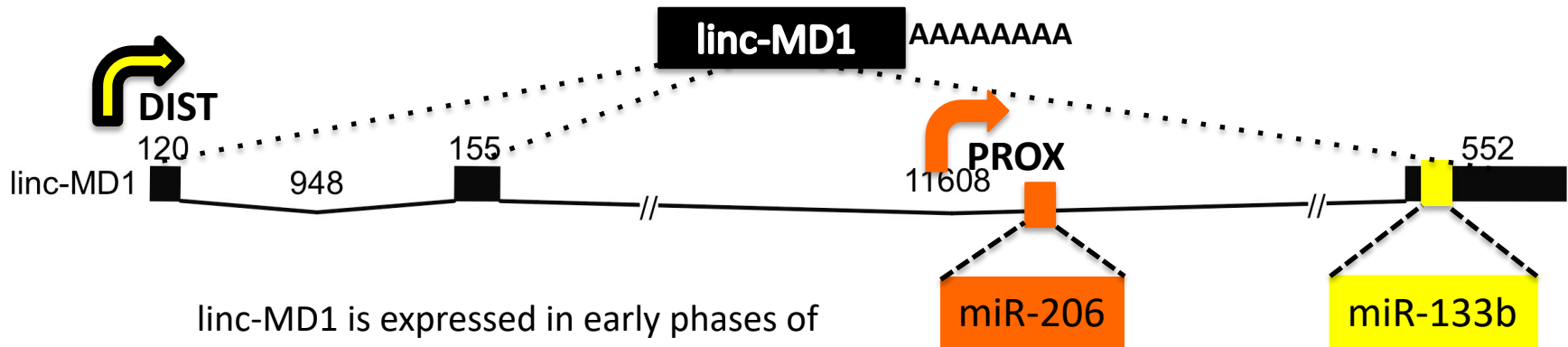




# linc-MD1 is a cytoplasmic polyA+ long non-coding RNA



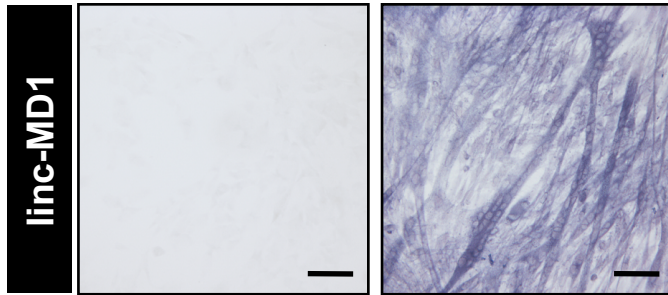
# linc-MD1 is a cytoplasmic polyA+ long non-coding RNA



linc-MD1 is expressed in differentiating myoblasts  
and not in mature fibers

GM

DM

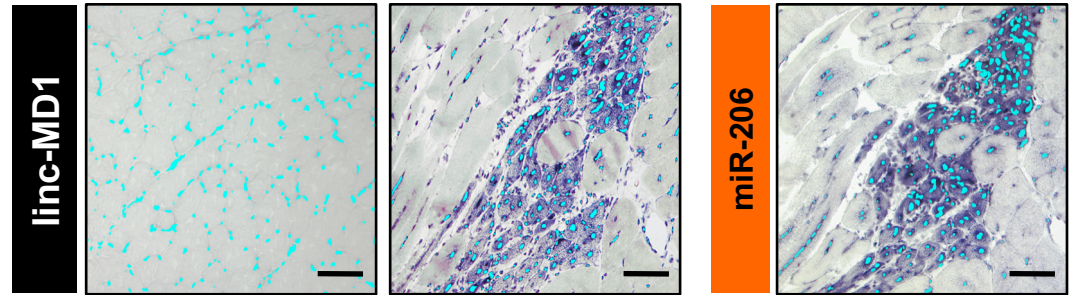


C2 myoblasts

WT

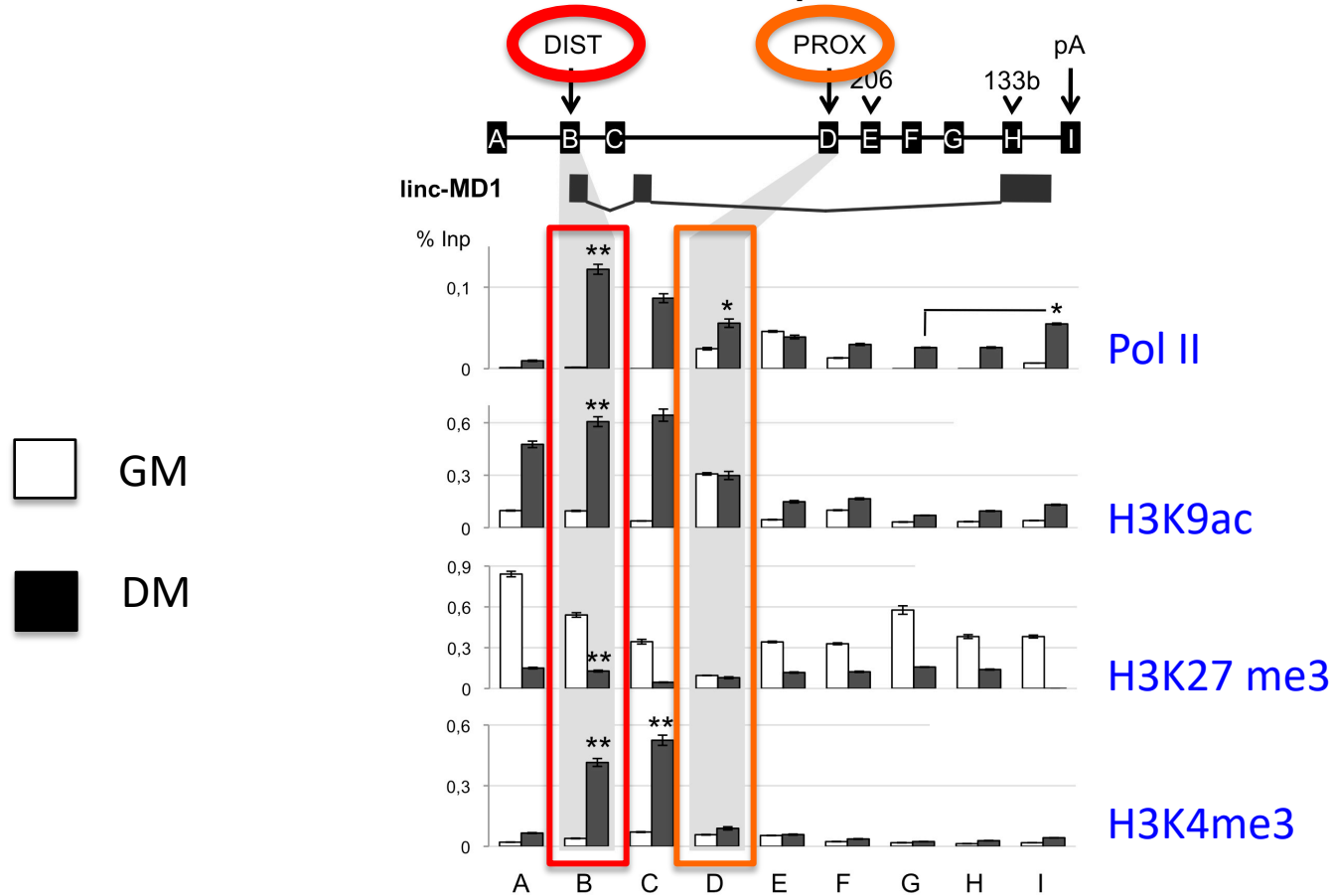
*mdx*

*mdx*



gastrocnemius

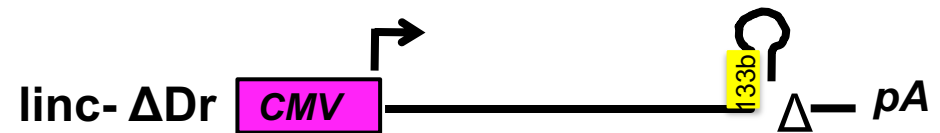
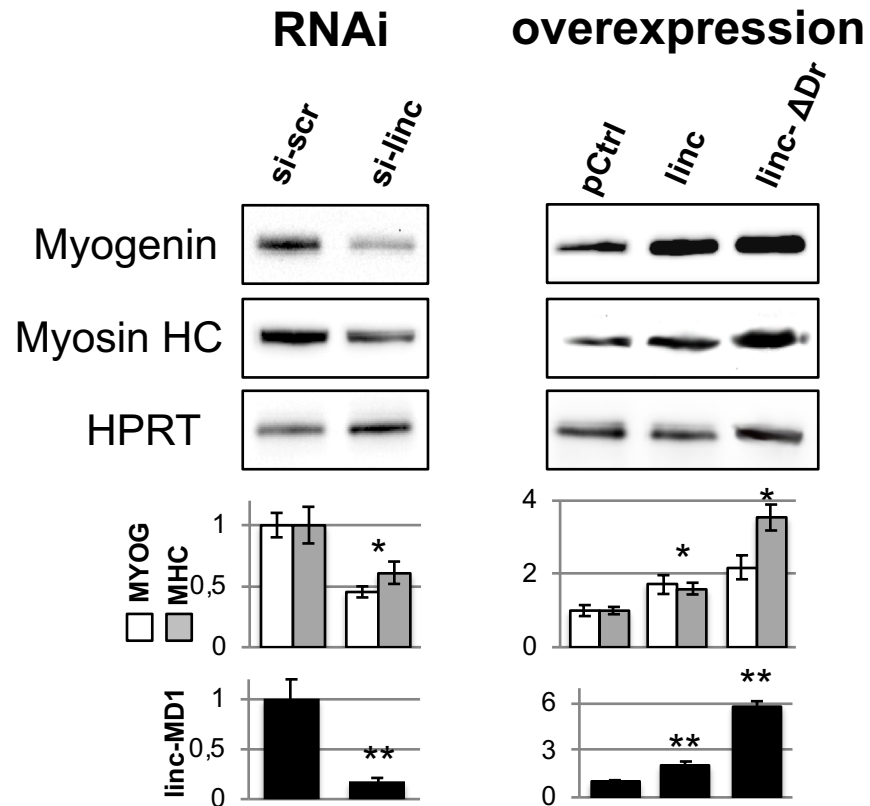
# Transcriptional and Epigenetic Regulation of linc-MD1 and miR-206/133b expression



PROX and DIST promoters act at different stages of muscle differentiation, promoting miR-206, miR-133b and linc-MD1 expression

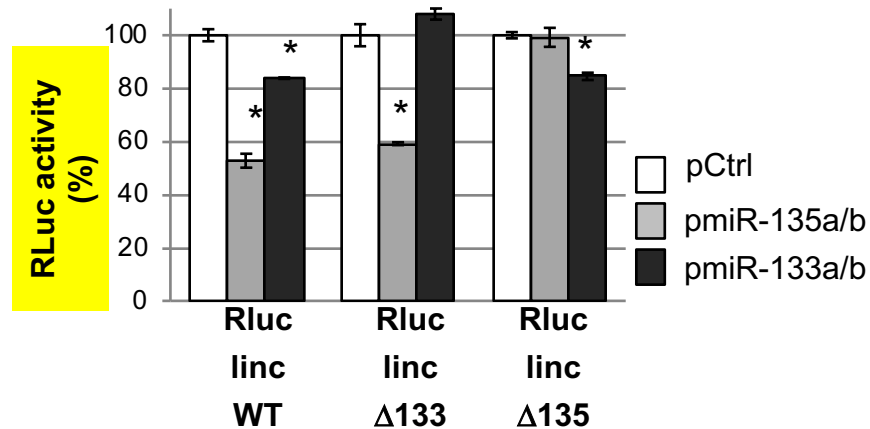
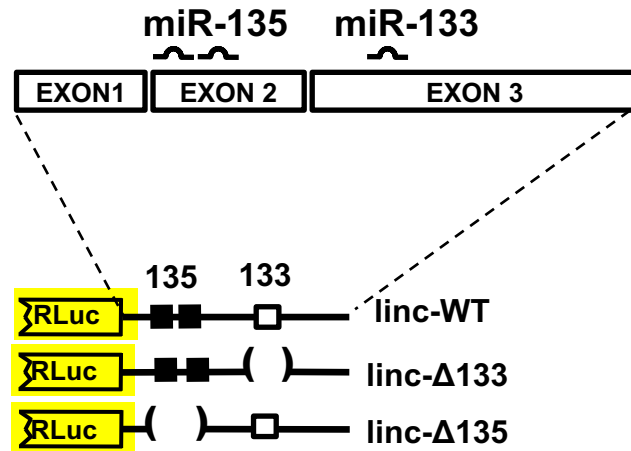


# linc-MD1 levels affect expression of early myogenic markers



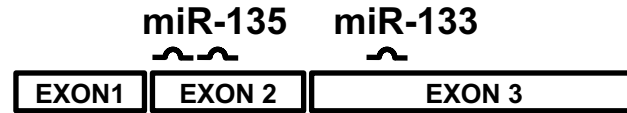
# linc-MD1 is target of miR-135 and miR-133

linc-MD1

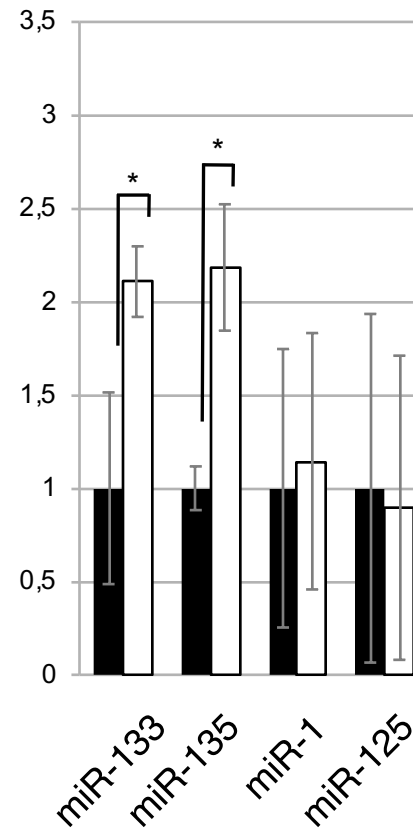
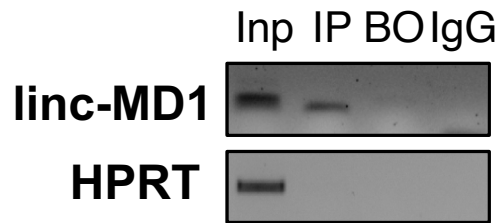


# *linc-MD1 binds Ago2 and miR-135 and miR-133*

**linc-MD1**



**RIP Ago2**



the 3' UTRs of the myogenic factors MAML1 and MEF2C contain miR-135 and miR-133 binding sites

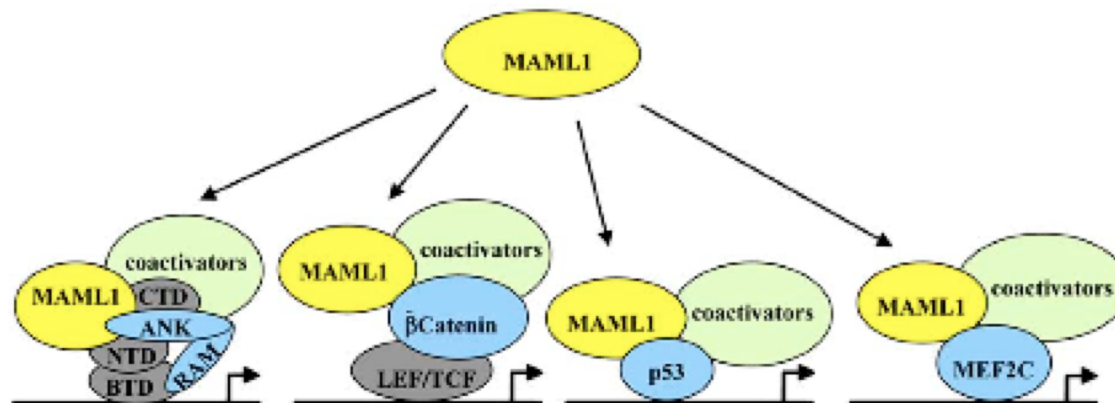
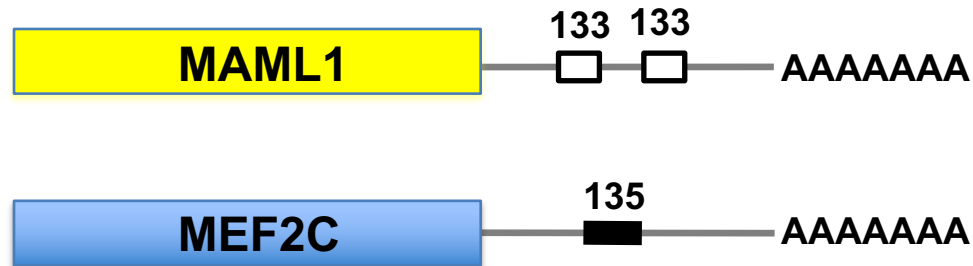
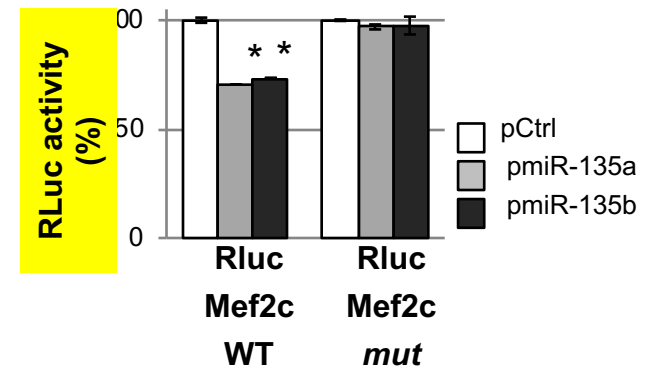
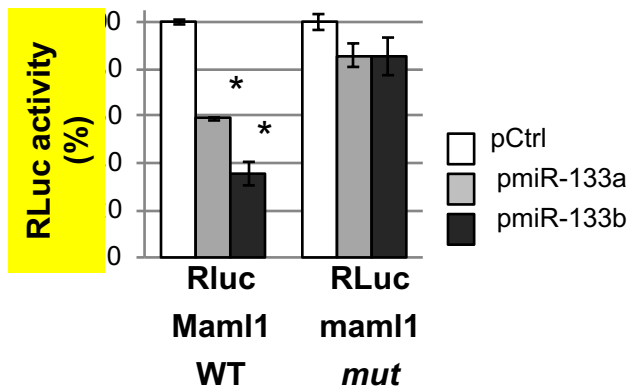
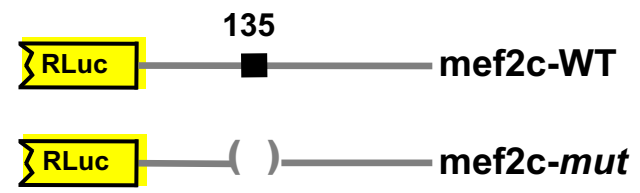
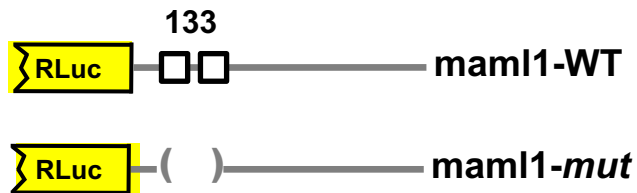


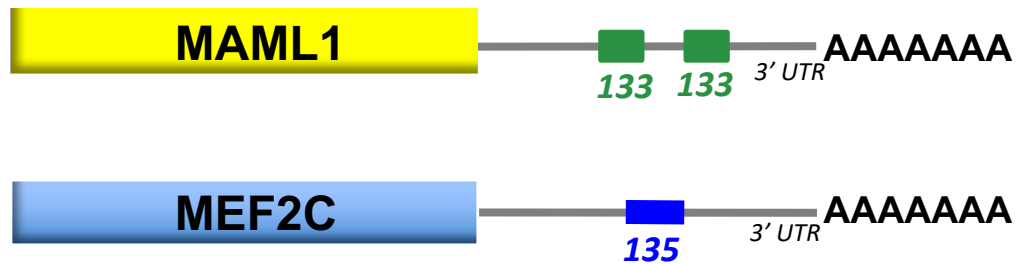
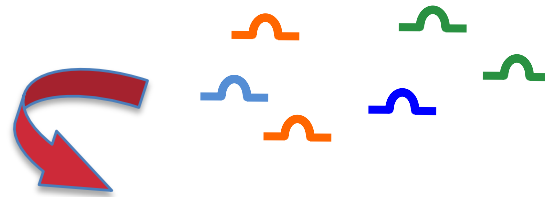
Fig. (1). The MAML1 protein acts as a coactivator for diverse transcription factors. MAML1 is recruited by the ANK (ankyrin repeats) domain of Notch and forms a complex with the DNA-binding protein CSL (composed of NTD (N-terminal domain), CTD (C-terminal domain), and BTD ( $\beta$ -trefoil domain)). The RAM (RBP-Jk associated molecule) domain of Notch interacts with the BTD of CSL. In addition, MAML1 is recruited by  $\beta$ -catenin, p53 and MEF2C to regulate various signalling pathways. Most likely, additional coactivators are working cooperatively with MAML1 in gene regulation.



# the myogenic factors MAML1 and MEF2C are target of miR-133 and miR-135

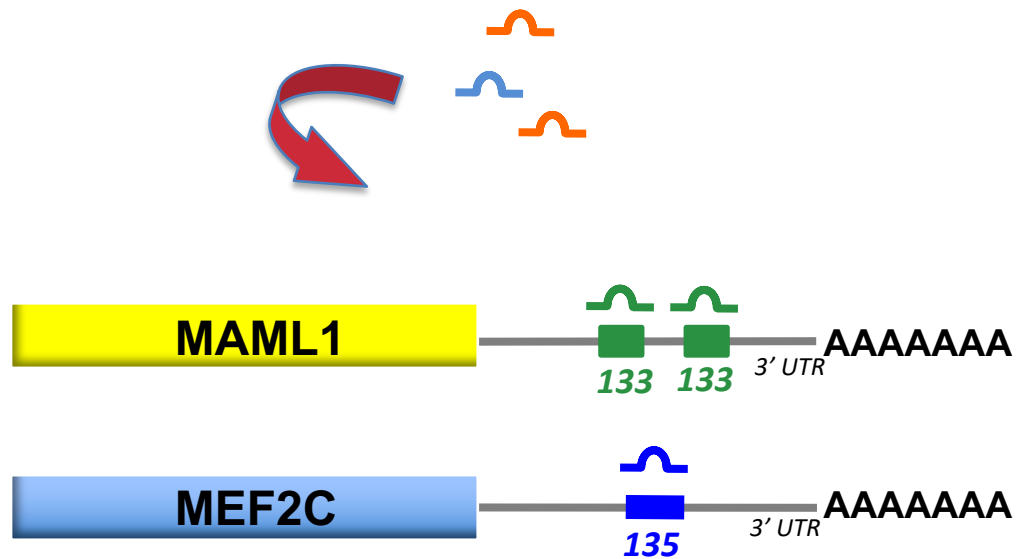


# Competition for miRNA binding



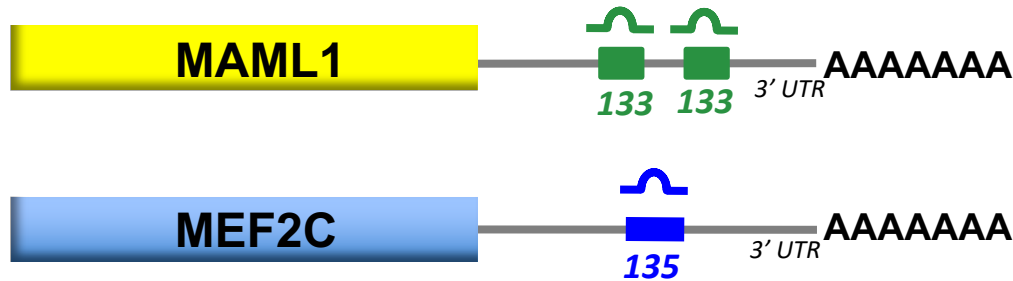
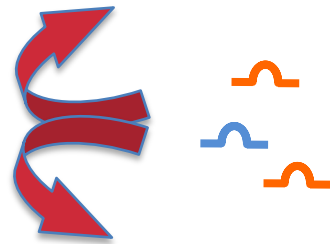
MAML1 and MEF2C are myogenic factors controlling muscle differentiation

# Competition for miRNA binding



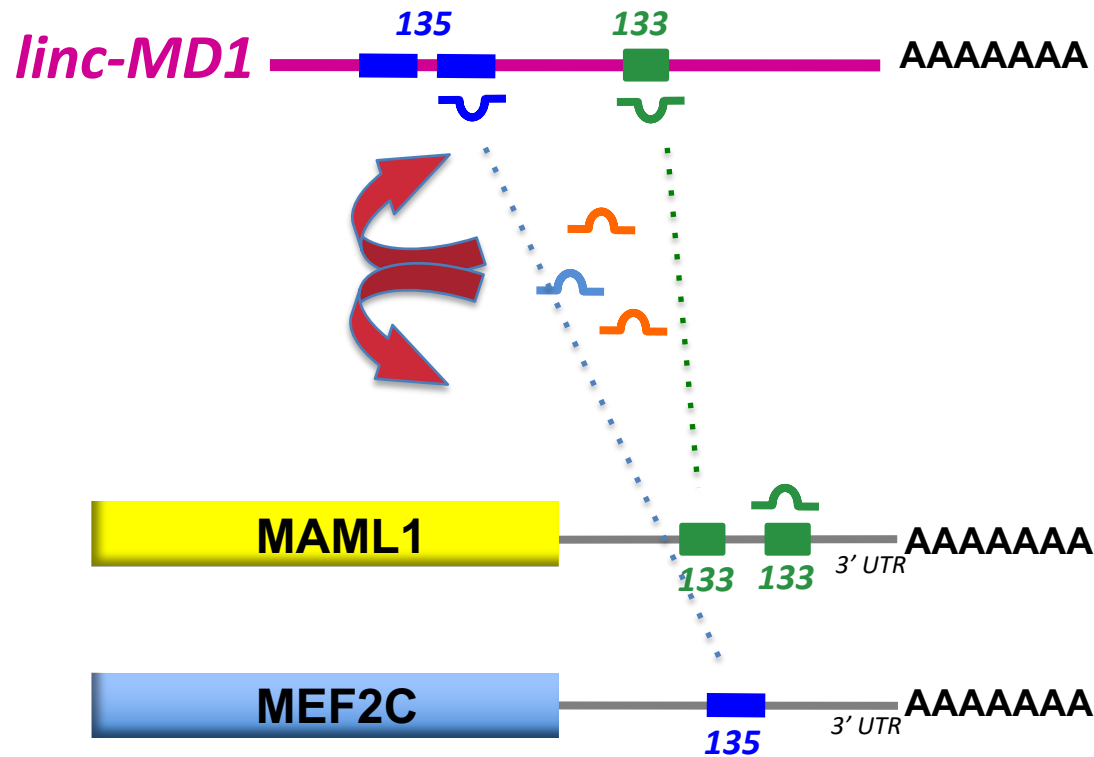
MAML1 and MEF2C are myogenic factors controlling muscle differentiation

# Competition for miRNA binding



MAML1 and MEF2C are myogenic factors controlling muscle differentiation

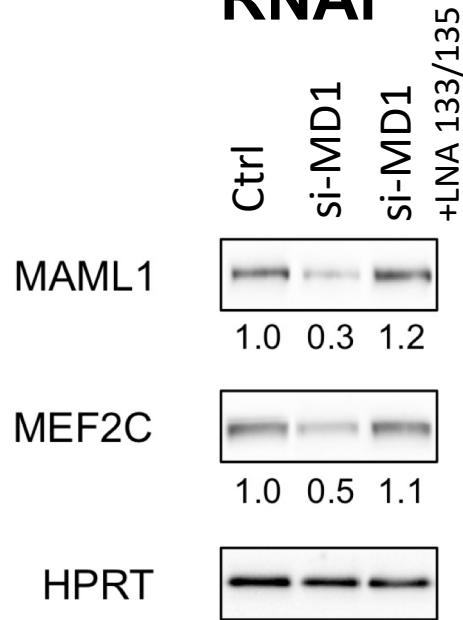
# Competition for miRNA binding



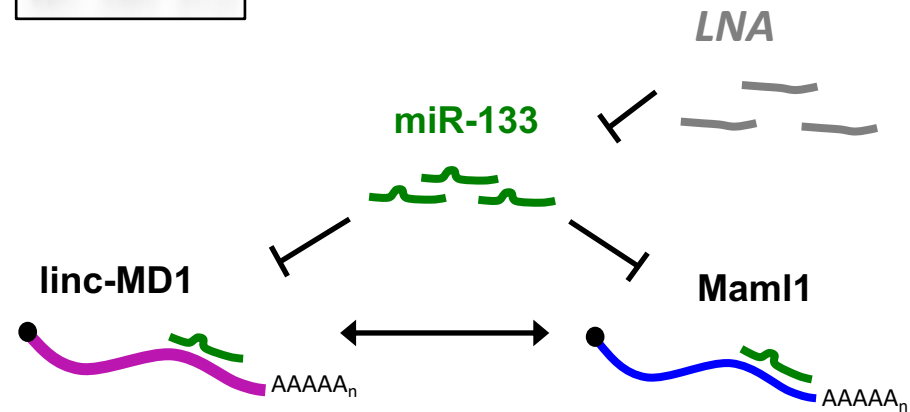
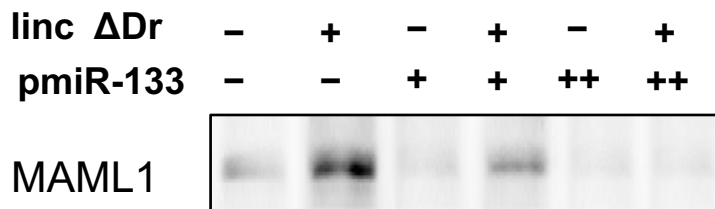
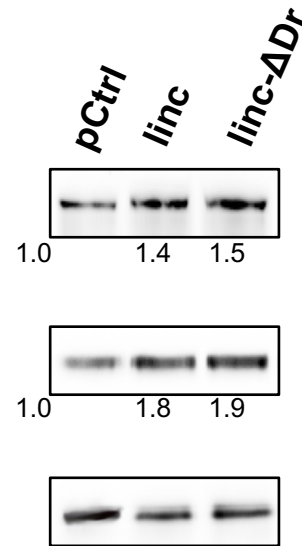
MAML1 and MEF2C are myogenic factors controlling muscle differentiation

# *linc-MD1* controls the levels of MAML1 and MEF2C

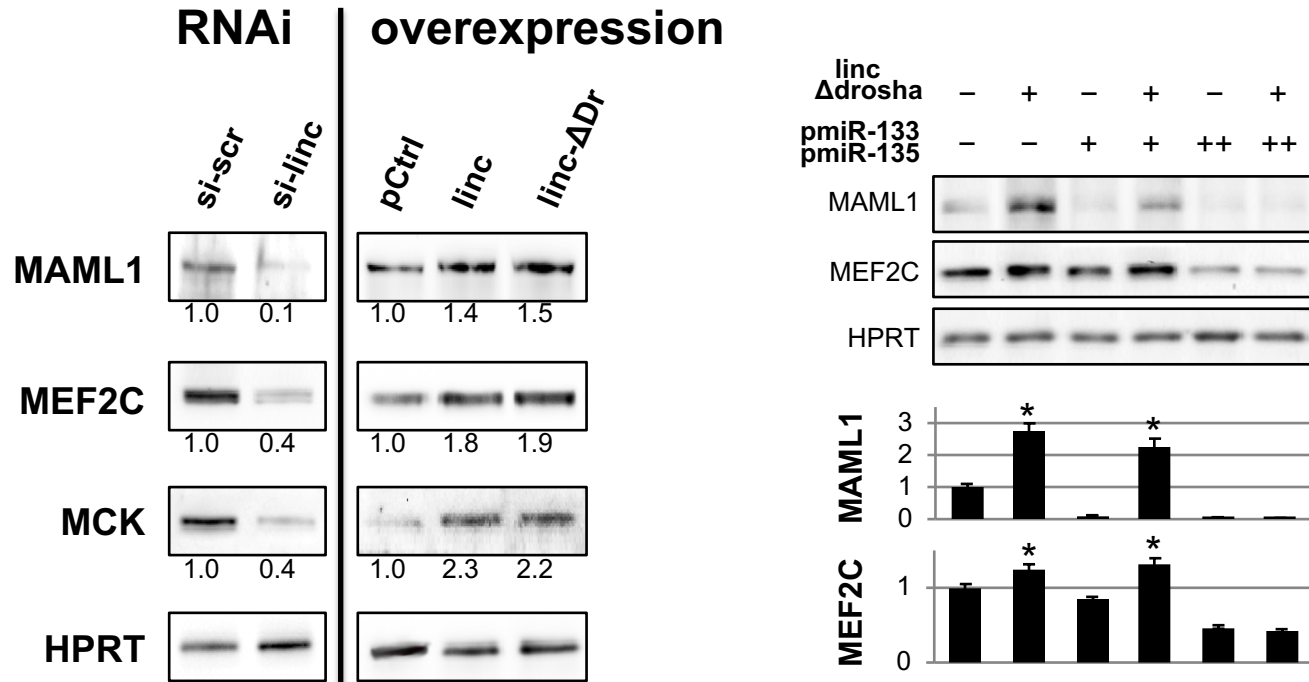
## RNAi



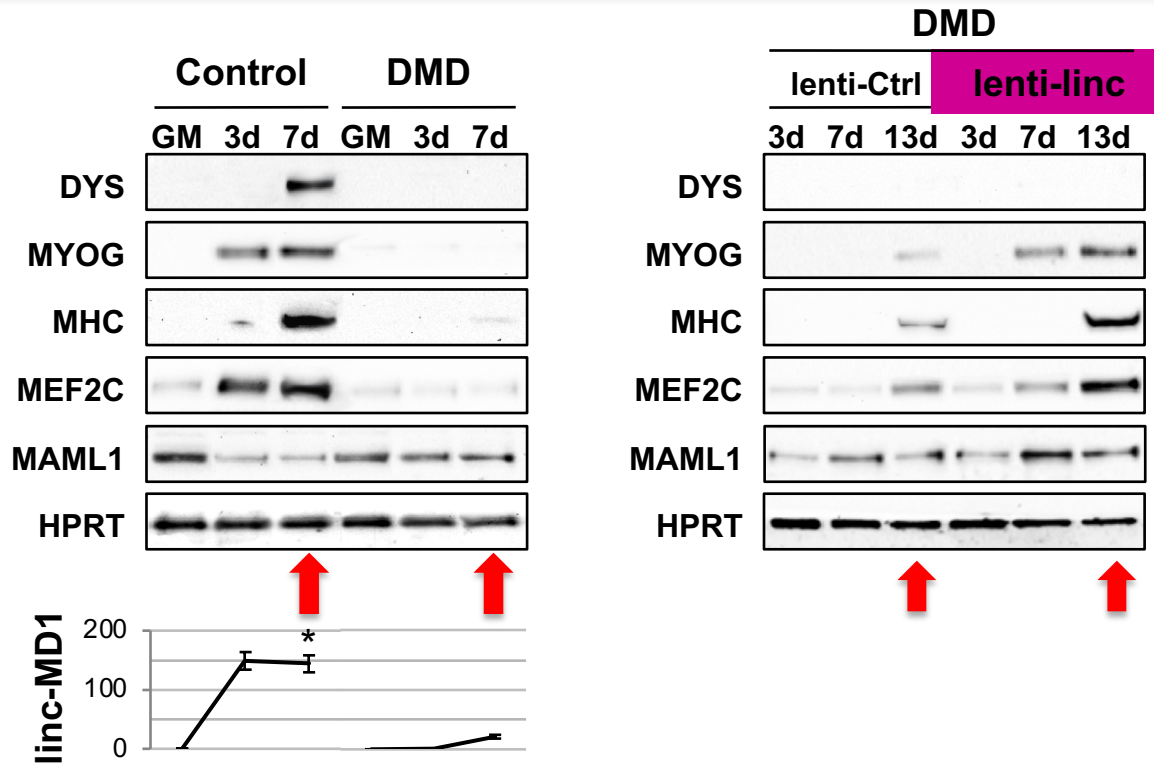
## overexpression



# linc-MD1 controls the levels of MAML1 and MEF2C



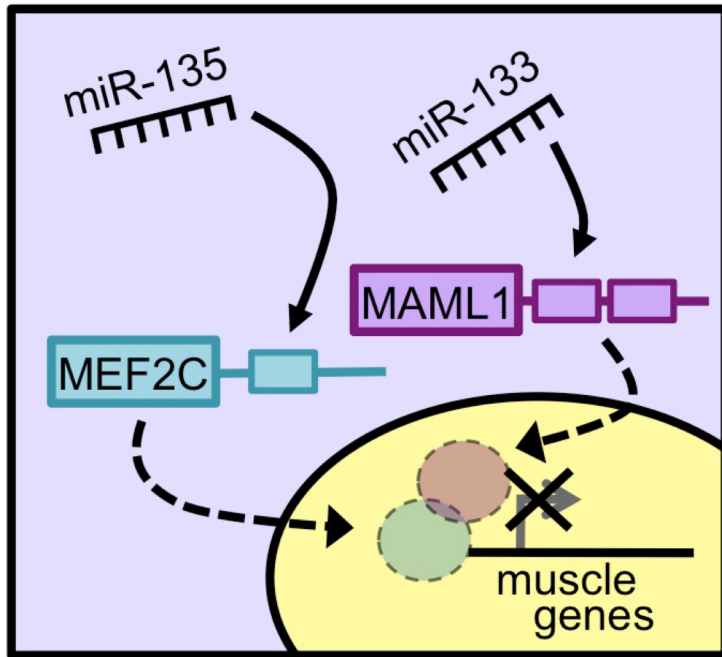
- linc-MD1 controls differentiation in human myoblasts
- it is down-regulated in Duchenne myoblasts



- low levels of linc-MD1 account for the delay in differentiation of DMD myoblasts
- rescue of linc-MD1 restores an almost wt differentiation timing

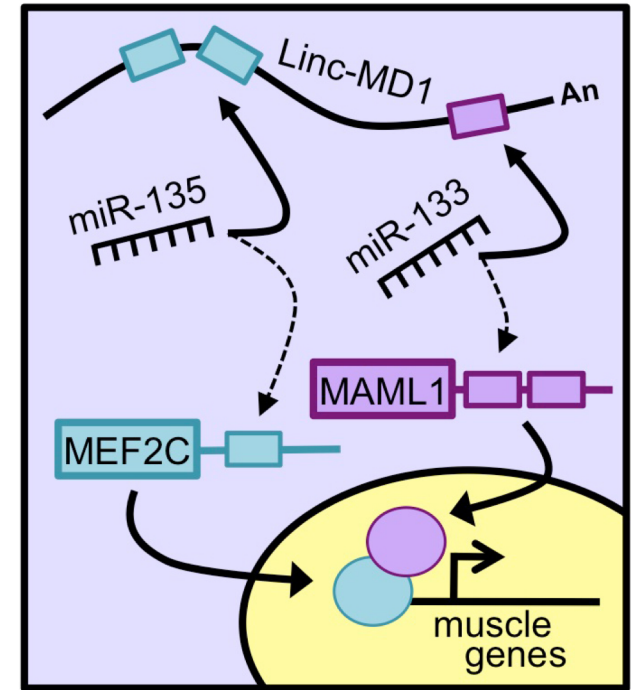


# linc-MD1 acts as a sponge for specific miRNAs



myoblasts

linc MD1

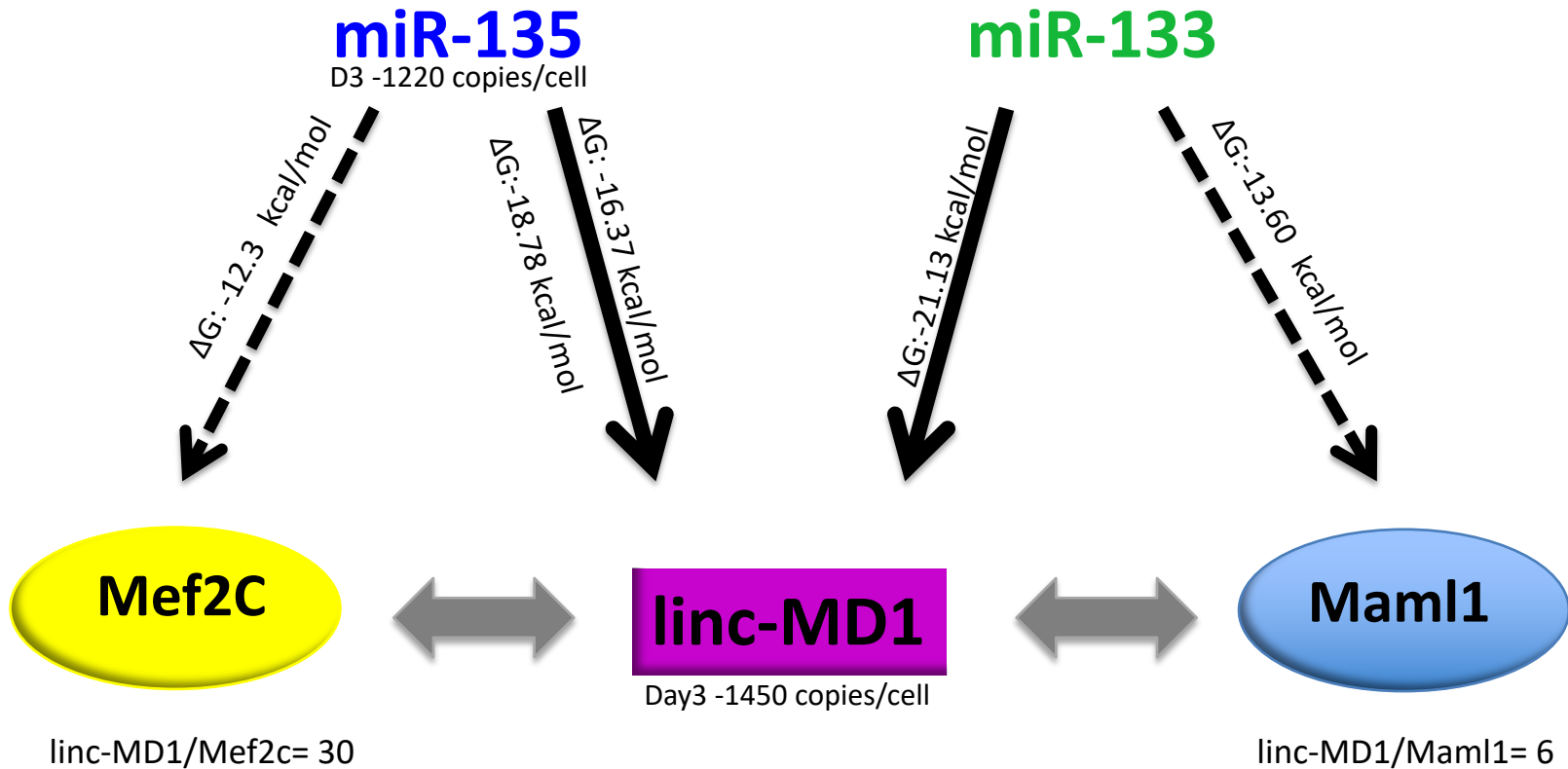


late muscle gene expression

differentiation

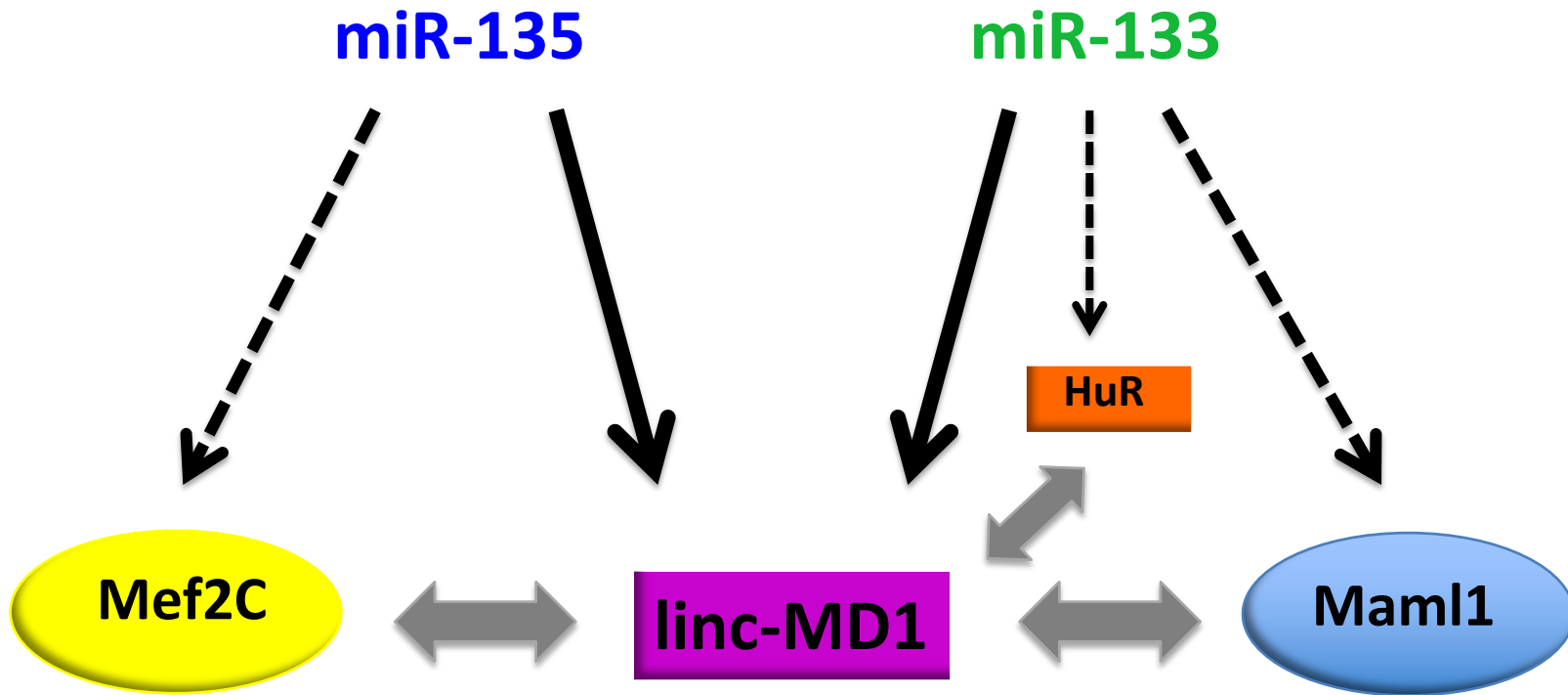


# Crosstalk between coding and non coding RNAs

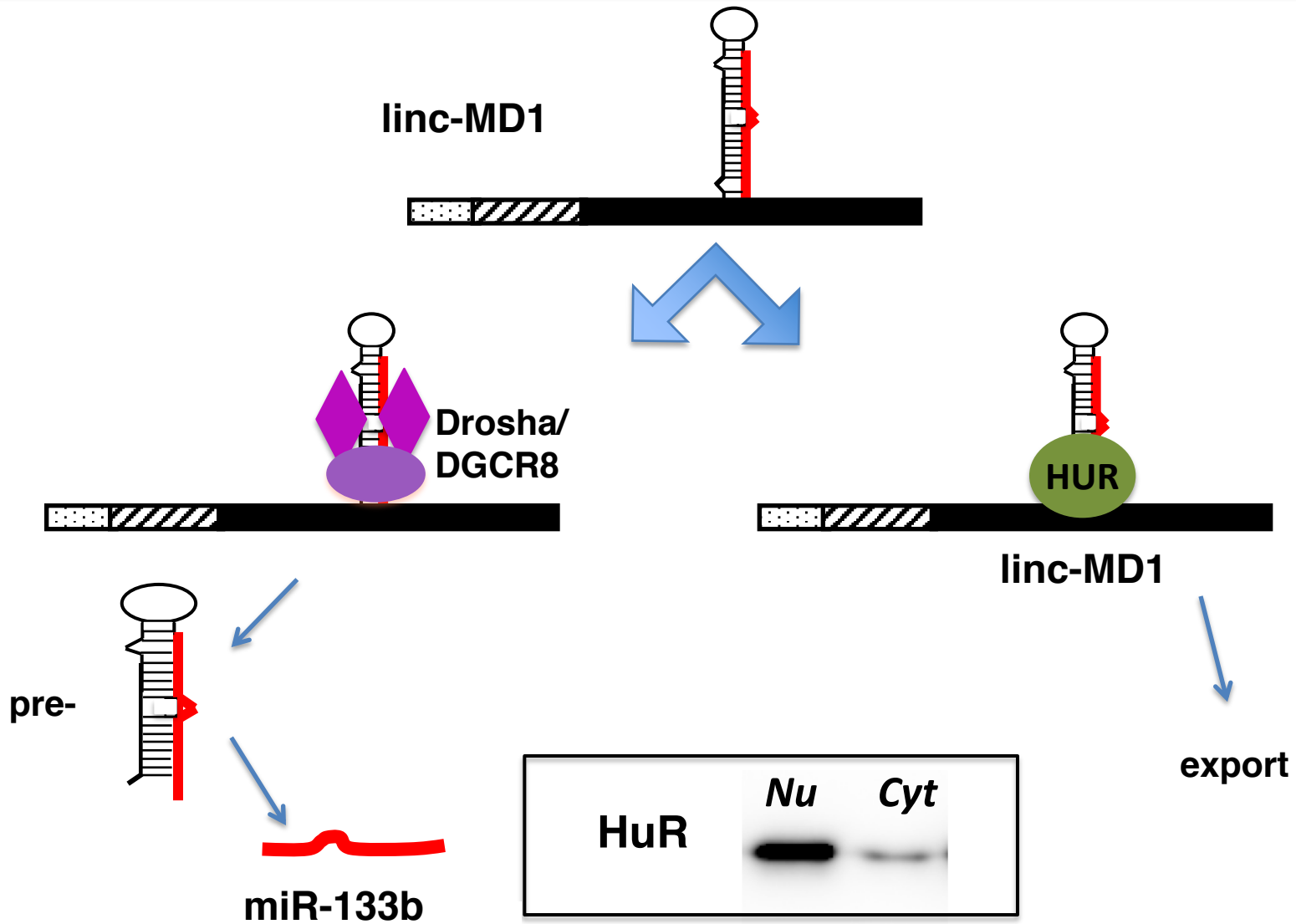


$\Delta G$  values were obtained from miRanda  
(Enright et al., 2003)

*Expanding the circuitry.....*

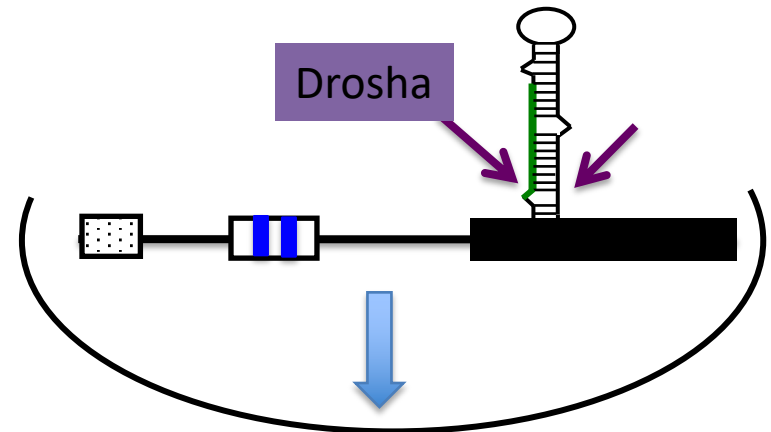
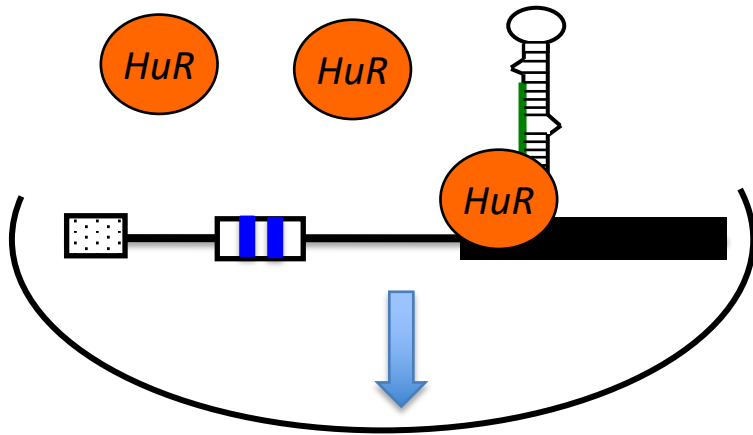


# HuR affects the alternative fate of linc-MD1

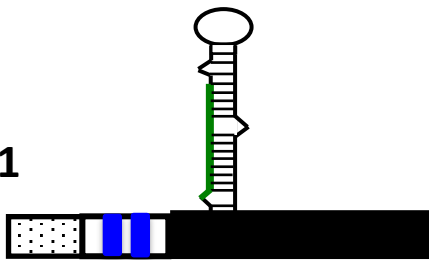


# Nuclear HuR controls the relative ratio of linc-MD1 versus miR-133b

nucleus



linc-MD1



pre-miR-133



cytoplasm

Nuclear HuR

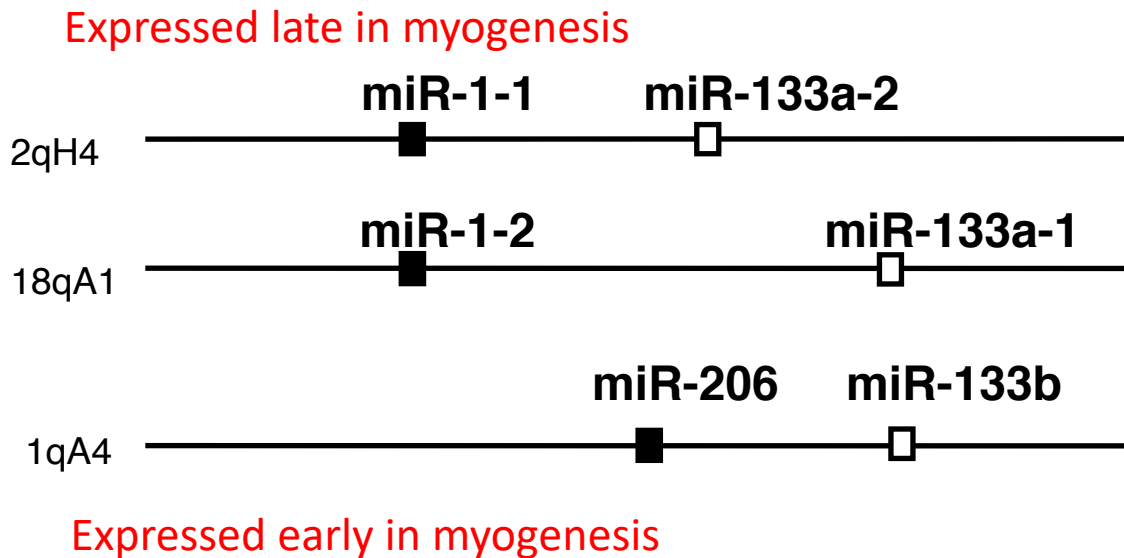
differentiation

## miR-206 is *present only in vertebrates*

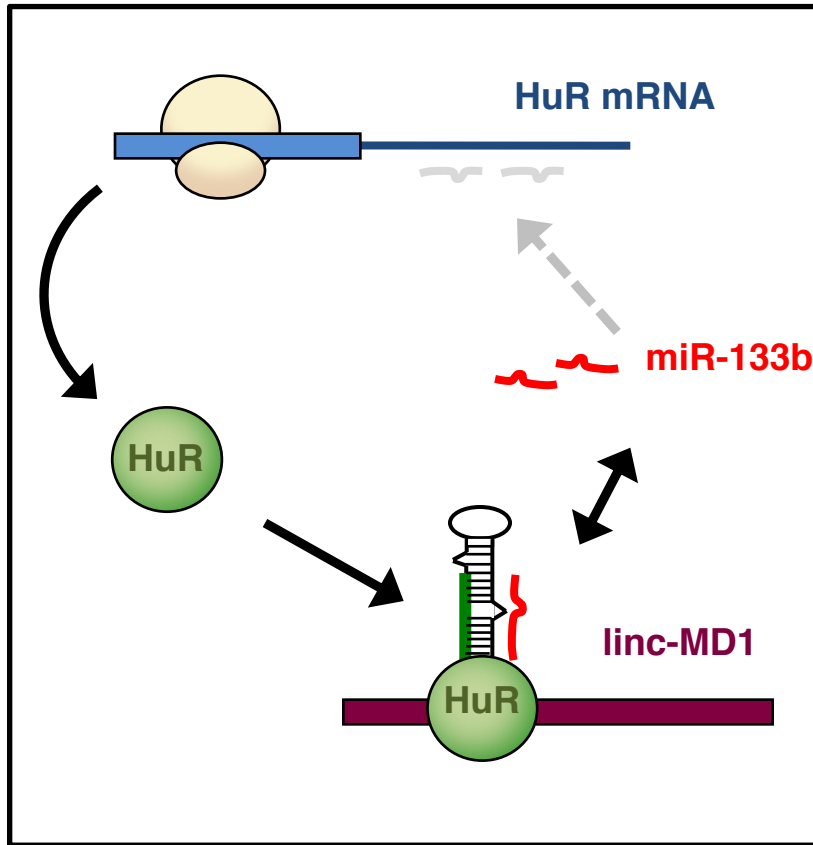
The origin of the miR-206/miR-133b locus correlates, indeed, with the increase in complexity of vertebrate skeletal muscle: in fact, fly and worm do not have satellite cells nor do they possess different fiber types such as slow-twitch and fast twitch fibers as found in vertebrate skeletal muscle

*miR-206 – in activated precursor cells*  
*miR1 – in differentiated myoblasts*

**Mouse genomic localization**

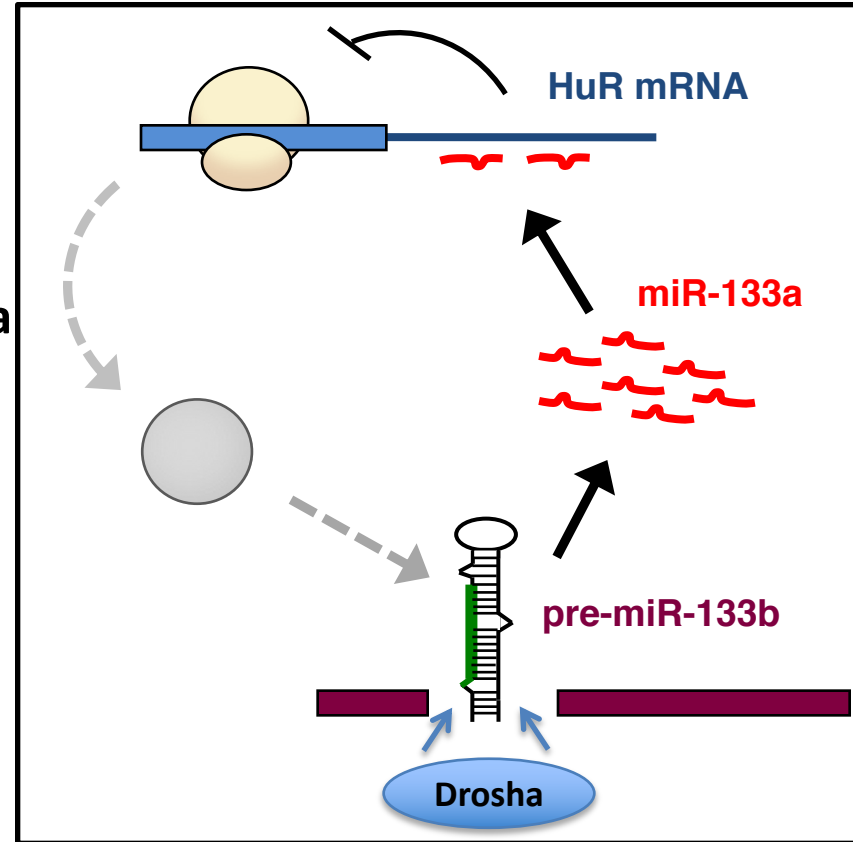


# A feed forward positive loop between HuR and linc-MD1



early phases of differentiation

miR-133a



late phases of differentiation

differentiation